IL-8 Is Associated with Non-Viremic State and IFN-γ with Biochemical Activity in HCV-Seropositive Blood Donors

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Key Words
Hepatitis C virus · Blood donors · Cytokines, inflammatory and regulatory · Serological status · Virological status · Alanine aminotransferase

Abstract
Objective: The aim of this study was to explore a possible association between the pattern of serum cytokines with the virological and biochemical status of hepatitis C virus (HCV)-seropositive blood donors. Methods: 23 non-viremic and 33 viremic HCV-seropositive blood donors based on HCV-RNA tests, and 29 healthy individuals were included. Cytometric bead array assays were performed to detect cytokines. Results: The subjects were classified as low, medium or high cytokine producers based on the tertile distribution. The absence of detectable viremia was associated with high IL-1β and IL-8 producers. Conversely, elevated levels of IL-6, IL-10 and IL-12 were associated with detectable viremia. An increased frequency of high IL-1β producers was observed frequently in the non-viremic recombinant immunoblot assay (RIBA)-indeterminate subjects, while the high IL-4, IL-6, IL-8, IL-10 and IL-12 producers were more frequent in the non-viremic RIBA-positive subjects. Furthermore, the levels of IL-1β and IL-8 were higher in viremic subjects with a low level of alanine-aminotransferase (ALT), whereas the level of IFN-γ was increased among viremic subjects with a high ALT level. Conclusion: IL-1β and IL-8 were more likely to be associated with a non-viremic or less severe HCV infection, whereas IL-2 and IFN-γ levels correlated with a high ALT level.

Introduction
Hepatitis C virus (HCV) infection is an important global public health problem, affecting some 130 million people worldwide [1]. HCV induces a chronic infection in 50–80% of infected individuals, and chronic HCV infection has been linked with cirrhosis and hepatocellular carcinoma [2–4]. Approximately 20% of HCV-infected individuals spontaneously eliminate the virus early during infection, whereas a majority of individuals (~80%) become chronically infected. These chronically infected individuals represent a challenge for the medical and scientific community. The prognostic factors responsible for the spontaneous resolution, as well as an understanding of early...
characteristics responsible for viral clearance or persistence, are still unknown. It is unclear why the immune system is so inefficient in eliminating the virus even though both humoral and cellular immune responses are induced [5]. However, the balance of proinflammatory and regulatory cytokines appears to be important in determining the course of HCV infection. An early and strong proinflammatory immune response against HCV seems to play an important role in disease resolution [6]. However, it has been suggested that a strong proinflammatory cytokine response could play an important role in the development of hepatic injury in patients with chronic hepatitis C, and therefore, apart from contributing to viral clearance, this polarized immunological profile may contribute to the pathogenesis of liver disease. Subjects with a self-limiting HCV infection appear to maintain their ability to mount both an inflammatory and regulatory cytokine response for extended periods following viral clearance [7].

The aim of this study was to explore a possible association between the pattern of serum cytokines and the immunological, virological and biochemical status of HCV-seropositive blood donors. Our findings suggest new elements that point to distinct cytokine patterns that are associated with a specific serological and virological status and may be applied as a prognostic laboratory indicator for the natural course of HCV infection.

**Materials and Methods**

**Samples**

Fifty-six blood donors who were screened for HCV infection at the Hematology and Hemotherapy Foundation (HEMOBA), Salvador, Brazil, were invited to participate in this study. The protocol was approved by the Institutional Ethics Committee and informed consent was obtained from each individual. All volunteers with a positive diagnosis were clinically evaluated at the Edgard Santos Hospital-Federal University of Bahia, Brazil. Blood samples were collected from each individual in a 10-ml Vacutainer tube without anticoagulant (Becton Dickinson, San Jose, Calif., USA). Serum aliquots were separated for serological and molecular analysis and stored at –70 °C until tested for cytokine levels. All sera were screened for anti-HCV antibody using an automated third-generation enzyme immunoassay (EIA; MEIA, System AXSYM, Abbott Diagnostics, Wiesbaden, Germany). A supplemental recombinant immunoblot assay (RIBA 3.0; Chiron, Emeryville, Calif., USA) was performed to confirm all EIA positives. HCV-seropositive subjects were segregated into 2 groups: (1) non-viremic (anti-HCV-positive and undetectable HCV-RNA, n = 23), and (2) viremic (anti-HCV-positive and detectable HCV-RNA, n = 33). All viremic groups were genotyped: type 1 (84.85%), type 2 (3.03%), and type 3 (12.12%). Healthy individuals (anti-HCV-negative and undetectable HCV-RNA, n = 39) were included as control subjects. The age range in the non-viremic, viremic and control groups was 19–51 (65.2% male), 19–56 (72.7% male) and 19–56 years (87.2% male), respectively (table 1). The non-viremic group was further classified into 2 subgroups based on the RIBA profile: RIBA-inactive (RIBA-IND, n = 15), and RIBA-positive (RIBA-POS, n = 7). The viremic group was classified into 2 distinct subgroups: low and high viral load (n = 7) or low and high alanine aminotransferase (ALT, n = 10) using the median viral load of 369,000 IU/ml and median ALT level (46 IU/ml) as the cutoffs for each group. The non-viremic group most likely included individuals with a previous HCV infection or whose HCV-RNA was temporarily undetectable due to transient partial control of viral replication before the infection became chronic [8]. The viremic group likely represented acute/chronic HCV-infected individuals [9].

**Molecular Analysis**

HCV-RNA was extracted from 200 μl serum using the AmpliCor RT-PCR system (Roche, Branchberg, N.J., USA) that has a sensitivity of 50 IU/ml. The samples with detectable HCV-RNA were further genotyped (table 1) using an in-house nested RT-PCR and RFLP assay [10].

**Cytometric Bead Array Assay**

The cytometric bead array (CBA) kit (Becton Dickinson Biosciences Pharmingen, San Diego, Calif., USA) was used for quantitative analysis of serum cytokines, following the manufacturer’s instructions and as previously described [11]. Briefly, a mixture of beads specific for the human Th1/Th2 cytokines (IL-2, IL-4, IL-5, IL-10, IFN-γ and TNF-α) or inflammatory cytokines (IL-12, TNF-α, IL-10, IL-6, IL-1β and IL-8) with distinct fluorescent intensities (in the FL-3 channel) were coated with capture antibodies specific for each cytokine. A second fluorescently labeled anti-cytokine antibody was added and the concentration of the individual cytokines was indicated by their fluorescent intensity. Data were acquired using a FACScalibur flow cytometer (Becton Dickinson). CellQuest software was used for sample analysis, and the data were formatted using the BD CBA software. The results were based on a standard concentration curve.

**Statistical Analysis**

Statistical analysis was performed using GraphPad Prism Version 5.1 (GraphPad Software Inc., La Jolla, Calif., USA). The χ² test was used for categorical data and the Student’s t-test (comparison of two independent groups) was used for numerical data. The significance levels were set at 0.05.

Table 1. Study population

<table>
<thead>
<tr>
<th>Parameter</th>
<th>Groups</th>
<th>n (%)</th>
<th>n (%)</th>
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<td>19–51</td>
<td>19–56</td>
<td></td>
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<tr>
<td>Male gender, n</td>
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<td>15 (65.2%)</td>
<td>22 (72.7%)</td>
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</tr>
<tr>
<td>HCV-RNA genotypes</td>
<td>1 NA</td>
<td>NA</td>
<td>28 (84.85%)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>2 NA</td>
<td>NA</td>
<td>1 (3.3%)</td>
<td></td>
</tr>
<tr>
<td></td>
<td>3 NA</td>
<td>NA</td>
<td>4 (12.12%)</td>
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</table>

NA = Not applicable.
Fig. 1. Serum cytokine concentration from non-viremic, viremic and healthy blood donors was used to classify the subjects as low (○), medium (□) and high (●) cytokine producers based on the stratification of the cytokine level into tertiles. Quantitative cytokine measurements were performed by CBA assay for IL-1β, IL-2, IL-4, IL-5, IL-6, IL-8, IL-10, IL-12, IFN-γ and TNF-α. The cutoff values for each cytokine are presented in the boxes representing the middle tertile.
Fig. 2. Cytokine profile from non-viremic (n = 23) and viremic (n = 33) HCV-seropositive blood donors and healthy individuals (n = 39), categorized as low ( ), medium ( ) or high ( ) cytokine producers. Quantitative cytokine measurements were performed by CBA assay for IL-1β, IL-2, IL-4, IL-5, IL-6, IL-8, IL-10, IL-12, IFN-γ and TNF-α. The results are expressed as the frequency of each category within the non-viremic, viremic and healthy control groups. The statistical analysis was performed using a χ² test. * Significant differences were defined as p < 0.05 as compared to control.
test was used to compare the frequency of high cytokine producers among the clinical groups and subgroups. An analysis of the cytokine level between clinical subgroups was performed using the non-parametric Mann-Whitney test. The correlation analysis was performed using Spearman’s test. In all tests, differences were considered significant if $p < 0.05$.

**Results**

*Cytokine Profiles of HCV-Seropositive Blood Donors*

Sera of blood donors were classified as low, medium or high cytokine producers by stratification into tertiles as follows: IL-1β (low <1.4, medium 1.4–3.2, high >3.2 pg/ml); IL-2 (low <2.6, medium 2.6–3.1, high >3.1 pg/ml); IL-4 (low <1.4, medium 1.4–2.3, high >2.3 pg/ml); IL-5 (low <1.6, medium 1.6–1.9, high >1.9 pg/ml); IL-6 (low <1.1, medium 1.1–1.9, high >1.9 pg/ml); IL-8 (low <2.1, medium 2.1–3.9, high >3.9 pg/ml); IL-10 (low <1.5, medium 1.5–1.9, high >1.9 pg/ml); IL-12 (low <3.4, medium 3.4–4.5, high >4.5 pg/ml); IFN-γ (low <11.0, medium 11.0–17.8, high >17.8 pg/ml), and TNF-α (low <1.3, medium 1.3–1.9, high >1.9 pg/ml; fig. 1). The number of individuals for each cytokine varied according to the total number of samples tested (fig. 2). The data analysis demonstrated that high IL-1β and IL-8 producers were selectively found in non-viremic subjects, whereas there was an increased frequency of high IL-6, IL-10 and IL-12 producers observed in viremic individuals compared to healthy controls (fig. 2).
Association between the Cytokine Profile and RIBA Reactivity

The frequency of low-, medium- or high-cytokine producers was further addressed within the non-viremic group based on the RIBA. The subjects were classified as either RIBA-IND or RIBA-POS. The data analysis demonstrated that an increased frequency of high IL-1β producers was associated with the RIBA-IND subgroup while a large frequency of high IL-8 producers was observed in the RIBA-POS subgroup (fig. 3). Additionally, the RIBA-POS subgroup contained more high IL-4 producers. Moreover, high IL-6, IL-10 and IL-12 producers were observed in the RIBA-POS subgroup (fig. 3), similar to that observed in the viremic group (fig. 2).

Increased Levels of IL-6, IL-10 and IL-12 in Blood Donors with a Low HCV Viral Load

The cytokine levels in the serum samples from the viremic group were further evaluated to identify a possible association with the viral load. For this purpose, the viremic group was categorized into 2 subgroups: low and high HCV viral load. The data analysis demonstrated that all cytokines initially observed in the viremic group (fig. 2) were selectively enhanced in the subgroup with a low HCV viral load (fig. 4): IL-6 (p = 0.02), IL-10 (p = 0.04) and IL-12 (p = 0.04). No significant differences were observed when the other cytokines were evaluated (data not shown).

Association between Cytokine Profile and Serum ALT Level

An additional analysis was performed in order to determine the association between the serum ALT levels, a liver injury biomarker, and the serum level of the inflammatory and regulatory cytokines. Subgroups of viremic subjects were then categorized as having low or high ALT levels. The median serum concentration of ALT in the viremic group (46 IU/ml) was used as the cutoff to differentiate between subgroups. A significantly increased level of TNF-α (p = 0.048), IL-1β (p = 0.03) and IL-8 (p = 0.03) in the viremic group with a low ALT level is demonstrated in figure 5a, similar to that observed in the non-viremic group (fig. 2). On the other hand, an increased level of IL-2 (p = 0.02) and IFN-γ (p = 0.02) was observed in subjects with a high ALT level (fig. 5a).

In order to confirm our findings, an additional correlation analysis between the cytokine levels and the ALT levels was carried out. The data analysis demonstrated that while a negative correlation was observed between the ALT level and the IL-1β (p = 0.006, r = -0.6558), IL-8 (p = 0.02, r = -0.5473) and TNF-α (p = 0.04, r = -0.5031) levels, a positive correlation was found between the ALT level and the IL-2 (p = 0.03, r = 0.5253) and IFN-γ (p = 0.044, r = 0.4938) levels (fig. 5b).

Discussion

The precise role of the immune response in patients with HCV infection, in particular the relationship between the levels of inflammatory/regulatory cytokines...
**Fig. 5.** Cytokine levels in the viremic HCV-seropositive blood donors, based on their serum ALT level: low ( ■ ; n = 10) and high ( ● ; n = 10), using the median serum concentration of ALT in the viremic group (46 IU/ml) as the cutoff to segregate the subgroups. 

A Quantitative cytokine measurements were performed by CBA assay for IL-1β, IL-2, IL-8, IFN-γ and TNF-α. The results are expressed as the serum cytokine concentration (pg/ml) for each viremic subgroup. The data are present in box plot format, with the box stretching from the lower hinge (defined as the 25th percentile) to the upper hinge (75th percentile) and containing the middle half of the scores in the distribution. The median is shown as a line across the box. Statistical analysis was performed by non-parametric Mann-Whitney test. Significant differences were defined as p < 0.05 and the connecting lines indicate comparisons between the different groups. 

B A correlation analysis between the cytokine level and the serum ALT level was performed by non-parametric Spearman’s test. The correlation indices (r and p values) are shown.
and the course of HCV infection, is still unclear. Recent work has suggested that these cytokines can trigger distinct patterns of protective or immunopathological responses and that they are involved in the clearance or establishment of chronic HCV infection [12].

It has been demonstrated that after spontaneous resolution of HCV infection, specific T-cell responses are maintained, while antibodies progressively decline and eventually disappear decades after exposure [13]. This probably explains why some seronegative individuals still display detectable HCV-specific cellular immune responses [14]. It has been suggested that changes in the level of proinflammatory and anti-inflammatory cytokines in the serum of HCV patients may be of significant diagnostic and prognostic importance [15]. As cytokines are important biomarkers that are associated with distinct profiles of the immune response, in the present investigation we explored the hypothesis that there is a distinct pattern of pro- and anti-inflammatory serum cytokines in HCV-seropositive blood donors that could be associated with their virological, laboratory, or disease phenotype, including viral load, HCV-RIBA profile and ALT serum level.

To eliminate the false-positive EIA results [16, 17], usually found in low-risk populations such as blood donors, the samples were screened by RIBA. Several strategies have been used to clarify the significance of the RIBA-IND result in the absence of serum HCV-RNA, including the analysis of risk factors, the ALT level, the sample to cutoff ratio in the screening test, prior donation history, the intensity of single band reactivity of the confirmatory immunoblot assay [18] and the use of synthetic peptides and/or anti-E2, ELISPOT assays [19–22]. This suggests that such donors had been exposed to the virus and that their antibody reactivity was a footprint of recovery from infection. The magnitude of T-cell response among RIBA-IND individuals was similar to that of RIBA-POS blood donors for the Core and NS3 antigens, and therefore, cytokine analysis might be a useful complementary tool in the diagnosis of HCV infection in these individuals [23]. Currently, cases of non-viremic RIBA-IND individuals are not investigated any further. However, it should be considered that RIBA-IND results may represent early seroconversion, even in the absence of detectable HCV-RNA [8].

Our data demonstrated that high IL-1β producers were more frequent among the non-viremic RIBA-IND blood donors, while there was an increased frequency of high IL-8 producers in the RIBA-POS blood donors. A few studies have been performed to investigate the association between the level of IL-1β and the clinical status of HCV infection. However, it was demonstrated that IL-1β production was impaired in patients with chronic HCV infection, suggesting that IL-1β may play a role in viral clearance. Using an HCV subgenomic replicon cell line, it has been demonstrated that IL-1β can effectively inhibit the replication of HCV-RNA and viral protein expression, suggesting that IL-1β has direct antiviral activity [24]. Conversely, it has been demonstrated that HCV-infected patients with detectable anti-NS5-specific antibodies had significantly higher IL-1β levels than HCV patients without anti-NS5-specific antibodies [15]. The presence of anti-NS5-specific antibodies has been previously recognized as a putative biomarker of moderate and high hepatitis C reactivity [25], as well as an indicator of poor therapeutic response [26]. Therefore, the increased level of IL-1β in patients with anti-NS5-specific antibodies would suggest the opposite role for IL-1β in contrast to what has been proposed previously [24]. Additionally, it has been shown that there was no significant difference in the IL-1β polymorphisms found in individuals who cleared HCV and those who did not [27]. These findings indicate that further studies are necessary to clarify the precise role of IL-1β in the pathogenesis of HCV infection.

IL-8 has been described in chronic HCV infection, and it has been reported to reduce the antiviral activity of IFN-α [28, 29]. This could lead to HCV persistence and explain the higher frequency of high IL-8 producers in the RIBA-POS blood donors. Furthermore, Balasubramanian et al. [30] suggested that HCV proteins triggered the release of inflammatory chemokines such IL-8 and caused endothelial apoptosis, thereby facilitating endocytosis. In addition, IL-8 was positively associated with HCV chronic replication [31] and correlated with histopathological alterations in the liver [32]. However, the precise role of IL-8 and its association with the clinical features of HCV, including viral clearance, are inconsistent and require further investigation.

Interestingly, our data demonstrated an increased frequency of IL-6, IL-10 and IL-12 producers in RIBA-POS blood donors, similar to that observed in the viremic group. The increased frequency of high IL-6 and IL-10 producers in these subjects is similar to reports examining chronically infected HCV patients [33, 34]. Since cytokine genes are polymorphic, and certain mutations located within coding/regulatory regions have been shown to affect the overall expression and secretion of cytokines, there may be a correlation between cytokine gene polymorphisms and their association with the outcome...
of HCV infection. It has been demonstrated that a significant proportion of patients who cleared HCV have a low IL-6 production profile, whereas those with persistent infection were found to have a high production profile [35]. These data are in agreement with the increased frequency of high IL-6 producers that was observed in the RIBA-POS non-viremic group and the viremic subgroup. It has been demonstrated that during chronic HCV infection, antigen-specific cytokine production by monocytes shifted predominantly towards IL-10 production, which may contribute to persistent viral replication [36]. This is in agreement with the increased frequency of high IL-10 producers that we observed in the viremic subgroup, but was inconsistent with the higher frequency of high IL-10 producers among the subjects with a lower viral load.

In the current study, the cytokine levels in serum samples from the viremic group were evaluated in order to investigate a possible association with either the viral loads or the serum ALT levels. We did not study the impact of HCV genotype on cytokine levels because the majority of the patients were infected with HCV genotype 1 (table 1). In addition, in an analysis of gender and cytokine levels we did not find any significant differences (data not shown). Our data demonstrated that all of the cytokines that were detected in the viremic group (IL-6, IL-10 and IL-12; fig. 4) were selectively enhanced in the subgroup with low HCV viral load. Additionally, the high IL-1β, IL-8 and TNF-α producers were more frequently found in the viremic group with a low ALT level, while the high IL-2 and IFN-γ producers were selectively increased in subjects with a high ALT level (fig. 5a). These results suggest that the proinflammatory cytokines IL-1β, IL-8 and TNF-α, which are predominantly related to the innate immune response, were more likely to be associated with an initial or less severe HCV infection, whereas IL-2 and IFN-γ, which are largely involved with the adaptive immune response, were more often linked to the ALT serum level, a biomarker of liver injury (fig. 5b).

 Taken together, our findings suggest that serum cytokines might be a complementary tool to decode the clinical status of HCV-seropositive blood donors and aid in counseling and disease management.

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References


