

Life Sciences, the Research Fund Divry, and Innogenetics, Belgium; Dr van Duijn was sponsored by the Netherland Organisation for Scientific Research (NWO), the Eurodem Concerted Action of Dementia, and the Nestor Stimulation Program of Research into the Elderly of the Dutch Ministry of Health and the Ministry of Education, the Netherlands, and the National Institute of Health Sciences (NIHS) and the Alzheimer Association, United States. Dr Van Broeckhoven is a research associate of the Belgian National Fund for Scientific Research.

References

1. Mahley RW. Apolipoprotein E: cholesterol transport protein with expanding role in cell biology. *Science* 1988;240:622-640
2. Hallman DM, Boerwinkle E, Saha N, et al. The apolipoprotein E polymorphism: a comparison of allele frequencies and effects in nine populations. *Am J Hum Genet* 1991;49:338-349
3. Strittmatter WJ, Saunders AM, Schmechel D, et al. Apolipoprotein E: high-avidity binding to β -amyloid and increased frequency of type 4 allele in late-onset familial Alzheimer disease. *Proc Natl Acad Sci USA* 1993;90:1977-1981
4. Saunders AM, Strittmatter WJ, Schmechel D, et al. Association of apolipoprotein E allele ϵ 4 with late-onset familial and sporadic Alzheimer's disease. *Neurology* 1993;43:1467-1472
5. Corder EH, Saunders AM, Strittmatter WJ, et al. Gene dose of apolipoprotein E type 4 allele and the risk of Alzheimer's disease in late onset families. *Science* 1993;261:921-923
6. Saunders AM, Schmechel K, Breitner JCS, et al. Apolipoprotein E ϵ 4 allele distributions in late-onset Alzheimer's disease and in other amyloid forming diseases. *Lancet* 1993;2:710-711
7. Hardy J, Houlden H, Collinge J, et al. Apolipoprotein E polymorphism and Alzheimer's disease. *Lancet* 1993;2:738
8. Namba Y, Tomonaga M, Kawasaki H, et al. Apolipoprotein E immunoreactivity in cerebral amyloid deposits and neurofibrillary tangles in Alzheimer's disease and kuru plaque amyloid in Creutzfeldt-Jakob disease. *Brain Res* 1991;541:163-166
9. Wisniewski T, Golabek A, Matsubara E, et al. Apolipoprotein E: binding to soluble Alzheimer's β -amyloid. *Biochem Biophys Res Commun* 1993;192:359-365
10. Strittmatter WJ, Weisgraber KH, Huang D, et al. Binding of human apolipoprotein E to synthetic amyloid β peptide: isoform-specific effects and implications for late-onset Alzheimer disease. *Proc Natl Acad Sci USA* 1993;90:8098-8102
11. Schmechel DE, Saunders AM, Strittmatter WJ, et al. Increased amyloid β -peptide deposition as a consequence of apolipoprotein E genotype in late-onset Alzheimer's disease. *Proc Natl Acad Sci USA* 1993;90:9649-9653
12. Bakker E, Van Broeckhoven C, Haan J, et al. DNA-diagnosis for hereditary cerebral hemorrhage with amyloidosis (Dutch type). *Am J Hum Genet* 1991;49:518-521
13. Hendriks L, Van Duijn CM, Cras P, et al. Presenile dementia and cerebral haemorrhage linked to a mutation at codon 692 of the β -amyloid precursor protein gene. *Nature Genet* 1992;1:218-221
14. De Knijff P, Van Den Maagdenberg AMJM, Stalenhoef AFH, et al. Familial dysbetalipoproteinemia associated with apolipoprotein E3-Leiden in an extended multigeneration pedigree. *J Clin Invest* 1991;88:643-655
15. Durlinger ETL, Haan J, Roos RAC. Hereditary cerebral hemorrhage with amyloidosis—Dutch type. *Neurology* 1993;43:1626-1627
16. Smit M, De Knijff P, Rosseneu M, et al. Apolipoprotein E polymorphism in the Netherlands and its effect on plasma lipid and apolipoprotein levels. *Hum Genet* 1988;80:287-292
17. Haan J, Roos RAC, Briet PE, et al. Hereditary cerebral hemor-

rhage with amyloidosis—Dutch type. *Clin Neurol Neurosurg* 1989;81:285-290

18. Haan J, Algra PR, Roos RAC. Hereditary cerebral hemorrhage with amyloidosis—Dutch type: clinical and CT analysis of 24 cases. *Arch Neurol* 1990;47:649-653

Immune Complex Deposition in the Choroid Plexus of Patients with Acquired Immunodeficiency Syndrome

M. F. Falangola, MD,* B. G. Castro-Filho, MD,† and C. K. Petito, MD*

We identified immune complex deposits in the choroid plexus of approximately 75% of patients with acquired immunodeficiency syndrome (AIDS) who had either normal brains or human immunodeficiency virus encephalitis. Since circulating immune complexes are common in AIDS patients, and since local choroid plexus pathology usually was absent, their likely origin is from the bloodstream. Choroid plexus deposits of immune complexes have been implicated in altering the function of this structure or in enhancing its vulnerability to infection. Therefore, immune complex deposition in the choroid plexus of AIDS patients may be responsible for some of the common alterations in the cerebrospinal fluid and for the frequency of opportunistic infections in this region.

Falangola MF, Castro-Filho BG, Petito CK. Immune complex deposition in the choroid plexus of patients with acquired immunodeficiency syndrome. *Ann Neurol* 1994;36:437-440

Circulating immune complexes (ICs) are found in more than 80% of both adults and children with the acquired immunodeficiency syndrome (AIDS) [1, 2] and in chil-

From the *Department of Pathology, University of Miami School of Medicine, Miami, FL, and †Centro de Pesquisas Goncalo Moniz-Fiocruz, Bahia, Brazil.

Received Jan 20, 1994, and in revised form Mar 10. Accepted for publication Mar 10, 1994.

Address correspondence to Dr Petito, Department of Pathology (Neuropathology—R-5), University of Miami School of Medicine, Papanicolaou Research Building, Room 417, 1550 N.W. 10th Avenue, Miami, FL 33136.

Patient Data

Patient No.	Sex	Age (yr)	Risk Factors	Duration of AIDS ^a (mo)	AIDS-related Systemic Diseases ^b	Brain Disease	Choroid Plexus IC Deposits	Choroid Plexus Inflammation
1	M	46	Homosexual	10	<i>Cryptococcus</i>	HIVE	+	—
2	M	34	IVDA	36	PCP/CMV	HIVE	+	—
3	F	33	IVDA	12	PCP/HS/ITP	HIVE	+	—
4	F	27	IVDA	60	PCP/ <i>Mycobacterium avium</i> /CMV	HIVE	+	+
5	M	30	IVDA	24	Kaposi's sarcoma/Tb	HIVE	+	+
6	M	41	IVDA	>3	ITP	HIVE	+	—
7	M	54	Homosexual	1	Lymphoma/ Kaposi's sarcoma	HIVE	+	+
8	M	37	IVDA	22	PCP		+	—
9	M	42	Homosexual	9	Lymphoma		+	+
10	M	36	Homosexual	7	PCP/ <i>Aspergillus</i>	Focal vasculitis	+	—
11	M	52	Unknown	NA	HIV myositis/ candidiasis		+	—
12	M	39	Unknown	5	PCP/ <i>M. avium</i> / Kaposi's sarcoma	HIVE	+	—
13	F	45	Heterosexual	24	CMV	HIVE/Wernicke's	+	—
14	M	32	Homosexual	0.56	CMV/histoplasmosis		+	—
15	M	46	Homosexual	12	<i>Cryptococcus</i> / Kaposi's sarcoma	HIVE/Wernicke's	—	+
16	M	49	Promiscuity	8	Lymphoma/CMV		—	—
17	M	34	Homosexual	48	PCP/CMV/Herpes zoster	HIVE	—	+
18	F	28	Heterosexual	NA	<i>M. avium</i>	HIVE	—	—
19	M	50	Homosexual	2	PCP/Kaposi's sarcoma		—	—

^aInterval between initial presentation with AIDS-defining illness and death.

^bIncludes those diagnosed during life and at autopsy.

IC = immune complex; HIVE = HIV encephalitis; PCP = *Pneumocystis carinii* pneumonia; CVM = cytomegalovirus; HS = herpes simplex; ITP = idiopathic thrombocytopenic purpura; Tb = *Mycobacterium tuberculosis*; NA = information not available; IVDA = intravenous drug abuse.

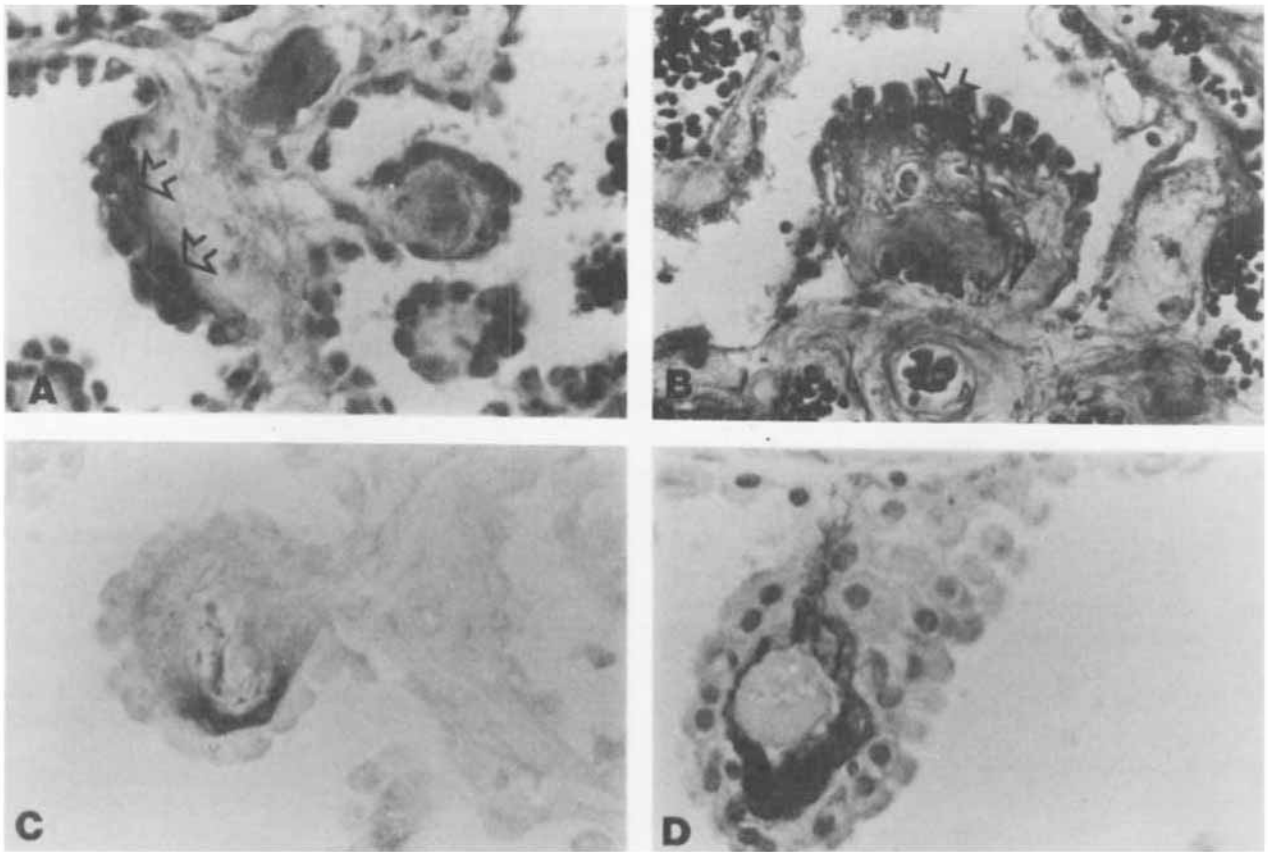
dren, the levels of ICs generally correlate with a more severe disease course [2]. These ICs contain specific immunoglobulin isotypes, complement, and antigens of human immunodeficiency virus (HIV) and other infectious agents [3]. Deposition of ICs into tissue may damage systemic organs such as lung or kidney but the brain usually is protected by the blood-brain barrier, which prevents circulating macromolecules, including ICs, from entering.

The choroid plexus, however, is vulnerable to IC deposition since its capillaries have open or "gap" junctions that allow free passage of macromolecules. In addition, choroid plexus receptors for immunoglobulins and complement [4, 5] may increase its vulnerability to IC deposition. Accordingly, we examined postmortem choroid plexus of patients with AIDS for the presence of IC deposition. In order to rule out local IC formation, patients with opportunistic infections and

lymphomas of the central nervous system (CNS) were excluded, as the choroid plexus frequently is involved [6] (personal observations, 1994). A preliminary report of this study was published previously [7].

Materials and Methods

Formalin-fixed choroid plexus specimens from 19 AIDS patients, 12 of whom had HIV encephalitis (HIVE), were examined. Material for frozen sections or electron microscopy was unavailable. Paraffin-embedded sections cut 5 to 7- μ m thick were stained with hematoxylin and eosin (H&E), periodic acid-Schiff (PAS), and Heidenhain's azan. Immunohistochemical studies were performed by the avidin-biotin method using overnight incubation with monoclonal antibody to HIV gp41 (1:500 dilution; Genetics Systems, Seattle, WA), polyclonal antibody to IgG-Fc fragment (1:1,000 dilution; Dako, Carpinteria, CA), and polyclonal antibody to C3d complement (α 2D) (1:2,000; Dako). Following incubation with the primary antisera, sections were washed and



Subepithelial immune complex deposits (arrows) in the choroid plexus are stained with periodic acid-Schiff (A) and azan (B) and are immunoreactive for IgG (C) and complement (D). ($\times 600$)

sequentially incubated with biotinylated secondary antibody, the avidin-biotin complex (Vector Labs, Burlingame, CA), and hydrogen peroxide with 3,3'-diaminobenzidine. A positive control was a sample from a patient previously diagnosed with HIV (for anti-HIV gp41) and the identification of intravascular serum immunoreactivity (for anti-IgG and anti-C3d) in each slide examined. Saline solution was substituted for all primary antibody as a negative control. The choroid plexus was blindly evaluated for the presence or absence of inflammation, endothelial cell hypertrophy, and mesenchymal cell proliferation—each graded as negative, focal, moderate, and intense—and for the presence of IC deposition (PAS and IgG/C3d deposits).

Results

Fifteen patients were men and four were women; their average age was 40 years. Risk factors for HIV infection included homosexuality (in 8), intravenous drug abuse (in 6), heterosexual contact (in 2), and promiscuity (in 1), and were unknown in 2 patients (Table). Neither age, sex, HIV risk factors, duration of AIDS, AIDS-related infections or tumors in systemic organs,

nor HIV correlated with the presence of inflammation or IC deposits in the choroid plexus. None of the patients had detectable infectious agents in the choroid plexus as determined by routine microscopic examination.

Inflammation was present in 6 (31%) of the 19 patients and consisted of focal infiltrates of lymphocytes and plasma cells in the connective tissue, predominantly in the perivascular areas. It was more frequent with HIV (41%) than without HIV (14%). Similarly, marked endothelial hypertrophy, characterized by enlarged vesicular nuclei, and mesenchymal cell proliferation of the choroid villi stroma were seen more frequently with HIV (67 and 58% vs 43 and 29%). The relatively small number of patients ($n = 19$) did not allow for adequate statistical analyses and thus the significance of these comparisons is not clear.

Fourteen of the 19 patients had IC deposits located immediately beneath the basement membrane of the choroid plexus epithelium. In 12 patients, homogeneous, acidophilic, PAS-positive deposits were seen (Fig A) and were azan positive (Fig B) in 3 and immunoreactive for IgG and C3d in 10 (Figs C, D). The choroid plexus in an additional 2 patients (Patients 13

and 14) contained IgG and C3d-immunoreactive deposits, although there was no staining. IC deposits were absent in 5 patients although 2 demonstrated foci of C3d immunoreactivity in the subepithelial regions. ICs were equally as common in patients with (75%) as in those without (71%) HIVE. All were immunonegative for HIV gp41.

Discussion

Circulating ICs develop from the interaction between antigen-antibody complex and fractions of complement, and thus are found with a large number of infectious, neoplastic, and autoimmune diseases. Their deposition may lead to tissue injury, which has been associated with systemic vasculitis and glomerulonephritis. Tissues that are highly vascularized and have capillaries with gap junctions such as the kidney and the choroid plexus are at increased risk for IC deposition. Not surprisingly, ICs in choroid plexus have been found with autoimmune nephritis, systemic lupus erythematosus, hepatosplenic schistosomiasis, and cirrhosis [8–11] but not with diseases such as cancer, atherosclerosis, or bronchopneumonia [5].

IC disease is a known complication of AIDS. In addition to the high incidence of circulating ICs, IgA-related IC-mediated renal disease [12] and IC-related vasculitis in peripheral nerve and muscle of HIV-positive patients [13] have been described. The present demonstration of frequent IC deposition in the choroid plexus of AIDS patients is additional evidence that HIV infection has a component of IC disease. The deposits were associated with endothelial hypertrophy and mesenchymal proliferation but not with local inflammation. It is likely that they were deposited from circulating ICs, as local choroid plexus inflammation was not required for their formation. The incidence of IC deposition of approximately 75% was similar in patients with otherwise normal brains and choroid plexus as well as in patients with HIVE and choroid plexus inflammation.

IC deposition in the choroid plexus in AIDS patients may have at least two important functions. First, it could damage the blood–cerebrospinal fluid (CSF) barrier and alter the selective permeability of the choroid plexus and composition of the CSF. Indeed, the CSF of AIDS patients frequently has elevated protein levels, IgG, and pleocytosis, for which choroid plexus abnormalities could be responsible [14]. Second, IC deposition could render the choroid plexus vulnerable to blood-borne infections including opportunistic infections such as toxoplasmosis (6) and HIV infection. Precedent for this hypothesis is seen in an experimental model of serum sickness in which IC deposition in the choroid plexus renders the animal more vulnerable to CNS infections [15].

This study was supported by grants from the Conselho Nacional de Desenvolvimento Científico e Tecnológico (to M. F. F.) and the National Institutes of Health (grant RO1 NS27416, to C. K. P.).

The authors appreciate the helpful critique of Dr Philip Ruiz, the technical help of Ms Brenda Roberts, and the secretarial assistance of Ms Lee Ann Moffett.

References

1. McDougal JS, Hubbard M, Nicholson JKA, et al. Immune complexes in the acquired immunodeficiency syndrome (AIDS): relationship to disease manifestation, risk group, and immunologic defect. *J Clin Immunol* 1985;5:130–138
2. Ellaurie M, Calvelli T, Rubinstein A. Immune complexes in pediatric human immunodeficiency virus infection. *Am J Dis Child* 1990;144:1207–1209
3. McHugh TM, Srites DP, Busch MP, et al. Relation of circulating levels of human immunodeficiency virus (HIV) antigen, antibody to p24, and HIV-containing immune complexes in HIV-infected patients. *J Infect Dis* 1988;158:1088–1091
4. Braathen LR, Forre OT, Husby G, Williams RC. Evidence for IgG receptors and complement factor C3b receptors in human choroid plexus. *Clin Immunol Immunopathol* 1979;14:284–291
5. Peress NS, Roxburgh VA, Gelfand MC. Binding sites for immune components in human choroid plexus. *Arthritis Rheum* 1981;24:520–626
6. Falangola MF, Petito CK. Choroid plexus infection in cerebral toxoplasmosis in AIDS patients. *Neurology* 1993;43:2035–2040
7. Falangola MF, Petito CK, Castro-Filho BG. Immune complex deposition in the choroid plexus of AIDS patients. *Ann Neurol* 1993;34:282 (Abstract)
8. McIntosh RM, Copack P, Chernack WB, et al. The human choroid plexus and autoimmune nephritis. *Arch Pathol* 1975;99:48–50
9. Atkins CJ, Kondon JJ, Quismorio FP, Friou GJ. The choroid plexus in systemic lupus erythematosus. *Ann Intern Med* 1972;76:65–72
10. Pittella JE, Bambirra EA. Histopathological and immunofluorescence study of the choroid plexus in hepatosplenic schistosomiasis mansoni. *Am J Med Hyg* 1989;41:548–552
11. Pittella JE, Bambirra EA. Immune complexes in the choroid plexus in liver cirrhosis. *Arch Pathol Lab Med* 1991;115:220–222
12. Kimmel PL, Phillips TM, Ferreira-Centeno A, et al. Brief report: idiopathic IgA nephropathy in patients with human immunodeficiency virus infection. *N Engl J Med* 1992;327:702–706
13. Said G, Lacroix-Ciaudo C, Fujimura H, et al. The peripheral neuropathy of necrotizing arteritis: a clinico-pathological study. *Ann Neurol* 1988;23:461–465
14. Marshall DW, Brey RL, Cahill WT, et al. Spectrum of cerebrospinal fluid findings in various stages of human immunodeficiency virus infection. *Arch Neurol* 1988;45:954–958
15. Peress NS, Gruenewald R, Carioto LA, Krause CH. Increased risk of experimental central nervous system listeriosis in rats with chronic serum sickness: an immunohistopathological study. *J Neuropathol Exp Neurol* 1983;42:409–420