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## Genomic taxonomy of vibrios

Cristiane C Thompson\*<sup>1</sup>, Ana Carolina P Vicente<sup>1</sup>, Rangel C Souza<sup>2</sup>,  
Ana Tereza R Vasconcelos<sup>2</sup>, Tammi Vesth<sup>3</sup>, Nelson Alves Jr<sup>4</sup>,  
David W Ussery<sup>3</sup>, Tetsuya Iida<sup>5</sup> and Fabiano L Thompson\*<sup>4</sup>

Address: <sup>1</sup>Laboratory of Molecular Genetics of Microorganisms, Oswaldo Cruz Institute, FIOCRUZ, Rio de Janeiro, Brazil, <sup>2</sup>National Laboratory for Scientific Computing, Department of Applied and Computational Mathematics, Laboratory of Bioinformatics, Av. Getúlio Vargas 333, Quitandinha, 25651-070, Petropolis, RJ, Brazil, <sup>3</sup>Center for Biological Sequence Analysis, Department of Biotechnology, Building 208, The Technical University of Denmark, DK-2800 Kgs. Lyngby, Denmark, <sup>4</sup>Department of Genetics, Institute of Biology, Federal University of Rio de Janeiro, UFRJ, Brazil and <sup>5</sup>Laboratory of Genomic Research on Pathogenic Bacteria, International Research Center for Infectious Diseases, Research Institute for Microbial Diseases, Osaka University, Suita, Osaka 565-0871, Japan

Email: Cristiane C Thompson\* - thompson@ioc.fiocruz.br; Ana Carolina P Vicente - anapaulo@ioc.fiocruz.br; Rangel C Souza - rangel@lncc.br; Ana Tereza R Vasconcelos - atrv@lncc.br; Tammi Vesth - tammivesth@hotmail.com; Nelson Alves - nelsonjrr@yahoo.com.br; David W Ussery - dave@cbs.dtu.dk; Tetsuya Iida - iida@biken.osaka-u.ac.jp; Fabiano L Thompson\* - fabiano.thompson@biologia.ufrj.br

\* Corresponding authors

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### Abstract

**Background:** *Vibrio* taxonomy has been based on a polyphasic approach. In this study, we retrieve useful taxonomic information (i.e. data that can be used to distinguish different taxonomic levels, such as species and genera) from 32 genome sequences of different *vibrio* species. We use a variety of tools to explore the taxonomic relationship between the sequenced genomes, including Multilocus Sequence Analysis (MLSA), supertrees, Average Amino Acid Identity (AAI), genomic signatures, and Genome BLAST atlases. Our aim is to analyse the usefulness of these tools for species identification in vibrios.

**Results:** We have generated four new genome sequences of three *Vibrio* species, i.e., *V. alginolyticus* 40B, *V. harveyi*-like IDA3, and *V. mimicus* strains VM573 and VM603, and present a broad analyses of these genomes along with other sequenced *Vibrio* species. The genome atlas and pangenome plots provide a tantalizing image of the genomic differences that occur between closely related sister species, e.g. *V. cholerae* and *V. mimicus*. The *vibrio* pangenome contains around 26504 genes. The *V. cholerae* core genome and pangenome consist of 1520 and 6923 genes, respectively. Pangenomes might allow different strains of *V. cholerae* to occupy different niches. MLSA and supertree analyses resulted in a similar phylogenetic picture, with a clear distinction of four groups (*Vibrio* core group, *V. cholerae*-*V. mimicus*, *Aliivibrio* spp., and *Photobacterium* spp.). A *Vibrio* species is defined as a group of strains that share > 95% DNA identity in MLSA and supertree analysis, > 96% AAI, ≤ 10 genome signature dissimilarity, and > 61% proteome identity. Strains of the same species and species of the same genus will form monophyletic groups on the basis of MLSA and supertree.

**Conclusion:** The combination of different analytical and bioinformatics tools will enable the most accurate species identification through genomic computational analysis. This endeavour will culminate in the birth of the online genomic taxonomy whereby researchers and end-users of taxonomy will be able to identify their isolates through a web-based server. This novel approach to microbial systematics will result in a tremendous advance concerning biodiversity discovery, description, and understanding.

## Background

Taxonomy is at the basis of the biological sciences, being one of its oldest branches. It deals with identification, classification (*i.e.* creation of new taxa) and nomenclature. In the early 1970s, a seminal work done by Colwell put forward the concept of polyphasic taxonomy that is currently still in use [1]. According to this concept, in order to achieve the most natural classification system, there should be an integration of information from the molecular to the ecological levels. DNA-DNA hybridization (DDH) data had a pivotal role to infer the species boundaries in the polyphasic taxonomy. Indeed DDH proved to be the most powerful tool to unambiguously identify prokaryotes, providing for the first time, a reliable means of categorizing microbes. However, this technique suffers from various limitations, including the need to include reference strains in each new experiment. In addition, it is not portable and requires special facilities available in a limited number of international laboratories. The introduction of 16S rRNA analysis as an alternative method for use in taxonomy allowed the development of an evolutionary framework [2,3]. Boundaries for species identification and a species definition were set on the basis of extensive empirical data [4]. A bacterial species is defined as a group of strains (including the type strain), having > 70% DDH similarity, < 5°C  $\Delta T_m$ , < 5% mol G+C difference of total genomic DNA, > 97% 16S rRNA identity [4,5].

The bacterial species definition is pragmatic and operational, aiming at the establishment of a rapid, reliable, reproducible, and useful taxonomic framework, based on microbial evolution, for a variety of applications (*e.g.* medicine and agriculture) [6]. This polyphasic definition is a consensus in microbiology, although it is not based on a concept (*i.e.* the biological processes behind speciation and species). It is crucial to highlight that the current polyphasic framework does not question if this definition corresponds to a biological reality [7]. Within the framework of polyphasic taxonomy, strains of the same species have similar phenotypes (*e.g.* expression of different types of enzymes, ability to using different types of compounds as energy source, and growth in different temperatures and concentrations of acid and salt), genotypes (*e.g.* rep-PCR and AFLP), and chemotaxonomic features (*e.g.* FAME and polyamines), forming distinguishable tight groups [7]. Ideally, these groups should be readily identifiable and differentiated from closely related species. However, currently there is not a consensus on the definition of a bacterial genus [7].

The most recent ideas on the species concepts corroborate polyphasic taxonomy [8]. According to these authors, a bacterial species concept is a framework that explains how bacterial strains share common features and how they

maintain genomic cohesion. The cohesion is observed as recurrent patterns or groups of strains that are recognized in nature and are termed species for practical reasons. However, the biological process giving rise and maintaining cohesion of these groups is what matters in a species concept. Homologous recombination appears to be a major force leading to genomic cohesion of strains of the same species. It is more frequent between strains of the same species than between strains of different species simply because this genetic process depends on sequence similarity. Horizontal gene transfer (HGT) between unrelated strains would cause an increase in phenotypic variation, but would not be frequent enough to hamper the formation and recognition of species. An alternative species concept is the stable ecotype in which cohesion of bacterial strains of a given species is maintained by accumulation of advantageous mutations and periodic purging of allelic variability [9]. In contrast with the polyphasic species definition which is widely accepted, there is not a consensus on a bacterial species concept. It is of course possible that one single concept is not sufficient to explain the complexity of bacterial diversity. In addition, mutation, homologous recombination and HGT may be detected at varying levels in a single strain, making the scenario even more complex.

Whole microbial genome sequencing studies launched microbial taxonomy into a new era, with the possibility of establishing systematics on the basis of complete genomes [10]. How does one go about using whole genome sequences (WGS) for establishing a genomic taxonomy? And more specifically, how can one taxonomically define and identify species by means of WGS? WGS may contain taxonomic information in the form of gene content, genome wide signatures, phylogenetic markers, amino acid identity and overall genetic composition that might be useful for building novel taxonomic schemes [11]. Pioneer computational and mathematical studies performed in the 1990s suggested that genomes contain species-specific signatures [12]. Genome signature is a compositional parameter reflecting the dinucleotide relative abundance, which is similar between closely related species, and dissimilar between non-related species. Genome signatures appear to allow the identification of isolates and metagenomes into known species [13,14]. Whole genome sequences also permit the reconstruction of more robust taxonomic trees (*i.e.* supertrees) based on all genes of the core genome [15-17]. A good congruence was obtained by the traditional 16S rRNA based trees and the novel supertree methods [18] proposed that the average amino acid identity (AAI) could be used to distinguish closely related sister species. Subsequently, a close relationship between DDH and AAI was shown [19]. Some studies have suggested that the effective number of codons (Nc) could also be a species-specific marker [20].

In *Vibrios*, the birth of the genomic taxonomy occurred with a series of papers that attempted to use multilocus sequence analysis (MLSA) [21-26]. These studies allowed the establishment of rapid and powerful identification systems through the internet. Currently there are MLSA schemes for most of the human pathogens available for free access in the internet. Establishing a universal MLSA will not be possible though. Studies accomplished so far have shown that the resolution of different markers varies according to the taxonomic groups. For instance, the *recA* gene is very useful to differentiate closely related species of *Burkholderia* [27], but it is not appropriate for vibrios [24]. Clearly, genes have different molecular clocks in different microbes, indicating the need of a multigene approach. With the advent of ultra-rapid genome sequencing, it is now possible to sequence one almost complete microbial genome in less than a day [28,29]. The new generation of DNA sequencers will enable sequencing of more than a dozen prokaryotic genomes in less than an hour, possibly making it cheaper and faster to sequence a whole genome than several genes for MLSA. In future, MLSA might be used simply as a rapid screen methodology [30].

Vibrios are an excellent test model for genomic taxonomy because they are ubiquitous in the marine environment, associated with a wide range of marine life (some species such as *V. cholerae*, *V. parahaemolyticus* and *V. vulnificus* cause serious disease in man) and experiencing a variety of environmental conditions and selection forces, leading to high genomic plasticity [31,32]. Consequently, differentiation of sister species becomes very difficult. For instance, *V. cholerae* and *V. mimicus* have nearly indistinguishable phenotypes. Among the phenotypic tests used in the Bergey's manual, only sucrose fermentation and lipase activity may discriminate the two species. According to the most recent version of the Bergey's manual, a *Vibrio* species is defined as a group of strains forming small (0.5-0.8 × 1.4-2.6 μm) comma-shaped rods with polar flagella enclosed in a sheath, facultative anaerobic metabolism, capable of fermenting D-glucose and growth at 20°C [33]. Primarily aquatic, most species are oxidase positive, reduce nitrate to nitrite, require Na<sup>+</sup> for growth, and ferment D-fructose, maltose, and glycerol. Each vibrio species is further identified by an array of over 100 phenotypic tests. There is not an operational definition for genera within the vibrios [33]. In our hands, vibrio species may be better defined on the basis of amplified fragment length polymorphism (AFLP) and MLSA [21,22,25,34]. Strains of the same species (including the type strain) share more than 60% mutual AFLP band pattern similarity and more than 95% similarity in MLSA (using the loci *rpoA*, *recA*, *pyrH*, *ftsZ*, *topA*, *mreB*, *gyrB* and *gapA*). More importantly, strains of the same species and species of the same genus will form monophyletic groups on the basis

of MLSA. This was the main argument used to propose the newly described genus *Aliivibrio* [35].

In order to test the feasibility of the genomic taxonomy in vibrios, several markers were analysed in a collection of 32 genomes, including four newly pyrosequenced genomes. Several *Vibrio* strains had the genome completely sequenced and are available on the web. Eleven *V. cholerae* and two *V. mimicus* genomes formed an ideal test case for taxonomy because of their close relatedness as sister species. These sister species have nearly identical 16S rRNA sequences and around 70% DDH. Disclosing species-specific patterns for the different genome-wide markers would reinforce their usefulness in prokaryotic taxonomy. The aim of this study was to extract taxonomic information from vibrio genome sequences by means of a detailed analysis of MLSA, supertree, Nc, AAI, genomic signatures, Genome BLAST atlas and pangenome plot that would allow species identification.

## Methods

### Genome sequence data

We used 32 genomes of vibrios in this study unless otherwise stated. The genomic sequences of 28 vibrios were obtained from the National Center for Biotechnology Information (NCBI) (Table 1). We have sequenced the genome of *V. alginolyticus* 40B, *V. harveyi*-like 1DA3, and *V. mimicus* strains VM573 and VM603. *V. alginolyticus* 40B and *V. harveyi*-like 1DA3 were isolated from Brazilian corals (*Mussismilia hispida* and *Phyllogorgia dilatata* in 2007 at the Abrolhos reef bank, respectively). *V. mimicus* VM573 (CT and TCP positive) was isolated from a patient with diarrhea in 1990s in the US, whereas *V. mimicus* VM603 was isolated from riverine water in the Brazilian Amazonia region in 1990s. These genomes were sequenced by the Roche-454 pyrosequencing method. Genomic DNA was extracted using the method of Pitcher [36]. The pyrosequencing technique was performed according to [37]. Briefly, genomic DNA was randomly sheared to small fragments and ligated to common adaptors. Single fragments were attached to beads in an emulsion. Amplification by PCR was done in the emulsion and produced ~10<sup>7</sup> copies of the fragments per bead. After removal of the emulsion, the beads were deposited on a fiber optic slide. The DNAs were sequenced using a pyrosequencing protocol. Sequencing of *V. mimicus* VM603 genome was performed on the prototype Roche 454 Genome Sequencer 20™ system, whereas sequencing of *V. alginolyticus* 40B, *V. mimicus* VM573 and *V. harveyi*-like 1DA3 genomes was performed on a Roche 454 Genome Sequencer FLX™ system. The reads were assembled using the Newbler software of the 454/Pyrosequencing. These genomes were annotated automatically using the software SABIÁ [38] and have been deposited at DDBJ/EMBL/GenBank under the project accession number [GenBank:ACZB00000000]

**Table 1: Genomic features of the vibrios genomes.**

Organism	Accession no.	Genome size (nt)	G+C (mol%)	No. of CDS	% coding region	Nc*
<i>Aliivibrio salmonicida</i> FLI1238	<u>FMI78379</u>	3325164	39		77	48
Chromosome I	<u>FMI78380</u>	1206461	38	2820	77	
Chromosome II				984		
<i>Photobacterium profundum</i> SS9						
Chromosome I	<u>CR354531</u>	4085304	41	3416	82	51
Chromosome II	<u>CR354532</u>	2237943	41	2006	80	
<b><i>Vibrio alginolyticus</i> 40B</b>	<u>ACZB00000000</u>	5234286	45	4341	81	53
<i>Vibrio alginolyticus</i> 12G01	<u>AAPS00000000</u>	5160431	44	4732	86	53
<i>Vibrio angustum</i> S14*	<u>AAOJ00000000</u>	5101447	39	4558	84	48
<i>Vibrio campbellii</i> AND4	<u>ABGR00000000</u>	4255798	44	3935	85	53
<i>Vibrio cholerae</i> N16961	<u>AE003852</u>	2961149	47		87	52
Chromosome I	<u>AE003853</u>	1072315	46	2742	84	
Chromosome II				1093		
<i>Vibrio cholerae</i> 0395	<u>CP000627</u>	3024069	47		88	52
Chromosome I	<u>CP000626</u>	1108250	46	2742	86	
Chromosome II				1133		
<i>Vibrio cholerae</i> 1587	<u>AAUR00000000</u>	4137501	47	3758	82	52
<i>Vibrio cholerae</i> 2740-80	<u>AAUT00000000</u>	3945478	47	3771	87	52
<i>Vibrio cholerae</i> 623-39	<u>AAWG00000000</u>	3975259	47	3777	86	52
<i>Vibrio cholerae</i> B33	<u>AAWE00000000</u>	4026835	47	3677	83	53
<i>Vibrio cholerae</i> MAK757	<u>AAUS00000000</u>	3917446	47	3501	82	52
<i>Vibrio cholerae</i> MZO-2	<u>AAWF00000000</u>	3862985	47	3425	83	52
<i>Vibrio cholerae</i> MZO-3	<u>AAUJ00000000</u>	4146039	47	3897	86	52
<i>Vibrio cholerae</i> NCTC8457	<u>AAWD00000000</u>	4063388	47	3975	86	53
<i>Vibrio cholerae</i> V52	<u>AAKJ00000000</u>	3974495	47	3815	86	52
<i>Vibrio fischeri</i> ES114*	<u>CP000020</u>	2897536	38			45
Chromosome I	<u>CP000021</u>	1330333	37	2586	86	
Chromosome II				1175	87	
<i>Vibrio fischeri</i> MJ11*						45
Chromosome I	<u>CP001139</u>	2905029	38	2590	86	
Chromosome II	<u>CP001133</u>	1418848	37	1254	87	
<b><i>Vibrio harveyi</i>-like IAD3</b>	<u>ACZC00000000</u>	5989646	46	4954	66	51
<i>Vibrio harveyi</i> ATCC BAA-1116	<u>CP000789</u>	3765351	45		85	53
Chromosome I	<u>CP000790</u>	2204018	45	3546	86	
Chromosome II				2374		
<i>Vibrio harveyi</i> HY01	<u>AAWP00000000</u>	5400985	45	4327	75	51
<b><i>Vibrio mimicus</i> VM573</b>	<u>ACYV00000000</u>	4373300	46	3744	86	53
<b><i>Vibrio mimicus</i> VM603</b>	<u>ACYU00000000</u>	4421792	46	3790	86	53
<i>Vibrio parahaemolyticus</i> RIMD2210633						
Chromosome I	<u>BA000031</u>	3288558	45	3080	86	52
Chromosome II	<u>BA000032</u>	1877212	45	1752	86	
<i>Vibrio parahaemolyticus</i> AQ3810	<u>AAWQ00000000</u>	5771228	45	5509	80	53
<i>Vibrio shilonii</i> AK1	<u>ABCH00000000</u>	5701826	43	5360	88	54
<i>Vibrio</i> sp Ex25	<u>AAKK00000000</u>	4844262	44	4240	84	53
<i>Vibrio</i> sp MED222	<u>AAND00000000</u>	4891901	43	4590	85	52
<i>Vibrio splendidus</i> 12B01	<u>AAMR00000000</u>	5596386	44	5231	85	53
<i>Vibrio vulnificus</i> CMCP6						
Chromosome I	<u>AE016795</u>	3281944	46	2915	83	53
Chromosome II	<u>AE016796</u>	1844853	47	1557	86	
<i>Vibrio vulnificus</i> YJ016	<u>BA000037</u>	3354505	46		87	53
Chromosome I	<u>BA000038</u>	1857073	47	3259	89	
Chromosome II				1696		

The genomes sequenced by this study are in bold. \**Vibrio angustum* and *Vibrio fischeri* were reclassified as *Photobacterium angustum* [67] and *Aliivibrio fischeri* [35], respectively. \*calculated using concatenated chromosome sequences.

(*V. alginolyticus* 40B), [GenBank:ACZC000000000] (*V. harveyi*-like 1DA3), [GenBank:ACYV000000000] (*V. mimicus* VM573) and [GenBank:ACYU000000000] (*V. mimicus* VM603). The version described in this paper is the first version. The genomes are also available online <http://www.vibrio.lncc.br>. The DNA G+C content of *V. alginolyticus* 40B, *V. mimicus* VM573 and *V. harveyi*-like 1DA3 genomes was calculated using MEGA version 4.0 [39]. We used concatenated genomic sequences of the two chromosomes of vibrios for our analyses.

### Genome BLAST Atlas, proteome matrix, and pangenome plot

The BlastAtlas plots were constructed as described previously [40,41]. The pangenome plot, and proteome matrix were constructed as described [42]. For building the atlas, the genomes were automatically annotated and were compared to the reference chromosome (*V. cholera* strain N16961 in this case). The BLAST matrix perl script performs an all-against-all BLAST comparison of genomes from multiple organisms. For every combination, a protein blast is carried out, finding all homologous proteins. For our purposes, we use the "50-50 rule", which requires both of the following characteristics: 1.) at least 50% of the query protein must overlap in the alignment, and 2.) at least 50% of the residues within the alignment must be identical. After the homologous proteins are identified, the proteins are clustered into protein families and the number of families containing proteins from both strains are counted. The fraction of these shared families out of the total number of families is the number reported in the BLAST matrix. Since the direction of comparison of the two organisms will give identical results under these conditions, one redundant half of the square matrix plot is left out. Thus, we use a triangular shaped diagram where the hypotenuse corresponds to the paralogs (red), which are the internal homologous proteins (e.g., repeated genes). Since this is a comparison of all the proteins in a genome, compared to the pan-genome, it is possible to see related organisms, in terms of their similar composition of gene families.

### 16S rRNA tree, Multilocus Sequence Analysis (MLSA) and Supertree approach

MLSA and supertree approach were based on the concatenated sequences of house-keeping genes [15,21]. The 16S rRNA gene sequences, the gene sequences used for MLSA (i.e. *ftsZ*, *gyrB*, *mreB*, *pyrH*, *recA*, *rpoA* and *topA*) and the gene sequences used for supertree (i.e. aminopeptidase P, *alaS*, *aspS*, *ftsZ*, *gltX*, *gyrB*, *hisS*, *ileS*, *infB*, *metG*, *mreB*, *pntA*, *pheT*, *pyrH*, *recA*, *rpoA*, *rpoB*, *rpsH*, signal recognition particle protein, threonyl-tRNA synthetase, *topA*, *valS* and 30S ribosomal protein S11) were obtained from the NCBI. The concatenated sequences were aligned by CLUSTALX. Phylogenetic analyses were conducted using MEGA ver-

sion 4.0 [39] and PAUP version 4.0b10 [43]. The phylogenetic inference was based on the maximum-parsimony character method (MP), the neighbour-joining genetic distance method (NJ) [44], and the maximum likelihood method (ML). Distance estimations were obtained according to the Kimura-2-parameter for 16S rRNA gene and Jukes-Cantor [45] for MLSA and supertree for NJ. The program Modeltest was used to select the GTR+I+G as the model for MLSA and supertree and Tamura-Nei+I+G as the model for 16S rRNA in the ML analysis. The reliability of each tree topology was checked by 2000 bootstrap replications [46].

### Average amino acid identity (AAI)

The AAI was calculated according to [18]. Genes conserved between a pair of genomes were determined by whole-genome pairwise sequence comparisons using the BLAST algorithm release 2.2.5 [47]. For these comparisons, all protein-coding sequences (CDSs) from one genome were searched against the genomic sequence of the other genome. CDSs that had a BLAST match of at least 40% identity at the amino acid level and an alignable region with more than 70% of the length of the query CDS were considered as conserved genes [48]. This cutoff is above the twilight zone of similarity searches, where inference of homology is error prone due to low similarity between aligned sequences. Thus, query CDSs were presumably homologous to their matches. The genetic relatedness between a pair of genomes was measured by the average amino acid identity of all conserved genes between the two genomes as computed by the BLAST algorithm.

### Codon usage

Codon usage bias was calculated for each genome. The effective number of codons used in a sequence ( $N_c$ ) [20], was calculated using CHIPS (EMBOSS).  $N_c$  values range from 20 (in an extremely biased genome where only one codon is used per amino acid) to 61 (all synonymous codons are used with equal probability) [20]. The rose plot of codon usage was constructed as described previously [42].

### Determination of dinucleotide relative abundance values

We determined the dinucleotide relative abundance value for each genome. Sequences were concatenated with their inverted complementary sequence using REVSEQ, YANK and UNION (EMBOSS). Mononucleotide and dinucleotide frequencies were calculated using COMPSEQ (EMBOSS). Dinucleotide relative abundances ( $\rho^*_{XY}$ ) were calculated using the equation  $\rho^*_{XY} = f_{XY}/f_X f_Y$  where  $f_{XY}$  denotes the frequency of dinucleotide XY, and  $f_X$  and  $f_Y$  denote the frequencies of X and Y, respectively [12]. Statistical theory and data from previous studies [12,49] indicate that the normal range of  $\rho^*_{XY}$  is between 0.78 and

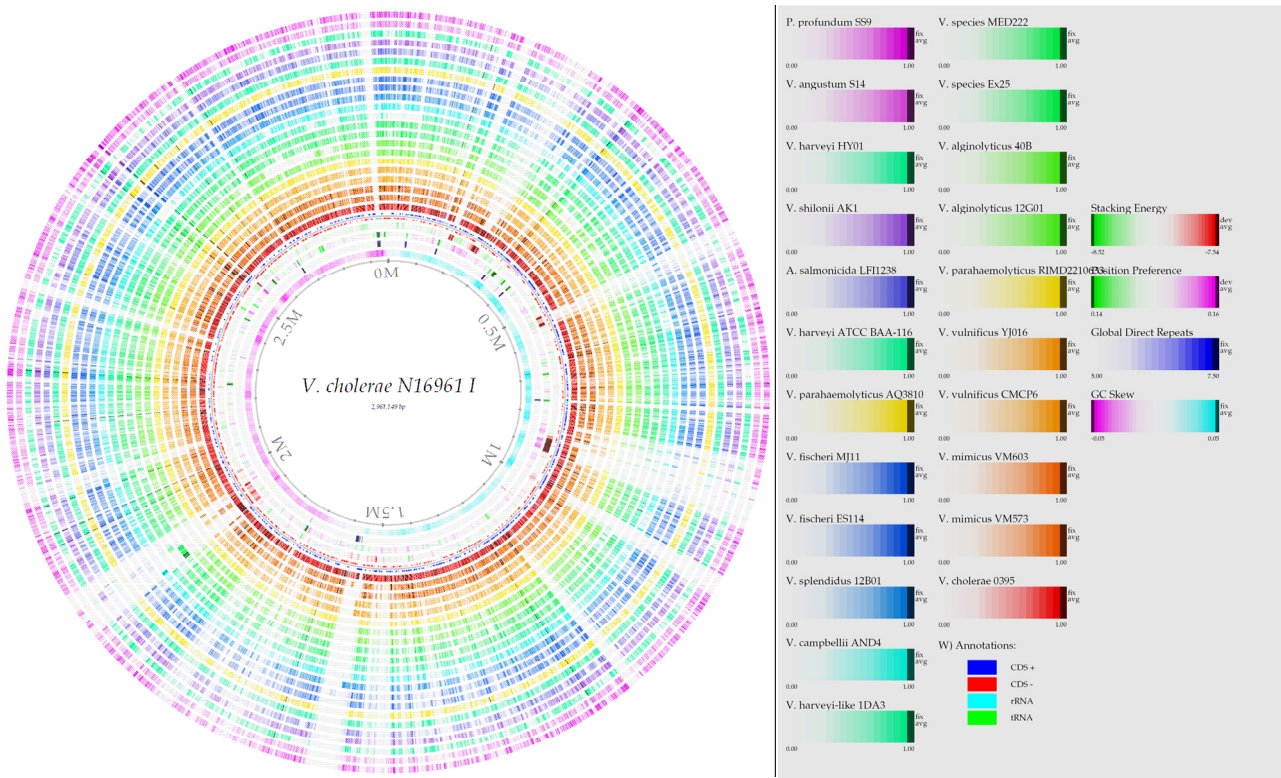
1.23. The difference in genome signature between two sequences is expressed by the genomic dissimilarity ( $\delta^*$ ), which is the average absolute dinucleotide of relative abundance difference between two sequences. The dissimilarities in relative abundance of dinucleotides between both sequences were calculated using the equation described by [12]:  $\delta^*(f,g) = 1/16\sum|\rho^*_{XY}(f) - \rho^*_{XY}(g)|$  (multiplied by 1000 for convenience), where the sum extends over all dinucleotides.

**Results**

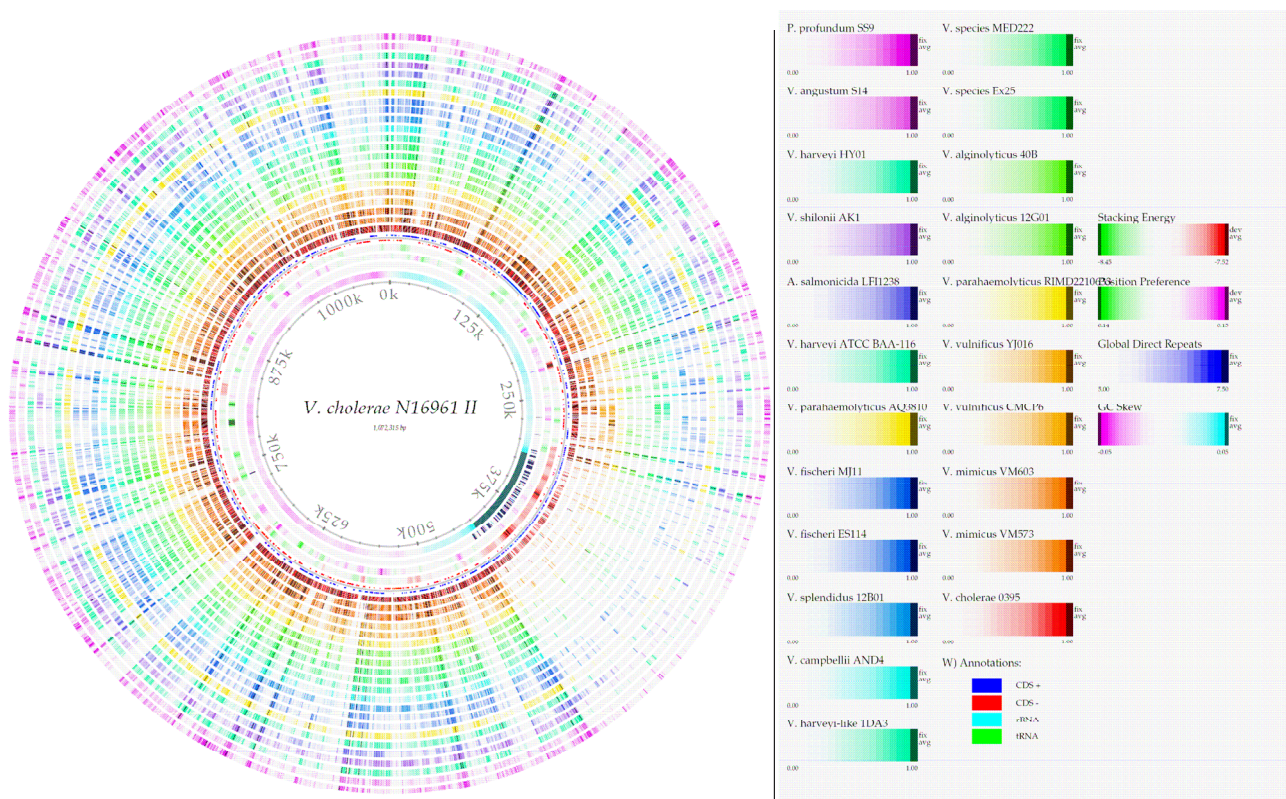
**General features of the sequenced genomes**

The new genomic sequences generated in this study for *V. alginolyticus* 40B (ACZB00000000), *V. harveyi-like* 1DA3 (ACZC00000000), *V. mimicus* strains VM573 (ACYV00000000), and VM603 (ACYU00000000), had 290, 229, 82 and 488 contigs with a total length of approximately 5,234,286, 5,989,646, 4,373,300 and 4,321,792 bp, respectively. The estimated coverage depth was 18, 22, 24 and 20x, respectively. The average GC content for the draft genomes were 45%, 46%, 46%, and

46%, respectively (Table 1). A first attempt to have a global visualization of the differences in gene content between the reference genome *V. cholerae* N16961 and the genomes of the other vibrios was obtained by the genome BLAST atlas which *per se* is not meant to be a taxonomic tool (Figure 1 and Figure 2). There are several regions (lightly colored) of low conservation throughout the chromosomes 1 and 2. In chromosome 2 there is a large region in the low right area that is poorly conserved within the other vibrios. This region corresponds to the superintegron [50]. We can observe in chromosome 1 and 2 that there are regions which contain genes that are conserved only in *V. cholerae*, missing in the other vibrio genomes. These regions might encode for some sort of environmental niche-specific genes. *V. cholerae* strains have little mutual gene content variation even in the hypervariable superintegron region (see chromosome II midpoint 375 Kb). The two *V. mimicus* genomes were the closest to the N16961 according to the atlas.



**Figure 1**  
**Genome BLAST atlas. The chromosome I of vibrios.** The Atlas was constructed using the genome of *V. cholerae* N16961 as the reference strain on which the genes of the other strains are mapped. Genomic regions unique to this strain and not appearing in other vibrio strains are lightly colored. The position of the genes in the different replicons may not be the same.



**Figure 2**  
**Genome BLAST atlas. The chromosome II of vibrios.** The Atlas was constructed using the genome of *V. cholerae* N16961 as the reference strain on which the genes of the other strains are mapped. Genomic regions unique to this strain and not appearing in other vibrio strains are lightly colored. Notice the hyper-variable region (midpoint at 375 K) in the chromosome II of vibrios (the super-integron), corresponding to 1/6 of this replicon. The position of the genes in the different replicons may not be the same.

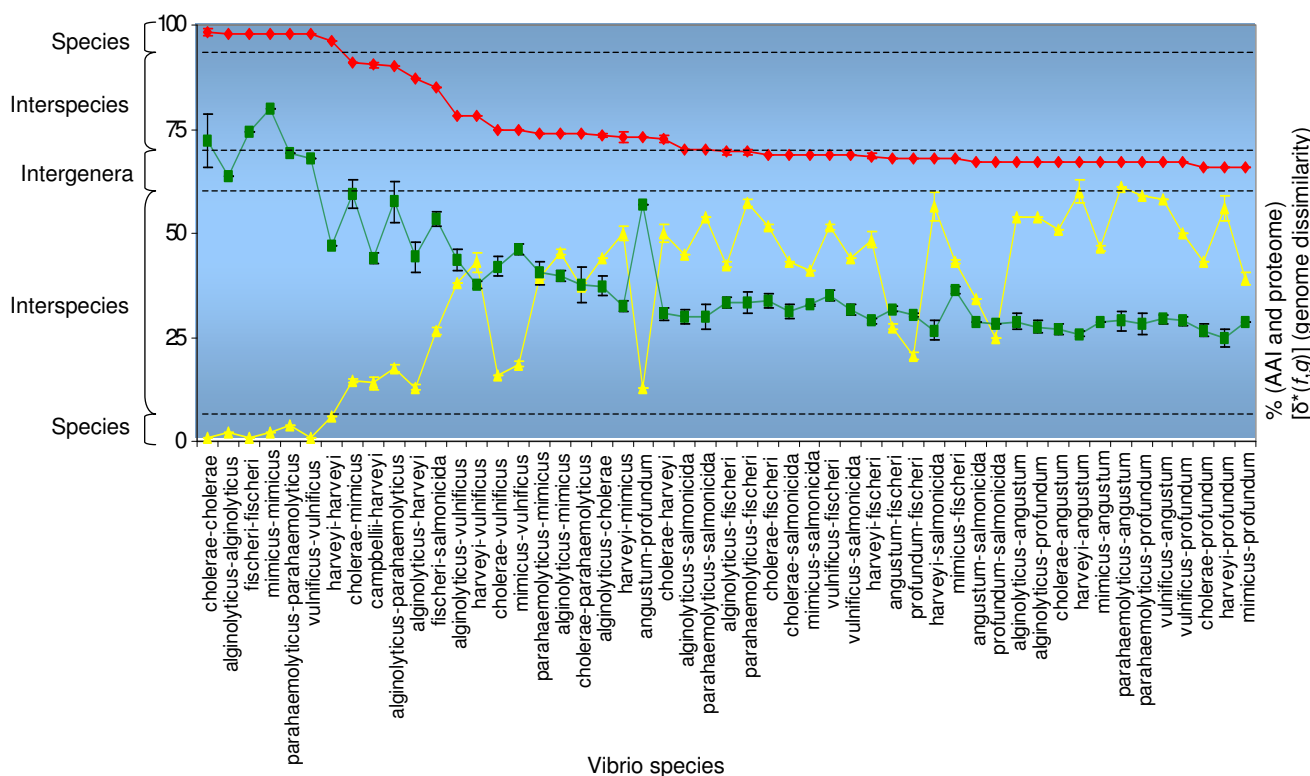
**Proteome BLAST**

The BLAST proteome for all vibrio strains varied between 23.1% (*V. harveyi*-*P. profundum*) and 79.8% (*V. mimicus*-*V. mimicus*) similarity, whereas the percentage of paralogs varied between 1.8% (*V. mimicus*-*V. mimicus*) to 9.3% (*V. parahaemolyticus*-*V. parahaemolyticus*). The BLAST comparison indicated that *V. cholerae* genomes had mutual proteome identity at minimum 61.8% and at maximum 78.4% (Figure 3 and see Additional file 1; Table S1). The intraspecific proteome identity in *V. cholerae* varied between 61.8% and 78.3%, whereas the paralogs in *V. cholerae* genomes varied from 2.8% (99 proteins) to 3.8% (130 proteins). The sister species *V. cholerae*-*V. mimicus*, *V. parahaemolyticus*-*V. alginolyticus* and *V. harveyi*-*V. campbellii* had proteome identity at maximum 65.7%, 64.4% and 45%, respectively. The maximum proteome identity between the genera *Vibrio* and *Aliivibrio* was 38.6% (i.e. *V. splendidus* and *A. fischeri*), whereas the identity between *Vibrio* and *Photobacterium* was 31.8% (i.e. *V. splendidus* and

*P. angustum*). *Aliivibrio* and *Photobacterium* had at maximum 32.3% identity.

**Phylogenetic reconstructions by 16S rRNA, MLSA and supertree**

We selected both conserved and variable single copy genes belonging to different functional groups, from both chromosomes of vibrios and that have been used in several taxonomic studies [15,21,22,24,25,51]. Phylogenetic trees based on 16S rRNA gene sequences, MLSA and the supertree approach were constructed using the ML (Figure 4), MP (see Additional file 2; Figure S1) and NJ methods (see Additional file 3; Figure S2). The trees based on 16S rRNA gene sequences, MLSA and supertree showed similar topology in the three methods. Bootstrap analysis indicated that, most branches were highly significant. The phylogenetic reconstruction indicated a clear separation of groups (i.e. genera) within the vibrio clade. The genera *Photobacterium* and *Aliivibrio* were clearly separated from the genus *Vibrio*. The sister species of vibrios, *V. cholerae*-



**Figure 3**  
**Taxonomic resolution of AAI, BLAST proteome and genome dissimilarity [ $\delta^*(f,g)$ ] of vibrios.** Mean and standard deviations. Red = AAI; Green = proteome; yellow = [ $\delta^*(f,g)$ ]. The taxonomic resolution of AAI is down to the intergenera level, whereas [ $\delta^*(f,g)$ ] has a resolution at interspecies level. The dashed lines delimit ( $p < 0.001$ ) the different taxonomic levels for AAI and [ $\delta^*(f,g)$ ] but not for the proteome. The proteome did not completely fit this figure (and dashed lines limits), showing some noise signal for *V. harveyi-V.harveyi*.

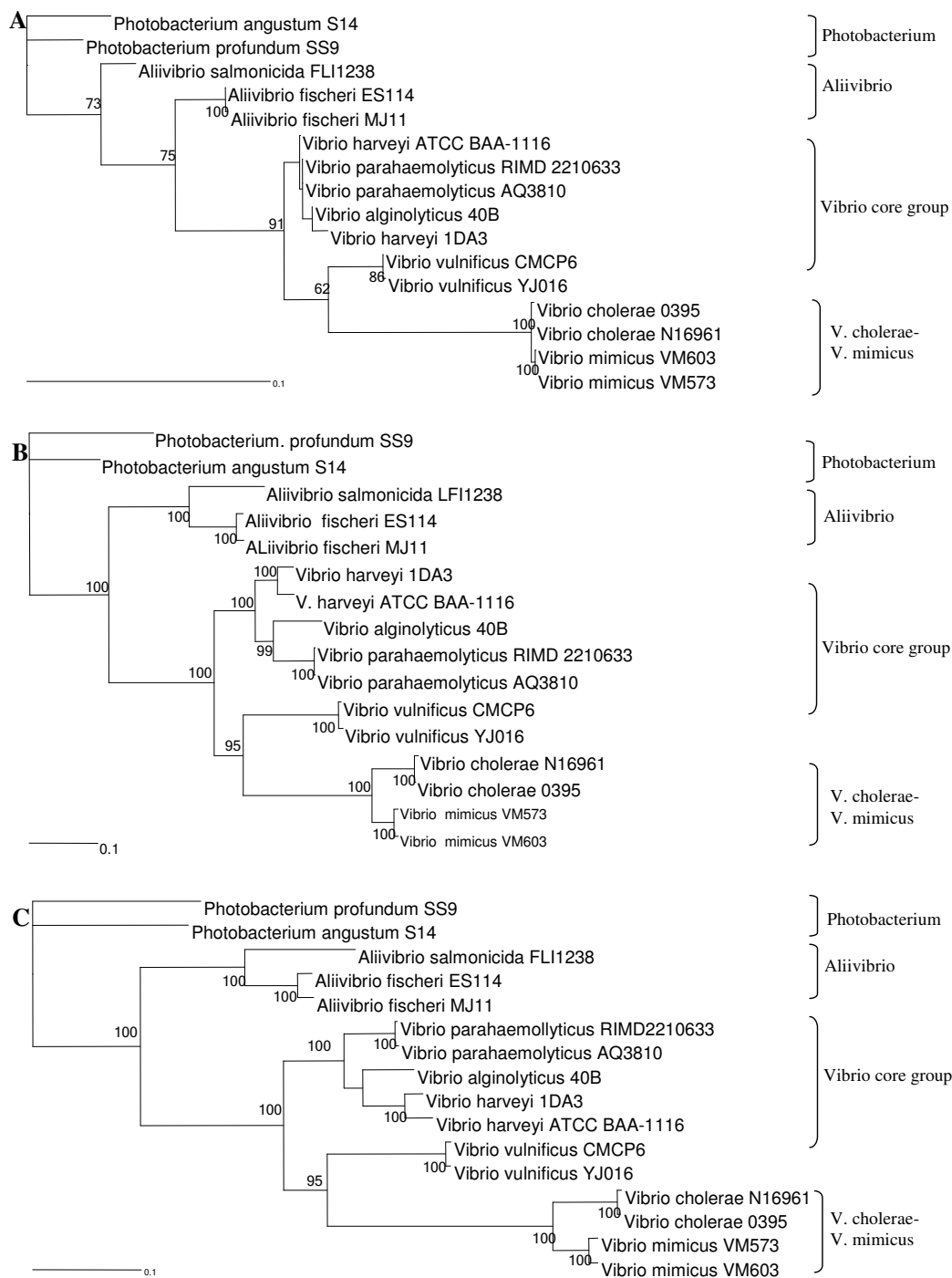
*V. mimicus*, *V. parahaemolyticus-V. alginolyticus* were separated from each other in the MLSA and supertree approaches in all three phylogenetic methods. These pairs of species had almost identical 16S rRNA gene sequences ( $\geq 99\%$  sequence identity) though. Slight grouping differences were observed. *V. alginolyticus* appeared to be at the outskirts of the *V. parahaemolyticus* branch in the MLSA tree while in the supertree *V. alginolyticus* appeared at the outskirts of the *V. harveyi* branch in the three phylogenetic methods, simply because the number of genes used for each analysis was different. The difference may be due to different molecular clocks of the different genes. In the ML analysis, *V. vulnificus* appeared between *V. cholerae* and the vibrio core group. In all three phylogenetic methods, MLSA and supertree had the same taxonomic resolution to discriminate between species.

**Average amino acid identity (AAI)**

The percentage of identity of putative orthologous protein-encoding genes detected in the pairwise comparison is shown in Figure 3 (and see Additional file 4; Table S2).

The identity of protein-encoding genes between different genera of vibrios varied considerably. The mutual AAI for the pairs *Vibrio* and *Photobacterium*, *Vibrio* and *Aliivibrio*, and *Photobacterium* and *Aliivibrio* were at most 67%, 70% and 68%, respectively. The AAI within the genus *Photobacterium* (represented by *P. angustum* S14 and *P. profundum* SS9) was only 73%, whereas the AAI within the genus *Aliivibrio* was 85%. The AAI within the genus *Vibrio* varied between 70 and 91%. The *Vibrio* core group (i.e. *V. alginolyticus*, *V. campbellii*, *V. harveyi* and *V. parahaemolyticus*) shared at most 75% of their protein-encoding genes. The wider range of variation is explained by the higher number of representatives in the latter genus. The sister species *V. cholerae-V. mimicus*, *V. harveyi-V. campbellii*, *V. parahaemolyticus-V. alginolyticus* shared 90-91% AAI, whereas the intra-species AAI in *V. cholerae* varied between 98 and 99.5%. *Vibrio* sp. EX25 and *V. alginolyticus* 40B had 95% identity, suggesting that EX25 belongs to the species *V. alginolyticus*. The rather low AAI within the species *V. harveyi* (i.e. 90%) may be due to the incomplete genome sequences and to unresolved taxonomic issues. *V. harveyi-*





**Figure 4**

**A-C. Phylogenetic trees based on the maximum likelihood method using 16S rRNA gene, MLSA (i.e. *ftsZ*, *gyrB*,  *mreB*, *pyrH*, *recA*, *rpoA* and *topA*; 10,141 bp), and supertree (i.e. aminopeptidase P, *alaS*, *aspS*, *ftsZ*, *gltX*, *gyrB*, *hisS*, *ileS*, *infB*, *metG*, *mreB*, *pntA*, *pheT*, *pyrH*, *recA*, *rpoA*, *rpoB*, *rpsH*, signal recognition particle protein, threonyl-tRNA synthetase, *topA*, *valS* and 30S ribosomal protein S11; 41,617 bp). Bootstrap percentages after 2000 replications are shown. Because some genomes used in this study are not completely sequenced, for the comparison of 16S rRNA, MLSA and supertree, we used 16 genomes of vibrios. The genes used in MLSA and supertree were found only in these 16 genomes.**

like 1DA3 had 70% DDH in previous experiments and formed a separate genomic group on the basis of molecular fingerprinting [52].

#### Dinucleotide relative abundance values ( $\rho^*$ ) and species-specific genome signatures [ $\delta^*(f,g)$ ]

$\rho^*$  values were in the normal range for all dinucleotides in all taxa investigated except for CG (over-represented in almost all genomes except in *V. campbellii*, *V. harveyi*, *V. shilonii*, *V. splendidus* and *Vibrio* sp. MED222) and TA (under-represented in almost all genomes except in *V. shilonii*, *P. angustum*, *P. profundum*, *A. fischeri* and *A. salmonicida*) (data not shown). The genomic dissimilarity value [ $\delta^*(f,g)$ ] of the genus *Vibrio* towards the genera *Photobacterium* and *Aliivibrio* was 38-66 and 35-59, respectively. The  $\delta^*(f,g)$  value between the genera *Photobacterium* and *Aliivibrio* were in the range of 20 to 34.  $\delta^*(f,g)$  values within the genera *Aliivibrio* and *Photobacterium* were 26.5 and 13, respectively.

$\delta(f,g)$  values within each vibrio species were between 1 and 4, whereas the interspecies  $\delta(f,g)$  were between 10 and 61 (Figure 3 and see Additional file 5; Table S3). Thus, the interspecies value was higher than the intergenus value. The  $\delta(f,g)$  values among the *Vibrio* core group members and *V. cholerae/V. mimicus* were at least 38. The  $\delta(f,g)$  values between the sisters species *V. cholerae-V. mimicus*, *V. harveyi-V. campbellii* and *V. parahaemolyticus-V. alginolyticus* were 14, 13 and 17, respectively. As vibrio species contain two chromosomes (one larger ca. 2.9 Mb and one smaller chromosome ca. 1.0 Mb) we also calculated the  $\delta(f,g)$  between the two chromosomes of the same strain. The  $\delta(f,g)$  values of the intragenomic comparison of the two chromosomes of complete vibrio genomes were between 10 and 18. The two chromosomes are essential for the cell survival and persistence, but yet they showed distinct patterns, suggesting a high genomic plasticity.

#### Codon usage bias

Overall codon usage bias was very similar among the vibrio species investigated (Table 1). There was little variation in  $N_c$  among the different genomes, with  $N_c$  ranging from 45 to 54. The  $N_c$  within the genera *Vibrio*, *Photobacterium* and *Aliivibrio* were 51-54, 48-51, and 45-48, respectively. Sister vibrio species had similar  $N_c$  values. Thus, there was not a clear differentiation of closely related taxa using the  $N_c$ .

The rose plot shows the difference of codon usage for representative vibrios. The frequency of each codon is plotted in red. Distinguishable signatures for each genus are apparent (Figure 5). For instance, the genus *Vibrio* represented by *V. cholerae* and *V. harveyi* do not have a preferential codon usage, while the genera *Photobacterium* and *Aliivibrio* use codons that tend to end in either A or U. For

instance, the frequency of UUA in *Photobacterium* and *Aliivibrio* genomes is higher than in *Vibrio* genomes, while the frequency of GCG is higher in *Vibrio* than *Photobacterium* and *Aliivibrio*.

## Discussion

### Genomic taxonomy

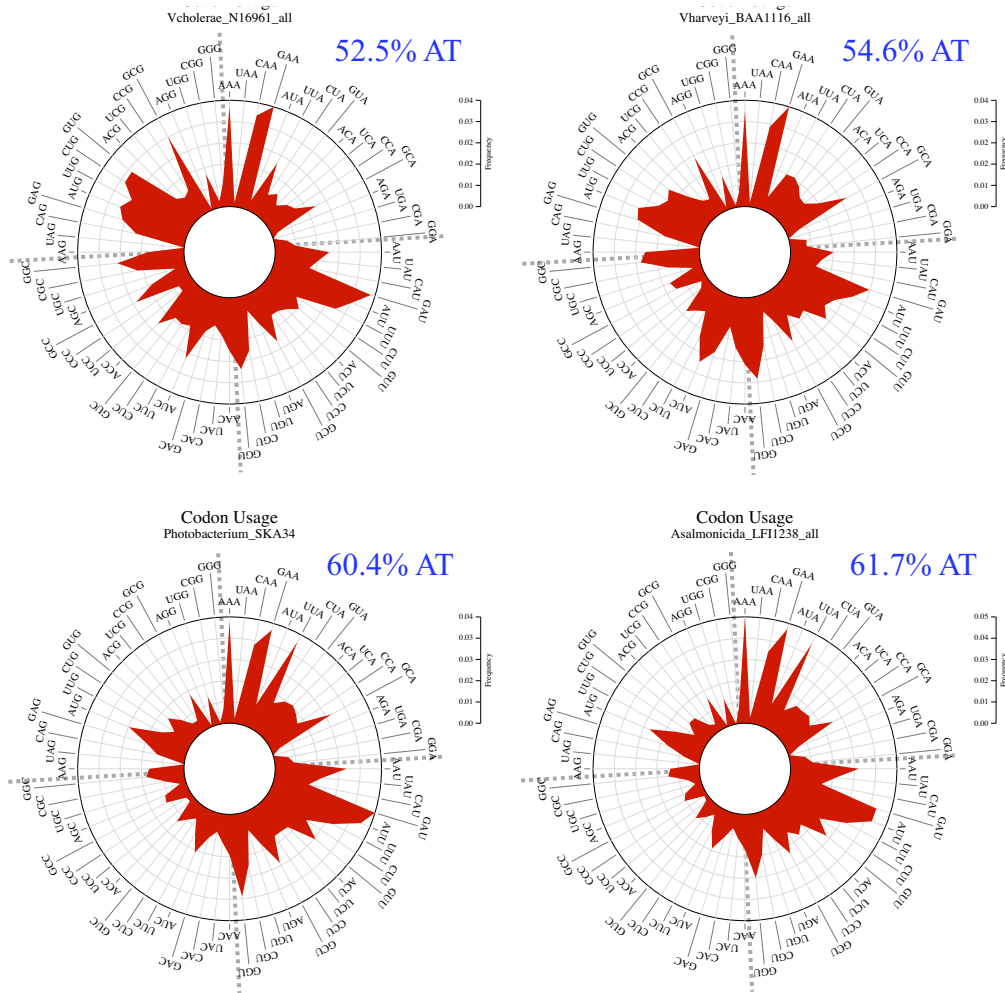
This study aimed at providing the underpins for the establishment of an online genomic taxonomy of vibrios. The methods used to extract taxonomic information from vibrios genomes are freely available in the web, have complementary taxonomic resolutions and are all amenable to automation for species and genera identification. Species identification is the major goal of microbial taxonomy. The identification of closely related sister species *V. cholerae - V. mimicus*, *V. alginolyticus - V. parahaemolyticus*, and *V. harveyi - V. campbellii* were evident in our study. The methods with the higher resolution for species and genera identification were, in order, MLSA, supertrees, and AAI. Karlin's genomic signature ( $\delta(f,g)$ ) performed well for species identification, whereas  $N_c$  appeared to be useful for differentiating genera.

### AAI

According to our analyses the AAI is one of the most useful genomic features for figuring out vibrio taxonomy. With the ever growing number of whole-genome sequences, this new method could be incorporated in a future re-valuation of the bacterial species definition. It is important to bear in mind that the number of orthologous genes shared between species depends on genome size and phylogenetic relationship [53,54]. *Vibrio* genomes have a wide variation in genome size, varying from 4 to 6 Mb (Table 1). As the genome size may influence the AAI, possibly the cut-off for species delineation will vary slightly when additional vibrio species are analysed.

### Genome signature dissimilarity $\delta^*(f,g)$

Karlin's genomic signature dissimilarity  $\delta^*(f,g)$  can be used for species identification in vibrios. Overall, the genomic signature of vibrios was more similar between closely related species than to distantly related species. However, species from different genera may have similar signatures. For instance, *V. mimicus* and *Photobacterium* spp. shared values of genomic signature dissimilarity in the range of 38-40, whereas *V. mimicus* and *V. splendidus* had a genomic signature dissimilarity of 57. *V. campbellii* and *Aliivibrio fischeri* had a genomic signature dissimilarity of 35, indicating that the taxonomic resolution of Karlin's genomic signature  $\delta^*(f,g)$  is lower for discriminating genera. Similar results were found in other studies concerning the resolution of this type of signature [55]. Genome signatures alone have significant limitations when used as phylogenetic markers for higher taxonomic



**Figure 5**  
**Codon usage rose plots for four representatives of different vibrio genera.** The vibrios with a higher AT content (bottom rose plots) have a codon usage bias of A or U at the third position of the codon. The frequency scale is represented at the right side of the rose plots.

levels *e.g.* genera to phyla. Apparently, the primary limitation is the lack of divergence in some phylogenetically distant related species that could result from absence of molecular clock. The equilibrium between mutational biases and selective constraints results in equilibrium in the oligonucleotide composition of a genome. Similar genome signatures between phylogenetically distant related species could arise from coincidental convergence due to crowding of the genome signature space derived from dinucleotide frequencies, which may not capture sufficient information to differentiate between distant taxa *e.g.* genera to phyla.

**Concordance between the methods**

The vibrio genome BLAST atlas was an useful tool for depicting compositional differences between genomes of different species. Using this tool, differences between the

sister species *V. cholerae* and *V. mimicus* in terms of gene content and DNA features were observed. Overall there was a significant correlation between the different methods (Table 2), but some methods had a stronger evolutionary signal and different taxonomic resolution than others. For instance, AAI and supertree showed the closest correlation with MLSA. All methods, except 16S rRNA and codon usage, provided significant ( $P < 0.001$ ; T test) taxonomic resolution for differentiation of species and genera of vibrios. In general, the taxonomic resolution of 16S rRNA and codon usage was restricted to differentiation of genera.

The AAI and the proteome matrix correlated well, yet the latter is measuring the fraction of proteins that are the same in both genomes, and the former is measuring the average identity of the amino acids of the proteins in

**Table 2: Pearson correlation coefficient (expressed as percentage) between different methods**

	1	2	3	4	5	6
1. 16S rRNA gene identity	100					
2. Identity in MLSA	86.5	100				
3. Identity in supertree analysis	91.1	98.4	100			
4. Average aminoacid identity (AAI)	85.9	97.7	96.9	100		
5. Karlin genome signature dissimilarity	71.5	85.3	82.3	84.9	100	
6. BLAST proteome identity	77.1	89.0	86.4	92.5	85.5	100

those matches. In addition, AAI uses 40% amino acid identity and > 70% of the aligned length of a protein which is stricter than the settings used (50% identity-50% length) to construct the proteome matrix. Because the Karlin's genomic signature dissimilarity indexes genome wide variation, its phylogenetic resolution is distinct of individual genetic marker genes. In addition, this signature considers variation in both coding and non coding genomic regions. This may explain why the correlation between the signature and the gene sequence based methods obtained in this study is slightly lower.

#### **Towards a new species definition in vibrios**

A new species definition is mandatory if one aims to establish an automatic identification of vibrios through a web-based server. So far, the 16S rRNA gene analysis has been applied for species definition and identification [2]. Its value for these purposes in vibrios is limited because of its low taxonomic resolution. This study showed enough WGS-based evidence to propose a new species definition in vibrios. In our hands, a vibrio species is defined as a group of strains that share > 95% DNA identity in MLSA and supertree gene sequence, > 96% AAI,  $\leq 10$  genome signature dissimilarity, and > 61% proteome identity. Strains of the same species and species of the same genus will form monophyletic groups on the basis of MLSA and supertree.

#### **Ecology and genomic features**

Each *Vibrio* species appear to have a specific ecologic niche. Genomes exhibit diverse patterns of species-specific compositional bias, *i.e.* GC content, GC and AT skews, codon bias, and mutation bias. The exact mechanisms that generate and maintain the genome signatures are complex, but possibly involve differences in species-specific properties of DNA replication and repair machineries [49,56]. In *Borrelia burgdorferi*, there is a bias related to the speed of the replication [57], whereas in Proteobacteria, DNA repair enzymes co-evolve with the genome signature [58]. The evolutionary distances between DNA repair and recombination orthologous enzymes (mainly those involved in the nucleotide excision repair system) were highly correlated with genome signature distances. On the other hand, there was a significantly lower correlation

between the evolutionary distances of the structural and metabolic enzymes and genome signature.

Environmental temperature and oxygen appear to influence the GC content of bacteria [59]. The frequencies of AA, TA, and TT dinucleotides were higher than the frequencies of AT, GC, and CG dinucleotides in the vaccine strain of *Pasteurella multocida* compared to the virulent strain. Although the vaccine strain is cultured at higher temperature, its GC content is lower than the virulent strain. The AA + TT dinucleotide increased significantly in the vaccine strain, which may represent an adaptation to increased culturing temperature because AA/TT dinucleotides are conformationally very stable. Higher culturing temperature increases spontaneous hydrolytic deamination of cytosine and 5-methylcytosine which, in turn, tend to decrease GC content [60,61]. Deamination and methylation favour nucleotide changes from G and C to A and T in a variety of microbial genomes [62].

Horizontal gene transfer may influence the genomic features of vibrios. The three genetic processes that mediate HGT often occur in vibrios and may cause phenotypic variation [63,64]. Such variation may confound a phenotype based identification. Another interesting feature of vibrios genomes is the presence of two chromosomes. The intragenomic dissimilarity between the two chromosomes of each vibrio strain is higher than the genomic dissimilarity between chromosome I of two strains of the same species. For instance, *V. cholerae* N16961 chromosomes I and II genomic dissimilarity  $\delta(f,g)$  was 12 and *V. cholerae* N16961 chromosome I and *V. cholerae* O395 chromosome I was 1. The fact that the two chromosomes of vibrios are dissimilar, with chromosome II less conserved than the chromosome I might support the hypothesis that the chromosome II was acquired by horizontal gene transfer [50]. The chromosome II has only a few essential housekeeping protein coding genes. This chromosome might have been a megaplasmid acquired by an ancestor prior diversification of the vibrios. Nearly 1/6 of the chromosome II (ca. 150 Kb) corresponds to a super-integron, a rapidly evolving region specialized in capture and loss of genes, and gene expression [65]. It is important to highlight that the fact that the two chromosomes

of the same strain are more dissimilar than the chromosomes of different strains does not hamper the use of genome signatures for identification. Chromosomes I of *V. cholerae* and *V. mimicus* have signature dissimilarity higher than the signature dissimilarity between chromosome I of two *V. cholerae* strains. The same holds true for the chromosome II.

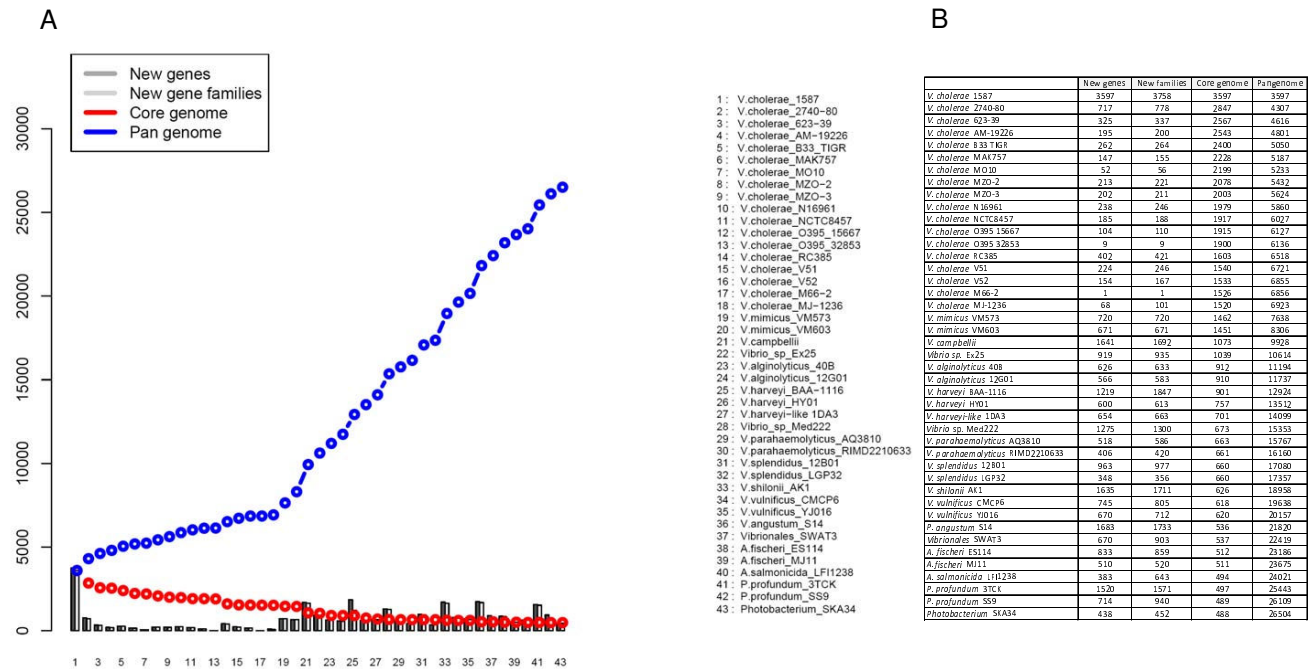
The pangenome (N = 26504 genes) and the core genome (N = 488 genes) of all vibrios correspond to a vast reservoir of genetic diversity (Figure 6). The core genome of *V. cholerae* (N = 1520 genes) might represent the minimum set of genes that allow survival of the species in the environment, whereas the *V. cholerae* pangenome (N = 6923 genes) reflects the ability of this species to occupy different niches in the environment. The increase in the pangenome is due to new strain-specific genes which were found in each new *V. cholerae* strain analysed. Unique genes, e.g. the sensor kinase *rscS*, found in the *A. fischeri* allow this strain to occupy a specific niche in the environment (*i.e.* to colonize its squid host) [66]. The major toxin genes (CT and TCP) of *V. cholerae* toxigenic strains allow these strains to cause disease, but they were also found in *V. mimicus*. However, *V. mimicus* has not caused epidemics so far.

### Conclusion

The availability of new technologies for ultra-rapid whole genome sequencing and the development of concepts in comparative genomics will allow for rapid and reliable automatable identification of microbial isolates through a web-based server. The concept of an online electronic taxonomy based on whole genome features as illustrated in this study will improve microbial taxonomy. Environmental biodiversity surveys and ecologic studies on vibrios will also benefit from this new approach to identification. In this new context, traditional molecular approaches (*i.e.* DDH, MLSA, AFLP, rep-PCR) may still be useful for the screen of large collections of strains that will subsequently be used in whole genome based identification schemes.

### Abbreviations

DDH: DNA-DNA hybridization; AFLP: Amplified Fragment Length Polymorphism; HGT: Horizontal Gene Transfer; HR: Homologous Recombination; WGS: Whole Genome Sequencing; T<sub>m</sub>: is the melting temperature of a double strand DNA molecule; ΔT<sub>m</sub>: is the difference between the T<sub>m</sub> of a given double strand DNA molecule and the T<sub>m</sub> of a hybrid of this molecule formed under controlled experimental conditions; CT: Cholera Toxin; TCP: Toxin Co-regulated Pilus.



**Figure 6**  
**Pangenome plot of vibrios.** Pangenome consists of panel A and panel B. The vibrio pangenome is around 26504 genes. *V. cholerae* has a pangenome of 6923 genes with clear increment of genes of its sister species *V. mimicus* (8306 genes).

## Authors' contributions

CCT carried out the computational analyses, phylogenetic and statistical analyses, analysed the results and wrote the manuscript. ACPV participated in the discussion and in the draft of the manuscript. RS and ATRV participated in the database construction and genomic annotation. TI carried out the pyrosequencing of the *V. mimicus* VM603 genome and helped writing the paper. NAJr obtained the vibrio samples and carried out the preliminary taxonomic identification of the genomes. DU carried out the pyrosequencing of the *V. alginolyticus* 40B, *V. harveyi*-like 1DA3 and *V. mimicus* VM573 genomes. DU and TV performed the BLAST atlas and matrix and the pangenome plot, and drafted the manuscript. FLT conceived the study, analysed the data and wrote the manuscript. All the authors have read and approved the final manuscript.

## Additional material

### Additional file 1

**Table S1.** BLAST matrix. The matrix lists the identity between proteomes of different strains of vibrios. The number of proteins and gene families in each genome are shown directly beneath the strain number. The hypotenuse (red) corresponds to the paralogs. The data provided the identity between proteomes of different strains of vibrios. The number of proteins and gene families in each genome are shown directly beneath the strain number. The hypotenuse (red) corresponds to the paralogs.

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### Additional file 2

**Figure S1A-C.** Phylogenetic trees based on the maximum parsimony method using 16S rRNA gene, MLSA (i.e. *ftsZ*, *gyrB*, *mreB*, *pyrH*, *recA*, *rpoA* and *topA*; 10,141 bp), and supertree (i.e. *aminopeptidase P*, *alaS*, *aspS*, *ftsZ*, *gltX*, *gyrB*, *hisS*, *ileS*, *infB*, *metG*, *mreB*, *pntA*, *pheT*, *pyrH*, *recA*, *rpoA*, *rpoB*, *rpsH*, *signal recognition particle protein*, *threonyl-tRNA synthetase*, *topA*, *valS* and 30S ribosomal protein S11; 41,617 bp). Bootstrap percentages after 2000 replications are shown. Because some genomes used in this study are not completely sequenced, for the comparison of 16S rRNA, MLSA and supertree, we used 16 genomes of vibrios. The genes used in MLSA and supertree were found only in these 16 genomes. The data provided the phylogenetic relationship between vibrio strains

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### Additional file 3

**Figure S2A-C.** Phylogenetic trees based on the neighbour-joining method using 16S rRNA gene, MLSA (i.e. *ftsZ*, *gyrB*, *mreB*, *pyrH*, *recA*, *rpoA* and *topA*; 10,141 bp), and supertree (i.e. *aminopeptidase P*, *alaS*, *aspS*, *ftsZ*, *gltX*, *gyrB*, *hisS*, *ileS*, *infB*, *metG*, *mreB*, *pntA*, *pheT*, *pyrH*, *recA*, *rpoA*, *rpoB*, *rpsH*, *signal recognition particle protein*, *threonyl-tRNA synthetase*, *topA*, *valS* and 30S ribosomal protein S11; 41,617 bp). Bootstrap percentages after 2000 replications are shown. Because some genomes used in this study are not completely sequenced, for the comparison of 16S rRNA, MLSA and supertree, we used 16 genomes of vibrios. The genes used in MLSA and supertree were found only in these 16 genomes. The data provided the phylogenetic relationship between vibrio strains

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### Additional file 4

**Table S2.** Percentage of average amino acid identity (AAI) between vibrio species. Representative genomes were used for the calculations. The data provided the percentage of average amino acid identity (AAI) between vibrio species.

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### Additional file 5

**Table S3.** Genomic dissimilarity [ $\delta(f,g)$ ] values between vibrio species. Representative genomes were used for the calculations. The data provided the genomic dissimilarity [ $\delta(f,g)$ ] values between vibrio species.

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## References

- Colwell RR: **Polyphasic taxonomy of the genus vibrio: numerical taxonomy of *Vibrio cholerae*, *Vibrio parahaemolyticus*, and related *Vibrio* species.** *J Bacteriol* 1970, **104(1)**:410-433.
- Wang Q, Garrity GM, Tiedje JM, Cole JR: **Naive Bayesian classifier for rapid assignment of rRNA sequences into the new bacterial taxonomy.** *Appl Environ Microbiol* 2007, **73(16)**:5261-5267.
- Woese CR, Fox GE: **Phylogenetic structure of the prokaryotic domain: the primary kingdoms.** *Proc Natl Acad Sci USA* 1977, **74(11)**:5088-5090.
- Stackebrandt E, Goebel BM: **A place for DNA-DNA reassociation and 16S ribosomal-RNA sequence analysis in the present species definition in bacteriology.** *Int J Syst Bacteriol* 1994, **49**:846-849.
- Wayne LG, Brenner DJ, Colwell RR, Grimont PAD, Kandler O, Krichevsky MI, Moore LH, Moore WEC, Murray RGE, Stackebrandt

- E, et al.: **Report of the ad hoc committee on reconciliation of approaches to bacterial systematics.** *Int J Syst Bacteriol* 1987, **37**:463-464.
6. Gevers D, Cohan FM, Lawrence JG, Spratt BG, Coenye T, Feil EJ, Stackebrandt E, Peer Y Van de, Vandamme P, Thompson FL, et al.: **Opinion: Re-evaluating prokaryotic species.** *Nat Rev Microbiol* 2005, **3(9)**:733-739.
  7. Gillis M, Vandamme P, De Vos P, Swings J, Kersters K: **Polyphasic taxonomy.** In *Bergey's Manual of Systematic Bacteriology: The Proteobacteria: Introductory Essays Volume 2*. Edited by: Brenner DJ, Krieg NR, Staley JT, Garrity GM. New York: Springer; 2005:43-48.
  8. Lawrence JG, Retchless AC: **The interplay of homologous recombination and horizontal gene transfer in bacterial speciation.** *Methods Mol Biol* 2009, **532**:29-53.
  9. Cohan FM, Koeppl AF: **The origins of ecological diversity in prokaryotes.** *Curr Biol* 2008, **18**:1024-1034.
  10. Coenye T, Gevers D, Peer Y Van de, Vandamme P, Swings J: **Towards a prokaryotic genomic taxonomy.** *FEMS Microbiol Rev* 2005, **29(2)**:147-167.
  11. Coenye T, Vandamme P: **Extracting phylogenetic information from whole-genome sequencing projects: the lactic acid bacteria as a test case.** *Microbiology* 2003, **149(Pt 12)**:3507-3517.
  12. Karlin S, Mrazek J, Campbell AM: **Compositional biases of bacterial genomes and evolutionary implications.** *J Bacteriol* 1997, **179(12)**:3899-3913.
  13. Phillipy AM, Mason JA, Ayanbule K, Sommer DD, Taviani E, Huq A, Colwell RR, Knight IT, Salzberg SL: **Comprehensive DNA signature discovery and validation.** *PLoS Comput Biol* 2007, **3(5)**:e98.
  14. van Passel MWJ, Kuramae EE, Luyf ACM, Bart A, Boekhout T: **The reach of the genome signature in prokaryotes.** *BMC Evol Biol* 2006, **6(84)**.
  15. Brown JR, Douady CJ, Italia MJ, Marshall WE, Stanhope MJ: **Universal trees based on large combined protein sequence data sets.** *Nat Genet* 2001, **28(3)**:281-285.
  16. Daubin V, Gouy M, Perriere G: **Bacterial molecular phylogeny using supertree approach.** *Genome Inform* 2001, **12**:155-164.
  17. Wolf YI, Rogozin IB, Grishin NV, Tatusov RL, Koonin EV: **Genome trees constructed using five different approaches suggest new major bacterial clades.** *BMC Evol Biol* 2001, **1**:8.
  18. Konstantinidis KT, Tiedje JM: **Towards a genome-based taxonomy for prokaryotes.** *J Bacteriol* 2005, **187(18)**:6258-6264.
  19. Goris J, Konstantinidis KT, Klappenbach JA, Coenye T, Vandamme P, Tiedje JM: **DNA-DNA hybridization values and their relationship to whole-genome sequence similarities.** *Int J Syst Evol Microbiol* 2007, **57(Pt 1)**:81-91.
  20. Wright F: **The 'effective number of codons' used in a gene.** *Gene* 1990, **87(1)**:23-29.
  21. Sawabe T, Kita-Tsukamoto K, Thompson FL: **Inferring the evolutionary history of vibrios by means of multilocus sequence analysis.** *J Bacteriol* 2007, **189(21)**:7932-7936.
  22. Thompson FL, Gevers D, Thompson CC, Dawyndt P, Naser S, Hoste B, Munn CB, Swings J: **Phylogeny and molecular identification of vibrios on the basis of multilocus sequence analysis.** *Appl Environ Microbiol* 2005, **71(9)**:5107-5115.
  23. Thompson CC, Thompson FL, Vicente AC, Swings J: **Phylogenetic analysis of vibrios and related species by means of atpA gene sequences.** *Int J Syst Evol Microbiol* 2007, **57(Pt 11)**:2480-2484.
  24. Thompson FL, Gomez-Gil B, Vasconcelos AT, Sawabe T: **Multilocus sequence analysis reveals that *Vibrio harveyi* and *V. campbellii* are distinct species.** *Appl Environ Microbiol* 2007, **73(13)**:4279-4285.
  25. Thompson CC, Thompson FL, Vicente AC: **Identification of vibrio cholerae and vibrio mimicus by multilocus sequence analysis (MLSA).** *Int J Syst Evol Microbiol* 2008, **58(Pt 3)**:617-621.
  26. Thompson FL, Austin B, Swings J: **The biology of vibrios.** Washington: ASM; 2006.
  27. Payne GW, Vandamme P, Morgan SH, Lipuma JJ, Coenye T, Weightman AJ, Jones TH, Mahenthalingam E: **Development of a recA gene-based identification approach for the entire Burkholderia genus.** *Appl Environ Microbiol* 2005, **71(7)**:3917-3927.
  28. Andersson AF, Lindberg M, Jakobsson H, Backhed F, Nyren P, Engstrand L: **Comparative analysis of human gut microbiota by barcoded pyrosequencing.** *PLoS ONE* 2008, **3(7)**:e2836.
  29. Eid J, Fehr A, Gray J, Luong K, Lyle J, Otto G, Peluso P, Rank D, Baybayan P, Bettman B, et al.: **Real-time DNA sequencing from single polymerase molecules.** *Science* 2009, **323(5910)**:133-138.
  30. Mahenthalingam E, Baldwin A, Drevinek P, Vanlaere E, Vandamme P, Lipuma JJ, Dowson CG: **Multilocus sequence typing breathes life into a microbial metagenome.** *PLoS ONE* 2006, **1**:e17.
  31. Hunt DE, David LA, Gevers D, Preheim SP, Alm EJ, Polz MF: **Resource partitioning and sympatric differentiation among closely related bacterioplankton.** *Science* 2008, **320(5879)**:1081-1085.
  32. Thompson FL, Klose KE: **Vibrio the First International Conference on the Biology of Vibrios.** *J Bacteriol* 2005, **188(13)**:4592-4596.
  33. Farmer JJ III, Janda M, Brenner FW, Cameron DN, Birkhead KM: **Genus I. *Vibrio Pacini 1854*.** In *Bergey's Manual of Systematic Bacteriology: The Proteobacteria: The Gamma-Proteobacteria Volume 2*. Edited by: Brenner DJ, Krieg NR, Staley JT, Garrity GM. New York: Springer; 2005:494-545.
  34. Thompson FL, Iida T, Swings J: **Biodiversity of vibrios.** *Microbiol Mol Biol Rev* 2004, **68(3)**:403-431. table of contents
  35. Urbanczyk H, Ast JC, Higgins MJ, Carson J, Dunlap PV: **Reclassification of *Vibrio fischeri*, *Vibrio logei*, *Vibrio salmonicida* and *Vibrio wodanis* as *Aliivibrio fischeri* gen. nov., comb. nov., *Aliivibrio logei* comb. nov., *Aliivibrio salmonicida* comb. nov. and *Aliivibrio wodanis* comb. nov.** *Int J Syst Evol Microbiol* 2007, **57(Pt 12)**:2823-2829.
  36. Pitcher DG, Saunders NA, Owen RJ: **Rapid extraction of bacterial genomic DNA with guanidium thiocyanate.** *Lett Appl Microbiol* 1989, **8**:151-156.
  37. Margulies M, Egholm M, Altman WE, Attiya S, Bader JS, Bemben LA, Berka J, Braverman MS, Chen YJ, Chen Z, et al.: **Genome sequencing in microfabricated high-density picolitre reactors.** *Nature* 2005, **437(7057)**:376-380.
  38. Almeida LG, Paixao R, Souza RC, Costa GC, Barrientos FJ, Santos MT, Almeida DF, Vasconcelos AT: **A System for Automated Bacterial (genome) Integrated Annotation--SABIA.** *Bioinformatics* 2004, **20(16)**:2832-2833.
  39. Tamura K, Dudley J, Nei M, Kumar S: **MEGA4: Molecular Evolutionary Genetics Analysis (MEGA) software version 4.0.** *Mol Biol Evol* 2007, **24(8)**:1596-1599.
  40. Hallin PF, Binnewies TT, Ussery DW: **The genome BLAST atlas-a GeneWiz extension for visualization of whole-genome homology.** *Mol Biosyst* 2008, **4(5)**:363-371.
  41. Pedersen AG, Jensen LJ, Brunak S, Staerfeldt HH, Ussery DW: **A DNA structural atlas for Escherichia coli.** *J Mol Biol* 2000, **299(4)**:907-930.
  42. Ussery DW, Borini S, Wassenaar T: **Computing for comparative genomics: bioinformatics for microbiologists.** London: Springer-verlag; 2009.
  43. Swofford DL, Olsen GJ, Waddell PJ, Hillis DM: **Phylogenetic inference.** In *Molecular Systematics* Edited by: Hillis DM, Moritz C, Mable BK. Sunderland: Sinauer; 1996:407-514.
  44. Saitou N, Nei M: **The Neighbor-Joining Method - a New Method for Reconstructing Phylogenetic Trees.** *Japanese Journal of Genetics* 1986, **61(6)**:611-611.
  45. Jukes TH, Cantor CR: **Evolution of protein molecules.** In *Mammalian protein metabolism* Edited by: Munro HN. New York: Academic Press; 1996:21-132.
  46. Felsenstein J: **Confidence limits on phylogenies: An approach using the bootstrap.** *Evolution* 1985, **39**:783-791.
  47. Altschul SF, Madden TL, Schaffer AA, Zhang J, Zhang Z, Miller W, Lipman DJ: **Gapped BLAST and PSI-BLAST: a new generation of protein database search programs.** *Nucleic Acids Res* 1997, **25(17)**:3389-3402.
  48. Rost B: **Twilight zone of protein sequence alignments.** *Protein Eng* 1999, **12(2)**:85-94.
  49. Karlin S: **Global dinucleotide signatures and analysis of genomic heterogeneity.** *Curr Opin Microbiol* 1998, **1(5)**:598-610.
  50. Heidelberg JF, Eisen JA, Nelson WC, Clayton RA, Gwinn ML, Dodson RJ, Haft DH, Hickey EK, Peterson JD, Umayam L, et al.: **DNA sequence of both chromosomes of the cholera pathogen *Vibrio cholerae*.** *Nature* 2000, **406(6795)**:477-483.
  51. Gonzalez-Escalona N, Martinez-Urtaza J, Romero J, Espejo RT, Jaykus LA, DePaola A: **Determination of molecular phylogenetics of *Vibrio parahaemolyticus* strains by multilocus sequence typing.** *J Bacteriol* 2008, **190(8)**:2831-2840.
  52. Thompson FL, Hoste B, Vandemeulebroecke K, Swings J: **Genomic diversity amongst *Vibrio* isolates from different sources**

- determined by fluorescent amplified fragment length polymorphism. *Syst Appl Microbiol* 2001, **24(4)**:520-538.
53. Bansal AK, Meyer TE: **Evolutionary analysis by whole-genome comparisons.** *J Bacteriol* 2002, **184(8)**:2260-2272.
  54. Snel B, Bork P, Huynen MA: **Genome phylogeny based on gene content.** *Nat Genet* 1999, **21(1)**:108-110.
  55. Mrazek J: **Phylogenetic Signals in DNA Composition: Limitations and Prospects.** *Mol Biol Evol* 2009, **26(5)**:1163-1169.
  56. Foerster KU, von Mering C, Hooper SD, Bork P: **Environments shape the nucleotide composition of genomes.** *EMBO Rep* 2005, **6(12)**:1208-1213.
  57. Vetsigian K, Goldenfeld N: **Genome rhetoric and the emergence of compositional bias.** *Proc Natl Acad Sci USA* 2009, **106(1)**:215-220.
  58. Paz A, Kirzhner V, Nevo E, Korol A: **Coevolution of DNA-interacting proteins and genome "dialect".** *Mol Biol Evol* 2006, **23(1)**:56-64.
  59. Xia X, Wei T, Xie Z, Danchin A: **Genomic changes in nucleotide and dinucleotide frequencies in *Pasteurella multocida* cultured under high temperature.** *Genetics* 2002, **161(4)**:1385-1394.
  60. Horst JP, Fritz HJ: **Counteracting the mutagenic effect of hydrolytic deamination of DNA 5-methylcytosine residues at high temperature: DNA mismatch N-glycosylase Mig.Mth of the thermophilic archaeon *Methanobacterium thermoautotrophicum* THF.** *EMBO J* 1996, **15(19)**:5459-5469.
  61. Lindahl T: **Instability and decay of the primary structure of DNA.** *Nature* 1993, **362(6422)**:709-715.
  62. Wang RF, Campbell W, Cao WW, Summage C, Steele RS, Cerniglia CE: **Detection of *Pasteurella pneumotropica* in laboratory mice and rats by polymerase chain reaction.** *Lab Anim Sci* 1996, **46(1)**:81-85.
  63. Meibom KL, Blokesch M, Dolganov NA, Wu CY, Schoolnik GK: **Chitin induces natural competence in *Vibrio cholerae*.** *Science* 2005, **310(5755)**:1824-1827.
  64. Vidgen M, Carson J, Higgins M, Owens L: **Changes to the phenotypic profile of *Vibrio harveyi* when infected with the *Vibrio harveyi* myovirus-like (VHML) bacteriophage.** *J Appl Microbiol* 2006, **100(3)**:481-487.
  65. Mazel D: **Integrans: agents of bacterial evolution.** *Nat Rev Microbiol* 2006, **4(8)**:608-620.
  66. Mandel MJ, Wollenberg MS, Stabb EV, Visick KL, Ruby EG: **A single regulatory gene is sufficient to alter bacterial host range.** *Nature* 2009, **458(7235)**:215-218.
  67. Reichelt JL, Baumann P, Baumann L: **Study of genetic relationships among marine species of the genera *Beneckeia* and *Photobacterium* by means of in vitro DNA/DNA hybridization.** *Arch Microbiol* 1976, **110(1)**:101-120.

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