Melia azedarach L. extracts and their activity on Musca domestica L. (Diptera: Muscidae)

Marise M. O. Cabral,^{*,1,2} Esteban R. F. Crescente,⁴ Paloma M. Mendonça⁴ Celma M. S. Gomes,³ Vanderleia C. Oliveira,⁴ Alphonse Kelecom⁵

¹Laboratório de Diptera, Instituto Oswaldo Cruz, Fundação Oswaldo Cruz, Av. Brasil 4365, Pavilhão Carlos Chagas, 413, 21045-900 Rio de Janeiro-RJ, Brazil,

²Laboratório de Insetos Vetores, Universidade Severino Sombra, Av. Expedicionário Oswaldo de Almeida Ramos, 280, 27700-000 Vassouras-RJ, Brazil,

³Laboratório de Hematozoários, Instituto Oswaldo Cruz, Fundação Oswaldo Cruz, Av. Brasil 4365, Pavilhão Carlos Chagas, 4º andar, 21045-900 Rio de Janeiro-RJ, Brazil,

⁴Laboratório de Ecoepidemiologia da Doença de Chagas, Instituto Oswaldo Cruz, Fundação Oswaldo Cruz, Av. Brasil 4365, 21045-900 Rio de Janeiro-RJ, Brazil,

⁵Departamento de Biologia Geral, Universidade Federal Fluminense, Outeiro de São João Batista s/n, Cx. Postal 100.436, 24001-970 Niterói-RJ, Brazil

RESUMO: "Extratos de *Melia azedarach* **L. e sua atividade sobre** *Musca domestica* **L.** (**Diptera: Muscidae**)". Os extratos brutos e as frações obtidas das sementes de *Melia azedarach* L. (Meliaceae) foram testados em *Musca domestica* Linnaeus, 1758 (Diptera: Muscidae). Os bioensaios mostraram inibição no desenvolvimento pós-embrionário das moscas e um significativo aumento do período larva recém eclodida- adulto. Além disso, o peso pupal foi reduzido e a proporção sexual alterada. Foi observada toxicidade para os ovos das moscas.

Unitermos: Extratos de planta, Diptera, *Melia azedarach*, regulação do desenvolvimento, atividade biológica.

ABSTRACT: Crudes extracts and fractions from seeds of *Melia azedarach* L. (Meliaceae) have been assayed on *Musca domestica* Linnaeus, 1758 (Diptera: Muscidae). Thus, the postembryonic development of the flies was reduced and the delay from newly hatched larvae to adults had significant increase. In addition, the pupal weights were reduced and the sexual ratio altered. Toxicity to fly eggs was also observed.

Keywords: Plant extract, Diptera, Melia azedarach, growth development, biological activity.

INTRODUCTION

The Meliaceae Melia azedarach L. is a huge tree, natural from South Asia and very common in Brazil, where it is known as "Cinamomo". A number of biological activities have already been described for crude extracts, fractions and metabolites isolated from this plant such as giardicial activity (Amaral et al., 2006), inhibitors of the enzyme acetylcholinesterase (Barbosa-Filho et al., 2006a), inhibitors of the angiotensin converting enzyme (Barbosa-Filho et al., 2006b) and antileprotic activity (Barbosa-Filho et al., 2007). Antitumor (Hartwell, 1971), anti-rheumatic, sedative and antiulcer properties (Duke & Wain, 1981; List & Horhammer, 1979) and used against flu and hypertension (Agra et al., 2008) are reported in traditional medicine. The plant also possesses insecticide (Cabral et al., 1995; Lepage et al., 1946) and vermicide activities (Zhao, 1984). Among secondary metabolites found in

M. azedarach, one should mention triterpenes (Cabral et al., 1996; Mulholland et al., 2000), steroids, aromatic compounds (Mulholland et al., 2000) and the interesting limonoid meliaternin (Carpinella et al., 2003). In previous studies, we described a new euphane triterpene (Kelecom et al., 1996) and four lignans (Cabral et al., 1995) from the methanol extract of the seeds of *M. azedarach* collected in the city of Niterói, RJ. This was the first report of lignans in the family Meliaceae. We also reported anti-moulting activity of this crude extract against the bloodsucking bug *Rhodnius prolixus* (Cabral et al., 1996). The study described the biological activity of extracts and fractions of *M. azedarach* L. on the postembryonic development of *Musca domestica* Linnaeus, 1758 (Diptera: Muscidae).

MATERIAL AND METHODS

Seeds of *M. azedarach* were dried, powdered

699

Artigo

and successively extracted with n-hexane, AcOEt (A) and MeOH (B). The oily hexane crude extract was then partitioned between cold hexane and 5% aqueous methanol to yield a hexane fraction (C) and a methanol fraction (D). Aliquots of the extracts A and B, and of the fractions C and D were carefully dried, traces of organic solvents eliminated under vacuum, and the organic residues bioassayed on *M. domestica*, a cosmopolitan diptera with high degree of synanthropy that acts as a mechanical and biological vector for enteroviruses, enterobacteria, protozoan cysts, helminth eggs and

larvae, and fungi (Cox et al., 1912, Coutinho et al., 1957, Greenberg, 1971, 1973, Harwood & James, 1979; Oliveira, 1999; Oliveira et al., 2002). Insects used in this study were brought up in the colony maintained in laboratory (Queiroz et al., 1991). Samples were tested in triplicate by topic application at the concentration of 100 μ g/ μ L on groups of 50 larvae (L1) of *M. domestica* and on egg masses (5 mg) at the concentration of 200 μ g/mg (1 μ L/mg). Insect viability, duration periods of larval, pupal and newly hatched larvae to the adult stage, were observed as well as sexual ratio and pupal weight.

Table 1. Viability (%) and sex ratio of post-embryonic development of *M. domestica* treated with extracts from *M. azedarach*, topic application on L1 larvae.

Treatment	Larval stage viability (%)	Pupal stage viability (%)	Larvae to adult viability (%)	Sex ratio
Control	85 ^a	85^{a} wt (mg) = 22.10 ±3.90 a	73 ^a	0.44
А	79 ^c ***	$68^{b} **$ wt (mg) = 20.17 ±3.28 b***	53 ^b **	0.50
В	77 ^c ***	$65^{b} *$ wt (mg) =20.88 ±3.31 a	50 ^b ***	0.55
С	73 ^c ***	68 ^b *** wt (mg) =21.52 ±3.47X a, c*	50 ^b ***	0.49
D	54 ^b ***	$69^{b} **$ wt (mg) = 20.41 ±3.69 b, c**	37 ^c ***	0.38

Triplicate experiments with groups of 50 larvae of *M. domestica*, each. Numbers followed by the same letter did not differ among themselves and those followed by different letters have a significant difference (* P < 0.05, ** P < 0.01, ***P < 0.001) when the Tukey test was used; wt = pupal weight (mg), values are mean ± standard deviation (X ± SD); sex ratio refer to ratio number of females *versus* number of males.

Table 2. Duration in days of post-embryonic development of *M. domestica* treated with extracts from *M. azedarach*, topic application on L1 larvae.

Treatment	Larval stage (days)	Pupal stage (days)		Larvae to adult (days)		
	$\mathrm{X}\pm\mathrm{SD}$	VI	$X \pm SD$	VI	$\mathbf{X} \pm \mathbf{S}\mathbf{D}$	VI
Control	6.58 ± 0.77 a	5-10	5.56 ± 0.69 a	2-7	12.08 ± 0.62 a	11-14
А	6.45 ± 0.66 a	5 -8	$5.89 \pm 0.52 \text{ b**}$	4-8	12.25 ± 0.68 a	10-14
В	$6.43 \pm 0.76 \text{ a*}$	5-10	$5.96 \pm 0.52 \text{ b***}$	5-7	12.35 ± 0.86 a	11-14
С	$6.72 \pm 0.81 \text{ b*}$	5-10	$5.96 \pm 0.64 \text{ b***}$	5-8	$12.63 \pm 0.83 \text{ b***}$	11-14
D	6.65 ± 0.74 a	5-10	$5.98 \pm 0.65 \text{ b***}$	2-7	$12.59 \pm 0.65 \text{ b***}$	11-14

Triplicate experiments with groups of 50 larvae of *M. domestica*, each. VI= variation interval. Means followed by the same letter did not differ among themselves and those followed by different letters have a significant difference (* P < 0.05, ** P < 0.01, ***P < 0.001), when the Tukey test was used. Values are mean \pm standard deviation (X \pm SD).

Table 3. Viability of eggs (%) of *M. domestica* treated with fraction D from *M. azedarach*, topic application on eggs mass at the concentration of 200 µg/mg.

Treatment	Number of eggs	Number of larvae	Egg viability (%)
Control	167	152	91 a
Acetone control	266	233	88 b *
Fraction D	245	54	22 c ***

Triplicate experiments with groups of eggs mass (5mg) of *M. domestica*, each. Numbers followed by the same letter did not differ among themselves and those followed by different letters have a significant difference (* P < 0.05, ***P < 0.001) when the test χ^2 was used.

The results were expressed by their means and standard deviations, and the significance established statistically using the χ^2 and the Tukey tests.

RESULTS

The results are summarized in Tables 1-3. Thus, the post-embryonic development of the flies appeared to be drastically reduced on treatment with samples B, C and D, showing a viabilility of 50, 50 and 37% for each sample, respectively (Table 1). The viability of the larval stage was reduced by all tested samples of *M. azedarach*, mainly by fraction D that induced a reduction of 31% when compared with those of the control groups. The pupal weight was reduced by all the samples as compared to the control group (22 mg) with concomitant reduction of the flies size; thus, mean weights of 20 mg (spreading: 10-28 mg) were observed when larvae were treated with A (P < 0.001) and D (P < 0.001) 0.01) respectively (Table 1). In addition, the sexual ratio (number females/number males) showed a reduction of the number of females (ratio = 0.38) in the group treated with fraction D (Table 1).

The duration of the pupal stage showed a significant increase when *M. domestica* were treated with samples A (P < 0.01), B (P < 0.001), C (P < 0.001) and D (P < 0.001). The duration of the newly hatched larvae to adults also had significant increase when treated with samples C (P < 0.001) and D (P < 0.001) (Table 2).

Treatment of egg mass with D, at the concentration of 200 μ g/mg egg mass, resulted in only 22% of hatched larvae (Table 3) thus proving the high toxicity of this fraction from *M. azedarach* seeds on *M. domestica* eggs.

DISCUSSION AND CONCLUSION

This study describes preliminary results of the analysis of the biological activity of M. azedarach extracts focusing toxicity, pupation inhibition and outbreak of M. domestica, looking for fractions and substances that might be efficient to control dipterans vectors of pathogens. Inhibition of the post-embryonic development of *M. domestica* was observed in extract D that induced 46% and 63% larval and outbreak inhibitions respectively. Some authors described that an extract of the seeds of this tree is able to exert a repellent activity on Locusta migratoria (Pradhan et al., 1962), and, that the acetone extract interferes on the pupal development of Culex pipiens (Al Sharook et al., 1991). Similarly, anti-molting activity was found in M. azedarach against the triatomine insect R. prolixus, vector of Chagas's disease (Cabral et al., 1995, 1996).

High toxicity against larvae and pupae of *M*. *domestica* was evidenced in fractions C and D, both obtained by partition of the hexane crude extract from the Meliaceae. Moderate to high mortality of *M*. *domestica*

was observed when these flies were exposed to extracts from Gymnospermae from New Zeeland (Singh & Upadhyay, 1993). A similar result was found for the AcOEt (80%) and hexane (85%) crude extracts on R. prolixus (Cabral et al., 1996). The same authors assayed extracts of *M. azedarach* and demonstrated molting inhibition and toxicity on the Hemiptera O. fasciatus and on the hematophagous triatomine R. prolixus (Cabral et al., 1999). Purifications of the crude extracts increased the activities. On fractionation, the anti-molting activity was found in fraction B, that inhibited 80% of the ecdysis of R. prolixus (Cabral et al., 1996), the active principle being identified as pinoresinol, that showed 90% (25 μ g/ μ L) and 65% (25 μ g/ μ L) molting inhibition on O. fasciatus and R. prolixus, respectively (Cabral et al., 1999).

Studies on species of Meliaceae of the genus *Trichilia* also appeared promising for insecticide use on soil plagues and on the armyworm *Spodoptera frugiperda* (Hernández et al., 1983). Similarly in the health area, several vegetal extracts are under study aiming at the control of insects, vectors of illnesses to man, such as hematophagous mosquitoes, domestic flies, cockroaches and earwigs (Lagunes et al., 1984, Simas et al., 2004).

In conclusion, the seed extracts of *M. azedarach* showed potent bioactivity against *M. domestica*, which is an important urban pest all over the world. Sample D demonstrated the highest activity. This seems to be the first report of biological activities of *M. azedarach* extracts against Muscidae insects.

ACKNOWLEDGEMENTS

This work was supported by grants from the Fundação de Amparo à Pesquisa do Estado do Rio de Janeiro (FAPERJ).

REFERENCES

- Agra MF, Silva KN, Basílio IJLD, França PF, Barbosa-Filho JM 2008. Survey of medicinal plants used in the region Northeast of Brazil. *Rev Bras Farmacogn 18*: 472-508.
- Al-Sharook Z, Balan K, Jiang Y, Rembold H 1991. Insect growth inhibitors from two tropical meliaceae. Effect of crude seed extracts on mosquito larvae. J Appl Entomol 111: 425-430.
- Amaral FMM, Ribeiro MNS, Barbosa-Filho JM, Reis AS, Nascimento FRF, Macedo RO 2006. Plants and chemical constituents with giardicidal activity. *Rev Bras Farmacogn 16 (Supl.)*: 696-720.
- Barbosa-Filho JM, Medeiros KCP, Diniz MFFM, Batista LM, Athayde-Filho PF, Silva MS, Cunha EVL, Almeida JRGS, Quintans-Júnior LJ 2006a. Natural products inhibitors of the enzyme acetylcholinesterase. *Rev Bras Farmacogn 16*: 258-285.
- Barbosa-Filho JM, Martins VKM, Rabelo LA, Moura MD,

Silva MS, Cunha EVL, Souza MFV, Almeida RN, Medeiros IA 2006b. Natural products inhibitors of the angiotensin converting enzyme (ACE). A review between 1980-2000. *Rev Bras Farmacogn 16*: 421-446.

- Barbosa-Filho JM, Nascimento-Júnior FA, Tomaz ACA, Athayde-Filho PF, Silva MS, Cunha EVL, Souza MFV, Batista LM, Diniz MFFM 2007. Natural products with antileprotic activity. *Rev Bras Farmacogn 17*: 141-148.
- Cabral MMO, Garcia ES, Kelecom A 1995. Lignanes from the Brazilian *Melia azedarach*, and their activity in *Rhodnius prolixus* (Hemiptera, Reduviidae). *Mem Inst Oswaldo Cruz 90:* 759-763.
- Cabral MMO, Garcia ES, Rembold H, De Simone SG, Kelecom A 1996. Antimoulting activity in Brazilian Melia azedarach. Mem Inst Oswaldo Cruz 91: 117-118.
- Cabral MMO, Kelecom A, Garcia ES 1999. Effects of the lignan, pinoresinol on the moulting cycle of the bloodsucking bug, *Rhodnius prolixus*, and of the milkweed bug, *Oncopeltus fasciatus*. *Fitoterapia 70:* 561-567.
- Carpinella MC, Defago MT, Valladares G, Palacios SM 2003. Antifeedant and insecticide properties of a limonoids from of Melia *azedarach* (Meliaceae) with potential use for pest management. *J Agric Food Chem 51*: 369-374.
- Coutinho J, Taunay A, Lima LDC 1957. Importância de Musca domestica como vetores de agentes patogênicos para o homem. Rev Inst Adolfo Lutz 17: 5-23.
- Cox GL, Lewis FC, Glynn EE 1912. The number and varieties of bacteria carried by the common housefly in sanitary and unsanitary city areas. *J Hyg 12*: 290-312.
- Duke JA, Wain KK 1981. Medicinal Plants of the World. 3 vol. Computer index with more than 85,000 entries. Plants genetics and germplasm Institute. Agriculture Research Service, Beltsville, Maryland.
- Greenberg B 1971. Flies and Disease. vol 1: Ecology, Classification and Biotic Association. Princeton Univ. Press. Princeton, NI.
- Greenberg B 1973. Flies and Disease. Vol 2: Biology and disease transmission. Princeton Univ. Press. Princeton, NI.
- Harwood RF, James MT 1979. Entomology in human and animal health. Macillan Publishing Co. NY.
- Hartwell JL 1971. Plants used against cancer. A survey. Lloydia 34: 103.
- Hernández XE, Inzunza MF, Solano SCB 1983. Insectos de control de plagas y enfermedades identificadas en la agricultura tradicional en México. *Rev Chapingó 40*: 55-56.
- Kelecom A, Cabral MMO, Garcia ES 1996. A new euphane from the Brazilian Melia azedarach. J Braz Chem Soc 7: 39-41.
- Lagunes TA, Arenas JC, Rodríguez HC 1984. Extractos acuosos y polvos vegetales con propiedades insecticidas. Chapingo, Colégio de Postgraduados, Centro de Entomologia y acarologia, 203p.
- Lepage HS, Giamotti O, Orlando A 1946. Proteção de culturas contra gafanhotos por meio de extratos de *Melia azedarach. Biológico 12*: 265-271.
- List PH, Horhammer L 1979. Hager's handbuch der

pharmazeutischen praxis. vol 6. Springer-Verlag, Berlin.

- Mulholland DA, Parel B, Coombes PH 2000. The Chemistry of the Meliaceae and Ptaeroxylaceae of Southern and Eastern Africa and Madagascar. *Curr Org Chem* 4: 1011-1054.
- Oliveira VC 1999. Avaliação do Potencial de Dípteros Caliptrados como veiculadores de ovos de helmintos na Fundação Rio-Zoo. Tese de Doutorado. Universidade Federal Rural do Rio de Janeiro, 86p.
- Oliveira VC, Mello RP, d'Almeida JM 2002. Dípteros Muscóides como Vetores Mecânicos de Ovos de Helmintos em Jardim Zoológico, Brasil. *Rev Saúde Pública 36*: 614-620.
- Pradhan HS, Yotwani MG, Raí BK 1962. The neem seed deterrent to Locust. Ind Farming12: 7-11.
- Queiroz MMC, Milward-de-Azevedo EMV 1991. Técnicas de criação e alguns aspectos da biologia de *Chrysomya albiceps* Wiedemann (Diptera, Calliphoridae), em condições de laboratório. *Rev Bras Zool 8:* 75-84.
- Simas NK, Lima EC, Conceição SR, Kuster RM, Oliveira Filho AM, Lage CLS 2004. Produtos naturais para o controle da transmissão da dengue-atividade larvicida de Myroxylon balsamum (óleo vermelho) e de terpenóides e fenilpropanóides. Quim Nova 27: 46-49.
- Singh G, Upahyay RK 1993. Essential oil: a potent source of natural pesticides. J Sci Ind Res 52: 676-683.
- Zhao CX 1984. Effect of *Melia azedarach* extract on Schistosomiasis in mice. *Chinese Med J* 97: 910-912.