



Early View

Research letter

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Higher IL-6 Levels and Changes in TGF- β are Associated with Lung Impairment in Pulmonary Tuberculosis.

Akshay N Gupte^{1,2}, Sriram Selvaraju³, Sanjay Gaikwad⁴, Vidya Mave^{1,2}, Pavan Kumar⁵, Subash Babu⁵, Bruno B. Andrade^{6,7,8}, William Checkley⁹, Robert Bollinger^{1,2} and Amita Gupta^{1,2} for the CTRIUMPH study team.

¹ Division of Infectious Diseases, Johns Hopkins University School of Medicine, Baltimore, USA

² Center for Clinical Global Health Education, Johns Hopkins University School of Medicine, Baltimore, USA

³ Department of Clinical Research, National Institute for Research in Tuberculosis, Chennai, India

⁴ Department of Pulmonary Medicine, Byramjee-Jeejeebhoy Government Medical College, Pune, India

⁵ National Institutes of Health – National Institute for Research in Tuberculosis – International Center for Excellence in Research, Chennai, India

⁶ Instituto Gonçalo Moniz, Fundação Oswaldo Cruz (FIOCRUZ), Salvador, Brazil

⁷ Multinational Organization Network Sponsoring Translational and Epidemiological Research (MONSTER) Initiative, Salvador, Brazil

⁸ Universidade Salvador (UNIFACS), Laureate Universities, Salvador, Brazil

⁹ Division of Pulmonary Medicine, Johns Hopkins University School of Medicine, Baltimore, USA

Corresponding author:

Akshay N Gupte, agupte1@jhmi.edu

Division of Infectious Diseases, Johns Hopkins University School of Medicine

Introduction

Pulmonary tuberculosis (PTB) is associated with granuloma formation, necrosis and cavitation in lung tissue. Lung injury in PTB can persist despite microbiological cure and is associated with chronic obstructive pulmonary disease (COPD) independent of smoking exposure[1]. Furthermore, pulmonary sequelae of PTB are an under-recognized cause of respiratory disability and excess mortality[2].

Our prior work in India found up to 50% of PTB cases had impaired lung function post-treatment[3]. We also found a positive correlation between the duration of symptomatic illness prior to PTB treatment and severity of lung impairment post-treatment. These data suggest that acute lung injury in PTB likely occurs during the pre- or early-treatment period, and that lung tissue repair and remodeling during treatment may play an important role in post-PTB lung disease.

However, the pathogenesis of PTB-associated lung disease is unclear. Pre-clinical studies have suggested a pro-fibrotic role of interleukin (IL)1- β , IL-6, IL-4, IL-17 and transforming growth factor (TGF)- β s in the lungs[4, 5]; markers which also play an important role in the host immune response in PTB. Further, tumor necrosis factor alpha (TNF- α) and matrix metalloproteinases (MMPs) have been implicated in lung tissue destruction and cavitation in PTB[6].

There is growing interest in the utility of host directed therapies (HDTs); adjunctive therapies that modulate immune mechanisms in the host; for improving clinical outcomes. HDTs with anti-inflammatory properties could potentially prevent or limit the extent of lung injury in PTB. Therefore, we measured the association of

potentially modifiable inflammatory markers implicated in lung tissue destruction and fibrosis, with respiratory morbidity and impaired lung function in a prospective cohort of adults with drug sensitive PTB.

Methods

We randomly selected adults (18 years and older) with microbiologically confirmed PTB receiving standard multi-drug therapy in the CTRIUMPH study in India[7]. Participants with drug-resistant disease, prior PTB, or prior chronic lung diseases were excluded. PTB cases were prospectively evaluated at initiation, 2 months and completion of treatment for respiratory health status using the Saint Georges Respiratory Questionnaire (SGRQ). Pre- and post-bronchodilator spirometry was performed at treatment completion according to ATS/ERS guidelines[8]. Forced expiratory volume in the first second (FEV₁), forced vital capacity (FVC) and the FEV₁ to FVC ratio (FEV₁/FVC) were z-score standardized for analysis using Global Lung Initiative reference equations[9].

Plasma samples collected at initiation, 2 months and completion of treatment were tested, in duplicates, for TNF- α , IL-1 β , IL-4, IL-6, IL-17, TGF- β 1, TGF- β 2, TGF- β 3, MMP-1, MMP-3, MMP-7 and tissue inhibitors of MMPs (TIMP)-1, TIMP-2, TIMP-3 and TIMP-4 concentrations at the National Institutes of Health – National Institute for Research in Tuberculosis – International Center for Excellence in Research, Chennai, India using multiplex enzyme-linked immunosorbent assay (Bio-Rad Laboratories, CA, USA) on Luminex platform using manufacturer recommended protocols. We used uni- and multi-variable random effects regression to measure the pooled association between log₂-transformed cytokine concentrations and total SGRQ scores during

treatment. A difference of 4-points or more in the total SGRQ score was considered clinically relevant[10]. Uni- and multi-variable linear regression and Spearman's correlation coefficient were used to measure the association of cytokine concentrations, and their change during treatment, with post-bronchodilator lung function at treatment completion. Multivariable regression analyses accounted for confounding by age, sex, smoking exposure, diabetes and markers of PTB disease severity, including body mass index (BMI), smear grade, cavitory disease and duration of symptomatic illness prior to treatment initiation. P-values were adjusted for multiple comparisons using the Benjamini-Hochberg procedure and a 10% false discovery rate.

Results

We enrolled 30 PTB cases contributing 90 person-visits from the CTRIUMPH study. Participants selected for this analysis were comparable to those not selected in terms of their baseline characteristics. Overall, 20 (74%) were male, 9 (31%) ever-smoked, 2 (7%) had HIV coinfection, 7 (26%) had diabetes and 11 (37%) had cavitation on chest X-ray. The median (IQR) age and BMI was 36 (28-50) years and 18.1 (16.0-20.0) kg/m², respectively. Cytokine concentrations did not differ significantly by cavitory disease, sputum smear grade or BMI.

Cytokine concentrations at treatment initiation were not associated with lower lung function at treatment completion. All cytokine concentrations declined during treatment. However, greater declines in TGF- β 2 during treatment were associated with higher FEV1/FVC z-scores post-treatment. Specifically, greater declines in TGF- β 2 during the first two months of treatment, but not the last four months, were associated with higher FEV1/FVC post-treatment (0.78-point higher z-score per 2-fold decline in

TGF- β 2, 95%CI 0.28 to 1.29, $p=0.005$) in the univariable analysis. After adjusting for potential confounders, including markers of disease severity, a 2-fold decline in TGF- β 2 was associated with a 0.80-point higher z-score (95%CI 0.30 to 1.30, $p=0.005$). Interestingly, PTB cases in the lowest tertile of FEV1/FVC z-scores at treatment completion had a paradoxical increase in TGF- β 2 levels during the first two months of treatment compared to PTB cases with FEV1/FVC z-scores in the highest tertile ($p=0.005$) (Figure-1, Panel A).

The median (IQR) SGRQ score was 43 (29-55) points at treatment initiation and declined to 6 (3-19) points at treatment completion ($p<0.001$) (Figure-1, Panel B). Higher levels of IL-6 were associated with higher SGRQ scores during treatment (3-point higher SGRQ score per 2-fold higher IL-6 concentrations, 95%CI 2 to 5, $p=0.002$) in the univariable analysis. After adjusting for potential confounders, including markers of disease severity, a 2-fold higher IL-6 concentration was associated with a clinically relevant 4-point higher SGRQ score (95%CI 1 to 5, $p=0.004$) during treatment.

We did not find a statistically significant association between TNF- α , IL-1 β , IL-4, IL-17, MMP-1, MMP-3, MMP-7, TIMP-1, TIMP-2, TIMP-3 and TIMP-4, and total SGRQ scores or impaired lung function in our cohort.

Discussion

TGF- β has been implicated in pulmonary fibrosis and airway remodeling through cellular growth and differentiation[11]. Pre-clinical studies have described the role of TGF- β in granuloma formation in PTB[12]. A novel finding of our study was the association between greater declines in TGF- β 2 levels during early PTB treatment and better lung function post-treatment. Importantly, PTB patients in the lowest tertile of lung function

post-treatment had a paradoxical increase in TGF- β 2 levels during the first two months of treatment. These data support the role of TGF- β in post-PTB lung impairment and suggest that facilitating rapid resolution of TGF- β through immunomodulatory HDTs during early treatment may mitigate post-PTB lung sequelae.

Prior studies in COPD patients have demonstrated an inverse relationship between IL-6 and functional capacity [13]. However, the role of IL-6 in PTB-associated respiratory morbidity has not been studied. We have previously shown that poor respiratory health status was associated with all-cause mortality in PTB[14]. Here, we report that PTB patients with higher IL-6 levels had worse respiratory health status during treatment. While these data suggest a role of IL-6 modulation for reducing respiratory morbidity in PTB, the impact of elevated IL-6 levels on long-term lung impairment and mortality needs further investigation.

Our single-cohort study is limited by its small sample size and we are likely underpowered to detect clinically relevant associations of a smaller magnitude. Well-powered validation studies in independent cohorts are needed to confirm our study findings. We additionally did not measure lung function at PTB treatment initiation or assess progression of post-PTB lung impairment. Despite these limitations, we report a novel association of elevated IL-6 and slow-to-resolve TGF- β , with lung impairment in PTB. Given the availability of US Food and Drug Administration and European Medicines Agency approved IL-6 and TGF- β inhibitors, our study findings have potentially important implications for the optimal timing and immune targets of future HDT trials to prevent PTB-associated lung disease.

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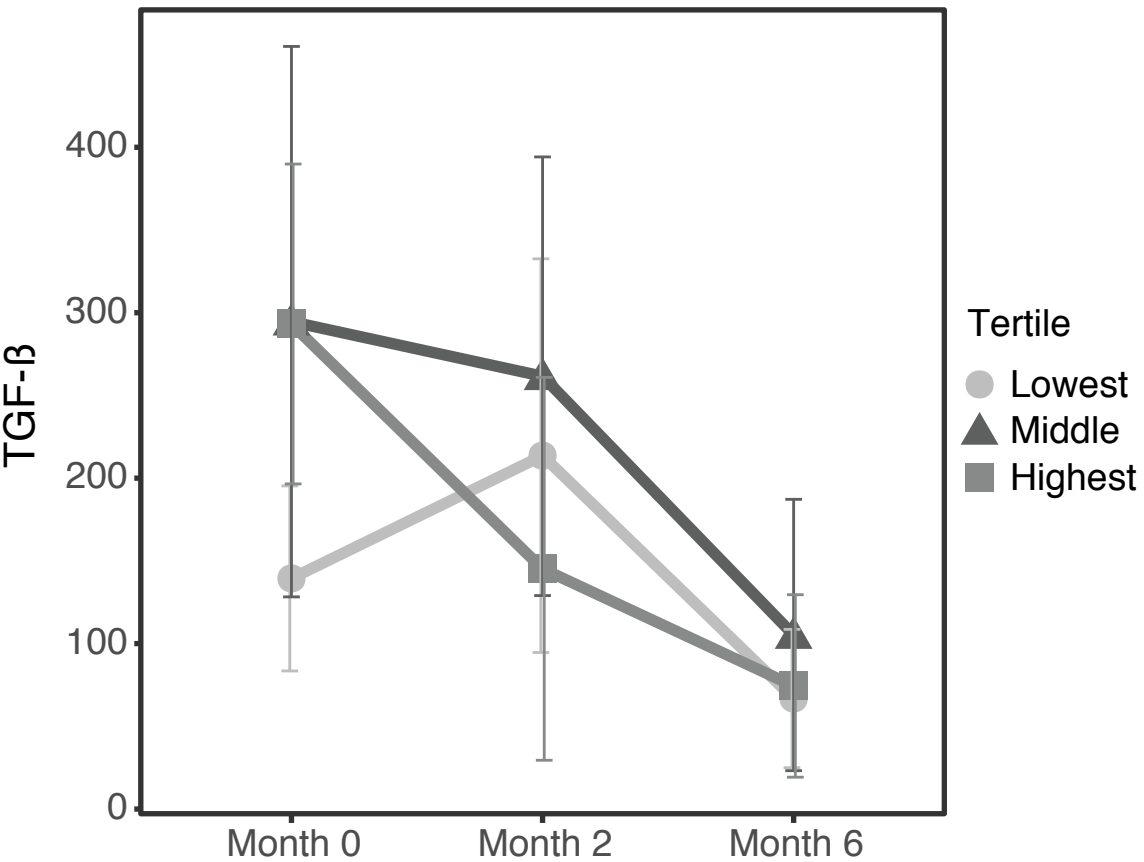
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Figure-1, Panel A: Trends in TGF- β 2 during PTB treatment stratified by tertiles of FEV1/FVC z-scores at treatment completion.

Y-axis depicts the concentration of TGF- β 2 in pg/mL. X-axis depicts the duration of PTB treatment. FEV1/FVC z-score thresholds for lowest and middle tertile were -0.62 and 0.10, respectively. Relative to PTB cases with FEV1/FVC z-scores in the highest tertile at treatment completion, those in the lowest tertile had a paradoxical increase in TGF- β 2 concentrations during the first two months of treatment.

Figure-1, Panel B: Trends in IL-6 and total SGRQ scores during PTB treatment.

Y-axis (left) depicts IL-6 concentrations while the Y-axis (right) depicts median SGRQ scores. X-axis depicts the duration of PTB treatment. The median SGRQ score declined from 43 points at treatment initiation to 6 points at treatment completion, and correlated with IL-6 concentrations during PTB treatment.

A**B**