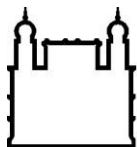


INSTITUTO CARLOS CHAGAS
DOUTORADO EM BIOCÊNCIAS E BIOTECNOLOGIA

FERNANDA TOMIOTTO PELLISSIER

**CARACTERIZAÇÃO DE CÉLULAS E MEDIADORES ENVOLVIDOS NA
PATOGÊNESE DA LESÃO CAUSADA POR *LEISHMANIA AMAZONENSIS* EM
MODELO EXPERIMENTAL**

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FIOCRUZ
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Tese apresentada ao Programa de Pós-Graduação em Biociências e Biotecnologia do Instituto Carlos Chagas como parte dos requisitos para obtenção do título de Doutora em Biociências e Biotecnologia.

Orientador: Dr. Wander Rogério Pavanelli

Co-orientador: Dr. Juliano Bordignon

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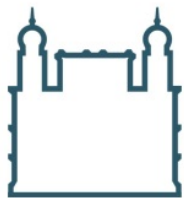
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**Ata da Sessão Pública de exame de tese para obtenção do grau de Doutora em
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Aos 03 dias do mês de março de dois mil e vinte um, às 9:00 horas, através de teleconferência <https://us02web.zoom.us/j/5953384663?pwd=NEo3T09MTEFZSIFHeEJHMzhMTEFhQT09>, reuniu-se a Banca Examinadora designada pelo Colegiado do Programa de Pós-Graduação em Biociências e Biotecnologia, composta pelos Professores: Dra. Letusa Albrecht, Dra. Carolina Panis e Dra. Ivete Conchon Costa, com a finalidade de julgar a tese da candidata **Fernanda Tomiotto Pellissier**, intitulada: **“CARACTERIZAÇÃO DE CÉLULAS E MEDIADORES ENVOLVIDOS NA PATOGÊNESE DA LESÃO CAUSADA POR LEISHMANIA AMAZONENSIS EM MODELO EXPERIMENTAL”**, sob a orientação de Dr. Wander Rogério Pavanelli e Co -orientação de Dr. Juliano Bordignon, para obtenção do grau de **Doutora** em Biociências e Biotecnologia. A candidata teve até 45 (quarenta e cinco) minutos para a apresentação, e cada examinador teve um tempo máximo de arguição de 30 (trinta) minutos, seguido de 30 (trinta) minutos para resposta do (a) candidato(a) ou de 60 (sessenta) minutos quando houve diálogo na arguição. O desenvolvimento dos trabalhos seguiu o roteiro de sessão de defesa, estabelecido pela Coordenação do Programa, com abertura, condução e encerramento da sessão solene de defesa feito pelo(a) Presidente **Dra. Letusa Albrecht**. Após haver analisado o referido trabalho e arguido a candidata, os membros da banca examinadora deliberaram pela:

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Na forma regulamentar foi lavrada a presente ata que é abaixo assinada pelos membros da banca, na ordem acima determinada, e pela candidata.

Dra. Dra. Letusa Albrecht – ICC/PR

Curitiba, 03 de março de 2021

Dra. Dra. Carolina Panis – UNIOESTE

Dra. Dra. Ivete Conchon Costa – UEL

Candidato(a): Fernanda Tomiotto Pellissier

À minha eternamente amada avó
Ilma Rossi Tomiotto.

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Filipenses 4:6,7

RESUMO

A leishmaniose cutânea é uma doença infecciosa zoonótica com ampla distribuição mundial. Parasitos do gênero *Leishmania* são os agentes etiológicos da doença, sendo que diferentes espécies podem desencadear manifestações clínicas que variam desde infecções autocuráveis à lesões crônicas desfigurantes. Os macrófagos são as células hospedeiras finais dos parasitos, e células chave para o direcionamento da resposta imune que culmina em manifestações clínicas. M1 e M2 são os dois principais fenótipos de macrófagos. M1 é um subtipo pró-inflamatório com propriedades microbicidas, e M2 é um subtipo antiinflamatório/regulador que está relacionado à resolução da inflamação e reparo tecidual. No entanto, o entendimento da resposta imune efetiva à infecção por esses protozoários ainda apresenta muitas lacunas, cuja compreensão pode ser fundamental para o potencial desenvolvimento de drogas e vacinas. Assim, investigamos as moléculas e mediadores locais envolvidos no desenvolvimento da lesão cutânea em modelos experimentais de suscetibilidade (BALB/c) e resistência parcial (C57BL/6) à infecção por *Leishmania amazonensis*. Verificou-se que camundongos BALB/c apresentaram pior evolução da doença em comparação com camundongos C57BL/6, com maior carga parasitária e níveis elevados de IL-6, enquanto C57BL/6 apresentaram maiores níveis de INF- γ e O₂⁻ após 11 semanas de infecção. Um pico de macrófagos locais foi observado após 24h de infecção em ambas as linhagens de camundongos estudadas, seguido por um novo aumento após 240h apenas em camundongos C57BL/6. Quando os marcadores de macrófagos M1 e M2 (iNOS, MHC-II, CD206 e arginase-1 [Arg-1]) foram analisados, foi identificado um aumento pronunciado nos níveis de Arg-1 em BALB/c após 11 semanas de infecção, enquanto em C57BL/6 houve predomínio de marcadores M2 coincidindo com um segundo pico de infiltração macrofágica em 240h, e com maior deposição de colágeno tipo III e resolução da lesão. Para confirmar que os resultados obtidos estavam relacionados à presença de macrófagos e sua capacidade de controlar a infecção, foi realizada a transferência adotiva de macrófagos peritoneais de C57BL/6 para BALB/c infectados. Os dados obtidos na 11^a semana revelaram que a transferência de macrófagos promoveu redução do edema e do número de parasitos no local da lesão, além de reduzir os níveis de Arg-1 sem afetar os níveis de iNOS. Assim, pode-se concluir que camundongos C57BL/6 têm uma melhor evolução da infecção causada por *L. amazonensis*, baseada em um balanço entre inflamação e resposta antioxidante, coincidindo com maior deposição de colágeno e melhor reparo tecidual, enquanto camundongos BALB/c apresentam elevados níveis de Arg-1 nos tempos tardios de infecção, marcador relacionado ao desenvolvimento e gravidade da doença. A pior evolução parece estar envolvida com o recrutamento de macrófagos ricos em Arg-1, uma vez que a transferência adotiva de macrófagos de camundongos C57BL/6 para BALB/c resultou em melhor evolução de lesão, com menores níveis de Arg-1.

Palavras-chave: Leishmaniose; Macrófagos; M1; M2; Arginase-1; iNOS; Colágeno.

ABSTRACT

Cutaneous leishmaniasis is a zoonotic infectious disease with broad world distribution. *Leishmania* parasites are the etiologic agents of the disease, and different species can trigger clinical manifestations ranging from self-curing infections to disfiguring chronic lesions. Macrophages are the final host cells for the parasites and key cells for the immune response and clinical manifestations. M1 and M2 are the two main macrophage phenotypes. M1 is a pro-inflammatory subtype with microbicidal properties, and M2 is an anti-inflammatory/regulatory subtype related to inflammation resolution and tissue repair. However, the understanding of the effective immune response to *Leishmania* infection still has many gaps, which may be fundamental to the potential development of drugs and vaccines. Thus, we investigated the molecules and local mediators involved in the development of the skin lesion in experimental models of susceptibility (BALB/c) and partial resistance (C57BL/6) to *Leishmania amazonensis* infection. BALB/c mice had a worse disease outcome compared to C57BL/6 mice, with almost 15 times higher parasitic load, ulcerated lesion formation, and higher levels of IL-6, while C57BL/6 presented higher levels of INF- γ and O₂⁻ after 11 weeks of infection and no lesion ulcerations. A peak of local macrophages was observed after 24h of infection in both studied mice strains, followed by a further increase after 240h only in C57BL/6 mice. When the M1 and M2 macrophage markers (iNOS, MHC-II, CD206, and arginase-1 [Arg-1]) were analyzed, we found that in BALB/c there was a pronounced increase in Arg-1 levels after 11 weeks of infection, whereas in C57BL/6 there was an initial predomination of markers from both profiles, followed by an M2 predominance, that coincided with the second peak of macrophage infiltration, 240h after the infection, greater deposition of type III collagen and lesion resolution. To confirm that the results obtained were related to the presence of macrophages and their ability to control the infection, we carried out an adoptive transfer of these cells from C57BL/6 to infected BALB/c. The data obtained at the 11th week revealed that the adoptive transfer of macrophages promoted a reduction in edema and number of parasites at the lesion site, besides inducing lower levels of Arg-1 without affecting the iNOS levels. Thus, C57BL/6 mice have a more effective response against *L. amazonensis*, based on a balance between inflammation and antioxidant response, coinciding with better tissue repair, while BALB/c mice have an excessive Arg-1 production in late infection times, which is related to the development of disease severity. The worst evolution seems to be involved with the recruitment of Arg-1 related macrophages, since the adoptive transfer of macrophages from C57BL/6 mice to BALB/c resulted in better outcomes, with lower levels of Arg-1.

Keywords: Leishmaniasis; Macrophages; M1; M2; Arginase-1; iNOS; Collagen.

LISTA DE ILUSTRAÇÕES

Capítulo 1

- Figura 1 – *Role of saliva vectors on macrophage polarization.* 8
Figura 2 – *Role of M1 and M2 macrophages in Leishmania infection.* 10

Capítulo 2

- Figura 1 – *Evolution of L. amazonensis infection in BALB/c and C57BL/6 mice ..* 27
Figura 2 – *Cytokines levels of BALB/c and C57BL/6 mice infected with L. amazonensis* 28
Figura 3 – *Superoxide anion measured at the paws of BALB/c and C57BL/6 mice infected with L. amazonensis* 29
Figura 4 – *Characterization of local macrophages in infected mice paws* 31
Figura 5 – *Analysis of collagen in mice paws infected with L. amazonensis.* 32
Figura 6 – *Adoptive transfer of macrophages (ATM).*..... 33
Figura 7 – *Immunomodulation of ATM mice.*..... 34
Figura Suplementar 1 – *Vacuolated macrophages in paws of BALB/c mice infected with L. amazonensis for 11 weeks.*..... 42
Figura Suplementar 2 – *Flow cytometry gating strategy for macrophages.*..... 42
Figura Suplementar 3 – *Analysis of intravenous ATM.*..... 43

LISTA DE TABELAS

Tabela 1 – <i>Chemokines differentially produced by M1 and M2 macrophages and their role in cell recruitment.</i>	6
Tabela 2 – <i>Salivary compounds and their effects on Leishmania infection</i>	7
Tabela 3 – <i>M1/M2 macrophages in leishmaniasis</i>	8

SUMÁRIO

1. INTRODUÇÃO	1
1.1. CAPÍTULO 1	
<i>Macrophage Polarization in Leishmaniasis: Broadening Horizons</i>	1
1.1.1. <i>Introduction</i>	3
1.1.2. <i>M1 and M2 macrophages: an overview</i>	4
1.1.3. <i>Interaction of vector saliva with immune cells</i>	6
1.1.4. <i>M1 and M2 macrophages in <u>Leishmania</u> infection</i>	7
1.1.5. <i>Conclusion</i>	10
1.1.6. <i>References</i>	11
2. OBJETIVOS	15
2.1. OBJETIVOS GERAIS	15
2.2. OBJETIVOS ESPECÍFICOS	15
3. RESULTADOS	16
3.1. CAPÍTULO 2	
<i>Murine susceptibility to <u>Leishmania amazonensis</u> is dependent on arginase-1 of macrophages</i>	16
3.1.1. <i>Introduction</i>	20
3.1.2. <i>Materials and methods</i>	22
3.1.3. <i>Results</i>	26
3.1.4. <i>Discussion</i>	35
3.1.5. <i>Conclusion</i>	37
3.1.6. <i>References</i>	38
4. CONCLUSÕES	44
5. REFERÊNCIAS	45
6. ANEXOS	56
6.1. COMPROVANTE DE ACEITE	56
6.2. OUTRAS PRODUÇÕES REALIZADAS DURANTE O DOUTORADO	57
6.2.1. Artigo 1	57
6.2.2. Artigo 2.....	100
6.2.3. Capítulo de livro	122
6.3. PARECER DA COMISSÃO DE ÉTICA DE NO USO DE ANIMAIS	165

1. INTRODUÇÃO

1.1. Capítulo 1

Polarização de Macrófagos na Leishmaniose: Ampliando os Horizontes
Macrophage Polarization in Leishmaniasis: Broadening Horizons

Fernanda Tomiotto-Pellissier, Bruna Taciane da Silva Bortoleti, João Paulo Assolini, Manoela Daiele Gonçalves, Amanda Cristina Machado Carloto, Milena Menegazzo Miranda-Sapla, Ivete Conchon-Costa, Juliano Bordignon, Wander Rogério Pavanelli

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As Leishmanioses são um grupo de doenças causadas por parasitos de mais de 20 espécies do gênero *Leishmania*, transmitidas aos seres humanos pela picada de flebotomíneos infectados. Mais de 700.000 novos casos são registrados anualmente, com 20.000 a 30.000 mortes anuais. As doenças se manifestam como diferentes formas clínicas, refletindo principalmente a diversidade de espécies do parasito e a resposta imune do hospedeiro.

Macrófagos são as principais células hospedeiras do parasito e a chave no desencadeamento de uma resposta protetora ou prejudicial do hospedeiro frente à infecção. Nas últimas décadas foi descrita a plasticidade de macrófagos em relação ao tipo de resposta desenvolvida, sendo subdivididos em dois grupos, denominados M1 e M2. Macrófagos M1, ou ativados classicamente, são aqueles detentores de características pró-inflamatórias, com grande potencial microbicida, enquanto macrófagos M2, ou alternativamente ativados, são células com características anti-inflamatória/regulatória.

Apesar dos avanços no conhecimento do papel de macrófagos M1 e M2 em diferentes doenças infecciosas, ainda pouco é descrito sobre suas funções no

desenvolvimento da Leishmaniose. Neste sentido, foi realizada uma extensa busca na literatura, visando a compilação dos dados envolvendo leishmanioses e polarização macrofágica, subdividida nas seguintes seções:

- 1) Introdução – consistindo em uma apresentação das Leishmanioses, suas manifestações clínicas, epidemiologia, ciclo biológico do parasito e resposta imune desenvolvida pelo hospedeiro.
- 2) Uma visão geral sobre macrófagos M1 e M2 – tratando das funções básicas dos macrófagos e interação com células da imunidade inata e adaptativa, seguida por discussão da plasticidade de macrófagos e polarização em M1 e M2. Além disso, esta seção trata das funções de macrófagos M1 e M2, bem como as citocinas, quimiocinas, fatores de transcrição, receptores, genes transcritos e interação celular de cada subtipo.
- 3) Interação da saliva do vetor com as células imunes – descrevendo o papel da saliva do inseto vetor no estabelecimento e curso da infecção, modulando a resposta imune e o padrão de moléculas dos macrófagos.
- 4) Macrófagos M1 e M2 na Leishmaniose – onde são apresentados os estudos *in vitro* e *in vivo* que correlacionam diretamente a o papel de macrófagos polarizados (M1/M2) na infecção por *Leishmania*, ou indiretamente, tratando da importância de moléculas envolvidas na polarização frente à infecção. Além disso, são apresentados trabalhos que sugerem estratégias terapêuticas para modular moléculas-chave no controle da atividade celular, induzindo determinado padrão de resposta. Nesta seção foram considerados estudos envolvendo as espécies de *Leishmania* que causam as formas viscerais e cutânea da doença.

A referida revisão bibliográfica, publicada no periódico *Frontiers in Immunology* trata de maneira ampla o estado da arte da temática da presente tese de doutorado “Modulação Macrofágica na Leishmaniose”. Desta forma, a mesma, diante da aprovação do colegiado da Pós-Graduação em Biociências e Biotecnologia e da banca de Qualificação, constitui o capítulo primeiro da presente tese de doutorado, substituindo o capítulo tradicionalmente intitulado “Introdução”.



Macrophage Polarization in Leishmaniasis: Broadening Horizons

Fernanda Tomiotto-Pellissier^{1,2}, Bruna Taciane da Silva Bortoleti^{1,2}, João Paulo Assolini², Manoela Daiele Gonçalves³, Amanda Cristina Machado Carloto², Milena Menegazzo Miranda-Sapla², Ivete Conchon-Costa², Juliano Bordignon^{1,4*} and Wander Rogério Pavanelli^{1,2*}

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Leishmaniasis is a vector-borne neglected tropical disease that affects more than 700,000 people annually. *Leishmania* parasites cause the disease, and different species trigger a distinct immune response and clinical manifestations. Macrophages are the final host cells for the proliferation of *Leishmania* parasites, and these cells are the key to a controlled or exacerbated response that culminates in clinical manifestations. M1 and M2 are the two main macrophage phenotypes. M1 is a pro-inflammatory subtype with microbicidal properties, and M2, or alternatively activated, is an anti-inflammatory/regulatory subtype that is related to inflammation resolution and tissue repair. The present review elucidates the roles of M1 and M2 polarization in leishmaniasis and highlights the role of the salivary components of the vector and the action of the parasite in the macrophage plasticity.

Keywords: classical macrophage, non-classical macrophage, vector saliva, *Leishmania*, immunomodulation, chemokine

INTRODUCTION

Leishmaniasis is a broad term that is used for a group of vector-borne diseases caused by species of protozoan parasites of the *Leishmania* genus of which 18 spp. are pathogenic to humans (1). The disease presents in five main clinical forms: visceral leishmaniasis (VL, or kala-azar), cutaneous leishmaniasis (CL), mucocutaneous leishmaniasis (MCL), diffuse cutaneous leishmaniasis (DCL) and post-kala-azar dermal leishmaniasis (PKDL) (2). All types of leishmaniasis are transmitted to an animal or human reservoir through the bite of female infected phlebotomine sand flies, which infect a range of 70 animal species, including humans, rodents, and canids in their transmission cycle (3).

The World Health Organization (WHO) classifies leishmaniasis as a neglected tropical disease because it is directly linked to economically disadvantaged populations in tropical regions (2). A total of 700,000 to one million cases of leishmaniasis occur annually in 102 countries, areas or territories worldwide, with 20,000–30,000 deaths (3).

The high prevalence of this disease is directly influenced by the success of long host–parasite coevolutionary process in which parasites *Leishmania* have the ability to manipulate the vertebrate immune system in their favor, through the synthesis of parasites molecules, but also by vector saliva molecules, which are injected into the blood-feeding site during transmission (4).

The *Leishmania* parasites exhibit a biological digenetic life cycle with variable morphology that alternates between two main distinct developmental stages: the free-living flagellated promastigote form found in the midgut of phlebotomine sandfly vectors and the obligate intracellular aflagellated amastigotes in phagolysosomal vesicles of the vertebrate phagocytic cells, mainly into macrophages (5, 6).

During the blood feeding of the infected sandfly, which inoculates the host with metacyclic promastigotes and a large portion of the salivary content of the insect. Phlebotomine saliva is composed of pharmacologically active components with anti-hemostatic, chemotactic and immunomodulatory properties, that directly influence the parasite infection process modulating the local immune response (7). At the site of the bite occurs a rapid and intense neutrophil infiltration after inoculation, followed by monocytes/macrophages (8, 9).

Neutrophils primarily phagocytize most (80–90%) of the parasites and produce chemokines and cytokines that recruit and activate different cell types to regulate the development of the adaptive immune response during *Leishmania* sp. infection (8). Neutrophils are important components of the initial immune response against *Leishmania* parasites, even though there are currently contradictory findings on their role in the *Leishmania* infection.

Although the effective participation of neutrophils in the elimination of the parasite has been reported for *L. braziliensis*, *L. amazonensis* (10–15) and *Leishmania donovani* (16), collectively, most of these studies reported that the leishmanicidal action of neutrophils is clearly insufficient to control the establishment of infection and the development of the disease [reviewed in (17)].

Subversion of neutrophil killing functions by *Leishmania* is a strategy that allows parasite spreading in the host with a consequent infection evolution, transforming the primary protective role of neutrophils into a deleterious one. Neutrophils do not eliminate the parasite but act as “Trojan horses,” becoming late apoptotic and rapidly internalized by macrophages and dendritic cells, increasing the infectivity and persistence of the parasite (18, 19).

Macrophages play a dual role in *Leishmania* infection. These cells are responsible for the destruction of internalized parasites but also provide a safe place for *Leishmania* replication. Therefore, macrophages are key to disease progression and the success or failure of the infection depends on the interplay between infecting *Leishmania* species and the type and magnitude of the host's immune response. Both of these factors are closely related to the clinical forms of leishmaniasis (20, 21).

Abbreviations: Arg, arginase; CL, cutaneous leishmaniasis; IL, interleukin; IFN γ , interferon gamma; IGF1, insulin-like growth factor 1; iNOS, inducible nitric oxide synthase; LPS, lipopolysaccharide; Max, maxadilan; MCL, mucocutaneous leishmaniasis; mTOR, mammalian target of rapamycin; NO, nitric oxide; PKDL, post-kala-azar dermal leishmaniasis; PPAR, proliferator-activated receptors; PSG, promastigote secretory gel; ROS, reactive oxygen species; SGE, salivary gland extract; SGH, salivary gland homogenate; SGL, salivary gland lysate; TAC1, transmembrane activator and calcium modulator and cyclophilin ligand interactor; TGF β , transforming growth factor beta; Th, T helper; TLR, toll like receptor; TNF α , tumor necrosis factor alpha; Treg, T regulatory; and VL, visceral leishmaniasis.

Macrophages are normally at rest as naïve macrophages (M0), but the microenvironment in which these cells are found provides different signals that activate them and lead to the development of functionally distinct macrophage's phenotype, toward “classically activated” (M1) or “alternatively activated” (M2) with different disease outcomes (22, 23). Therefore, the activation of M1 macrophages by Th1 lymphocyte subpopulation, which produces various cytokines, primarily interferon gamma (IFN- γ) and tumor necrosis factor-alpha (TNF- α) is crucial for the elimination of this intracellular pathogen via the triggering of an oxidative burst. The host cells increase the production of reactive oxygen species (ROS), including superoxide, hydrogen peroxide, and hydroxyl radicals, and nitric oxide (NO), which exhibit high microbicidal capacity (20, 22).

In contrast, the activation of Th2 lymphocytes, which produce IL-4 and IL-13 cytokines, induces the M2 profile characterized by polyamine biosynthesis via activation of the enzyme arginase (arg) and production of urea and L-ornithine, which are beneficial for *Leishmania* intramacrophage growth favoring parasite survival in the infected macrophages and disease progression (22, 24).

Different *Leishmania* species trigger distinct immune responses (25). These responses are far beyond the classical Th1/Th2 paradigm (26) and increase the interest in understanding the role of M1 and M2 macrophages in the context of different *Leishmania* species infection. The immunomodulatory influence of the saliva of different leishmaniasis vectors should also be considered in the differential recruitment/activation of macrophages subtypes.

It is known how important macrophages are in the resistance or susceptibility to *Leishmania* infection, therefore we reviewed the impact of macrophage plasticity and M1 and M2 phenotypes on infection outcome. We also consider the role of vector saliva, which is a well-established immunomodulatory element in the *Leishmania* infection, in macrophage plasticity and phenotype.

M1 AND M2 MACROPHAGES: AN OVERVIEW

Macrophages are phagocytic cells that are found in several tissues. In innate immunity, macrophages are responsible primarily for the control of pathogens and in adaptive immunity, this cell participates in the recognition, processing, and presentation of antigens to T cells (27). Macrophages interact with T and B cells via intercellular contact and the production of molecules and mediators to participate in the inflammatory response and tissue repair (27).

Two main macrophages phenotypes are known, M1 and M2 (28). The M1, or the classically activated macrophage, is a pro-inflammatory subtype that exhibits microbicidal properties (29). The M2, or the alternatively activated macrophage, is an anti-inflammatory/regulatory subtype that plays a role in the resolution of inflammation and tissue repair (30). The polarization of macrophages into M1 and M2 phenotypes is

dependent on the signals provided by the microenvironment (28, 30, 31).

The designations M1 and M2 originated from Th1 and Th2 cytokine patterns and are associated with the change in macrophage phenotypes (32). Macrophage subtypes also differ in the production of cytokines, chemokines and other mediators and the expression of receptors, surface molecules, and transcription factors, which can act as specific markers to aid in the identification and function of these cells (27, 29, 32–34).

M1 macrophages are characterized by a high production of pro-inflammatory cytokines (TNF- α , IL-1 β , IL-6, IL-12, IL-18, IL-23, and Type 1 IFN), high phagocytosis rate, and the production of reactive oxygen and nitrogen species and act to control intracellular pathogens. This macrophage subtype plays a role in tumor control, and it may be involved in autoimmune diseases and tissue damage (7, 27, 34–39).

M1 macrophages constitute the first line of defense against intracellular pathogens and induce the development of the Th1 response via IL-12 secretion (40, 41). The polarization of M1 macrophages may be primarily due to the presence of lipopolysaccharide (LPS), IFN- γ or TNF- α . Granulocyte-macrophage colony-stimulating factor (GM-CSF) may also result in the differentiation and maintenance of the M1 phenotype (41, 42). Ruan et al. (43) demonstrated that complement system activation is also related to M1 polarization.

M1 polarization activates transcription factors, such as AP1, STATs, NF κ Bp65 and IRFs, which lead to the expression of pro-inflammatory genes, costimulatory molecules and chemokines to attract various immune cells (**Table 1**) (7, 27, 34–39). IRF4 and IRF5 are involved in the polarization of M1 and M2 macrophages, but the role of IRFs in M2 polarization is not completely clear (44).

The alternatively activated macrophages, or M2, exhibit an anti-inflammatory/immunoregulatory phenotype that is related to tissue remodeling and repair, resistance to some parasites and the promotion of tumor growth (30, 45, 46). M2 was initially characterized by the expression of mannose receptor (CD206), but a range of markers and mediators produced by these cells were described, including important chemokines for the recruitment of different cells (**Table 1**) (37, 45, 47).

Different stimuli, such as IL-4/IL-13, IL-10, TGF- β , M-CSF, vitamin D3, and immunocomplexes, induce M2 macrophages (48, 49). Other cytokines, such as IL-21 and IL-33, may also act on macrophage polarization and maintenance to an M2 phenotype (50–52). Li et al. (52) demonstrated that IL-21 reduced the expression of CD86, iNOS, TLR-4, and IL-6 and TNF- α production via STAT3 phosphorylation. IL-33 amplifies IL-13-induced M2 polarization (50, 51).

The classification of M2 macrophages was recently expanded and subdivides these cells into four subtypes, M2a, M2b, M2c, and M2d, according to stimulus and function (23, 45). M2a macrophages are polarized by macrophage colony-stimulating factor (M-CSF) and IL-4 or IL-13. This subtype is characterized by arg-1, IL-10, and SOCS3 expression and produces CCL24, CCL17, and CCL22, which are responsible for the recruitment of eosinophils, basophils and Th2 cells. These cells are involved

in allergic reactions, parasite death and encapsulation, the promotion of fibrogenesis, tissue repair and cell proliferation (53–55).

The M2b phenotype is induced by the combination of immunocomplexes with IL-1 β Ra/TLR ligands, apoptotic cells or LPS. These cells secrete a large amount of IL-10 and inflammatory mediators, such as TNF- α and IL-6, and express iNOS (30, 53). M2b macrophages secrete the CCL1 chemokine, which results in the infiltration of eosinophils, Th2 lymphocytes and regulatory T cells (Tregs) (56). Therefore, M2b macrophages act as an immunoregulator and trigger activation of the Th2 response (23, 53).

M2c macrophage polarization results from IL-10, TGF- β and glucocorticoids (38, 57, 58). This subtype produces IL-10, TGF- β , CXCL13, CCL16, and CCL18, which leads to the down-regulation of pro-inflammatory cytokines and an increase in the recruitment of eosinophils and naïve T cells. M2c cells express high levels of arg-1, CD163, CD206, scavenger receptors, TLR1, TLR8, FPR1, CCR2, and CCR5, and this subtype is involved in tissue regeneration and angiogenesis (53–55, 59).

The M2d macrophage phenotype is involved in the inhibition of the immune response and the promotion of angiogenesis (60). These cells are induced by IL-6, toll-like receptor (TLR) ligands and adenosine A_{2A} receptor agonists (60, 61). Stimulation with adenosine may result in the polarization of M1 macrophages to M2d (53, 61). This phenotype expresses high levels of IL-10, VEGF, and iNOS and secretes CCL5, CXCL10, and CXCL6 and low levels of IL-12 and TNF- α (53, 60–63).

Signals of the microenvironment are of great importance of the change in the polarization state of macrophages, and the programming from one phenotype to another is closely related to the activation of specific transcription factors and microRNAs (miRNAs) (64, 65). miRNAs are small molecules of non-coding RNAs which can act on gene expression at the post-transcriptional level (64), regulating some important transcription factors in the M1 and M2 phenotypes (66, 67).

Li et al. (67) reviewed the role of various miRNAs which participate in the regulation and polarization of macrophages in murine and human models. The miRNAs miRNA-9, miRNA-146a, miRNA-146b, miRNA-124, miRNA-181a, miRNA let-7c, miRNA-93, and miRNA-210 act to suppress the M1 and promote M2 phenotype. On the other hand, miRNA-27a, miRNA-130a, miRNA-130b, miRNA-155, miRNA-21, miRNA-19a-3p, miRNA-23a, miRNA-125a, miRNA-125b, miRNA-26a, miRNA-26b, and miRNA-720 act on transcription factors involved in the promotion of M1 phenotype and M2 suppression [reviewed in (67)].

Although we discussed above on the macrophage polarization of M0 to M1 or M2, it is known that macrophages have high plasticity and can be repolarized or reprogrammed under specific stimuli, in other words, M1 macrophages can differentiate into M2, and vice versa (40, 68). M2 macrophage may be repolarized more quickly than M1 macrophages, and this repolarization can occur through exposure of TLR ligands such as LPS and/or IFN- γ , or by expression of miRNAs such as miR-155 (69, 70). Van den Bossche et al. (71) showed that exposure of M1 macrophages

TABLE 1 | Chemokines differentially produced by M1 and M2 macrophages and their role in cell recruitment.

M1 macrophages		M2 macrophages	
Chemokines	Cell recruitment	Chemokines	Cell recruitment
CXCL1	Neutrophils	CXCL13	B cells
CXCL2	Granulocytes, polymorphonuclear	CCL1	Monocytes, Th2 and Treg cells
CXCL3	Neutrophils	CCL16	Monocytes and lymphocytes
CXCL5	Neutrophils	CCL17	Th2 cells
CXCL8	Neutrophils	CCL18	Th2 cells
CXCL9	Activated T cells	CCL22	Th2 and Treg cells
CCL2	Monocytes, memory T cells and NK	CCL24	Eosinophils and basophils
CCL3	Monocytes, T lymphocytes and polymorphonuclear cells		
CCL4	Monocytes, T lymphocytes and polymorphonuclear cells		
CCL11	Eosinophils		
CX3CL1	T cells and monocytes		

CCL, CC-chemokine ligand; CXCL, CXC-chemokine ligand; NK, natural killer cells.

to IL-4 is not able to reprogram the macrophages to M2 (71). However, the miRNAs can suppress the M1 phenotype and promote the polarization to M2 as commented above (67).

INTERACTION OF VECTOR SALIVA WITH IMMUNE CELLS

Among over 800 species of phlebotomines recorded, 98 are proven or suspected vectors of human leishmaniasis; these include 42 *Phlebotomus* species in the Old World and 56 *Lutzomyia* species in the New World (all: Diptera: Psychodidae) (72). It is known that through the insect bite, vector saliva plays an important role in the establishment of *Leishmania* infection by increasing the infectivity of the parasite and modulating the host immune response (73, 74). Arthropod saliva contains anti-inflammatory, chemotactic and anti-hemostatic components that influence the course of parasite transmission to the host (7, 75, 76).

Most studies of the role of vector saliva in disease course were performed prior to the establishment of the M1 and M2 macrophage concept. Therefore, these works do not use this nomenclature. However, they investigated molecules that are involved in the plastic response of macrophages. These molecules are discussed below.

Some groups produced extracts, homogenates, sonicates and salivary lysates to elucidate the function of vector saliva in *Leishmania* transmission and infection (Table 2). We review the literature and assemble the results of different groups to provide an overview of the function of saliva.

Saliva contains molecules that induce a long-lasting erythema, which facilitates the obtaining of blood from capillaries in the host tissue (79). Vector saliva may facilitate cell recruitment via the promotion of vasodilation (79, 80, 92). The recruited cells include neutrophils, eosinophils and macrophages. Neutrophils may be attracted by the presence of several mediators, such as LTB₄, and the role of these cells as a “Trojan horse” favors the establishment of infection (80, 92). The role of eosinophils is

controversial, and whether these cells promote or suppress the infection is not well known (80, 95).

Some studies observed that vector saliva increased macrophages recruitment to the site of infection because of the modulation of chemotactic factors, such as CCL2/MCP-1, CCR2 and PGE₂ (80, 92). Although some studies have shown the participation of lipid mediators such as PGE₂ and LTB₄ in cell migration (73, 92), the role of these lipid mediators acting in the polarization or recruitment of a specific profile of macrophages is uncertain, differing between cell types and models studied (28, 96–98).

Vector saliva plays an important immunomodulatory role and favors the M2 profile in different manners. Vector saliva induces IL-10 to promote a regulatory response (81, 89, 90), and it is related to the activation of a Th2 response via the increase in IL-4 and IL-6 synthesis (85, 87) Rohousov et al. (88). Besides that, the salivary components reducing the M1 related parameters, such as the pro-inflammatory cytokines IFN- γ and IL-12, iNOS and nitric oxide (NO) (85, 86, 88, 90).

However, some studies showed the action of saliva inducing M1 parameters. C57BL/6 mice immunized with plasmids encoding salivary proteins developed a Th1 response, resulting in protection against *L. major* infection, demonstrating that saliva may provide a protective effect and conferring characteristics for the development of a vaccine against *Leishmania* (90, 99). In a clinical study, was observed that individuals from an endemic area exposed to *P. duboscqi* bite presented high serum levels of INF- γ and decrease of IL-13, IL-5, directing a Th1 profile. Nonetheless, when PBMC of those individuals were exposed to *P. duboscqi* saliva, the most presented a mixed Th1/Th2 response, without a specific polarized profile (82).

In addition to complexes containing salivary components, the biological activity of pharmacological compounds isolated from vector saliva was examined. Adenosine and adenosine monophosphatase are active compounds found in *P. papatasi* saliva, and these factors inhibit the function of dendritic cells via a PGE₂/IL-10-dependent mechanism to promote a tolerogenic profile that is characterized by the induction of regulatory T cells, which is also related to M2 polarization (73).

TABLE 2 | Salivary compounds and their effects on *Leishmania* infection.

Compound	Immunomodulatory effect	References	
Promastigote secretory gel (PSG)	↑ Arg	(77)	
	↑ IL-1β	(78)	
	↑ IL-6		
	↑ IL-10		
	↑ TNF-α		
	↑ CCL2		
	↑ CCL4		
	↑ CCL3		
	↑ CXCL2		
	↑ FGFR2		
	↑ EGF		
	↑ EGFR		
	↑ IGF1		
Salivary Gland Homogenate (SGH)	↑ MCP-1 ↑ CCR2 ↑ IL-10 ↑ Eosinophils ↑ Macrophages ↑ IFN-γ ↑ IL-13 ↑ IL-5	↓ iNOS ↓ NO ↓ IFN-γ ↓ IL-13 ↓ IL-5	(79) (80, 81) (82)
Salivary Gland Lysate (SGL)	↑ IL-4 ↑ IL-6	↓ IFN-γ ↓ IL-12 ↓ iNOS	(83, 84) (85–88)
Salivary Gland Extracts (SGE)	↑ IL-10 ↑ IL-4 ↑ CD8 ↑ INF-γ ↑ CD4	↓ NO	(89) (90)
Salivary Gland Sonicate (SGS)	↑ IL-4 ↑ PGE ₂ ↑ Macrophages ↑ LTB ₄	↓ IFN-γ	(91, 92)
Maxadilan (max)	↑ IL-6 ↑ IL-10 ↑ TGF-β ↑ CD86	↓ IL-1β ↓ IL-12p70 ↓ TNF-α ↓ IFN-γ ↓ CD80 ↓ CCR7	(87, 93, 94)
Adenosine	↑ IL-10 ↑ PGE ₂		(73)

CCR, chemokine receptor; CD, cluster of differentiation; IFN, interferon; IL, interleukin; iNOS, inducible nitric oxide synthase; MCP-1, monocyte chemoattractant protein-1; NO, nitric oxide; PG, prostaglandin; TNF, tumor necrosis factor; TGF, transforming growth factor; CCL, chemokines; CXCL, motif chemokine ligand; FGFR, fibroblast growth factor receptor; EGF, epidermal growth factor; EGFR, epidermal growth factor; IGF, Insulin-like growth factor.

Maxadilan (Max) is a vasodilator peptide isolated from the saliva of arthropod vectors, and it reduces CD80 expression, which is responsible for T cell activation, and CCR7, which is involved in the development of adaptive immunity. Max increases CD86 expression on a subpopulation of dendritic cells, which leads to a preferential Th2 type response. Max also promotes an increased production of IL-6, IL-10, and TGF-β, and reduction of the Th1 cytokines, IL-1β, IL-12p70, TNF-α, and IFN-γ (87, 93). Max treatment of *L. major*-infected peritoneal exudate cells increased parasite load because of Th2 polarization

and decreased NO production (94). These results suggest that Max acts on M2 polarization, as demonstrated previously for total saliva.

Parasites also play an important role in vector saliva modulation because these pathogens secrete promastigote secretory gel (PSG) in the insect gut. The vector regurgitates PSG with the other salivary components at the moment of blood-feeding (77). *Leishmania mexicana*-PSG regurgitated by *L. longipalpis* exacerbates skin infection via an increase in the recruitment of macrophages to the site of infection (77). PSG also increases parasite load *in vitro* and *in vivo* via increasing arginase activity (77).

In more detail, Giraud et al. (78) demonstrated that PSG exacerbates the inflammatory phase of the early wound response (high levels of cytokines IL-1β, IL-6, IL-10, TNFα and chemokines CCL2, CCL3, CCL4, CXCL2), to induce insulin-like growth factor 1 (IGF1)-signaling and later IGF1-dependent expression of arg-1 in macrophages. As a result, the M2 macrophages promote the effective infection of the parasites in a PGS-dependent manner (78).

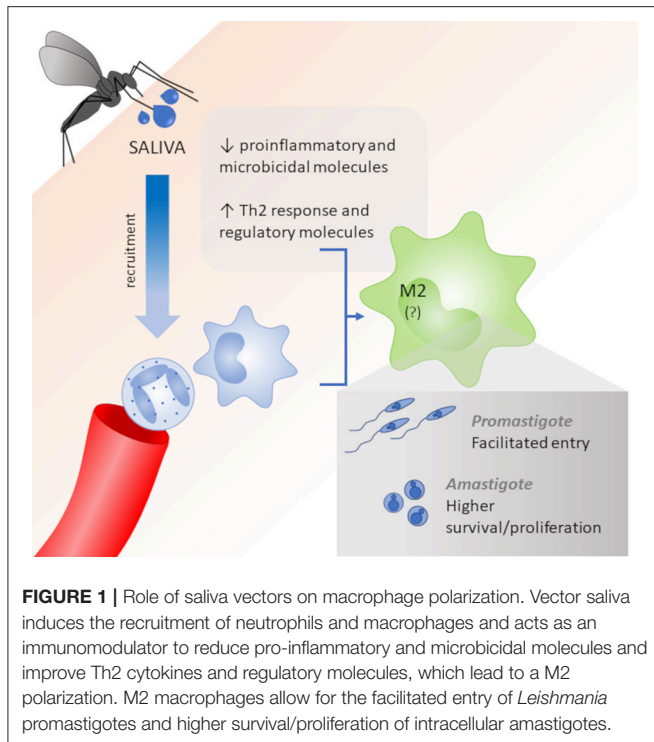
In this way, it is noteworthy that some works show that high levels of pro-inflammatory cytokines are induced by saliva components, leading to a Th1 profile, that can act in a host protective way. However, the most of studies inferred that the different salivary components are allied to the immunomodulatory capacity of the parasite, and these components are essential tools in the success of the infection, via the down-regulation of a pro-inflammatory response and reduction of macrophages M1, which are fundamental to parasite elimination and disease resolution. The saliva also up-regulates Th2-standard cytokines and regulatory molecules, which act in M2 polarization, facilitate the promastigote infection and increase the survival and proliferation of intracellular amastigotes (Figure 1).

M1 AND M2 MACROPHAGES IN LEISHMANIA INFECTION

The role of macrophage subsets in *Leishmania* infection was not investigated thoroughly. However, the fundamental role of these cells in the development of the lesion support an improved understanding of the M1 and M2 profiles as an important tool in the pathogenesis of leishmaniasis. *In vitro* and *in vivo* studies of the host response to *Leishmania* (Table 3) and therapeutic strategies for modulating key molecules that control cellular activity were performed. We discuss the studies of species that cause the visceral and cutaneous forms of the disease.

M1 and M2 Macrophages in Cutaneous Leishmaniasis (CL)

CL is the most common form of leishmaniasis, with an estimated 600,000 to 1 million new cases worldwide annually. CL causes skin lesions that leave life-long scars and serious disability, and it has become a serious public health problem (3). The primary etiological agents include the *Leishmania tropica* and *L. major*



species, in the Old World, and the *L. mexicana* species complex (e.g., *L. amazonensis*), and the subgenus *Viannia*, as the *L. (V.) braziliensis* species complex, in the New World (115).

Patients with CL have higher plasma levels of arg-1, TGF- β and PGE₂ (116), as well as, increased arginine in lesions (117), suggesting that M2 macrophages might play a role in the pathogenesis of the disease. In this sense, an *in vitro* study demonstrated that only M2 macrophages allow for *L. major* and *L. amazonensis* growth (114). These authors demonstrated that lipophosphoglycan (LPG) and glycoprotein GP63 of the parasites acted on M2 macrophages and suppressed non-coding RNAs, which left these cells permissive to infection (114). Lee et al. (118) also demonstrated that a non-healing strain of *L. major* efficiently interacted with M2 macrophages (CD206^{hi}), which phagocytized the parasite *in vitro* and *in vivo*. However, a strain that produced self-healing lesions was less phagocytosed by M2 macrophages. The authors stated that the preferential infection of M2 cells played a crucial role in the severity of the cutaneous disease.

The role of peroxisome proliferator-activated receptors (PPARs) was investigated in infected macrophages from mouse strains resistant and susceptible to *Leishmania* infection. PPARs are ligand-activated transcription factors that are expressed in macrophages, and regulate the expression of certain genes related to the inflammatory response (119). Odegaard et al. (120) demonstrated that PPAR γ -knockout mice have delayed in disease progress, with less footpad swelling and reduced parasitic burden. Importantly, genes preferentially expressed in alternatively activated macrophages, such as *arg-1*, *Mrc1*, and *Clec7a*, were also decreased in the tissue of PPAR γ -knockout mice. These data strongly suggest that PPAR γ is required for

TABLE 3 | M1/M2 macrophages in leishmaniasis.

	Model	Disease	<i>Leishmania</i> specie	References
Mouse	<i>in vivo</i>	–	<i>L. major</i>	(100)
Mouse	<i>in vitro</i>	–	<i>L. amazonensis</i>	(101)
Human	NI/ <i>in vitro</i>	VL	–	(102)
Mouse	<i>in vitro</i>	VL	–	(103)
Raw	<i>in vitro</i>	–	<i>L. amazonensis</i>	(104)
Dog	NI	VL	–	(105)
Dog	NI	VL	<i>L. infantum</i>	(106)
Mathematical	–	–	–	(107)
Mouse	<i>in vitro</i>	–	<i>L. major</i>	(108)
Human	NI	PKDL	–	(109)
Mouse	<i>in vitro/in vivo</i>	CL	<i>L. major</i>	(110)
Mouse	<i>in vitro</i>	–	<i>L. donovani</i>	(111)
Mouse	<i>in vitro</i>	–	<i>L. major</i>	(112)
Mouse	<i>in vitro</i>	–	<i>L. mexicana</i>	(113)
Mouse	<i>in vitro</i>	–	<i>L. major/L. amazonensis</i>	(114)

CL, cutaneous leishmaniasis; NI, natural infection; PKDL, post-kala-azar dermal leishmaniasis; VL, visceral leishmaniasis.

acquisition and maintenance of the M2 macrophages in *L. major* model (120). Besides that, Gallardo-Soler et al. (121) showed that PPAR γ and δ ligands promote intracellular *L. major* amastigote growth in infected macrophages, and this effect is dependent on both PPAR expression and arg-1 activity, namely suggesting that PPAR ligands promote amastigote growth in M2 macrophages in an arginase-dependent manner (121).

On the other hand, PPAR expression induced the activation of the murine macrophage cell line J774A.1 via polarization toward an M1 profile, with high production of pro-inflammatory mediators (TNF- α , IL-1 β , IL-6, TLR4, and ROS), and an increase in microbicide activity against *L. mexicana* (113).

The transmembrane activator and calcium modulator and cyclophilin ligand interactor (TACI) is a key molecule for plasma cell maintenance. This receptor is in the TNF family, and it is required in infections where protection depends on antibody response. Analysis of macrophage phenotype revealed that macrophages adapted the M2 phenotype in the absence of TACI. The levels of M2 markers, IL-4R α and CD206, were significantly higher in TACI-knockout macrophages than wild-type cells. TACI-knockout mice were unable to control *L. major* infection *in vitro*, which confirms their M2 phenotype, and intradermal inoculation of *Leishmania* resulted in a more severe manifestation of the disease than in the resistant C57BL/6 strain. The transfer of WT macrophages to TACI-knockout mice significantly reduced the disease severity (110).

One fundamental role of TNF was the induction of M1 differentiation and blockade of M2 polarization in the livers of *L. major*-infected mice (100). These authors also described the role of IL-6, which did not interfere with the macrophage phenotype alone, but it was highly expressed in M2 macrophages. The authors suggested that a balance between TNF and IL-6 mediated macrophage polarization in *L. major* infection (100).

Vellozo et al. (122) also demonstrated that the resistance of C57BL/6 mice to *Leishmania* infection was because of their ability to mature macrophages in the peritoneum from M0 to M1. Susceptible mice (BALB/c) exhibit immature peritoneal macrophages and succumb to infection. However, both mouse strains are resistant to *L. braziliensis* infection because in both strains convert M0 to M1 macrophages in this model, despite the incomplete M1 maturation and lower iNOS expression and NO production in BALB/c mice.

The co-culture of mesenchymal stem cells and *L. major*-infected macrophages also induced an event suggestive of M1 polarization, with the induction of inflammatory cytokines and reduction of IL-10 levels (108). This strategy provides new hope for stem cell therapy in the control of *L. major* (108).

SLPI (secretory leukocyte protease inhibitor) is a potent serine protease inhibitor that exhibits antimicrobial and anti-inflammatory functions. The role of SLPI in *L. major* infected-macrophage polarization was investigated (112). SPLI-knockout macrophages produced high levels of iNOS and IFN- γ but failed to contain cutaneous *L. major* infection. This study highlights that a very strong M1 response is detrimental in *L. major* models and causes tissue injury because of exacerbated inflammation. This study suggests that a balance of M1 and M2 macrophage responses influences the outcome of innate host defense against intracellular parasites, and SLPI is critical for the coordination of this balance and the resistance to chronic leishmaniasis (112).

One study of PKDL demonstrated that monocytes from patients exhibited decreased expression of TLR-2/4 and an attenuated generation of reactive oxidative/nitrosative species (109). Patients also exhibited an increased expression of classical M2 markers (arg-1, PPAR γ and CD206) in monocytes and lesional macrophages, which indicated the M2 polarization of macrophages. These subsets appeared to sustain disease chronicity, which is a hallmark of Indian PKDL (109).

Two studies showed the development of new drugs addressed the M1/M2 plasticity (101, 104). The first study demonstrated that high antimony dilutions, a homeopathic medicine, potentiated the *L. amazonensis*-induced reduction of pro-inflammatory cytokine (IL-6, IL-12, and IFN- γ) and chemokine (CCL-2 and CCL-4) production in RAW cells. The treatment also induced an increase of the parasites internalization, but there was a reduction of the acid vacuoles, which implied in less elimination of the amastigote forms. The authors suggest that this phenomenon is related to the M2 polarization and regulation of the chronic inflammation events (104).

The second study used crotoxin, which is the main component of *Crotalus durissus terrificus* venom, to treat peritoneal BALB/c mouse macrophages infected with *L. amazonensis* (101). The host cells exhibited an increase in nitric oxide, IL-6 and TNF- α production, that converged into an M1 activation profile, as suggested by their morphological changes (larger spreading), inducing leishmanicidal activity against intracellular parasites, which may be associated with a better prognosis for CL (101).

Siewe et al. (107) proposed a mathematical model that states that *Leishmania* parasite takes advantage of the immune system and invades M1 and M2 macrophages. Simulations of the model demonstrated that the number of M2 macrophages constantly

increased and M1 macrophages constantly decreased over the course of the infection, but the sum of the number of M1 and M2 cells reached a steady state, which was approximately the same as the healthy state of the host. The ratio of *Leishmania* parasites to macrophages depends homogeneously on their ratio at the time of the initial infection (107).

M1 and M2 Macrophages in Visceral Leishmaniasis (VL)

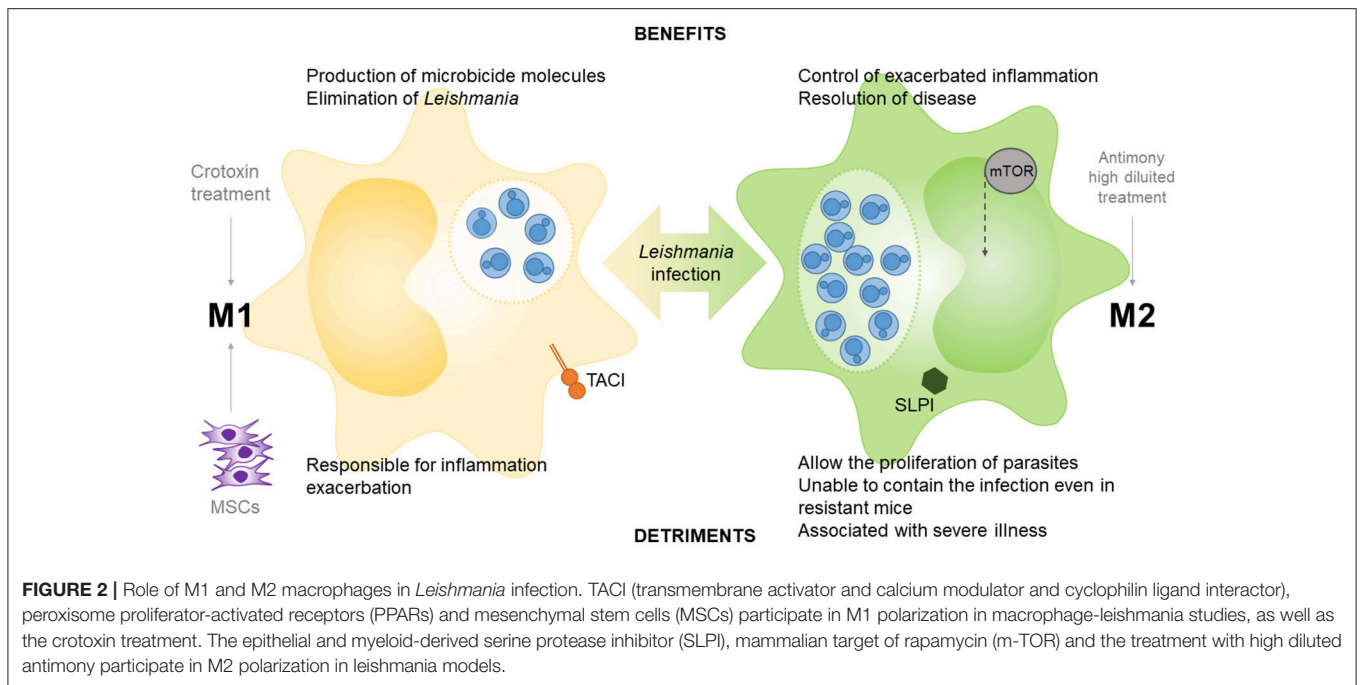
Visceral leishmaniasis is the most aggressive form of the disease, with high rates of mortality (3). Visceral leishmaniasis is generally caused by *L. donovani* and *L. infantum chagasi* species and affects internal organs, such as spleen, liver, and bone marrow (115, 123). It has been shown that blood arginase levels in VL patients are considerably increased (124), while NO levels are decreased (125). In this sense, a recent study demonstrated that monocytes/macrophages of VL patients present reduced oxidative burst and antigen presentation, with a M2 regulatory phenotype, characterized by high CD163, IL-10 and CXCL14 levels (126).

In the same direction, Silva et al. (102) demonstrated that the CD163s molecule was associated with the M2 macrophage phenotype, and it may be used as a biomarker of the clinical parameters of human VL severity. *L. donovani* also promotes the output of monocytes with a regulatory phenotype from the bone marrow that function as safe targets for the parasite (127). These data are consistent with analyses of dogs with VL, that demonstrated a higher number of M2-polarized macrophages compared to healthy dogs (106). The authors concluded that the predominance of the M2 phenotype (high CD163 labeling) in VL dogs favored the multiplication of *L. infantum* in the skin, spleen and lymph nodes (106).

Similarly, Chan et al. (128) suggest that the regulation of the macrophage phenotype is induced by the parasite, since PPAR expression is induced by parasitic infection. The authors showed that *L. donovani* activation of PPAR γ promotes survival, whereas blockade of PPAR γ facilitates removal of the parasite. Thus, *Leishmania* parasites harness PPAR γ to sustain the M2 phenotype and increase infectivity of visceral disease (128).

A recent study has also shown the role of mammalian target of rapamycin (mTOR) in M2 macrophage polarization for *Leishmania* survival (129). The authors observed that *L. donovani*-infection activated host mTOR pathway, resulting in reduced expression of M1 macrophage markers (ROS, NO, iNOS, NOX-1, IL-12, IL-1 β , and TNF- α), and increased expression of M2 macrophage markers (arg-1, IL-10, TGF- β , CD206, and CD163), favoring the *Leishmania* survival inside macrophages. Thus, they concluded that mTOR plays a crucial role in M2 macrophage polarization and parasite persistence in *L. donovani* infection (129).

These results correlated with studies performed in *L. donovani*-infected Syrian hamsters, in which the spleen environment was inflammatory with a high production of types I and II IFN. However, IFN- γ did not direct M1 macrophage polarization in this



model, which allowed for parasite growth and the expression of counter-regulatory molecules, such as arg-1 (103).

A different group demonstrated that macrophages cultured from the lymph nodes of VL dogs exhibited low arg-1 activity and high NO and PGE₂ production compared to uninfected dogs (105). These authors suggested that M1 macrophages were participating in the immune response in the lymph nodes of these animals (105). Attenuated *L. donovani* induced innate immunity via the classical activation of macrophages (M1), with upregulation of IL-1 β , TNF- α , IL-12 and downregulation of IL-10, YM1, Arg-1, and MRC-1 genes, which led to the generation of protective Th1 responses in BALB/c mice (111).

This controversy may be explained by the fact that different patterns of gene expression are found in macrophages at different time points (130). A recent study demonstrated that the early response against *L. donovani* infection was distinguished by the increase in of Th1 markers and M1-macrophage activation molecules (IFN- γ , Stat1, Cxcl9, Cxcl10, Ccr5, Cxcr3, Xcl1, and Ccl3). However, this activation was not protective because the parasitic burden increased over time. There was no marked overlap of macrophage phenotypes at intermediate times of infection, and the overexpression of these Th1/M1 markers was restored later in the chronic phase without parasitic burden control (130).

Together, these results demonstrate that there are no “good guys” and “bad guys” in the polarization of macrophages following infection by *Leishmania* parasites. Therefore, a balance between the potent microbicidal response of M1 macrophages and the potential regulation by M2 macrophages may be key to the success of overcoming leishmaniasis (Figure 2).

CONCLUSION

The collected works suggest that vector saliva plays an immunomodulatory effect on macrophages, which leads to the polarization of macrophages to an M2 phenotype. Therefore, vector saliva may contribute to an increase in the infectivity and persistence of the parasite during the initial periods of infection. Different *Leishmania* species exhibit virulence factors that can subvert the microbicidal mechanisms of the host to favor its proliferation because of the M2 polarization. However, M1 macrophages act during later periods of the infection and trigger an exacerbated immune response that leads to a worsening of the lesions, despite the role of these cells as powerful microbicidal agents. Therefore, a balance between the initial microbicidal response (M1) followed by a restorative response (M2) at later periods of time would provide the utmost benefit for the host. Further studies are necessary to fully elucidate the balance between these two main macrophages populations in the control of *Leishmania* infection.

AUTHOR CONTRIBUTIONS

FT-P study design, literature review, data collection and manuscript writing. BB, JA, MG, AC, and MM-S literature review, data collection and manuscript writing. IC-C and JB text correction and organization. WP study design, text correction and organization.

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Conflict of Interest Statement: The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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2. OBJETIVOS

2.1. OBJETIVOS GERAIS

Sabendo das lacunas existentes no entendimento da imunidade protetora desenvolvida frente à infecção por *Leishmania (Leishmania) amazonensis*, o objetivo deste trabalho foi demonstrar a resposta imune diferencial estabelecida por camundongos parcialmente resistentes (C57BL/6) e susceptíveis (BALB/c) à infecção por esta espécie de protozoário.

2.2. OBJETIVOS ESPECÍFICOS

- Caracterizar a evolução da infecção nas diferentes linhagens de camundongos;
- Quantificar a produção de citocinas dos padrões Th1, Th2 e Th17;
- Analisar a produção de ânion superóxido no local da infecção;
- Quantificar o número macrófagos no local da infecção e identificar marcadores relacionados aos padrões de macrófagos M1 e M2;
- Determinar o reparo tecidual no local da infecção;
- Avaliar a transferência adotiva de macrófagos como potencial interferente no curso da infecção.

3. RESULTADOS

3.1. Capítulo 2

Susceptibilidade murina à infecção por *Leishmania amazonensis* é dependente da arginase-1 de macrófagos

Murine susceptibility to Leishmania amazonensis infection is dependent on arginase-1 of macrophages

Fernanda Tomiotto Pellissier, Milena Menegazzo Miranda-Sapla, Taylon Felipe Silva, Bruna Taciane da Silva Bortoleti, Manoela Daisele Gonçalves, Virginia Márcia Concato, Ana Carolina Jacob Rodrigues, Idessania Nazareth Costa, Carolina Panis, Ivete Conchon-Costa, Juliano Bordignon, Wander Rogério Pavanelli

Artigo com resumo aceito para a edição especial “*Cutaneous Leishmaniasis: Exploring Pathogenesis and Immunomodulatory Approaches*” do periódico *Frontiers in Cellular and Infection Microbiology* - fator de impacto 4,12 (JCR 2020)/ Qualis A1 em fevereiro de 2021 (Anexo 6.1).

A partir dos resultados compilados no capítulo 1, percebe-se que apesar dos avanços no conhecimento do papel de macrófagos M1 e M2 em diferentes doenças infecciosas, ainda pouco é descrito sobre suas funções no desenvolvimento da Leishmaniose, em especial na doença causada por *Leishmania amazonensis*, espécie presente no Brasil, principalmente nas regiões norte e nordeste.

Em infecções por parasitos desta espécie, camundongos BALB/c desenvolvem lesões e uma resposta imune predominantemente do tipo Th2, enquanto camundongos da linhagem C57BL/6 apresentam edema local sem ulcerações, e manifestam resposta imune mista Th1 e Th2, sendo considerados parcialmente resistentes à infecção. Tal resposta mista é similar à observada na leishmaniose cutânea humana, validando a relevância biológica deste modelo experimental.

Sendo assim, avaliou-se a resposta diferencial estabelecida por camundongos BALB/c e C57BL/6 frente à infecção por *L. amazonensis*. O presente trabalho é constituído de duas etapas. Na primeira, foi verificado que camundongos C57BL/6 apresentaram marcadores de ambos os perfis imunes M1 e M2, e identificada a enzima arginase-1 (Arg-1) como um possível marcador de agravamento das lesões em período crônico de infecção em camundongos BALB/c.

Na segunda parte, foi observado que a transferência adotiva de macrófagos peritoneais de C57BL/6 para BALB/c infectados promove redução do edema e do número de parasitos no local da lesão, além de reduzir os níveis de Arg-1. Assim, a pior evolução das lesões parece estar envolvida com o recrutamento de macrófagos ricos em Arg-1. Esses resultados ampliam a compreensão sobre a imunidade protetora contra *L. amazonensis*, fornecendo novos conhecimentos que podem contribuir para o desenvolvimento de intervenções de tratamento ou prevenção da leishmaniose.

Os resultados brevemente descritos acima são detalhados no capítulo 2 da presente tese de doutorado, apresentado a seguir.

Murine susceptibility to *Leishmania amazonensis* infection is dependent on arginase-1 of macrophages

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Key-words: Leishmaniasis; M1; M2; IFN- γ ; wound healing; collagen; iNOS.

Abstract

Cutaneous leishmaniasis is a zoonotic infectious disease with broad world distribution that causes a range of diseases with clinical outcomes ranging from self-healing infections to chronic disfiguring disease. The understanding of the effective immune response to this infection still has many gaps, which comprehension is fundamental to the development of drugs and vaccines. Thus, we investigated the local profile of mediators involved in the development of cutaneous leishmaniasis in experimental models of susceptibility (BALB/c) and partial resistance (C57BL/6) to *Leishmania amazonensis* infection. It was found that BALB/c mice had a worse disease outcome compared to C57BL/6 mice, with higher parasitic load, ulcerated lesion formation, and higher levels of IL-6 in infected paws. On the other hand, C57BL/6 presented higher levels of IFN- γ and superoxide anion ($\bullet\text{O}_2^-$) after 11 weeks of infection and no lesion ulcerations. A peak of local macrophages was observed after 24h of infection in both studied mice strains, followed by a further increase after 240h detected only in C57BL/6 mice. Regarding M1 and M2 macrophage phenotype markers (iNOS, MHC-II, CD206, and arginase-1 [Arg-1]), we found that in BALB/c there was a pronounced increase in Arg-1 levels after 11 weeks of infection, whereas in C57BL/6 there was an initial predomination of markers from both profiles, followed by an M2 predominance, that coincided with the second peak of macrophage infiltration, 240h after the infection. Greater deposition of type III collagen and lesion resolution was also observed in C57BL/6 mice. The adoptive transfer of macrophages from C57BL/6 to infected BALB/c at the 11th week shown a reduction in both edema and the number of parasites at the lesion site, besides lower levels of Arg-1. Thus, C57BL/6 mice have a more effective response against *L. amazonensis*, based on a balance between inflammation and antioxidant response, coinciding with better tissue repair, while BALB/c mice have an excessive Arg-1 production in late infection times, which is related to the development of disease severity. The worst evolution seems to be involved with the recruitment of Arg-1 related macrophages, since the adoptive transfer of macrophages from C57BL/6 mice to BALB/c resulted in better outcomes, with lower levels of Arg-1.

1 Introduction

Leishmaniasis is a group of infectious diseases caused by species of protozoan parasites of the *Leishmania* genus, which are transmitted to animals and humans through the bite of female infected phlebotomine sand flies. The main clinical forms of the disease are cutaneous leishmaniasis (CL), visceral leishmaniasis, and mucocutaneous leishmaniasis. CL is the most common form, with estimates of 1 million new cases annually worldwide (WHO, 2020).

Considered a neglected tropical disease by the World Health Organization, leishmaniasis represents a great challenge in the pharmacological field with no experimental vaccine candidates with satisfactory progression in human trials until now (Lage et al., 2020). Additionally, the available treatment for CL present high toxicity and is not fully effective (Caridha et al., 2019). A range of factors could explain the lack of vaccines and therapeutic options against CL, like the diversity among *Leishmania* species and the complexity of the host's immune responses (Müller et al., 2018; Kaye et al., 2020). Thus, the understanding of the effective immune response to *Leishmania*-infection is fundamental to help drug discovery and vaccine development.

Since the discovery of T CD4⁺ helper 1 (Th1) and Th2 cells, experimental studies of CL have answered basic immunological questions related to the development of the lesions (Scott and Novais, 2016). It was postulated by experimental studies using *L. major* that C57BL/6 mice present a predominant Th1 response, associated with infection control, while BALB/c mice developed a Th2 response, favoring the disease progression (Sacks and Noben-Trauth, 2002; Scott and Novais, 2016). However, it is currently known that the Th1/Th2 paradigm is not valid for all *Leishmania* species.

In *L. amazonensis* infections, BALB/c mice also develop a lesion and a predominant Th2 response, however, C57BL/6 mice manifest a mixed immune response with mediators from both Th1 and Th2 patterns, being considered partially resistant to the infection (Soong, 2012; Pratti et al., 2016; Scott and Novais, 2016). This mixed response is similar to those observed in human infections, validating the biological relevance of these mice models for the study of cutaneous leishmaniasis (Soong, 2012).

Is noteworthy that *Leishmania* parasites can actively manipulate their hosts and subvert the microbicide mechanisms through the modification/delay of the development of a type 1 response (Gregory and Olivier, 2005; Rossi and Fasel, 2018; Aoki et al., 2019). These escape mechanisms have been extensively characterized and vary according to the parasite species and host genetic background (Alexander et al., 1980; Lira et al., 2000; Rosas et al., 2005; Velasquez et al., 2016; Muxel et al., 2018).

Besides this, the clinical course of *Leishmania* spp. infections is not only dependent on T cells response and involves a complex range of cells, including macrophages, the main host cells

Susceptibility to *L. amazonensis* is arginase-1 dependent

for parasite replication. The macrophages present a dual role during the infection, providing a safe place for parasites' survival inside the parasitophorous vacuole, but also triggering an inflammatory response (cytokines production and oxidative stress response) that can control the parasite replication. Therefore, macrophages are the key cell to disease progression, and their interaction with the parasites can dictate the success or failure of the infection (Liu and Uzonna, 2012).

There are two main macrophage types described, M1 or "classically activated" and M2 or "alternatively activated" (Mills et al., 2000). M1 macrophages are activated by the Th1 lymphocyte subpopulation and produce interferon-gamma (IFN- γ) and tumor necrosis factor-alpha (TNF- α), triggering the microbicide machinery and inducing the production of reactive species, especially superoxide anion (O_2^-) by NADPH oxidase enzyme, and nitric oxide (NO) by inducible nitric oxide synthase (iNOS), that eliminates *Leishmania* sp. parasites (Rossi and Fasel, 2018). However, intense activation of M1 macrophages in the attempt to control the infection can trigger inflammation with tissue damage and exacerbation of the lesion (Laskin et al., 2011).

On the other side, M2 macrophages are activated mainly by IL-13 and IL-4 produced by Th2 cells, which in turn activates the enzyme arginase-1 (Arg-1), culminating in the synthesis of polyamines, which allows *Leishmania* intramacrophagic replication, favoring parasite survival and disease progression (Pessenda and Santana da Silva, 2020). Meanwhile, despite the characteristics of permissiveness to infection, the M2 macrophage (and the Arg-1 activity) are also responsible for tissue remodeling and repair, and inflammation resolution (Laskin et al., 2011). As iNOS and Arg-1 share L-arginine as substrate, these enzymes represent two possible pathways of the immune response against *Leishmania* (Mills et al., 2000; Pessenda and Santana da Silva, 2020).

In this context, a balance between a potent microbicide response, combined with a resolutive and healing process seems to be the key to provide the utmost benefit for the host (Tomiotto-Pellissier et al., 2018). Nonetheless, little is known about this balance and the role of M1 and M2 related molecules in leishmaniasis, in which the understanding may be important to the potential development of drugs and vaccines.

Based on this, this work aimed to demonstrate the differential response established by susceptible (BALB/c) and partially resistant (C57BL/6) mice to *Leishmania amazonensis* infection. The results showed that BALB/c mice have a worse disease outcome compared to C57BL/6 mice, with higher levels of Arg-1. When the adoptive transfer of peritoneal macrophages from C57BL/6 to infected BALB/c was carried out, it was found a reduction in the parasite burden, lesion edema, and ulceration, concomitant with a reduction in the Arg-1 levels. Thus, the worst evolution of CL in BALB/c mice seems to be involved with the recruitment of Arg-1 related macrophages in the later times of *L. amazonensis* infection.

2 Materials and Methods

2.1. Culture of *Leishmania (L.) amazonensis*

Promastigote forms of *Leishmania (L.) amazonensis* (MHOM/BR/1989/166MJO) were maintained in 199 growth medium (GIBCO, Invitrogen, USA) supplemented with 10% fetal bovine serum (FBS, GIBCO Invitrogen), 1 M HEPES, 0.1% human urine, 0.1% L-glutamine, 10 µg/mL penicillin and streptomycin (GIBCO Invitrogen) and 10% sodium bicarbonate (CAQ, Brazil). The cell cultures were maintained in an incubator-type B.O.D. at 25 °C, in 25 cm² flasks. In all experiments, promastigote forms were used in the stationary growth phase.

2.2. Animals and Infection

BALB/c and C57BL/6 mice weighing approximately 25–30 g and aged 6–12 weeks were kept under sterile conditions and used according to protocols approved by the Institutional Animal Care and Committee. This study was approved by the Ethics Committee for Animal Experimentation of the State University of Londrina, number 8286.2016.60.

Mice were infected on the footpad of both hind paws, subcutaneously, with 10⁵ promastigote forms of *L. amazonensis*/paw. The animals were divided into groups and evaluated at 6, 24, 72, 144, and 240 hours, and 11 weeks post-infection (p.i.). Paw edema was measured weekly using a digital caliper (Starrett 799).

After the end of each time point, the animals were euthanized by intraperitoneal inoculation of 100 mg/kg ketamine (Ceva, Brazil) and 10 mg/kg xylazine (JA, Brazil) followed by cervical dislocation, and the footpads of infected paws were collected, weighed and processed for the next assays.

2.3. Parasite burden analysis

Quantitative PCR (qPCR) was performed to determine the tissue parasite load in each group. Briefly, hind paw samples were mechanically homogenized (Tissue-tearor, BioSpec) in TELT buffer (50 mM de Tris-HCl pH 8, EDTA 62,5 mM, Triton-X 4% e LiCl₂ 2.5 M) and DNA extraction was performed with the Easy-DNA kit (Invitrogen, USA, K1800-01) according to the manufacturer's instructions. After, it was added a solution of phenol: chloroform: isoamyl alcohol (25:24:1), and two volumes of cold ethanol (Merck, Germany) were added to the aqueous phase, and samples were stored at -20°C for 12 h. Samples were then centrifuged for 30 min at 10,000 g, washed with 70% ethanol, dried at room temperature, and resuspended in 10 mM Tris-HCl (pH 8.5). qPCR was performed by using GoTaq qPCR Master Mix (Promega, USA) with 100 ng total genomic DNA (gDNA). Parasite quantification was performed using AAP3 gene primer 5'-GGCGGCGGTATTATCTCGAT-3' (Forward) 5'-ACCACGAGGTAGATGACAGACA-3'

Susceptibility to *L. amazonensis* is arginase-1 dependent

(Reverse) *Leishmania*-specific primers at 10 pM (Tellevik et al., 2014). The samples were amplified with a StepOnePlus Real-Time PCR System (Applied Biosystems, USA) under the following PCR conditions steps: 2 min at 50°C, 2 min at 95°C, 40 cycles of 15 s at 95°C, 1 min at 55°C, followed by a dissociation step of 55 - 99°C (heating of 0.5°C/s). The results were based on a standard curve constructed with DNA from culture samples of *L. amazonensis* promastigote forms.

2.4. Cytokines determination

Th1, Th2, and Th17 cytokines were evaluated in the supernatant of the paw homogenates. The analyzes were performed using Cytometric Beads Array (CBA) assays using a commercial kit (BD Bioscience, USA) following the manufacturer's recommendations. The concentration of each cytokine was determined regarding the standard curve generated from reading the different dilutions of the recombinant cytokine. The limit of detection for each cytokine was 0.1, 0.03, 1.4, 0.5, 0.9, 0.8 and 16.8 pg/mL, respectively for IL-2, IL-4, IL-6, IFN- γ , TNF- α , IL-17 and IL-10. The data were obtained in BD Accuri C6 flow cytometer and analyzed using FCAP Array v. 3.0.1 software.

2.5. Superoxide anion measurement

The superoxide anion ($\bullet\text{O}_2^-$) production in the paw homogenates was evaluated by testing with nitroblue tetrazolium (NBT, Sigma-Aldrich, USA). For the assay, 10 μL of the sample were plated and 100 μL of the solution containing NBT (1 mg/mL) was added. After 15 minutes of reaction at room temperature, the samples were fixed with 10 μL of methanol, and the formazan, product of the reaction between NBT and $\bullet\text{O}_2^-$, solubilized by the addition of 120 μL of 2M KOH (Merck). The readings were performed on a spectrophotometer (GloMax Explorer instrument, Promega) at 660 nm and the results normalized per mg of tissue.

2.6. Determination of N-acetylglucosaminidase (NAG) activity

For the indirect quantification of the presence of macrophages at the infection site, the activity of the enzyme NAG was measured. The evaluation of the cellular infiltrate in the animals' footpad samples was performed by a colorimetric method described by (Bradley et al., 1982). Briefly, the samples were collected in 50 mM potassium phosphate buffer (pH 6.0) containing 13.72 mM HTAB (hexadecyl trimethyl ammonium bromide) and stored at -20 °C. The samples were then homogenized and centrifuged (21293 \times g, 2 min, 4 °C). The supernatant was used for the colorimetric reaction in a 96-well plate. Each aliquot of 15 μL of the sample was added to 200 μL of the reaction solution containing 52.64 mM of N-acetylglucosaminidase. The readings were

Susceptibility to *L. amazonensis* is arginase-1 dependent

performed on a spectrophotometer (GloMax Explorer instrument, Promega) at 450 nm, and the number of macrophages per mg of tissue was determined using a standard curve.

2.7. Histological processing

For the immunohistochemistry and collagen quantification, one of the paws of each animal was fixed in 10% formalin solution and after 24 hours they were decalcified for 21 days in 5% nitric acid (Anidrol, Brazil) and then processed for inclusion in paraffin. The cuts (5 µm thick) were adhered to slides treated with poly-L-lysine (Sigma-Aldrich), washed in xylol (FMAIA, Brazil) to remove excess paraffin, and rehydrated with decreasing concentrations of alcohol.

2.8. Immunohistochemistry of lesions for iNOS, MHC-II, Arg1, and CD206 labeling

The rehydrated histological sections of paws were submitted to antigenic recovery in citrate and acetate buffer solution (pH 6) for 10 minutes in a microwave (40W, 90°C). Endogenous peroxidase was blocked in a solution of 3% H₂O₂ (Anidrol), 10% methanol (Merck), and 0.1% tween-80 (Sigma-Aldrich) for 30 min and, subsequently, the non-specific sites blocked with PBS containing 1% FBS (GIBCO Invitrogen) for 30 min at room temperature. Then, the primary antibodies were added to the labeling of iNOS (NOS2, dilution 1:500, Santa Cruz Biotechnology, USA, Cat. SC7271), MHC-II (1:500, Santa Cruz, SC59318), arginase-1 (Arg-1, 1:600, Santa Cruz, SC18351) and CD206, 1:400, Santa Cruz, SC34577) for 2h at 37°C. Subsequently, the universal solution of secondary antibodies with biotinylated anti-rabbit, anti-mouse, and anti-goat IgG (LSAB + System-HRP, DAKO, Japan) was added for 1 hour in room temperature. After three washes with PBS, the slides containing the samples were incubated with the avidin-peroxidase (LSAB + System-HRP, DAKO, K069011-2) complex for 40 minutes at room temperature. Finally, it was added 3,3'-Diaminobenzidine (DAB, DAKO, K3468), and the counterstaining was performed with hematoxylin (Merck).

For determining a quantitative scoring, images were evaluated by using the color deconvolution tool from the Image J software (NIH, USA). Data are expressed in pixels (% of labeling in the image area).

2.9. Collagen quantification

The quantification of collagen in the paws lesion was performed by the Sirius-red staining method, assessed under polarized light through a photomicroscope (CARL ZEISS- Axio Imager A1) with a camera (HBO 100) coupled to a computer using the AxioVision software, with a 200x magnification. Five images of two sections of each animal were considered for analysis using the

Image-Pro Plus software (version 4.5). The results were expressed as a percentage of area with the presence of total collagen and type III collagen in the measured area (Miranda et al., 2015).

2.10. Adoptive transference of macrophages (ATM)

BALB/c and C57BL/6 mice were kept under the same conditions described in item 2.2.

Macrophages were recovered from the peritoneal cavity of non-infected C57BL/6 with cold PBS supplemented with 3% of FBS (GIBCO Invitrogen) and stained with antibodies F4/80-PE (dilution 1:100, Santa Cruz Biotech, SC377009) and CD11b-FITC (dilution 1:100, Santa Cruz Biotech, SC23937). The antibody staining profile was analyzed by flow cytometry (BD Accuri C6 flow cytometer) using FCAP Array v3.0.1 software.

BALB/c mice were infected on both hind paws, subcutaneously, with 10^5 promastigote forms of *L. amazonensis*/paw, and randomly divided into three groups: BALB/c ATM IP – animals who received the ATM with macrophages from C57BL/6 via intraperitoneal injection, BALB/c ATM IV – who received the ATM with macrophages from C57BL/6 via intravenous injection, and BALB/c – animals who not received ATM (n = 6/group).

At the ninth week of infection, the BALB/c ATM groups received the transference of 5×10^5 peritoneal macrophages from C57BL/6 mice via intraperitoneal (BALB/c ATM IP) or by retro-orbital injection (BALB/c ATM IV; under anesthesia with 100 mg/kg ketamine (Ceva) and 10 mg/kg xylazine (JA) (Figure 6A). At the end of the eleventh week, the animals were euthanized as described in 2.2. The edema was measured weekly as described in item 2.2, the parasite burden analysis was performed as described in item 2.3 and the cytokines measurement was performed as item 2.4.

2.11. Enzyme-Linked Immunosorbent Assay for iNOS and Arg-1 in paws homogenate

Radioimmunoprecipitation assay buffer (RIPA lysis buffer - 140 mM sodium chloride, 1% Triton X-100, 0.1% sodium deoxycholate, 0.5% sodium dodecyl sulfate, 1 mM EDTA, 10 mM Tris, pH 8.0 and 1 mM phenylmethanesulfonyl fluoride (PMSF) was added (1 mL) to the hind paws samples, mechanically homogenized (Tissue-tearor, BioSpec), and incubated for 2h, 4°C. The lysates were collected and centrifuged at $13,000 \times g$ for 20 min at 4°C, the supernatants were transferred to a new tube and the total protein concentration was quantified in NanoVue Plus GE Healthcare (Biochrom, USA). The protein concentration of all samples was normalized to 10 µg/mL and a 50 µL aliquot was added in a 96-well ELISA plate for adsorption of proteins overnight at 4°C, followed by incubation for 1h with blocking buffer (ELISA/ELISPOT, eBioscience, USA). The wells were washed 3x with wash buffer (PBS + 0.5% Tween 20 (Sigma-Aldrich)) and incubated with primary antibody anti-mouse iNOS (Santa Cruz Biotech, SC7271) and Arg-1 (Santa

Susceptibility to *L. amazonensis* is arginase-1 dependent

Cruz Biotech, SC18351) at 1:500 dilutions for 1h at room temperature. The wells were washed to remove unbound antibodies, followed by the addition of universal biotinylated secondary antibody (LSAB2 System-HRP, Dako), 1h incubation, washing, and addition of streptavidin-HRP (LSAB2 System-HRP, Dako) for 1h. After the incubation time, the wells were washed 5 times and 100 μ L of TMB Substrate Solution (eBioscience, 00-4201-56) was added, followed by incubation for 30 min and 100 μ L of stop solution (1N sulfuric acid) was added. The plate reading was performed in a microplate reader at 450 nm (GloMax, Promega), and the results are expressed as arbitrary units.

2.12. Statistical analysis

Data were expressed as a mean \pm SEM. Data were analyzed using the GraphPad Prism statistical software (GraphPad Software, Inc., USA, 500.288). Differences between the two groups were evaluated using the student's t-test. Comparisons between multiple groups were done using ANOVA, followed by Tukey's test. Differences were considered as statistically significant upon $p < 0.05$. At least five animals per group were evaluated.

3 RESULTS

3.1. BALB/c mice show worse evolution of *L. amazonensis* infection when compared to C57BL/6 mice

The evolution of *L. amazonensis* infection was analyzed by the paw lesion edema, measured weekly until the eleventh week after infection. The edema was similar on both BALB/c and C57BL/6 mice until the eighth week of infection, however, from the ninth week on, the BALB/c mice showed significantly higher edema than the C57BL/6 (Figure 1A). Also, after 11 weeks of infection, ulcerated lesions were observed in all BALB/c mice, while none of the C57BL/6 strain had ulcerations (Figure 1B).

Regarding the parasitic load, we observed that BALB/c mice showed a significantly greater amount of parasites in the infected paws than the C57BL/6 mice after 11 weeks of infection (Figure 1C).

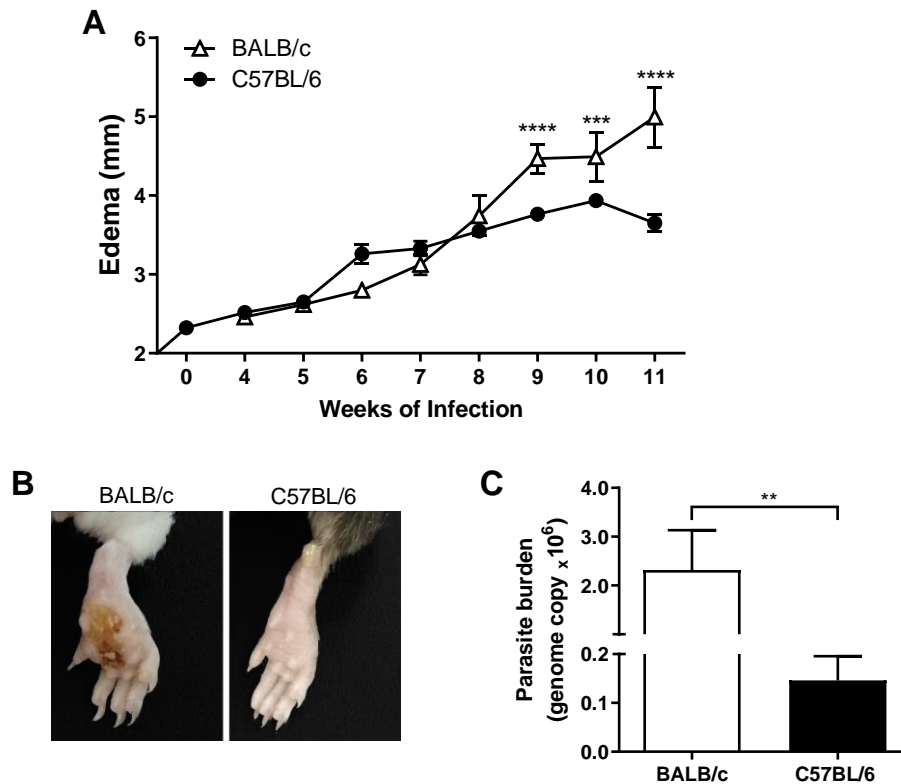


Figure 1 - Evolution of *L. amazonensis* infection in BALB/c and C57BL/6 mice. (A) BALB/c and C57BL/6 mice were infected on the hind paws with 10^5 promastigote forms of *L. amazonensis* and the edema was analyzed for 11 weeks. (B) Representative images of BALB/c and C57BL/6 mice paws 11 weeks after the infection with *L. amazonensis*. (C) Parasitic load (number of *Leishmania amazonensis* kDNA copies) determined in the mouse paw homogenate by quantitative real-time PCR after 11 weeks of infection. Data represent the mean \pm SEM of 6 mice group. **Significant difference in relation to the opposite strain infected by *L. amazonensis*, $p \leq 0.01$, *** $p \leq 0.001$, **** $p \leq 0.0001$.

3.2. Cytokines are differentially produced by *L. amazonensis*-infected BALB/c and C57BL/6 mice paw

Based on the knowledge that *L. amazonensis* infection leads to a differential inflammatory response in BALB/c and C57BL/6 mice we measured the production of inflammatory cytokines in paws homogenates of infected-mice after 11 weeks. C57BL/6 mice presented higher levels of IFN- γ (Figure 2A), while BALB/c exhibited higher levels of IL-6 (Figure 2C). TNF- α levels, 11 weeks after infection, did not differ in BALB/c and C57BL/6-infected mice (Figure 2B). IL-2, IL-4, IL-10, and IL-17 values, as well as IFN- γ , TNF- α , and IL-6 levels at the infection times from 6 to 240h, remained below the detection limit of the technique; therefore, were not interpreted in the study.

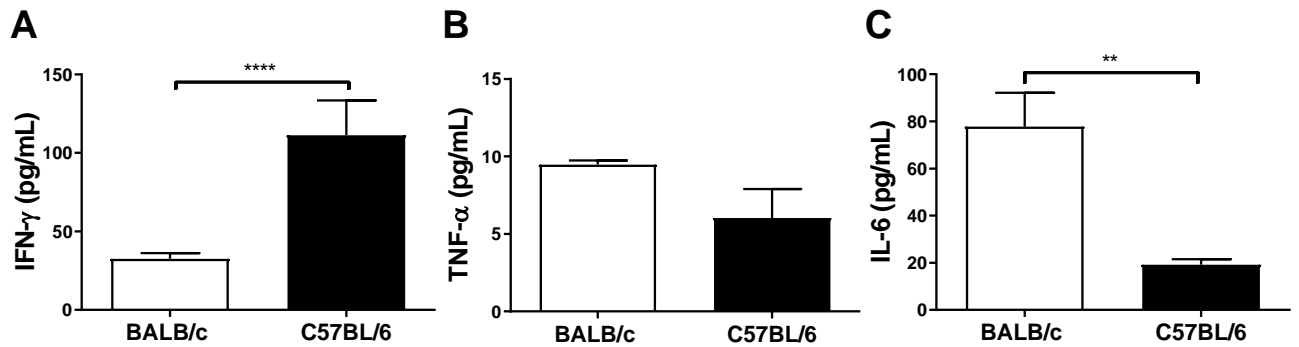


Figure 2 - Cytokines levels of BALB/c and C57BL/6 mice infected with *L. amazonensis*. Homogenates of *L. amazonensis*-infected BALB/c and C57BL/6 paws for 11 weeks were submitted to the CBA assay. (A) Measurement of IFN- γ , (B) TNF- α and (C) IL-6 levels after 11 weeks of infection. Data represent the mean \pm SEM of 5 mice group. * Significant difference in relation to the opposite strain infected by *L. amazonensis*, $p \leq 0.05$; ** $p \leq 0.01$; *** $p \leq 0.001$.

3.3. C57BL/6 mice produce higher levels of $\bullet\text{O}_2^-$ in the infection site when compared to BALB/c mice

Aiming to understand the mechanisms involved with the early and late-stage pathogenesis of *L. amazonensis* infection in C57BL/6 and BALB/c mice strains, it was measured the $\bullet\text{O}_2^-$ production (NBT assay) in paws homogenates. C57BL/6 mice had a higher basal production of $\bullet\text{O}_2^-$ (0h), when compared to BALB/c mice. After infection, these radical levels were drastically reduced in both strains of mice (6h) but were restored later in the C57BL/6 mice (72h to 11 weeks). In BALB/c mice, it was observed that there was no restoration of $\bullet\text{O}_2^-$ levels in the evaluated periods (no statistical difference between 6h and the later time points), which remained significantly lower when compared to C57BL/6 (72 to 240h) (Figure 3).

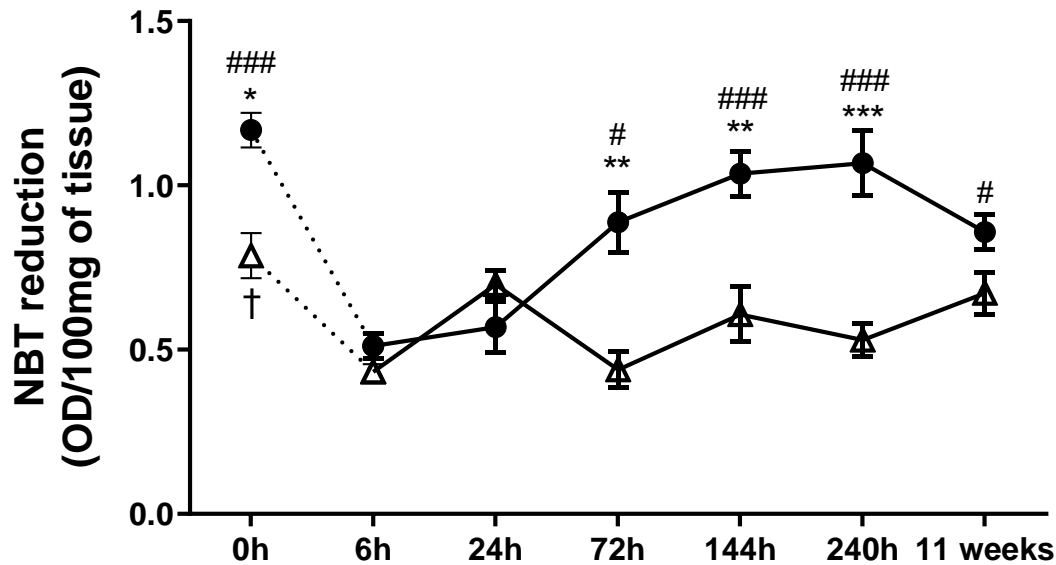


Figure 3 - Superoxide anion measured at the paws of BALB/c and C57BL/6 mice infected with *L. amazonensis*. Homogenates of BALB/c and C57BL/6 paws infected with *L. amazonensis* from 6h to 11 weeks submitted to the NBT assay. Uninfected mice were used as controls (0h). * Significant difference between mice strains infected by *L. amazonensis*, $p \leq 0.05$; ** $p \leq 0.01$, *** $p \leq 0.001$. #Difference in the time points vs. 6h within C57BL/6 mice, $p \leq 0.05$, ## $p \leq 0.01$, ### $p \leq 0.001$. †Difference in the time points vs. 6h within BALB/c mice, $p = 0.016$. Data represent the mean \pm SEM of 5 mice group.

3.4. Macrophage infiltration and M1 and M2 markers are different between BALB/c and C57BL/6 during *L. amazonensis* infection.

To understand the dynamics of macrophage infiltration in the lesion site, we analyzed the quantity of these cells in the paws throughout the infection. It was found that in BALB/c mice, a single peak of macrophage infiltration occurs after 24 hours post *L. amazonensis* infection and that the amount of local macrophages decreases later (144h to 11 weeks). On the other hand, C57BL/6 showed two peaks of macrophage infiltration, one more initial 24h after infection, and the later one at 240h p.i. (Figure 4A).

To identify the profile of macrophages at the infection site, it was performed the immunohistochemistry assay to identify the presence of MHC-II and iNOS labeling, related to the M1 profile, and CD206 and Arg-1 markers, related to the M2 phenotype. It was observed that at the beginning of infection (6h) the C57BL/6 mice show markers from both macrophages subpopulations when compared to BALB/c mice. After 24h, time point related to the first macrophage infiltration (Figure 4A), high CD206 marking was observed in C57BL/6. 72h p.i., Arg-1 was higher on C57BL/6 mice paws. After 144h, there was higher labeling for iNOS, Arg-1, and CD206 in C57BL/6 compared to BALB/c mice. In the time point that coincides with the second peak of infiltration, 240h, C57BL/6 mice presented higher marking of both Arg-1 and CD206 in

Susceptibility to *L. amazonensis* is arginase-1 dependent

the infection site. After 11 weeks of infection, this profile was altered, with C57BL/6 presenting a greater presence of M1 markers, and BALB/c showed significantly higher levels of Arg-1 marking when compared to the C57BL/6 (Figure 4B -E, G).

We also found that, after 240h of infection, C57BL/6 mice showed a higher Arg-1/iNOS ratio than BALB/c. However, after 11 weeks of infection, this balance was reversed, with a substantial increase in the proportion of marking for Arg-1 on the BALB/c paws compared to C57BL/6 mice (Figure 4F).

Furthermore, immunohistochemical images show intense labeling Arg-1 after 11 weeks of infection is coincident with areas with vacuolated macrophages filled with parasites (Figure 4G, Figure S1). This pattern was also verified in the heatmap (Figure 4H), where it was possible to identify prominent Arg-1 labeling in BALB/c, while the levels found in C57BL/6 were more evenly distributed among the analyzed markers and times.

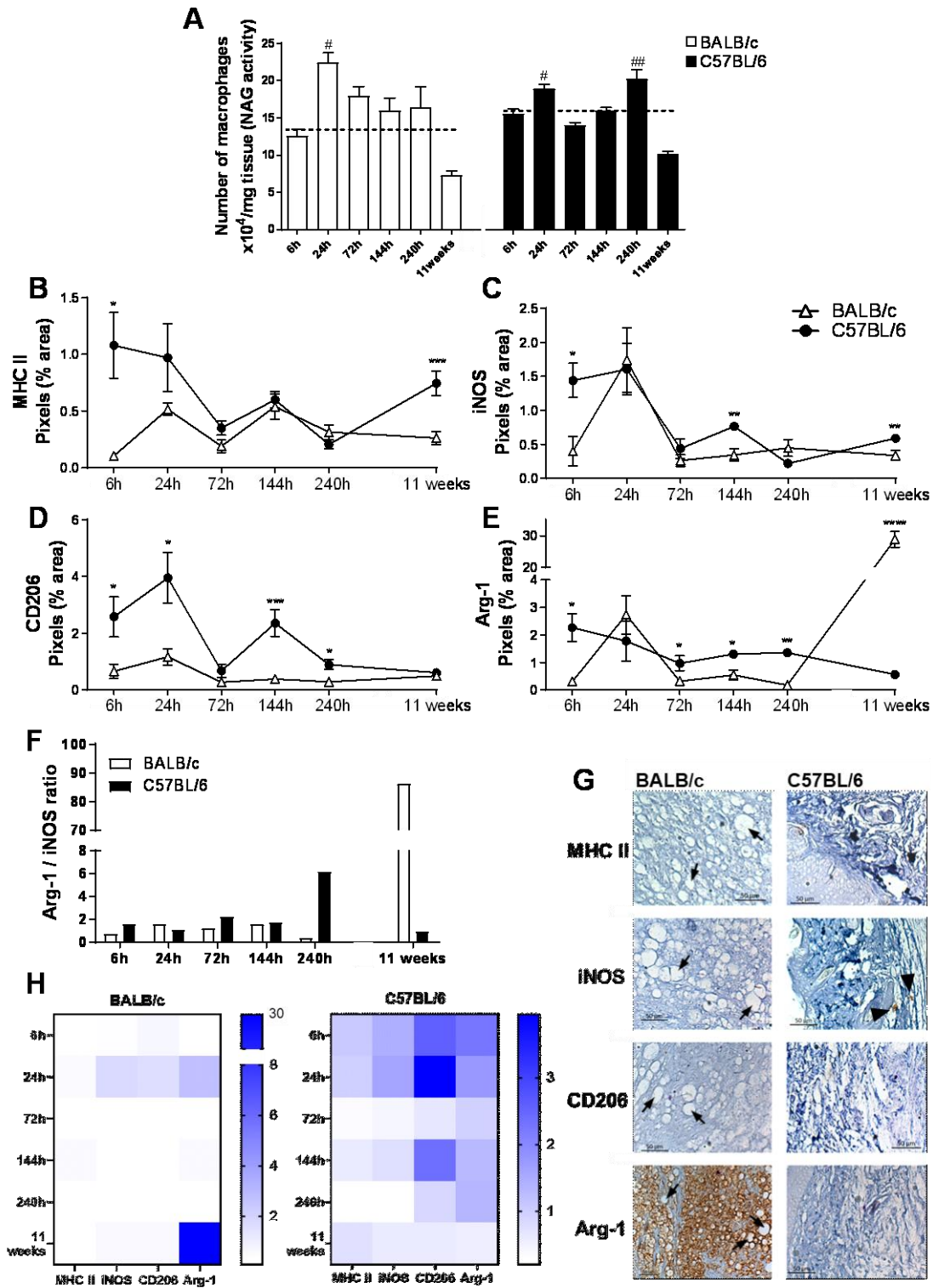


Figure 4 - Characterization of local macrophages in infected mice paws. (A) Indirect measurement of macrophages number by NAG activity. (B) Immunohistochemical quantification of MHC-II, (C) iNOS, (D) Arg-1, and (E) CD206 staining in the paw of infected mice. [#]Significant difference in relation to the non-infected mice (dashed lines), $p \leq 0.05$; ^{###} $p \leq 0.01$. ^{*}Significant difference between BALB/c and C57BL/6 mice infected with *L. amazonensis*, $p \leq 0.05$; ^{**} $p \leq 0.01$, ^{***} $p \leq 0.001$, ^{****} $p \leq 0.0001$. (F) Arg-1/iNOS ratio. (G) Representative figures of immunohistochemical labeling. The positive marking on immunohistochemistry sections

corresponds to the brown areas (arrowhead). Arrows indicate vacuolated macrophages filled with parasites. (H) Heatmap of MHC-II, iNOS, Arg-1, and CD206, in all evaluated time points. Data from A-E represent the mean \pm SEM of 6 mice group

3.5. Type III collagen deposition in the site of infection is higher in C57BL/6 mice in the late stages of infection

Knowing that tissue repair is closely related to the clinical course of leishmaniasis, it was performed the identification of total collagen and the newly deposited collagen (type III) in the paws of infected mice as a readout of tissue repair (Frahs et al., 2018). From the results obtained it was possible to verify that, at 240h and 11 weeks p.i., C57BL/6 mice had significantly more total and type III collagen than BALB/c (Figure 5A and B). Besides, in C57BL/6 the amount of total collagen increased from 240h to 11 weeks of infection (Figure 5A).

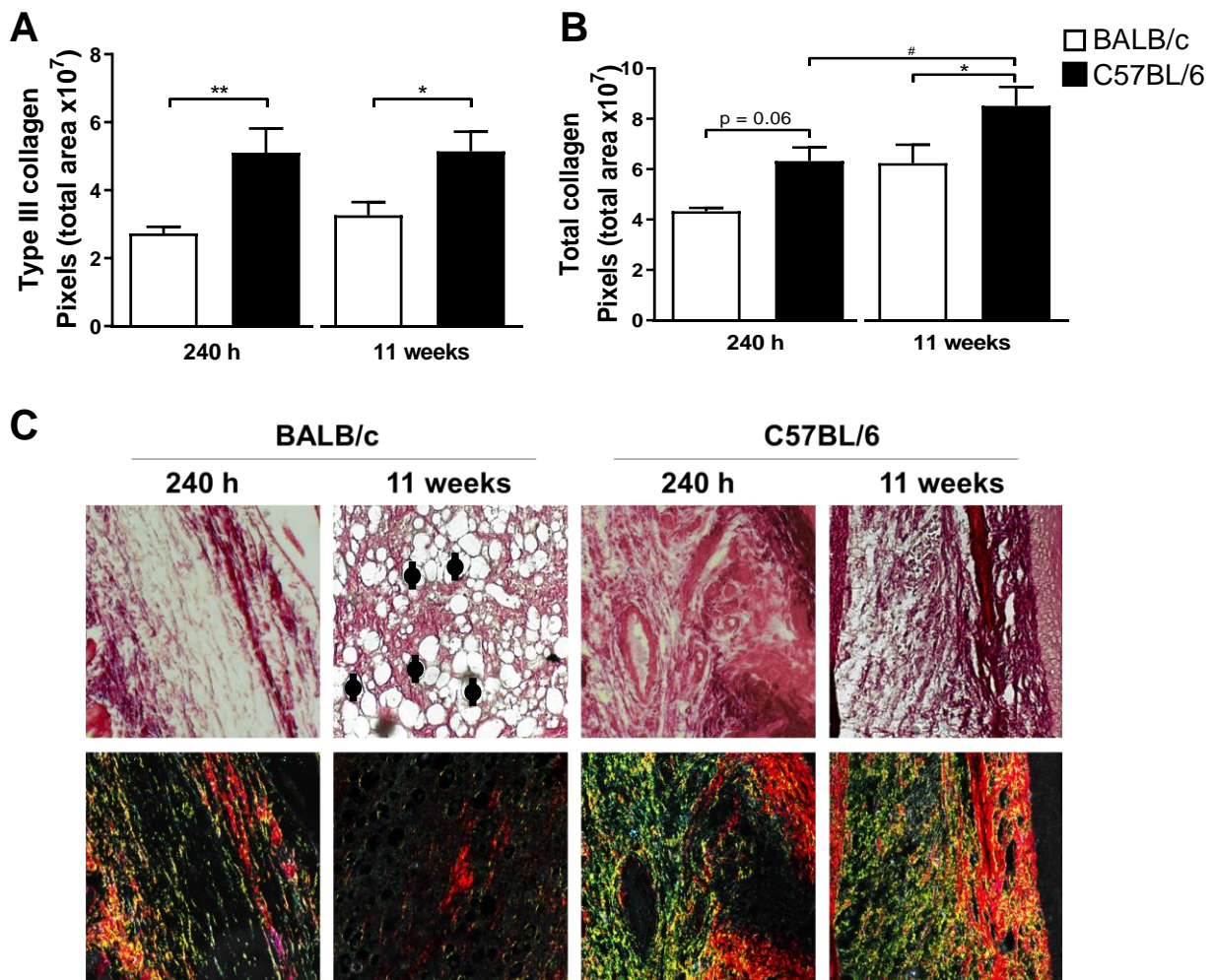


Figure 5 - Analysis of collagen in mice paws infected with *L. amazonensis*. Histological sections of the paws of *L. amazonensis*-infected BALB/c and C57BL/6 mice for 240h or 11 weeks p.i. were stained with Sirius Red to measure (A) total collagen and (B) type III collagen, using the Image-Pro Plus. * Significant difference between the mice strains infected by *L. amazonensis*, $p \leq 0.05$; ** $p \leq 0.01$, *** $p \leq 0.001$, **** $p \leq 0.0001$. #Significant difference of time points in C57BL/6 mice, $p \leq 0.05$. (C) Representative figures of histological sections of the paws of BALB/c and C57BL/6 mice infected by *L. amazonensis* for 240h and 11 weeks. ϕ - Indication of some

Susceptibility to *L. amazonensis* is arginase-1 dependent

parasitophorous vacuoles. The fibers stained in red correspond to type I collagen and in yellow/green correspond to deposited type III collagen. Data from A and B represent the mean \pm SEM of 6 mice group.

Figure 5C and Figure S1 show in BALB/c mice 11 weeks after infection the presence of vacuolated macrophages containing amastigotes (parasitophorous vacuoles), coinciding with areas of low labeling for type I and type III collagen. In C57BL/6 mice, after 240h, there were no visible vacuoles into macrophages and, consequently, reduction of parasites at the infection site. Also, it is notable the collagen deposition in the C57BL/6 sections, characterizing a tissue repair process.

3.6. The recruited macrophages to the site of the lesion are decisive in the evolution of edema and the arginase-1 presence

To verify the role of recruited macrophages on *L. amazonensis* lesion evolution, it was performed the adoptive transfer of macrophages (ATM) from partially resistant mice (C57BL/6) to susceptible ones (BALB/c) after 9 weeks of infection, time point in which the paw edema starts to significantly differ between the mice strains (Figure 1A and 6A). First, peritoneal cells from C57BL/6 were characterized as predominantly macrophages (F4/80⁺/CD11b⁺) (Figure 6B, gate strategy Figure S2). Then, we found that mice who received ATM intraperitoneal (ATM IP) presented lower paw edema and more than 250 times less parasite burden (Figure 6C, D). Besides, most ATM animals did not show any ulceration at the infection site, while BALB/c mice presented typical ulcerated lesions (Figure 6E).

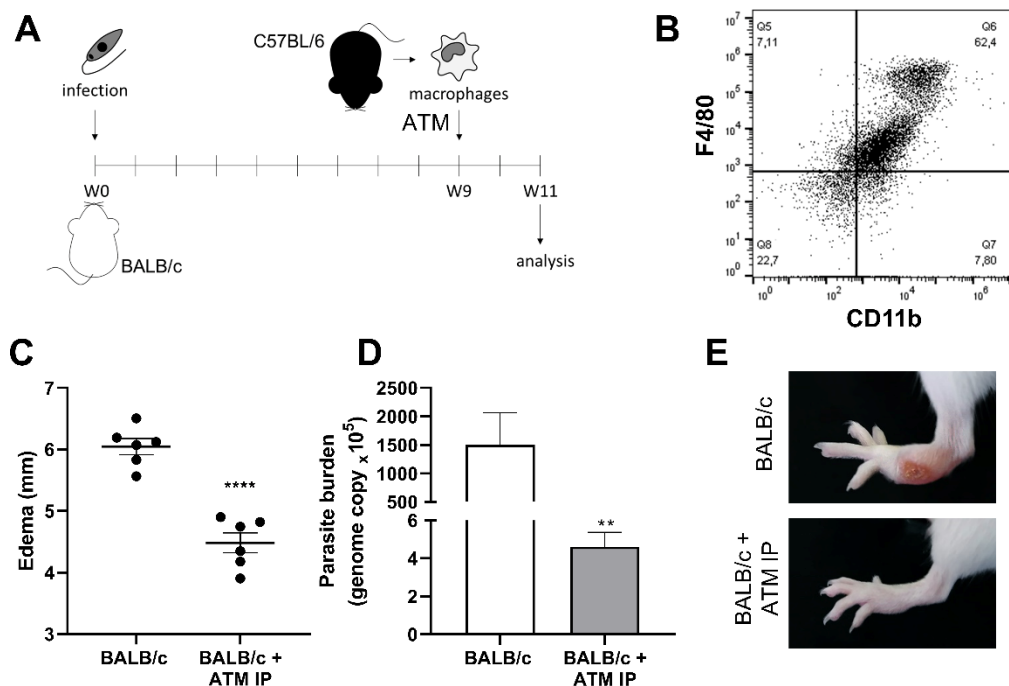


Figure 6 – Adoptive transfer of macrophages (ATM). (A) Representative scheme of the experimental design: BALB/c mice were infected with *L. amazonensis* and, after 9 weeks, they received peritoneal macrophages isolated from C57BL/6 mice intraperitoneally. (B) Dot plot showing the macrophage markers (F4/80⁺/CD11b⁺) in the peritoneal cells from non-infected C57BL/6 mice. (C) Paw edema of mice that did not receive the adoptive transfer of macrophages (BALB/c) and mice that received intraperitoneal ATM (BALB/c + ATM IP) after 11 weeks of infection. (D) Parasite burden and (E) representative photographs of mice paws after 11 weeks of infection. Data from C and D represent the mean ± SEM of 5 mice group* Significant difference in relation to the group that did not receive ATM, ** p ≤ 0.01, *** p ≤ 0.001, **** p ≤ 0.0001.

When analyzed the local cytokines at the eleventh week, ATM mice showed an increase of IFN- γ levels, while mice that did not receive ATM presented higher IL-6 levels (Figure 7A, B). It was also verified in ATM IP mice a reduction in Arg-1 levels, without significant variation in iNOS when compared to BALB/c who not received ATM (Figure 7C, D). Similar results were found when ATM was performed by intravenous route (ATM IV), with edema and Arg-1 reduction in ATM IV group, without significant difference in iNOS levels (Figure S3).

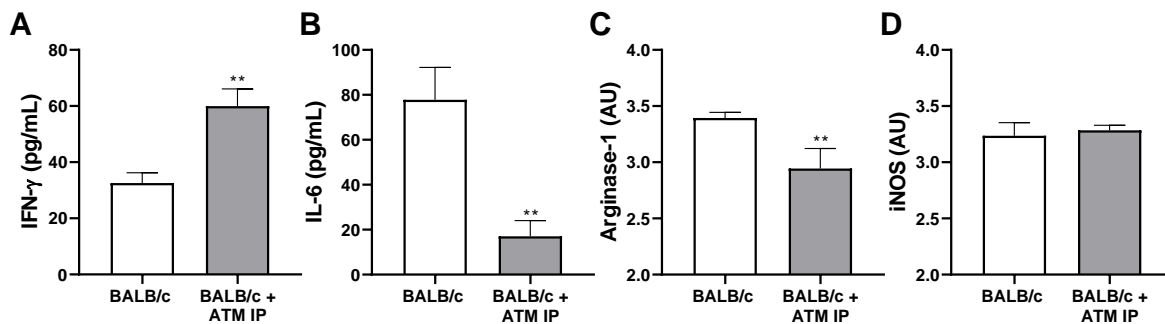


Figure 7 – Immunomodulation of ATM mice. Homogenates of *L. amazonensis*-infected BALB/c and BALB/c that received ATM via intraperitoneal paws were submitted to (A) IFN- γ and (B) IL-6 measurement by CBA assay; and (C) Arg-1 and (D) iNOS measurement by ELISA. Data represent the mean ± SEM of 5 mice group. ** Significant difference in relation to the opposite strain infected by *L. amazonensis*, p ≤ 0.01. ATM – adoptive transfer of macrophages. IP – intraperitoneal. AU – arbitrary units.

4 DISCUSSION

The pathological mechanisms that lead to cutaneous lesions caused by *L. amazonensis* infection are not yet fully understood. The present work aimed to elucidate the role of different molecules and cells, comparing the evolution of *L. amazonensis* infection in mice strains partially resistant and susceptible to infection

The most studied experimental model of cutaneous leishmaniasis is the *L. major* infection, in which the susceptibility of BALB/c mice has been correlated with the predominance of Th2 immune response, while C57BL/6 mice are resistant to infection, with a predominance of Th1 type response (Sacks and Noben-Trauth, 2002; Scott and Novais, 2016). However, this dichotomy is not identified in the disease induced by other *Leishmania* species, as *L. amazonensis* (Soong, 2012; Scott and Novais, 2016). Here, it was shown that BALB/c mice, had progressive edema from 0 to 11 weeks after *L. amazonensis* infection, with ulceration of the lesion and high local parasitic load 11 weeks p.i. On the other hand, C57BL/6 mice presented initial edema that was steadied at the later times analyzed, did not present ulcerated lesions, and had lower parasitic load compared to BALB/c.

It was also found that 11 weeks post-infection, BALB/c mice showed higher production of IL-6, which play an important role in the leishmaniasis pathogenesis, triggering the down-modulation of the microbicide molecules, and the polarization of macrophages to an M2-phenotype, that are more permissive to *Leishmania* proliferation (Hatzigeorgiou et al., 1993; Hu et al., 2018). On the other hand, C57BL/6 mice had a significant increase in IFN- γ levels after 11 weeks of infection, a key cytokine in the activation of inflammatory macrophages (M1), associated with protection in several experimental models of infection (Laskin et al., 2011), including *Leishmania* sp. (Wang et al., 1994; Sacks and Noben-Trauth, 2002).

Moreover, IFN- γ triggers the synthesis of reactive oxygen species (ROS), such as $\bullet\text{O}_2^-$. ROS are produced by M1 macrophages, causing oxidation of lipids, proteins, and nucleic acids, with the consequent elimination of intracellular parasites (Murray and Wynn, 2011; Jafari et al., 2014). C57BL/6 mice showed higher levels of $\bullet\text{O}_2^-$ over the infection, with lower parasitic load, explaining a better lesion evolution in this mice strain.

However, an exacerbated pro-oxidant response can be harmful to the host during *Leishmania* sp. infection, since the action of ROS are nonspecific and causes damage to the surrounding host cells, and consequently, inflammation and tissue injury (Morgado et al., 2018; Herb and Schramm, 2021). Therefore, in the presence of pro-oxidant molecules, such as $\bullet\text{O}_2^-$, adaptive homeostasis mechanisms must be activated as a way to protect against tissue damage (Pomatto et al., 2019). Mice of the C57BL/6 strain, despite producing $\bullet\text{O}_2^-$ in a more pronounced

Susceptibility to *L. amazonensis* is arginase-1 dependent

manner, showed higher collagen deposition, suggesting an ability to balance the pro- and antioxidant properties, and explaining the concomitant elimination of the parasite combined with the tissue protection capacity found in C57BL/6 mice (Morgado et al., 2018).

Regarding macrophages, it was verified a peak of infiltration in the mice paws of both C57BL/6 and BALB/c mice after 24h of infection, corroborating the expected kinetics of migration of these cells (Minutti et al., 2017). However, at the beginning of the infection, there was no clear polarization of macrophages for M1 or M2 phenotype in both mice strains. These data are in accordance with the mixed Th1/Th2 profile induced by *L. amazonensis* infection (Scott and Novais, 2016), and with the findings of Ontoria et al., (2018), that verified an inconstant distribution of M1 markers throughout the infection by *L. infantum* between 1 and 8 weeks.

The results also showed that after 240h of infection there was a second peak of macrophages infiltration in the C57BL/6 mice, with higher Arg-1 and CD206 marking compared to BALB/c. This indicates that macrophages recruited at this point are predominantly M2, which play an important role in tissue remodeling and wound healing (Tomiotto-Pellissier et al., 2018). Krzyszczyk et al. (2018) showed that as the tissue begins to recover, the general population of M2 macrophages induces the migration and proliferation of fibroblasts, keratinocytes, and endothelial cells to restore the dermis, epidermis, and vasculature, respectively, by remodeling the injured tissue. In CL wounds, this remodeling is marked by the control of inflammation and an increase in the amount of collagen at the sites where the parasites have been eliminated (Baldwin et al., 2007; Miranda et al., 2015). The data are in agreement with these studies and show that collagen deposition is higher in areas with low parasitic load and tissue reorganization in the C57BL/6 mice infection.

Although there was an infiltration of M2 macrophages in 240h in C57BL/6 mice, in the later period studied, 11 weeks, there is a shift for M1 macrophages, coinciding with higher levels of IFN- γ , with less parasitic load, and no apparent lesion. On the other side, BALB/c mice had worse lesion evolution and a marked increase in Arg-1 (an M2 macrophage marker) in the later period analyzed (Muraille et al., 2014). Arginase-1 plays a fundamental role in the survival and multiplication of intracellular amastigotes of *Leishmania* sp., bypassing the amino acid L-arginine for the preferential synthesis of polyamines, fundamental for the parasite's nutrition (Muxel et al., 2018).

In vitro studies showed that *L. amazonensis*-infected BALB/c macrophages present increased L-arginine uptake and expression of arginase and arginase-related genes (Muxel et al., 2018; Aoki et al., 2019). The detrimental role of Arg-1 from macrophages on CL lesions was further confirmed by the adoptive transference of macrophages from C57BL/6 to BALB/c mice, in which it was verified that mice that received the cells presented better disease outcome, with similar features to those found in partially resistant mice. The recruited macrophages have been described

Susceptibility to *L. amazonensis* is arginase-1 dependent

as important weapons in *Leishmania* control because the tissue-resident macrophages present oxidative deficiency compared to the recruited ones (Pessenda and Santana da Silva, 2020).

An important role for arginase in the susceptibility of mice to *L. major* infection has already been described (Kropf et al., 2005), as well as its action in *L. amazonensis* infection *in vitro*. Interestingly, skin biopsies and plasma from CL patients also present high levels of Arg-1 (reviewed in (Muxel et al., 2018)). However, until now little was known about Arg-1 in *L. amazonensis* infection *in vivo*. We showed that the participation of this enzyme in the susceptibility of animals is particularly important in later times of infection.

Peritoneal macrophages are a heterogeneous population of M0, M1, and M2 cells, which can differentiate or redifferentiate into both M1 and M2 phenotypes (Zhao et al., 2017). Besides, these cells are recruited to other tissue under infection or inflammation conditions (Cassado et al., 2015). Thus, we performed the adoptive transference of macrophages from C57BL/6 to BALB/c mice. Our results suggested the recruited macrophages have great importance in controlling susceptibility to *L. amazonensis*. The transfer of macrophages from partially resistant mice (C57BL/6) to a susceptible mouse (BALB/c) modulates the pro-*Leishmania* environment to an anti-leishmania state, with high IFN- γ and low Arg-1 levels, less parasite burden and edema, and better disease outcome.

5 CONCLUSION

Taking together the results suggest that a balance between a pro-inflammatory and microbicide M1-related and a tissue restorative M2-related response seems to provide the utmost benefit for the host. We also demonstrated the higher levels of Arg-1 in later periods of infection are related to a poor disease outcome in susceptible mice, but the transference of macrophages from partially resistant mice (C57BL/6) restore the host protection and disease control. These results expand our understanding of the protective immunity against *L. amazonensis*, providing new insights about potential interventions to treat or prevent the disease.

6 Conflict of Interest

The authors declare that the research was conducted in the absence of any commercial or financial relationships that could be construed as a potential conflict of interest.

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This is a provisional file, not the final typeset article

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SUPPLEMENTARY MATERIAL

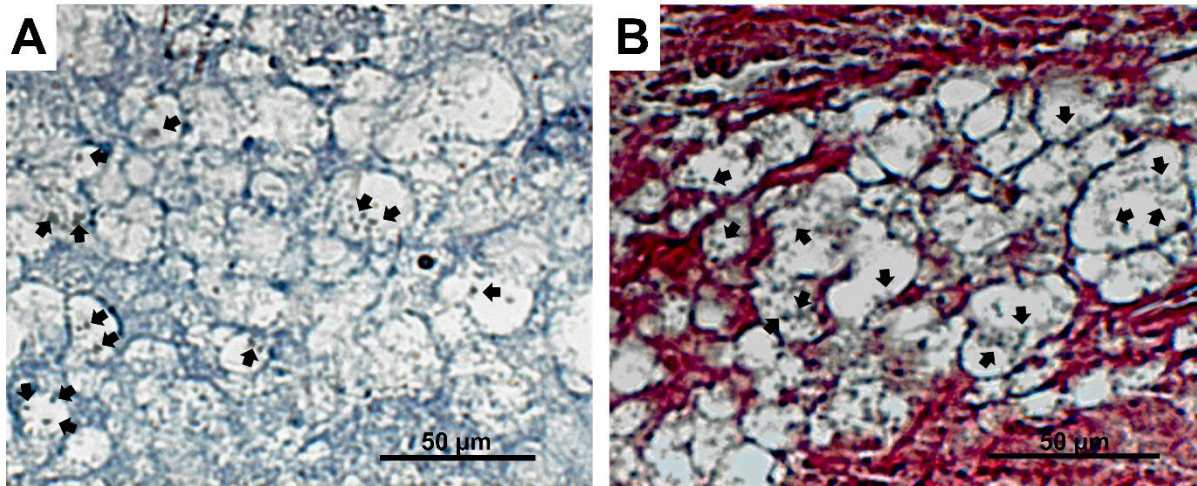


Figure S1 – Vacuolated macrophages in paws of BALB/c mice infected with *L. amazonensis* for 11 weeks. (A) Lesion areas of cuts stained with hematoxylin. (B) Lesion areas of cuts stained with Sirius Red. Both images show macrophage vacuoles filled with parasites. The arrows indicate *L. amazonensis* amastigote forms inside parasitophorous vacuoles. Scale bar indicates 50 μ m.

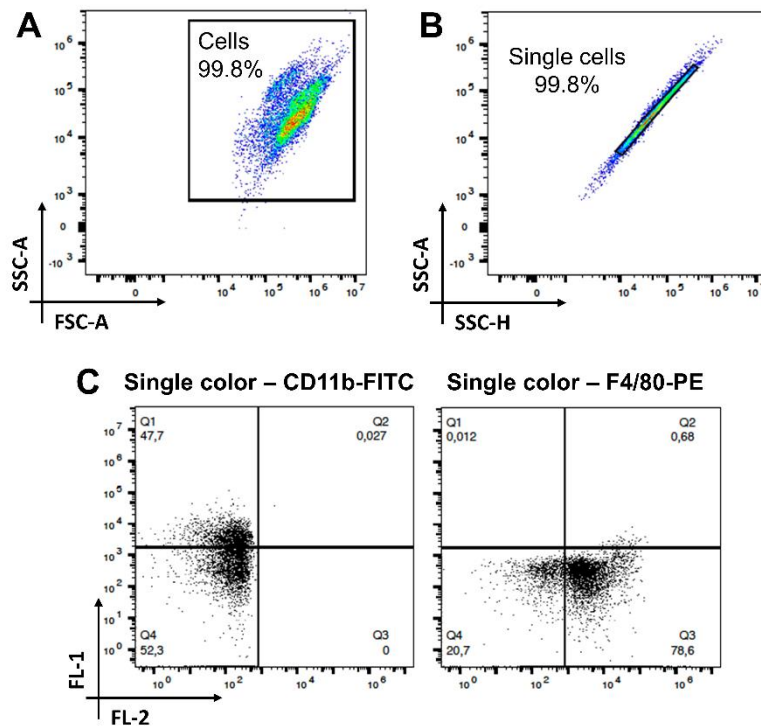


Figure S2 – Flow cytometry gating strategy for macrophages. (A) Cells were first identified by a forward scatter (FSC) and side scatter (SSC) gate. (B) Gating strategy for identifying singlets (single cells). (C) Single-color compensation controls for each fluorophore. Flow cytometry plots are shown in logarithmic format.

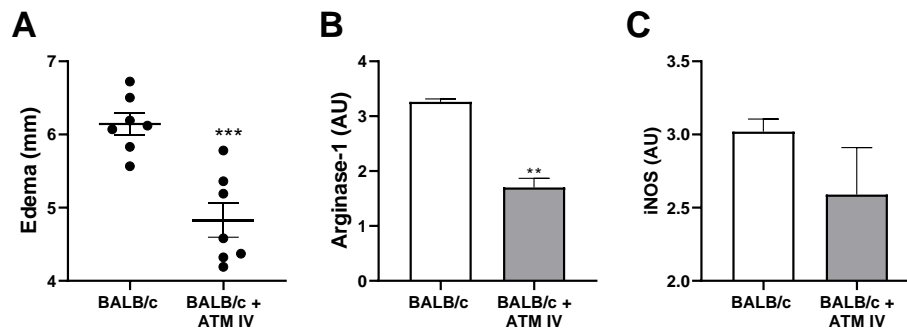


Figure S3 – Analysis of intravenous ATM. (A) Paw edema of mice that did not receive adoptive transfer of macrophages (BALB/c) and mice that received intravenous ATM (BALB/c + ATM IV) after 11 weeks of infection. Paws homogenates of were submitted to (B) Arg-1 levels. (C) iNOS levels. Data represent the mean \pm SEM of 5 mice group. ** Significant difference in relation to the group that did not receive ATM $p \leq 0.01$, *** $p \leq 0.001$. ATM – adoptive transfer of macrophages. IV – intravenous. AU – arbitrary units.

4. CONCLUSÕES

- Fatores de virulência dos parasitos atuam no favorecimento do desenvolvimento de macrófagos M2, que são permissivos à infecção e proliferação de *Leishmania*.
- Macrófagos M1, apesar de seu potencial leishmanicida, também podem contribuir com o desenvolvimento das lesões ao induzirem uma resposta inflamatória exacerbada que provoca danos ao tecido.
- Em modelos experimentais de suscetibilidade (BALB/c) e resistência parcial (C57BL/6) à infecção por *Leishmania amazonensis* não há um claro predomínio de macrófagos M1 ou M2 no local da infecção.
- A pior evolução das lesões parece estar envolvida com o recrutamento de macrófagos ricos em arginase-1 em camundongos suscetíveis.
- Um equilíbrio entre uma resposta pró-inflamatória e microbicida relacionada ao perfil M1 e uma resposta de reparo tecidual relacionada ao perfil M2 parecem fornecer o maior benefício para o hospedeiro.
- Esses resultados ampliam a compreensão sobre a imunidade protetora contra *L. amazonensis*, fornecendo novos conhecimentos que podem contribuir para o desenvolvimento de intervenções de tratamento ou prevenção da leishmaniose.

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6. ANEXOS

6.1. COMPROVANTE DE ACEITE

Izabel Galhardo Demarchi (Via FrontiersIn) <noreply@frontiersin.org>
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Para: Fernanda Tomiotto-Pellissier <fernandatomiotto@gmail.com>

1 de dezembro de 2020 18:24



Your Research Topic abstract was accepted

Dear Dr Tomiotto-Pellissier,

Many thanks for contributing to the Research Topic "**Cutaneous Leishmaniasis: Exploring Pathogenesis and Immunomodulatory Approaches**" with your abstract "**Mice susceptibility to Leishmania amazonensis is dependent on arginase-1 of macrophages**".

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6.2. OUTRAS PRODUÇÕES REALIZADAS DURANTE O DOUTORADO

6.2.1. Artigo 1

Inhibitory Activity of *Caryocar coriaceum* Wittm. Pulp and Peel Fruit Extract Against Promastigote and Amastigote forms of *Leishmania amazonensis*: *in-vitro* and *in-silico* approaches

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1 **Inhibitory Activity of *Caryocar coriaceum* Wittm. Pulp and Peel Fruit Extract Against**
2 **Promastigote and Amastigote forms of *Leishmania amazonensis*: an *in vitro* and *in silico***
3 **approach**

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27 **ABSTRACT**

28 Leishmaniasis is a group of neglected diseases caused by parasites of *Leishmania* genus.
29 The treatment of Leishmaniasis represents a great challenge, because the available drugs
30 present high toxicity and none of them has been shown to be fully effective. *Caryocar* is
31 a botanical genus rich in phenolic compounds, which leaves extracts have already been
32 described by its antileishmanial action. Thus, we investigated the effect of pulp and peel
33 extracts of the *Caryocar coriaceum* fruit on promastigote and amastigote forms of
34 *Leishmania amazonensis*. Both extracts had antipromastigote effect after 24, 48 and 72 h
35 and this effect was by apoptosis-like process induction, with reactive oxygen species
36 (ROS) production, damage to the mitochondria and plasma membrane, and
37 phosphatidylserine exposure. Knowing that the fruit extracts did not alter the viability of
38 macrophages, we observed that the treatment reduced the infection of these cells.
39 Thereafter, in the *in vitro* infection context, the extracts showed antioxidant proprieties,
40 by reducing NO, ROS and MDA levels. In addition, both peel and pulp extracts up-
41 regulated Nrf2/HO-1/Ferritin expression and increase the total iron bound in infected
42 macrophages, which culminates in a depletion of available iron for *L. amazonensis*
43 replication. *In silico*, the molecular modeling experiments showed that the three
44 flavonoids presented in the *C. coriaceum* extracts can act as synergistic inhibitors of
45 *Leishmania* proteins, and compete for the active site. Also, there is a preference for rutin
46 at the active site due to its greater interaction binding strength.

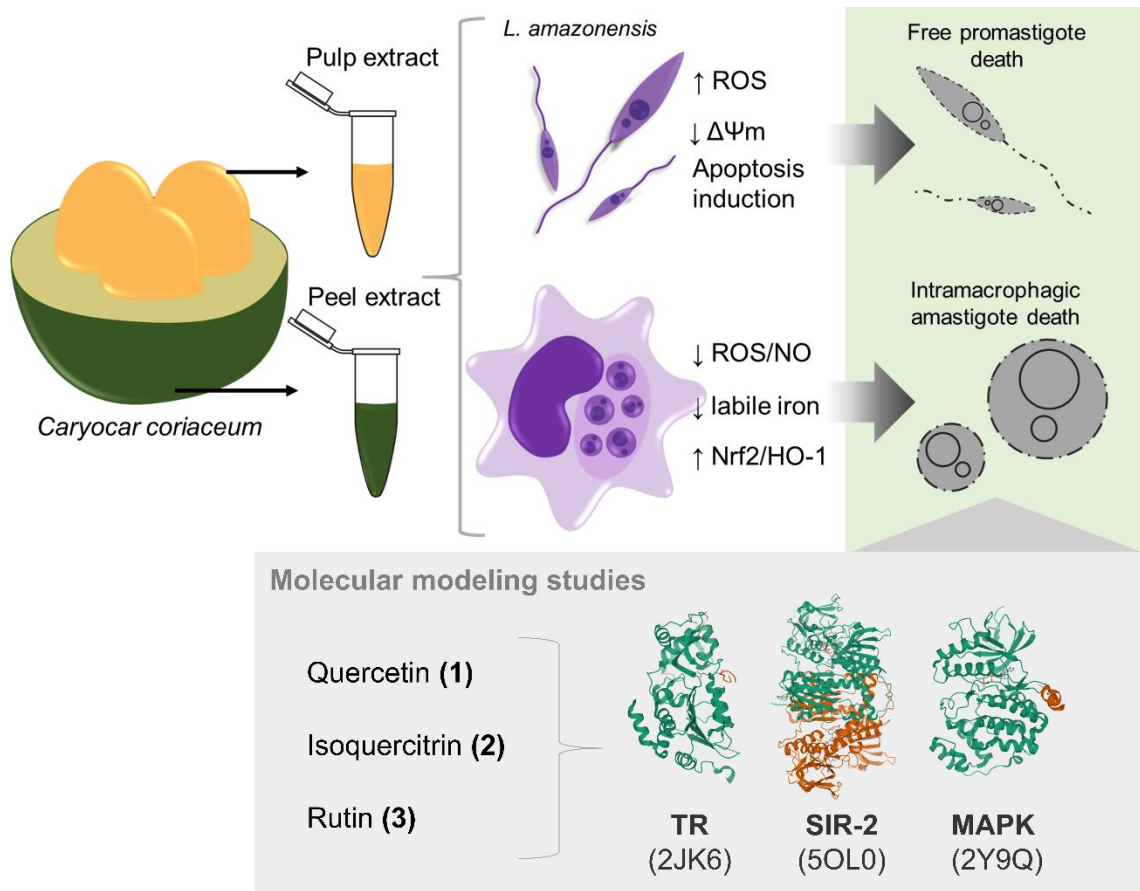
47

48 **Keywords:** Ferritin; Heme oxygenase-1; Leishmaniasis; Nrf2; Oxidative stress; Nitric
49 oxide; Docking molecular.

50

51

GRAPHICAL ABSTRACT



54 1 INTRODUCTION

55 Leishmaniasis is a group of diseases caused by parasitic protozoa of the
56 *Leishmania* genus transmitted through the bite of phlebotomine insects. The World
57 Health Organization (WHO) categorizes it as a neglected tropical disease since 350
58 million people in 102 countries worldwide are at risk of developing one of the disease
59 clinical forms (WHO, 2018). The macrophages are the main host cells to *Leishmania*
60 parasites whose machinery eliminates the parasites. However, the parasite can evade the
61 macrophage responses, persist on the host, and develop the disease (reviewed in: Rossi
62 and Fasel, 2018). Additionally, amastigote forms can survive inside macrophages
63 depending on the host's labile iron uptake regarding its metabolism, virulence, and
64 multiplication [3,4].

65 Nuclear factor transcription factor (erythroid-derived-2)-like 2 (NRF2) finely
66 controls iron metabolism in macrophages and acts on genes involved in heme synthesis,
67 hemoglobin catabolism, iron storage, and exportation [5]. Also, the NRF2/ heme
68 oxygenase-1 (HO-1)/ ferritin signaling cascade is important to *Leishmania amazonensis*
69 proliferation controlling by culminating in iron storage and, consequently, unavailability
70 of parasite uptake (Cataneo et al., 2019; Miranda-Sapla et al., 2019; Tomiotto-Pellissier
71 et al., 2018).

72 Leishmaniasis treatment represents a great challenge considering that the
73 available drugs have high toxicity and are yet to prove fully effective. Relapses,
74 therapeutic failure, and resistance to treatment are factors that motivate the search for
75 drugs with leishmanicidal actions that are more effective and less toxic to the patient [9].

76 *Caryocar* is a botanical genus belonging to the Caryocaraceae family distributed
77 through South and Central America in the vegetation type of Cerrado. The use of the
78 fruits encompasses the production of food, cosmetics as well as the scope of folk

79 medicine. Rich in phenolic compounds, these extracts have been described as potent
80 antioxidants, in addition to having anti-inflammatory, antineoplastic, and antimicrobial
81 effects [10–13]. The description of these extracts also appeared regarding its activity
82 against *Leishmania amazonensis* (Alves et al., 2017; Paula-Ju et al., 2006; Tomiotto-
83 Pellissier et al., 2018).

84 Previous studies demonstrated that leaf extracts of *Caryocar coriaceum* Wittm,
85 rich in flavonoids, have leishmanicidal activity (Tomiotto-Pellissier et al., 2018), but the
86 possible action mechanisms of fruit extracts on extra or intracellular parasite was not
87 evaluated.

88 Given this context, this study aimed at investigating the leishmanicidal effect of
89 *C. coriaceum*. fruit pulp and peel extracts on promastigote and amastigote *Leishmania*
90 (*Leishmania*) *amazonensis* forms, including an *in silico* experiment investigating the
91 main constituents of plant extracts quercetin, isoquercitrin and rutin.

92

93 **2 MATERIALS AND METHODS**

94 **2.1 Pulp and peel extracts of *Caryocar coriaceum* Wittm.**

95 The extracts of *C. coriaceum* Wittm. were kindly supplied by Dr. Selene Maia de
96 Morais of the State University of Ceará and obtained as previously described [15].
97 Briefly, peel and pulp of *C. coriaceum* Wittm. mature fruits were obtained at the Campus
98 of the State University of Ceará (UECE) (lat.: -3.792222; long.: -38.556111), Fortaleza,
99 Brazil, and were identified by Prisco Bezerra Herbarium under the code EAC57060. The
100 extracts were obtained by cold maceration with 96% ethanol, at 12 h cycle of light for 7
101 days. Filtration of the supernatant and evaporation of the solvent at reduced pressure in a
102 rotary evaporator led to crude ethanol extracts of *C. coriaceum*. fruit pulp and fruit peel.

103

104 **2.2 Extracts characterization by HPLC**

105 The identification and quantification of flavonoids on peel and pulp were
106 performed by high-performance liquid chromatography (HPLC) with SHIMADZU liquid
107 chromatography coupled to a SCL-10AVP controller system, UV-VIS detector SPD-
108 10AVP, and isocratic pump LC-10ATVP. The software LC Solution software was used
109 to record the chromatograms and measure peak areas. The column used was a Shimadzu
110 analytical CLC-ODS M (C-18) of 25 cm.

111 The calibration curve was constructed using the standards rutin, isoquercitrin, and
112 quercetin injected at different concentrations (0.25; 0.05; 0.025 and 0.005 mg.mL⁻¹) into
113 the liquid chromatography. The flow rate was 1.8 mL per minute for quercetin and 1.25
114 ml per minute to rutin both at a wavelength of 350 nm and a mobile phase of 20%
115 acetonitrile and 80% solution of H₃PO₄ pH buffer 2.8. The linear regression equation was
116 obtained by using the Microsoft Office Excel 2010 program.

117 The chromatographic profile of flavonoids rutin, quercetin, and isoquercitrin, the
118 chosen standards, was obtained preparing ethanolic solutions at a concentration of 0.5
119 mg.mL⁻¹. As a mobile phase, the same solution was used for the calibration curve at the
120 same wavelength and flow rate of 1.80 mL per minute.

121

122 **2.3 *Leishmania (Leishmania) amazonensis* maintenance**

123 *L. (L.) amazonensis* (MHOM/BR/1989/166MJO) promastigotes forms of were
124 maintained in culture medium 199 (GIBCO) pH 7.18-7.22 supplemented with 10% fetal
125 bovine serum (FBS) (GIBCO), 10 mM-HEPES buffer, 0.1% human urine, 0.1% L-
126 glutamine, 10U/mL-penicillin and 10µg/mL-streptomycin (GIBCO) and 10% sodium
127 bicarbonate. The cell culture was maintained at 25°C in a 25 cm² culture flask. All
128 experiments used promastigote forms at the stationary growth phase.

129

130 2.4 Antipromastigote assay

131 *L. amazonensis* promastigote forms (10^6 cells/mL) were treated with *C.*
132 *coriaceum*. extracts 25, 50 and 100 $\mu\text{g/mL}$. Parasites were counted on a Neubauer
133 chamber after 24, 48, and 72h of treatment. *L. amazonensis* promastigote maintained in
134 the culture medium was used as a control, DMSO 0.01% was used as a vehicle, and
135 amphotericin B (AMB) $1\mu\text{M}$ was used as a positive control.

136

137 2.5 Scanning electron microscopy of promastigote forms

138 Scanning electron microscopy of promastigotes forms was performed according
139 to [16]. Briefly, the parasites (10^6) were treated with 50 $\mu\text{g/mL}$ of pulp and peel extracts
140 for 24 h, processed, and analyzed through scanning electron microscopy (FEI QUANTA
141 200 scanning electron microscope). The length of the promastigote forms was analyzed
142 in the Image-Pro Plus software, based on the 8000 - 10500x magnification images
143 obtained. At least three images of each condition were subjected to measurement.

144

145 2.6 Mechanism of action determination on promastigote forms

146 To determine the mechanism of action of the *C. coriaceum*. extracts on
147 promastigote forms, the parasites (10^6 cells/mL) were treated for 24h with 50 $\mu\text{g/mL}$ of
148 the extracts, and different parameters were accessed as previously described by our group
149 (Tomiotto-Pellissier et al., 2018). Briefly, the inner mitochondrial membrane potential
150 was investigated by incubation with tetramethylrhodamine ethyl ester (TMRE) (Sigma);
151 phospholipids (PS) exposure was detected by Annexin-V FITC (Invitrogen); the cellular
152 membrane integrity was evaluated by propidium iodide (PI) (Sigma); and the reactive
153 oxygen species (ROS) were analyzed by a permeant probe diacetate 2',7'-

154 dichlorofluorescein (H₂DCFDA) (Sigma). All the analyses were performed on a
155 fluorescence microplate reader (Victor X3, PerkinElmer).

156

157 **2.7 Co-determination of annexin V and propidium iodide label**

158 Promastigotes (10⁶ cells/mL) were washed and resuspended in 100 μ L of assay
159 buffer 1x (Santa Cruz Biotechnology), followed by the addition of a mix containing 1 μ L
160 of annexin-V FITC and 5 μ L of PI (Santa Cruz Biotechnology). A total of 10,000 events
161 were acquired in a BD Accuri™ C6 flow cytometer. The labeling was analyzed as
162 previously described (Doroodgar et al., 2016; Tomiotto-Pellissier et al., 2018).

163

164 **2.8 Ethics Committee**

165 BALB/c mice were kindly provided by the Carlos Chagas Institute (ICC)/Fiocruz,
166 Curitiba, Brazil. The animals (25-30 g and 6-8 weeks) were kept under sterile conditions
167 and used according to protocols approved by the Institutional Animal Care and
168 Committee. This study was approved by the Ethics Committee for Animal
169 Experimentation of the State University of Londrina (13134.2016.62).170.

170

171 **2.9 Viability of peritoneal exudate cells**

172 The cytotoxic effects of *C. coriaceum* extracts on peritoneal exudate cells (PEC)
173 were tested based on mitochondrial oxidation of MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-
174 diphenyltetrazolium bromide) (Sigma) assay described by [16,18]. Briefly, 5x10⁵
175 cells/mL were recovered from the peritoneal cavity of BALB/c and the adherent cells
176 were incubated with 25, 50, and 100 μ g/mL of pulp or peel extract and cultured for 24 h.
177 After this period, MTT (5 mg/mL) was added for 3 h and the plates were read in a
178 spectrophotometer (Thermo Scientific, Multiskan GO) at 550 nm.

179

180 2.10 Anti-amastigote assay

181 PEC (5×10^5 cells/mL) were obtained as previously described. The adherent cells
182 were infected with *L. amazonensis* promastigotes of (1×10^6 cells/mL) for 2h. After, the
183 non-internalized parasites were washed with PBS and the infected cells treated with the
184 extracts (25, 50 or 100 $\mu\text{g/mL}$), RPMI 1640 medium (control), DMSO 0.01% (vehicle)
185 or AMB 1 μM (positive control) for 24h (37°C, 5% CO₂). After this, the cells were stained
186 with Giemsa (Laborclin) and 20 fields analyzed through an optical microscope (Olympus
187 BX41, Olympus Optical Co).

188

189 2.11 Promastigote recovery test

190 Promastigotes recovery assay was performed as previously described by
191 (Tomiotto-Pellissier et al., 2018). In brief, adherent PEC were infected with *L.*
192 *amazonensis* and treated with concentrations of *C. coriaceum* extracts. After 24 h of
193 treatment, it was induced the differentiation of intracellular viable amastigotes into
194 promastigote free forms. Free promastigotes recovered (FPR) were counted on a
195 Neubauer chamber for three consecutive days.

196

197 2.12 Malondialdehyde (MDA) Levels Measurement

198 MDA levels were determined through a High-Performance Chromatography
199 (HPLC) according to Tomiotto-Pellissier et al., 2018. The analyses were conducted using
200 an Alliance e2695 HPLC (Waters) equipped with a SecurityGuard ODS-C18 (4×3.0
201 mm, Phenomenex), C18 reverse-phase column (Eclipse XDBC18; 4.6×250 mm, 5 μm ,
202 Agilent) as well as a photodiode array detector (Photodiode ArrayDetector (PDA), 2998)
203 using Empower 2 software (Waters).

204

205 2.13 Determination of iron concentration and total bound iron

206 The determination of iron concentration in supernatants of the anti-amastigote
207 assay was performed utilizing the method adapted by our group [8]. Briefly, in pH 4.5
208 and in the presence of ascorbic acid, it occurs the release of iron bound to transferrin. The
209 resulting product (Fe^{2+}) forms a complex measured using a biochromatic endpoint
210 technique (600, 700nm). The test principle for the total bound iron is similar: all available
211 sites of iron-binding in transferrin were saturated. In pH 8.6, only saturated iron in excess,
212 unbound, is available to be reduced to ferric iron by ascorbic acid forming a complex.
213 The subsequent addition of acid (pH 4.5) releases the iron bound to transferrin, this
214 supplemental iron is reduced to ferric iron by ascorbic acid, forming an increased amount
215 of complex. The increase in absorbance during the change of pH 8.6 to pH 4.5 is
216 proportional to the concentration of iron-bound.

217

218 2.14 Relative quantification of Nrf2, ferritin and HO-1 mRNA

219 The mRNA expression levels quantification for nuclear factor erythroid 2-related
220 factor 2 (Nrf2), ferritin, and heme-oxygenase 1 (HO-1) genes were performed following
221 Tomiotto-Pellissier et al. (2018) using the Total RNA Isolation System kit (Promega)
222 following the manufacturer's procedure.

223

224 2.15 Statistical analysis

225 Data were expressed as a mean \pm SEM. At least three independent experiments
226 were performed, each with duplicate datasets. Data were analyzed using the GraphPad
227 Prism statistical software (GraphPad Software, Inc., USA, 500.288). Significant
228 differences between the groups were determined through one-way ANOVA, followed by

229 Tukey's test for multiple comparisons. Differences were considered statistically
230 significant upon $p \leq 0.05$.

231

232 **2.16 *In silico* antileishmanial evaluation**

233 2.16.1 Ligands preparation

234 The chosen ligands were: quercetin (1), isoquercitrin (2) and rutin (3). The control
235 molecules were phosphoaminophosphonic acid-adenylate ester (4), L-peptide linking
236 cellular tumor antigen p53 (5), flavin-adenine dinucleotide sulfate (6) and amphotericin
237 b (7). Their respective two-dimensional structures were obtained from the virtual
238 repository PubChem[®] (<https://pubchem.ncbi.nlm.nih.gov/>). In order to use molecules in
239 their best potential energy state for the simulation of molecular docking, they were
240 optimized by the semi-empirical parametric method 7 (PM7) of molecular mechanics
241 [19–21] with the MOPAC[®] software and converted to ligand.mol2.

242

243 2.16.2 Target protein preparation

244 Human ERK2 complexed with a Mitogen-Activated Protein Kinase docking
245 peptide (MAPK) (PDB: 2Y9Q) [22], Silent Information Regulator 2 related protein 1
246 (SIR) (PDB: 5OL0) [23] and Trypanothione Reductase (TR) (PDB: 2JK6) [24] biological
247 targets of *Leishmania* sp. and was reported from the RCSB[®] protein data bank
248 (<https://www.rcsb.org/>) in protein format .pdb. The structures were filed in the Protein
249 Data Bank, determined from X-ray diffraction, with a resolution of 1.55Å, 1.99Å and
250 2.95Å, respectively. Furthermore the file was loaded into UCSF Chimera[®] software [25]
251 for all residues removal and polar hydrogens additions for producing favorable
252 protonation states [26,27] and preparing the target for submission to the molecular
253 docking test.

254 2.16.3 Molecular docking study

255 After the ligand and protein preparation stage, the molecules were finally
256 subjected to molecular docking simulation by the SwissDock server
257 (<http://www.swissdock.ch/docking#>) [28,29] from the Molecular Modeling Group from
258 Swiss Institute of Bioinformatics (SIB) (<https://www.sib.swiss/>). The .zip files received
259 from the online server were downloaded and processed in the UCSF Chimera[®] software,
260 where the curcumins interaction and distance analyzes were performed, in comparison
261 with the catalytic sites occupied by the control ligands (4)-(6). 4 is the inhibitor described
262 from the characterization of MAPK (PDB: 2Y9Q) [22], 5 is the SIR (PDB: 5OL0)
263 inhibitor [23] and 6 is TR (PDB: 2JK6) inhibitor described [24], (7) is a drug considered
264 as positive controls described in the literature [30–32]. The detailed analysis of the
265 intermolecular interactions between the tested compounds and the residues of the MAPK,
266 SIR and TR catalytic sites was done by obtaining the theoretical values of H-bond donors
267 and acceptors of the ligands and their respective acid dissociation constants (pKa),
268 calculated in the software MarvinSketch[®] [33], where it was possible to predict the micro-
269 species formed from the ligands under physiological actions (approximately pH 7.4)
270 [34,35]. The data obtained from the analysis were plotted on the Morpheus[®] online
271 statistical tool (<https://software.broadinstitute.org/morpheus/>), where heat maps were
272 generated to identify the ligand-residue interaction and similarity profiles by the statistical
273 test Pearson. The types of linker-residue interactions, as well as the figures, were
274 generated using the Discovery Studio[®] [36].

275

276 **3 RESULTS**

277 **3.1 *Caryocar coriaceum* Wittm. composition**

278 The chromatographic analysis of the *C. coriaceum* fruit extracts showed that both
279 extracts, pulp, and peel, presented the flavonoids rutin and isoquercitrin. However, only
280 the pulp extract showed a small peak corresponding to quercetin (Figure 1).

281

282 **3.2 Peel and pulp extracts exert an anti-promastigote effect on *L. amazonensis***

283 We assessed the antileishmanial effect of pulp and peel extracts by determining
284 the number of viable parasites. All tested concentrations of the extracts proved a
285 significant reduction of *L. amazonensis* proliferation from 24h ($p \leq 0.01$), 48h ($p \leq 0.001$),
286 and 72h ($p \leq 0.001$) when compared to both the control and vehicle groups (Figure 2A).

287 Figure 2B shows the following difference between the concentrations: 100 $\mu\text{g/mL}$
288 dose was more effective at reducing *L. amazonensis* proliferation than 25 $\mu\text{g/mL}$ in 24
289 and 48h ($p \leq 0.05$ and $p \leq 0.01$, respectively) for pulp extract, and in 24, 48 and 72h for peel
290 extract ($p \leq 0.05$, $p \leq 0.01$ and $p \leq 0.01$, respectively). After 48h of pulp extract treatment,
291 100 $\mu\text{g/mL}$ had different results than 25 and 50 $\mu\text{g/mL}$ ($p \leq 0.01$); after 72h, no difference
292 appeared between the concentrations of this extract. In addition, the effect proved not
293 time-dependent for both extracts ($p > 0.05$). Considering the similar results, we conducted
294 further experiments by investigating the effect of treatments on promastigote forms using
295 the intermediate concentration (50 $\mu\text{g/mL}$) at 24h.

296

297 **3.3 Apoptosis-like mechanism induced by *C. coriaceum* extracts**

298 Upon the verification of the leishmanicidal action of the extracts, we sought to
299 elucidate the death mechanisms induced by these treatments and found that pulp and peel
300 extracts enhanced the ROS levels ($p \leq 0.001$), reduced the mitochondrial membrane
301 potential ($\Delta\Psi\text{m}$) ($p \leq 0.0001$), induced PS exposition ($p \leq 0.0001$), and damaged the plasma

302 membrane ($p \leq 0.001$) (Figure 3A-D). Enhanced ROS levels, induced PS exposition and
303 PI labeling proved higher in peel-treated than in pulp-treated parasites ($p \leq 0.0001$).

304 We performed an annexin V/ PI co-staining in treated promastigote forms to
305 differentiate the cell death mechanisms. The annexin V+ parasites showed an increase of
306 7.34 and 10.6% after treatment using pulp and peel, respectively, concerning the control,
307 while the PI+ cells enhanced in 7.8% for pulp and 1.48% for peel extract. Lastly, the
308 annexin V+/PI+ promastigotes proved 28.4 and 24.9% higher in the pulp and peel treated
309 parasites, respectively, when compared to the control, indicating that most of the treated
310 promastigote forms underwent late apoptosis-like death in both conditions (Figure 3F and
311 G).

312 Moreover, we found that extracts acted in the morphology of promastigote forms
313 since the extracts promoted a significant reduction on cell body ($p \leq 0.05$), flagellum
314 ($p \leq 0.05$), and total length ($p \leq 0.001$) (Figure 4A). Besides that, pulp extract induced
315 blebbing (apoptotic bodies) formation on the parasite membrane (Figure 4C), and both
316 extracts acted on the rounding, shrinking, and double flagella formation on *Leishmania*
317 (Figure 4D-F) after 24h of treatment using 50 $\mu\text{g/mL}$.

318

319 **3.4 *C. coriaceum* extracts act in intracellular amastigote forms without** 320 **cytotoxicity to host cells**

321 Considering the effects of *C. coriaceum*. pulp and peel extracts on promastigote
322 forms, we performed experiments to verify their possible action on intracellular
323 amastigotes. Initially, we found that the tested concentrations of both extracts (25 to 100
324 $\mu\text{g/mL}$) did not interfere with the viability of host cells (macrophages) (Figure 5A)
325 ($p \geq 0.99$), but they were subsequently able to significantly reduce the percentage of

326 infected macrophages (Figure 5B) and amastigote forms by macrophage (Figure 5C)
327 when compared to the control ($p \leq 0.05$) – not different between them or AMB.

328 To confirm the infection reduction, we performed a recovery assay of
329 promastigotes forms by subjecting the culture of amastigote-infected macrophages to
330 ideal conditions to differentiate viable amastigotes in free promastigote forms. All tested
331 concentrations reduced the percentage of recovery promastigotes forms after 48 and 72h
332 of culture (Figure 5D and E) ($p \leq 0.05$). After 48h, none of the treatments differed from
333 AMB and neither did the concentrations of 50 and 100 μ M of pulp and 100 μ M of peel
334 extract after 72h. Considering the similar effect of the different concentrations, we applied
335 the lowest concentration to carry out the following experiments.

336

337 **3.5 Pulp and peel *C. coriaceum* extracts act as an antioxidant on *L. amazonensis*-** 338 **infected macrophages**

339 Seeking to understand the mechanisms involved in the elimination of intracellular
340 parasites, we analyzed the production of reactive species which are important to the
341 response against *Leishmania* parasites. Surprisingly, our results demonstrated that
342 infected cells treated with *C. coriaceum*. pulp and peel extracts (25 μ g/mL) showed lower
343 NO and ROS production. The treatment using extracts acted as antioxidants, which is
344 reinforced with lower lipid peroxidation indicated when measuring the lipid peroxidation
345 metabolite MDA (Figure 6A, B, and C, respectively) in infected macrophages when
346 compared to the control ($p \leq 0.05$).

347

348 **3.6 *C. coriaceum* pulp and peel extracts modulate iron pool of infected** 349 **macrophages**

350 Still aiming at understanding the parasites death pathway, we investigated the
351 levels of iron and total binding iron capacity and found data indicating that the treatment
352 using pulp and peel extracts did not alter the total iron concentration in *L. amazonensis*-
353 infected macrophages (Figure 7A) ($p \geq 0.2$), but increased iron-bound (Figure 7B) when
354 compared to the control ($p \leq 0.001$) and decreased iron labile bioavailability. We found a
355 sharper increase in total iron-bound capacity for the peel extract treatment ($p \leq 0.001$).

356

357 **3.7 *C. coriaceum* fruit extracts induce Nrf2, HO-1 and Ferritin expression**

358 Considering that the treatments using the extracts induced an antioxidant action
359 and concomitant higher iron-bound, we investigated the cascade involving the
360 transcription factor Nrf2, HO-1 enzyme, and protein ferritin (Figure 8). Our results
361 showed that both pulp and peel extracts were able to up-regulate the expression of the
362 three genes investigated (Nrf2, HO-1, and ferritin) when compared to the control
363 ($p \leq 0.01$). Also, we found that the pulp extract caused more pronounced effects than the
364 peel extract in the expression analysis of HO-1 and Nrf2 ($p \leq 0.01$).

365

366 **3.8 Molecular docking**

367 The energy binding/affinity energy (ΔG) of the compounds (1), (2) and (3) were
368 strongest (lower than -6 kcal/mol) than the controls (4), (5) and (6) as it has an affinity
369 energy greater than -5 kcal/mol (Figure 9). Highlighting the bests results with all proteins
370 evaluated for the (3) compound, rutin, with high abundance in all extracts here proposed.
371 The (7) compound did it did not bind with the proteins evaluated here ($\Delta G > -3$), indicating
372 that its mechanism of action potentially acts on other pathways. The ligands proposed

373 here do not connect at the proposed active sites [22–24], but perform high molecular
374 strength interactions with their respective amino acids, as we will see below.

375 The ligand (4), MAPK's proposed inhibitor [22], strongly connects with 10 amino
376 acids (AA) by conventional hydrogen bonds with binding distances from 2.58 to 3.29 Å,
377 the strongest bonds are with ALA35, ASP106, ASP111, GLN105, GLY37, LYS114 and
378 LYS54. However, this compound has bonds with average interaction forces at G34
379 (carbon hydrogen bond; 3.69 Å), ARG67 and LYS151 (electrostatic salt bridges; 2.85 to
380 3.44 Å), and weak Pi hydrophobic bonds (4.15 to 5.44 Å) with ALA52, ILE31, LEU156
381 and VAL39.

382 As well, (5) compound, SIR's proposed inhibitor [23], has higher electrostatic salt
383 bridge binding (2.74 Å) with ASP196, strong conventional hydrogen bonds with 10 AA
384 distances up from 2.58 to 3.29 Å, the most important ones are ALA35, ASP106, ASP111,
385 GLY37, LYS54, SER153 and TYR36. Also, this compound poorly connects GLY34 AA
386 with carbon hydrogen bond (3.69 Å) and pi-alkyl bonds with ALA52, ILE31, LEU156
387 and VAL39 (3.87 to 5.44 Å).

388 The (6) compound, TR's proposed inhibitor [24], performs strong conventional
389 hydrogen bonds with 15 AA, most importantly at ALA47, ASP327, ASP35, GLY127,
390 GLY286, LYS60, THR39, THR51 and TYR221 of TR receptor distances up from 2.56
391 to 3.01 Å. Also, this ligand achieves average carbon hydrogen bonds with the residues of
392 ALA159, GLY11, GLY13, GLY161, GLY50, MET333, PRO336 and THR160 of TR
393 receptor from 3.11 to 3.73 Å. With PHE203, TYR198 and VAL36 fulfill hydrophobic pi-
394 alkyl interactions and with CYS57 pi-sulfur binding. Yet, amide-pi stacked connections
395 with GLY56 were performed.

396 Regarding the behavior of the binding energies of the compounds (1), (2) and (3)
397 evaluated in this work, despite all of it demonstrated better binding energy than all the

398 controls, its binding with AA of each target were also better than the controls, as we can
399 demonstrate at supplementary figures, in which the better compound (3) were
400 demonstrated.

401 Regarding the interaction between the ligands and the MAPK protein (Figure S1),
402 each interest compound occupies a different site, as seen. The better binding compound
403 was (3), with $\Delta G = -7.682$ kcal/mol, followed by (2) and (1) with -6.925 and -6.352 ,
404 respectively. For (3) the better bindings distances (lower values) ligand-residues were
405 ASP251, ASP291, PHE296 and SER246 with 1.88 to 2.84 Å, with conventional hydrogen
406 bonds; followed by carbon hydrogen bonds with LEU244 (2.54 Å), LYS272 (2.65 Å) and
407 LYS292 (2.72 Å). The weak bindings was with LYS300, pi-alkyl binding, with 3.57 Å.
408 The (2) compound also performs strong conventional hydrogen bonds with ARG91,
409 ASP20, GLN355 and GLY22 at distances from 2.05 to 3.01 Å. Also performing weak pi-
410 alkyl binding with ILE90, PRO23 and PRO356 at distances from 3.02 to 4.86 Å. Strong
411 conventional hydrogen bonds also are presents with interactions (1) compound and
412 MAPK residues with PHE329 at 2.75 Å and electrostatic pi-cation bond with LYS330 at
413 2.93 Å. Weak pi-alkyl binding were find with ALA325, ALA327 and ARG77 at distances
414 from 4.12 to 5.10 Å.

415 The interaction between the ligands and the SIR protein (Figure S2), each interest
416 compound also occupies a different site, as seen. Again, the better binding compound was
417 (3), with $\Delta G = -9.393$ kcal/mol, followed by (2) and (1) with -8.038 and -8.608 ,
418 respectively. For (3) the strongest bindings were conventional hydrogen bonds with
419 GLU243, ILE48, GLN124 and HIS144 with 1.96 to 3.03 Å. The weak bindings was with
420 ILE126, ALA40, PHE74, ASP50 and HIS204 acting as pi-pi, alkyl or pi-alkyl binding,
421 with 3.20 to 5.30 Å distances. Strong conventional hydrogen bonds also are presents with
422 interactions (2) compound and SIR residues with ASN186, ASN193, HIS156 and

423 LYS184 at 1.88 to 3.30 Å distances and carbon hydrogen bond with GLU192 at 2.32 Å.
424 Weak pi-alkyl binding were find with PRO195 and ALA197 at distances from 4.59 and
425 5.36 Å. The (1) compound also performs strong conventional hydrogen bonds with
426 ASP127 and PHE100 at distances from 1.96 and 2.33 Å. Also performing weak pi-sigma
427 and pi-alkyl binding with ALA132 and PRO97 at distances from 3.51 and 4.77 Å,
428 respectively.

429 After molecular docking simulation, it was found that all ligands (1) quercetin,
430 (2) isoquercitrin and (3) rutin occupy the same catalytic site of interaction of TR (Figure
431 S3). The better bind energy, or affinity energy, was find with this target. Once again, the
432 better binding compound was (3), with $\Delta G = -10.216$ kcal/mol, followed by (2) and (1)
433 with -7.701 and -6.969 , respectively. The strongest bindings relations between (3) and
434 TR residues were conventional hydrogen bonds with ASP453, THR457, CYS444 and
435 CYS469 with 2.47 to 3.62 Å; followed by carbon hydrogen bond at SER440 with 3.63 Å
436 distance. The (2) compound also performs strong conventional hydrogen bonds with
437 CYS469 at distances from 2.31 Å; and carbon hydrogen bonds with PHE454 and THR457
438 at distances from 2.50 and 3.04 Å. Also performing average force pi binding with
439 CYS444 at distance 2.75 Å. Also, (1) ligand interacts with strong conventional hydrogen
440 bond at SIR residues with VAL460 at 2.60 Å distance and weak pi-alkyl binding with
441 ALA465 at 4.22 Å distance.

442 For the construction of the analysis by heatmap, statistical Pearson's analyzes
443 were carried out in order to be able to show the topology of the amino acids that suffered
444 interaction with each proposed ligand. As evidenced, for the MAPK and SIR targets, each
445 ligand fixed in a different protein cavity (Figure 10). As the ligands (2) and (3) are
446 modifications of the ligand (1), these data demonstrate a high degree of their interaction
447 with the entire protein surface, corroborating the results obtained *in vitro*.

448 Still in Figure 10, with respect to the data shown in C, it can be seen that the
449 behavior of the compounds has a slight modification by formation of clusters. Thus, they
450 remain in another area than that of the proposed control, but are clustered in a region.
451 Interestingly, in relation to the control sites and the ligands there is a set of preserved AA
452 residues that only become evident when the residues are compared with each other, in
453 light of Pearson's statistical similarity. This data leads us to infer that, for compounds with
454 a flavonoid structure, potentially the most important preserved binding site of AA are
455 CYS444, CYS469, PHE454 and THR457.

456 When comparing the ligands in terms of their similarity, the formation of clusters
457 of AA residues can be identified. It is important to mention that the grouped substances
458 can act competitively with the molecules of the same site (Figure 10C) and in a synergistic
459 way with molecules of different sites (Figure 10A, B).

460

461 **4 DISCUSSION**

462 Over the past few decades, limitations have emerged regarding the use of synthetic
463 drugs – high toxicity, side effects, high costs – generating great interest in the
464 ethnobotanical research [37]. In the case of leishmaniasis, the current therapy can cause
465 serious side effects such as hepatotoxicity, nephrotoxicity, cardiotoxicity [9], which
466 arouses interest in natural compounds targeting minor damage to the patient.

467 Recent studies conducted by our group as well as other researchers have shown
468 that extracts from leaves of *Caryocar* genus plants act on different microorganisms,
469 including *Leishmania amazonensis* (Alves et al., 2017; Paula-Ju et al., 2006; Tomiotto-
470 Pellissier et al., 2018). Our group also demonstrated an inhibitory effect of both the pulp
471 and peel *C. coriaceum* extracts on *Leishmania amazonensis* promastigote forms [15].

472 However, until now no information has been achieved on the possible action mechanisms
473 of these extracts on the extra and intracellular parasite.

474 The present study showed the action of *C. coriaceum*. fruit ethanol extracts
475 ranging from 25 to 100 $\mu\text{g/mL}$ on *L. amazonensis* promastigotes. These results
476 corroborate with our previous work, which defined IC_{50} of 30 (± 5) and 38 (± 13) $\mu\text{g/mL}$
477 for pulp and peel extracts, respectively [15]. Furthermore, we demonstrated the ability of
478 the extracts to increase ROS levels, molecules with different cellular functions which in
479 excess can trigger organelles damage and consecutively cellular death [38].

480 In this sense, it was also found mitochondrial membrane damage upon lower
481 $\Delta\Psi\text{m}$. Mitochondrial integrity is essential for parasite survival since trypanosomatids
482 have a single mitochondrion [38]. Considering that mitochondrial dysfunction and
483 reactive oxygen species production can lead to cell death by apoptosis [39], we
484 investigated such process by applying annexin V labeling. Similarly to the extracts
485 (Tomiotto-Pellissier et al., 2018), the effect of fruit extracts also increased both annexin
486 V and PI marking.

487 Aiming at differentiating the death type in necrotic, apoptotic, and late-apoptotic,
488 we performed a co-staining analyzed through flow cytometry. We verified that most of
489 the parasites were in the double-positive zone indicating the predominance of the late-
490 apoptotic process. This death type was previously described for *Leishmania* parasites
491 treated with different compounds [16,17]. The morphological changes found in the
492 parasite reinforce such results, since bleb formation, cell length reduction, and cellular
493 shrinkage are typical signs of apoptotic death [39].

494 We used the next experimental set to elucidate whether the fruit extracts would be
495 able to act on intracellular amastigote forms, which is more resistant and represent a
496 challenging target since the compound needs to diffuse through the host cell structures to

497 act on the parasite [40]. Firstly, it was found that the pulp and peel of *C. coriaceum*.
498 extracts were not toxic to the macrophages, the major host cell for *Leishmania* parasites.
499 Previous results showed a selectivity index for the parasite (macrophage/ promastigote
500 forms) of approximately 9 and 12 for pulp and peel, respectively [15]. Subsequently, it
501 was demonstrated the ability of the extracts to improve the elimination of intracellular
502 amastigotes, similarly to those found in *C. coriaceum*. leaf extracts (Tomiotto-Pellissier
503 et al., 2018). Finally, it was investigated the mechanisms involved in such elimination as
504 follows below.

505 The main effector of molecules in the intracellular pathogen elimination is known
506 to be reactive nitrogen species (RNS) and reactive oxygen species (ROS). In contrast,
507 RNS and ROS are also responsible for tissue damage in leishmaniasis [2]. Given the
508 importance of these molecules to the infection course, it was investigated their effects in
509 the studied model and the levels of ROS and RNS were lower upon treatment.

510 This corroborates the previous results regarding the antioxidant effect of the *C.*
511 *coriaceum*. fruit extracts using 2,2-diphenyl-1-picrylhydrazyl (DPPH) methodology [15].
512 Additionally, antioxidant capacity is intrinsically related to the presence of phenols and
513 flavonoids, such as isoquercitrin, quercetin, and rutin, present in the extracts studied,
514 contributing to radical scavenging potential [41].

515 We investigated the iron metabolism after discarding the hypothesis of oxidative
516 stress is responsible for eliminating the parasite. *Leishmania* parasites use the host's labile
517 iron pool and depend on this metal uptake for its replication and survival [3,4]. Thus, the
518 higher iron-bound caused by *C. coriaceum*. fruit extracts treatment made iron unavailable
519 to the parasite and decreased macrophage infection rate.

520 Aiming at corroborating such hypothesis, it was verified the cascade involving
521 Nrf2/HO-1/ferritin complexes for being directly related to both the antioxidant capacity

522 and iron metabolism [42]. While on the one hand, Nrf2 activation may be a weapon of *L.*
523 *amazonensis* to block oxidative stress and persist in the host [43], on the other hand, it
524 can induce transcription of downstream genes, such as HO-1, acting on iron metabolism
525 by inducing ferritin expression. Finally, ferritin plays an important role in free iron
526 capture reducing its availability to intracellular parasites [42,44]. Still, overall antioxidant
527 balance can be a powerful ally to protect against tissue injury caused by leishmaniasis
528 [45].

529 Methanolic and ethanolic extracts of *C. coriaceum*. leaves up-regulate Nrf2
530 transcription, HO-1, and ferritin genes, however, when assessing oxidative mediators,
531 fruit and peel extracts appeared to be more effective as they reduced MDA and ROS
532 levels more effectively, as well as generating lower NO formation, whereas leaf extracts
533 did not influence this mediator production (Tomiotto-Pellissier et al., 2018).

534 Notably, similar results were found with compounds trans-chalcone (intermediate
535 in the synthesis of flavonoids) [7] and quercetin [8]. Both had a direct effect on
536 promastigote forms of *L. amazonensis* inducing an apoptosis-like death pattern and action
537 on intracellular amastigote forms by inducing NRF2 and iron unavailability to parasite
538 metabolism. Therefore, these effects can be due at least partially, to the presence of rutin,
539 isoquercitrin, and quercetin present in *C. coriaceum* extracts.

540 All three flavonoids presented in the *C. coriaceum* extracts can act as synergistic
541 inhibitors of MAPK and SIR proteins, and these compounds also tend to compete for the
542 active TR site. MAPK proteins in host cells are related to an induction of pro-
543 inflammatory response [46], while in *Leishmania*, they are involved in the regulation of
544 flagella length, playing an important role in disease transmission [47]. Both of these
545 features were found in the *in vitro* experiments, reinforcing the importance of the
546 flavonoids of *C. coriaceum* extracts in acting on these proteins.

547 The proteins belonging to the SIR family have been implicated in the regulation
548 of a number of biological processes, such as gene silencing, DNA repair, cell cycle,
549 metabolism, and apoptosis. In *Leishmania* parasites, SIR has emerged as a promising drug
550 target, due to its participation in the parasite survival, proliferation, drug resistance and
551 infectivity [23,48–50].

552 TR is considered to be one of the best targets to find new anti-Leishmanial drugs.
553 This enzyme is the main detoxifying system against oxidative damage found in
554 trypanosomatids and thus is important for parasite's infectivity and survival, but absent
555 in the human host [24,51]. In our study, it was found a competitive interaction of
556 flavonoids for the active TR site, with a preference for rutin due to its greater interaction
557 binding strength. The results found in the *in silico* analysis corroborate the results found
558 in the *in vitro* experiments, as well as the metabolic pathway for the elimination of the
559 pathogen proposed.

560 Together, our data demonstrate for the first time that pulp and peel extracts from
561 the *C. coriaceum* fruit can induce death of *L. amazonensis* promastigotes through
562 apoptosis-like mechanisms and act on intracellular amastigote forms by activating the
563 Nrf2/HO-1/ferritin pathway, consecutively reducing iron availability for parasite
564 survival,. Furthermore, both treatments induced an antioxidant response, which is
565 important to protect against the tissue damage caused by leishmaniasis. The *in silico*
566 evaluations corroborate the data found *in vitro*, showing a predictive action of the
567 flavonoids present in the extract on essential *Leishmania* proteins.

568

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571

572 **6 CONFLICT OF INTERESTS**

573 The authors declared that there is no conflict of interest.

574

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578

579 **8 AUTHOR CONTRIBUTION**

580 FTP - study design, carrying out experiments on *Leishmania* parasites, data
581 analysis and scientific writing. DRA, SMM, ESM and MMM - collecting and identifying
582 plants, performing extractions, characterization of extracts and *in silico* evaluations.
583 BTSB and MDG - microscopy experiments and antioxidant tests. TFS - data analysis and
584 scientific writing. ERT and LMY - conducting, interpreting and analyzing molecular
585 biology experiments. INC and ICC - critical data analysis and review of scientific writing.
586 MMMS and WRP - experimental design, critical data analysis and review of scientific
587 writing.

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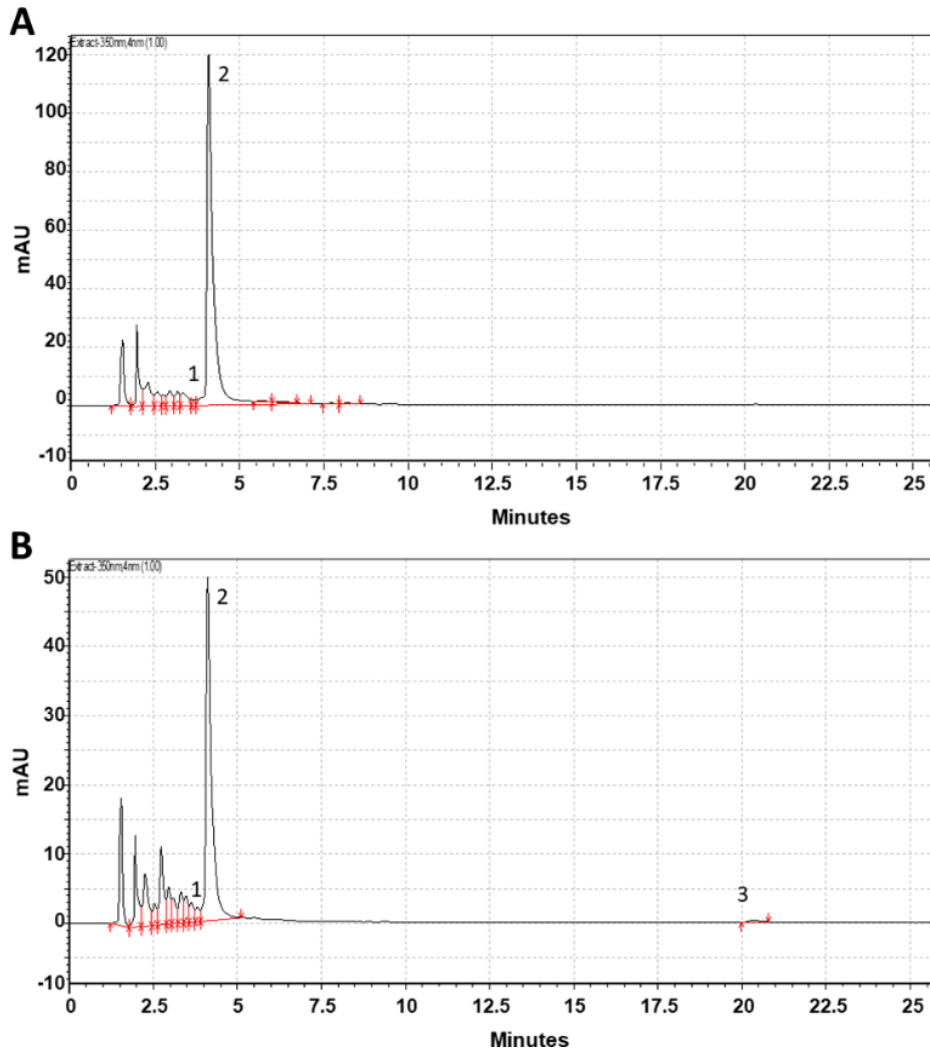
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FIGURE LIST

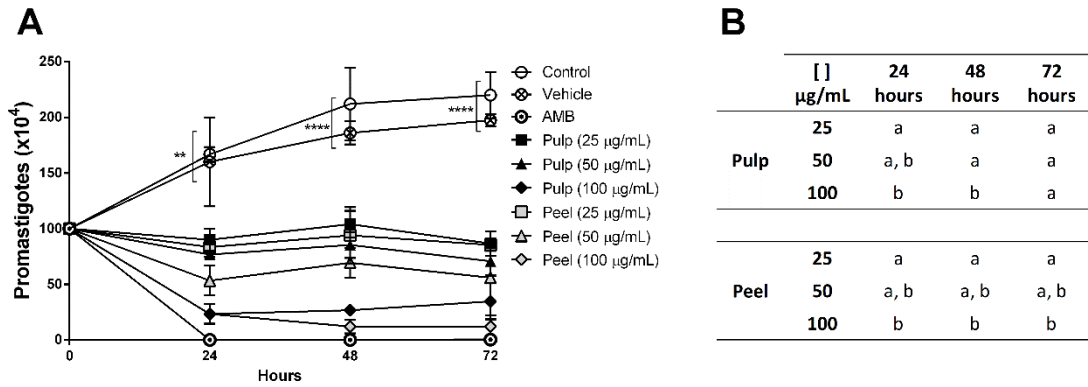


764

765 **Fig. 1 Characterization of *C. coriaceum* Wittm. fruit extracts.** The (A) pulp and (B)
 766 peel extracts were analysed by HPLC with Shimadzu analytical CLC-ODS M (C-18) of
 767 25 cm column. The calibration curve was constructed using the standards rutin (1),
 768 isoquercitrin (2) and quercetin (3) at 0.25; 0.05; 0.025 or 0.005mg/mL. The flow rate was
 769 1.8 mL per minute for quercetin and 1.25 ml per minute to rutin both in a wavelength of
 770 350 nm and a mobile phase of 20% acetonitrile and 80% solution of H₃PO₄ pH buffer
 771 2.8.

772

773



774

775 **Fig. 2 Antipromastigote effect of *C. coriaceum* Wittm. fruit extracts. *L. amazonensis***

776 promastigotes forms were subjected to *Caryocar coriaceum* Wittm. pulp or peel extracts

777 treatment (25–100 µg/mL). (A) The parasite viability was assessed at 0, 24 and 48h. As

778 control was used non-treated parasites, as vehicle control was used DMSO 0.02% and as

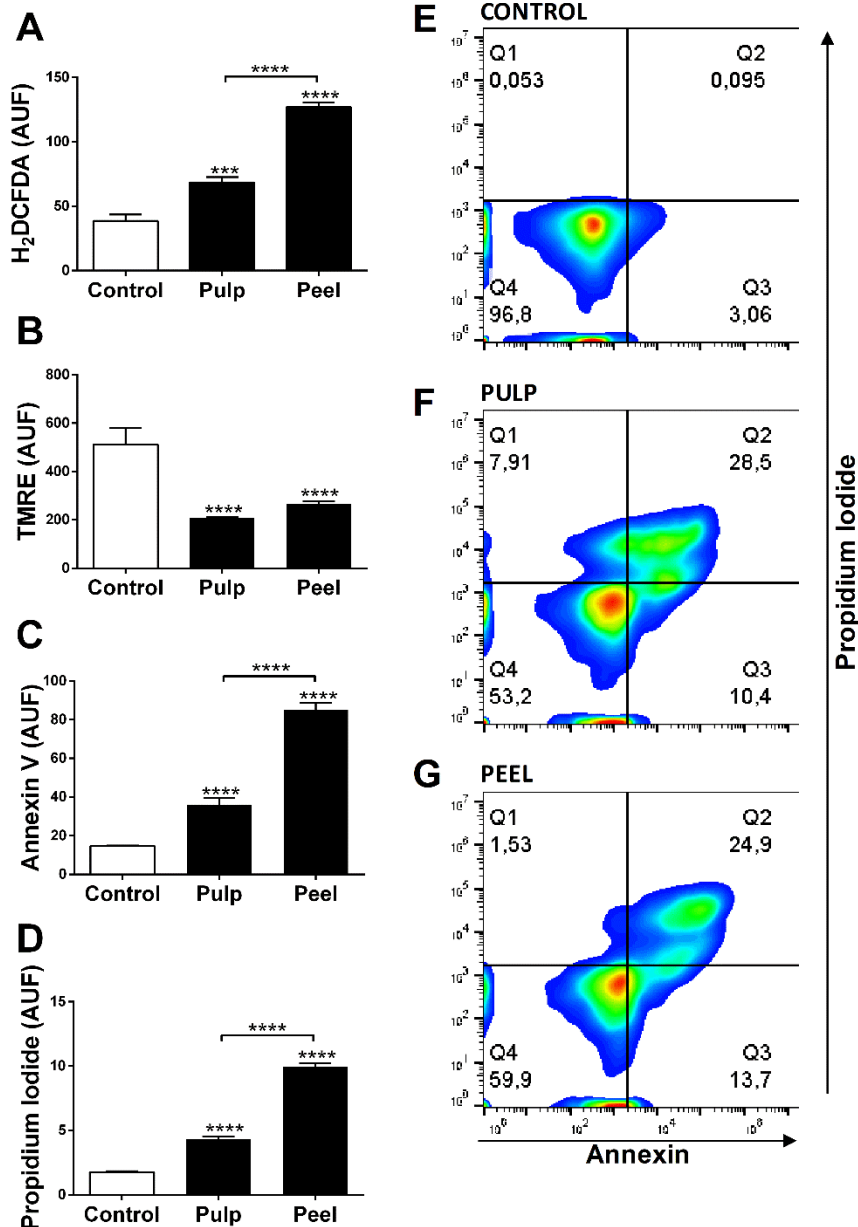
779 a positive control was used amphotericin B (AMB) 1 µM. (B) Detailed statistical analysis

780 between the treatments. Groups that not share the same letter are significantly different.

781 The values represent the mean ± SEM of three independent experiments performed in

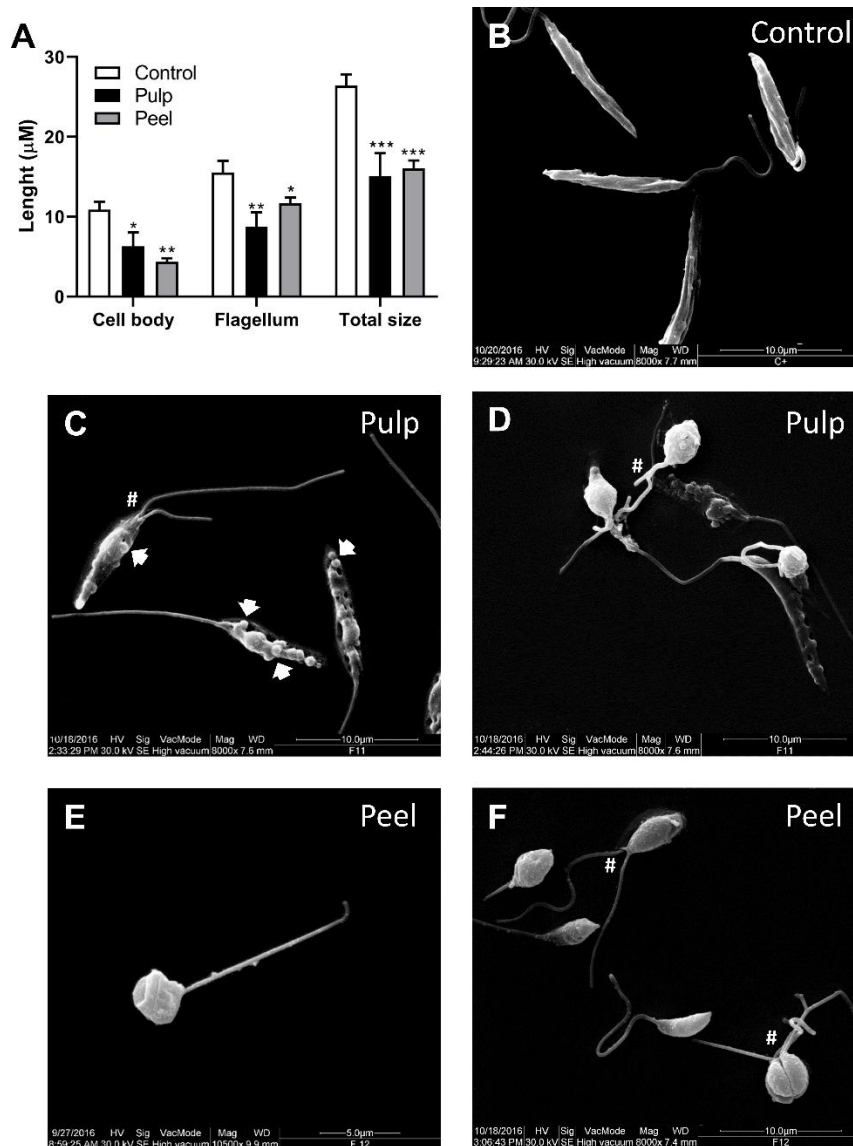
782 duplicate. ** Significant difference compared to the control group ($p \leq 0.01$),

783 ****($p \leq 0.001$).



784 **Fig 3** *C. coriaceum* Wittm. extracts induce late apoptotic-like death process in *L.*
 785 *amazonensis* promastigote forms. (A) H₂DCFDA probe was used for reactive species
 786 of oxygen measurement, (B) TMRE assay for fluorometric analysis of the mitochondrial
 787 membrane potential, (C) Annexin V labeling for phosphatidylserine exposition
 788 evaluation, and (D) propidium iodide staining for the analyses of plasma membrane
 789 integrity. Data represent the mean ± SEM of three independent experiments performed in
 790 duplicate. *** Significant difference in relation to control ($p \leq 0.01$), **** ($p \leq 0.01$). Co-

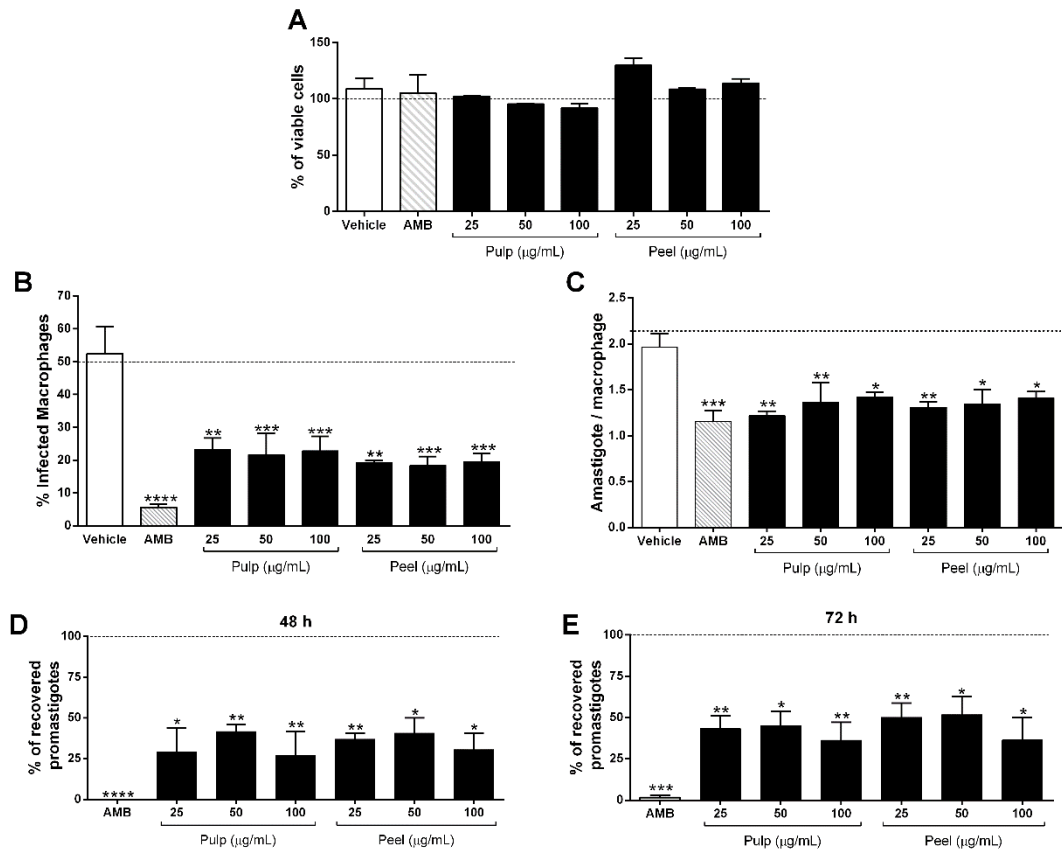
791 staining of Annexin V and PI of untreated (E), pulp-treated (F) and peel-treated (G)
 792 parasites. Typical pseudocolor plots of at least three independent experiments are shown.



793

794 **Fig 4 Morphological changes of treated *L. amazonensis* promastigote forms.** (A) Cell
 795 body, flagellum and total promastigote lengths were measured through scanning electron
 796 microscopy images by Image-Pro Plus software. At least three images of each condition
 797 were subjected to measurement. The values represent the mean \pm SEM of at least three
 798 independent measurements. *Significant difference compared to the control ($p \leq 0.05$),
 799 **($p \leq 0.01$), ***($p \leq 0.001$). (B) Scanning electron microscopy image of *L. amazonensis*
 800 promastigote forms without treatment (control), and promastigotes treated for 24h at 25°C

801 with pulp (C and D) or peel (E and F) extract (50 μ g/mL) (B). Arrows indicate the blebbing
 802 (apoptotic bodies) formation on parasite membrane. # indicate the double flagella
 803 formation.



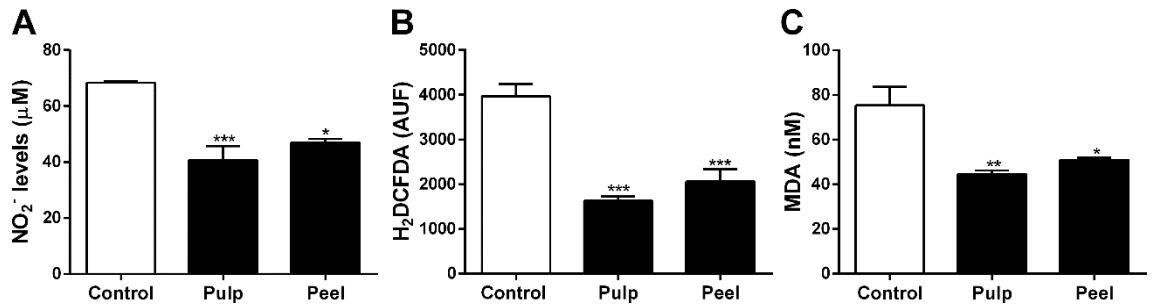
804

805 **Fig 5 Cytotoxic and antiamastigote effect of pulp and peel *C. coriaceum* Wittm.**

806 **extracts.** (A) Peritoneal BALB/c macrophages were submitted to a 24 h-treatment using
 807 25, 50 and 100 μ g/mL of pulp and peel extracts and viability analyzed through MTT
 808 assay. (B) *L. amazonensis*-infected macrophages submitted to 25, 50 and 100 μ g/mL of
 809 pulp and peel extracts or DMSO 0.02% (vehicle) were assessed as the percentage of
 810 infected macrophages and (C) amount of amastigotes per macrophage. Culture medium
 811 and AMB (1 μ M) were used as negative and positive controls. (D) 48h and (E) 72h
 812 incubation of the *L. amazonensis*-infected macrophages using medium 199. The number
 813 of recovered parasites was measured in Neubauer chamber. The dashed lines indicates
 814 the infected control group without treatment (100%). The values represent the mean \pm

815 SEM of three independent experiments performed in duplicate. *Significant difference
816 compared to the control ($p \leq 0.05$), **($p \leq 0.01$), ***($p \leq 0.001$).

817



818

819 **Fig 6** *Caryocar coriaceum* Wittm. extracts reduces the NO, ROS and MDA levels. *L.*820 *amazonensis*-infected macrophages after 24h treatment with *C. coriaceum* Wittm. pulp

821 and peel extracts at 25µg/mL were submitted to (A) nitrite measurement by Griess

822 method; (B) reactive oxygen species (ROS) measurement by fluorescent probe

823 H₂DCFDA; (C) malondialdehyde (MDA) measurement through HPLC. The values

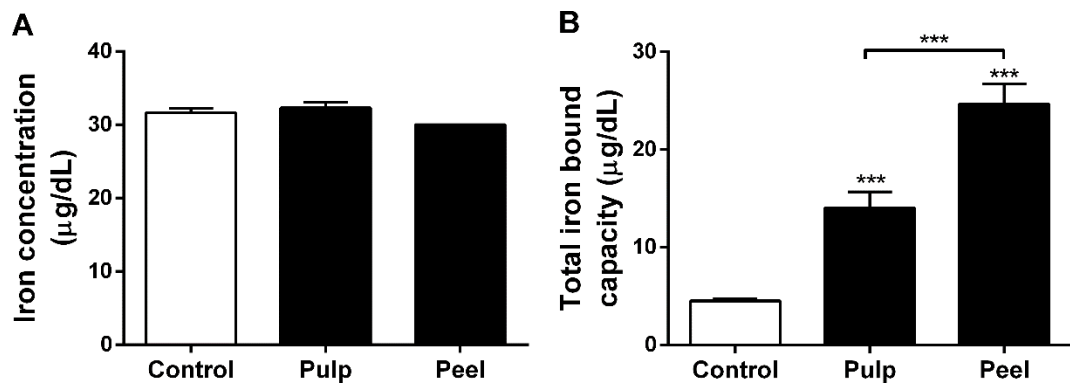
824 represent the mean ± SEM of three independent experiments performed in duplicate.

825 Control represents infected non-treated cells. *Significant difference compared to control

826 (p≤0.05), ** (p≤0.01), *** (p≤0.001).

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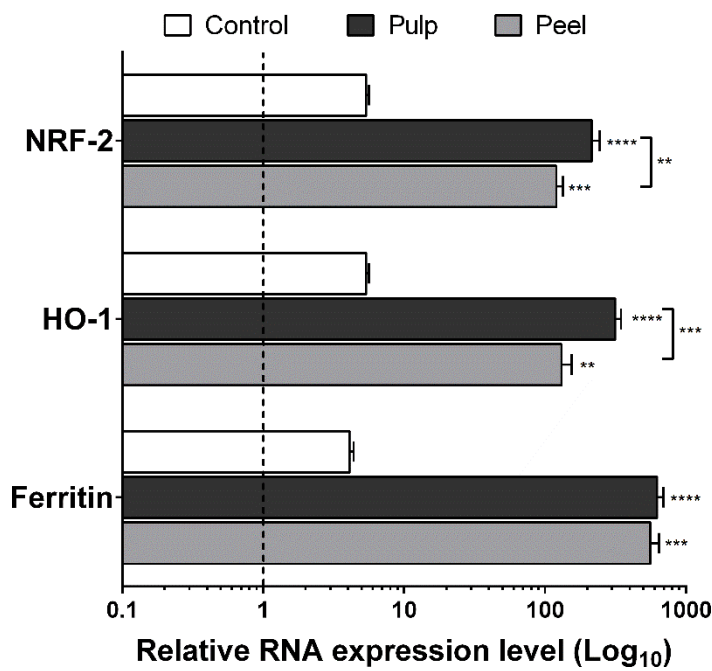
830 **Fig 7** *Caryocar coriaceum* Wittm. extracts treatment enhancing the total bound iron831 **capacity in *L. amazonensis*-infected macrophages.** The culture recovery supernatants

832 were used to determinate the (A) labile iron concentration and (B) total iron bound

833 capacity utilizing the Dimension® automated system. Data represent the mean ± SEM of

834 three independent experiments. *** Significant difference compared to control (p≤0.001).

835



836

837

838 **Fig 8** *Caryocar coriaceum* Wittm. extracts treatment upregulates Nrf2, heme839 **oxigenase-1, and ferritin RNAm expression.** Real-time RT-PCR quantitative mRNA

840 analyses were performed to quantify the expression of Nrf2, HO-1, and ferritin in

841 peritoneal macrophages infected with *L. amazonensis* and treated 25µg/mL of pulp and

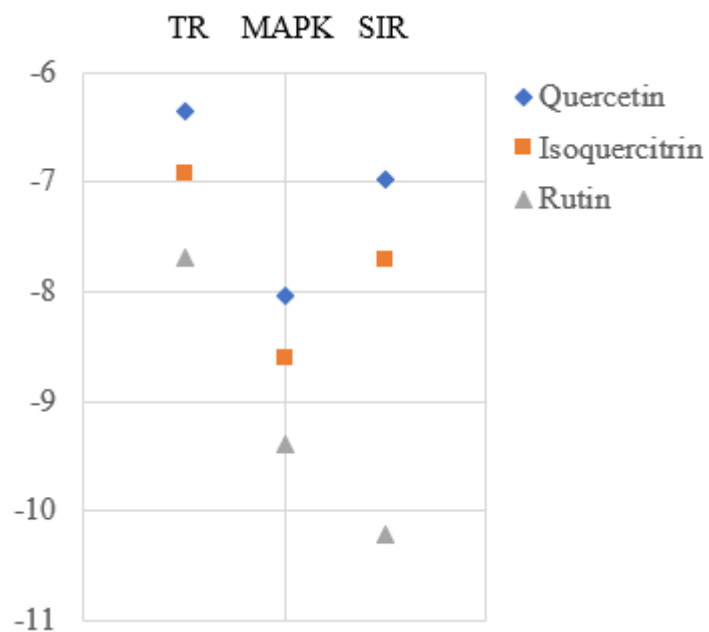
842 peel extracts. The dashed line represents the uninfected control. Data represent the mean

843 ± SEM of three independent experiments. **Significant difference compared to the

844 control ($p \leq 0.01$), ***($p \leq 0.001$),**** ($p \leq 0.0001$).

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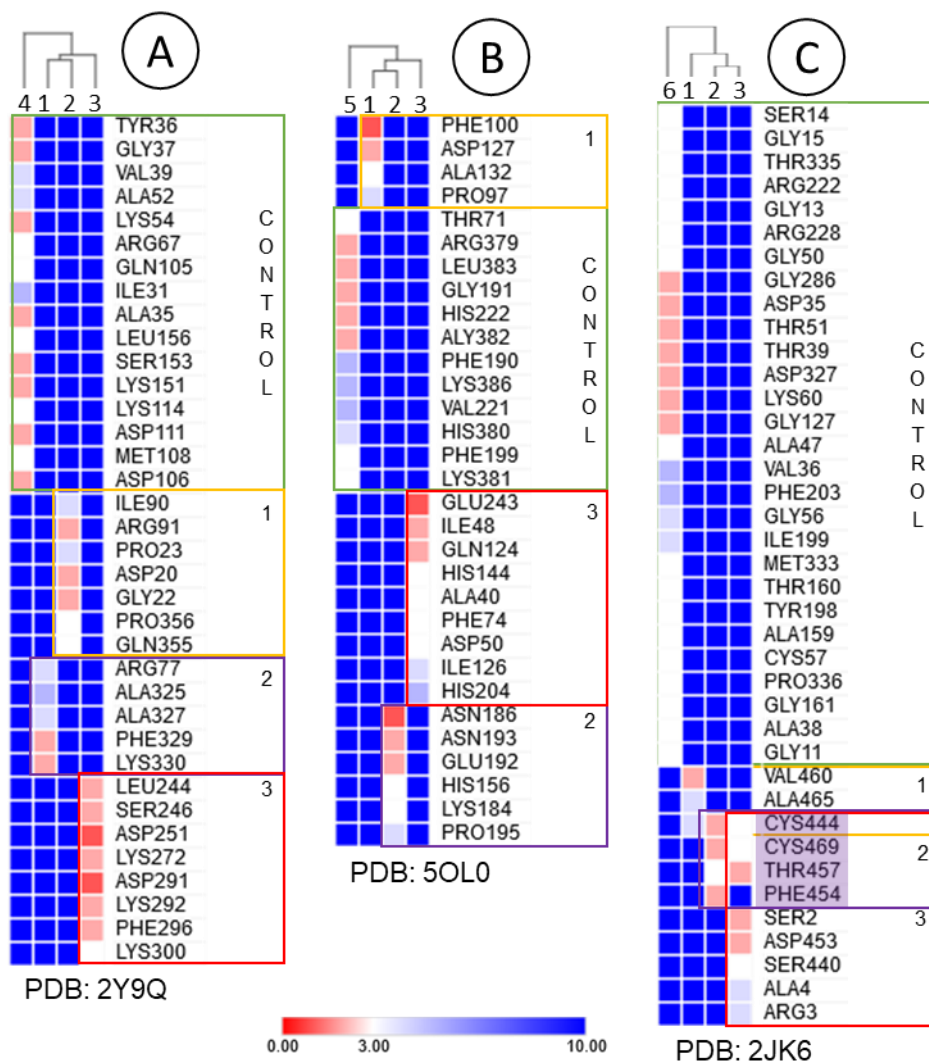
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847

848 **Fig 9 Energy binding (kcal/mol) of (1) quercetin (◆, blue diamond), (2) isoquercitrin**849 **(■, orange square) and (3) rutin (▲, grey triangle) with MAPK (PDB: 2Y9Q), SIR**850 **(PDB: 5OL0) and TR (PDB: 2JK6) biological targets of *Leishmania* sp.**

851



Nº	Ligand
1	Quercetin
2	Isoquercitrin
3	Rutin
4	Phosphoaminophosphonic acid-adenylate ester
5	L-peptide linking cellular tumor antigen p53
6	Flavin-adenine dinucleotide sulfate

852

853 **Fig 10 Heatmap panel illustrating statistically regarding interactions of minimum**
854 **binding energy of test ligands and parameters of molecular interaction with target**
855 **proteins MAPK (2Y9Q), SIR (5OL0) and TR (2JK6).** In all graphics, the closer to 0
856 (red) the interaction force is more determinant and intense, the closer to 10 (blue)
857 the greater the distance and the interaction force is negligible. Clusters of each ligand
858 evidenced by colors, (control) green; (1) yellow; (2) blue and (3) red. Preserved residues
859 highlighted by squares shaded by purple.

860

861

6.2.2. Artigo 2

Oregano (*Origanum vulgare*) essential oil presents drug-likeness proprieties and anti-*Leishmania* activity without triggering an inflammatory response

Artigo submetido à revista *Phytomedicine* (Qualis A1).

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2 **anti-*Leishmania* activity without triggering an inflammatory response**

3
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33

34 **ABSTRACT**

35 **Background:** Leishmaniasis is a neglected disease complex with high incidences in developing countries, for which
36 the available drugs present high toxicity and low efficacy. Thus, the search for new drugs is necessary and urgent.
37 The oregano essential oil (OEO) from *Origanum vulgare*, presents several biological effects, including the inhibitory
38 effect on *Leishmania amazonensis*. However, until now the mechanisms of action of OEO on promastigote or
39 intracellular amastigote forms of *Leishmania* species have not been studied.

40 **Purpose:** To investigate the effect of OEO on leishmaniasis through *in silico*, *in vitro*, and *in vivo* approaches.

41 **Methods:** This study assessed the drug-likeness prediction of OEO main components *in silico*, the leishmanicidal
42 activity against promastigotes and amastigote-infected macrophages, as well its microbicide and
43 immunomodulatory mechanisms *in vitro*, and the *in vivo* effect of OEO local treatment on *L. amazonensis*-infected
44 mice.

45 **Results:** We demonstrated that OEO main components are good candidates for the development of a drug according
46 to the *in silico* study. *In vitro*, OEO acted on promastigote forms, triggering a combination of autophagic, apoptotic,
47 and necrotic events, as well as on intramacrophagic amastigote forms, without trigger a pro-inflammatory response,
48 showing reduced levels of TNF- α , reactive oxygen species, and nitric oxide. *In vivo* studies confirmed the OEO
49 anti-leishmanial potential, once the treatment reduced the lesions of infected mice. Computational docking analysis
50 also suggested an interaction between *Leishmania* arginase and the main OEO component, carvacrol, proposing one
51 of the probable antileishmanial targets of OEO.

52 **Conclusion:** Our study provided new perspectives to the OEO treatment, showing its *in silico*, *in vitro*, and *in vivo*
53 effect against *L. amazonensis*, and providing satisfactory support for further studies of new prototypes of
54 antileishmanial drugs.

55 **Key-words:** Leishmaniasis; natural product; TNF- α ; reactive oxygen species; nitric oxide, *in vivo*.

56

57 **Abbreviations:** AMB – amphotericin B, AUF – arbitrary units of fluorescence, DCF – dichlorofluorescein, DMSO
58 - dimethyl sulfoxide, FBS – fetal bovine serum, FSC-A – forward scatter-area, H₂DCFDA – diacetate 2',7'-
59 dichlorofluorescein probe, IC₅₀ - the half-maximal inhibitory concentration, MTT – 3-(4,5-dimethylthiazol-2-yl)-
60 2,5- diphenyltetrazolium bromide, MDC – monodansylcadaverine, NO – nitric oxide, OEO - oregano essential oil,
61 PBS – phosphate buffered saline, PI – propidium iodide, ROS – reactive oxygen species, SEM – scanning electron
62 microscopy, SEM – scanning electron microscopy, TMRE – tetramethylrhodamine ethyl ester probe, TNF – tumor
63 necrosis factor.

64

65

66

67 INTRODUCTION

68 Leishmaniasis is a neglected disease with high incidences in developing countries, with more than 1 billion
69 people living in endemic areas at risk of infection, and with 700,000 to 1 million new cases annually. The disease is
70 caused by protozoa of the *Leishmania* genus and transmitted through the bite of female phlebotomine insects (WHO,
71 2020). These parasites have a digenetic life cycle where the free-flagellated procyclic promastigote form undergoes
72 differentiation to enter into an infective metacyclic stage in the sandfly, and finally, after infection, differentiates
73 into the disease-causing rounded amastigotes, the obligate intracellular form (Burza et al., 2018).

74 In humans, the clinical manifestation of the disease is dependent on the interplay between the parasite
75 species, vector biology, and the host's immune response ranging from the cutaneous (CL) to the visceral forms (VL)
76 (Aruleba et al., 2020). The parasite developed different immunomodulatory strategies that are essential for infection
77 establishment, hence, an appropriate inflammatory/immune response, defined by its ability to achieve the delicate
78 balance between the removal of *Leishmania* parasites and the minimization of pathology, is one of the major goals
79 for drug discovery [4].

80 Although there has been considerable progress in the development of treatment for VL, and several novel
81 compounds are currently in pre-clinical and clinical development for this clinical form, for CL there have been
82 limited advances in drug research and development, and the available treatment is based on drugs that present high
83 toxicity, relapses, therapeutic failure, and resistance (Alvar et al., 2006).

84 Several products of natural origin have been studied for their therapeutic potential in leishmaniasis (Singh
85 et al., 2014), with great interest in essential oils that possess a wide variety of hydrophobic compounds with
86 antimicrobial potential. The ability to diffuse across cell membranes certainly gives those molecules some advantage
87 in targeting intracellular microbes, being a valuable research option for the search of bioactive compounds (Bakkali
88 et al., 2008).

89 In this sense, the oregano essential oil (OEO), extracted from *Origanum vulgare*, has attracted attention due
90 to its biological effects as the broad antibacterial, antifungal, antiparasitic and antioxidant effect [9]. Regarding the
91 antiparasitic effect of OEO, it includes the inhibitory effect on *Leishmania amazonensis* (Sanchez-Suarez et al.,
92 2013; Teles et al., 2019). However, until now the mechanisms of action of OEO on promastigote or intracellular
93 amastigote forms of *Leishmania* species have not been studied.

94 The antimicrobial proprieties of OEO have been associated with the presence of different chemical classes
95 of compounds, and potent interactions between the OEO components have been reported, suggesting an improved
96 efficacy of the mixture when compared to the isolated compounds (Bassolé and Juliani, 2012). Moreover, a mixture
97 of molecules with antimicrobial activity could minimize the selection of resistant strains.

98 In the present work, we demonstrated that OEO containing carvacrol, thymol, γ -terpinene, p-cymene, and
99 β -caryophyllene as the main components is a good candidate for drug development according to the *in silico* study,
100 acting against *L. amazonensis* *in vitro* and *in vivo*. OEO presented action on promastigote forms, triggering a
101 combination of autophagic, apoptotic, and necrotic events, as well as in amastigote forms, without trigger a pro-
102 inflammatory response. The results were confirmed by the reduction of lesions on *in vivo* studies. Computational

103 docking analysis also suggested the arginase-carvacrol interactions by spontaneous binding proposing one of the
104 probable antileishmanial targets of OEO.

105

106 **MATERIALS AND METHODS**

107 **Animals and Ethics Committee**

108 BALB/c weighing approximately 25–30 g and aged 6–8 weeks were kept under sterile conditions and used
109 according to protocols approved by the Institutional Animal Care and Committee. This study was approved by the
110 Ethics Committee for Animal Experimentation of the State University of Londrina 8595.2018.89.

111

112 **Culture of *Leishmania (L.) amazonensis***

113 Promastigote forms of *Leishmania (Leishmania) amazonensis* ((MHOM/BR/1989/166MJO) were maintained
114 in culture medium 199 (GIBCO Invitrogen) supplemented with 10% fetal bovine serum-FBS (GIBCO Invitrogen),
115 1 M HEPES, 0.1% human urine, 0.1% L-glutamine, 10 µg/mL penicillin and streptomycin (GIBCO® Invitrogen)
116 and 10% sodium bicarbonate. The cell cultures were maintained in an incubator-type B.O.D. at 25 °C, in 25 cm²
117 flasks. In all experiments, promastigote forms were used in the stationary growth phase.

118

119 **Oregano essential oil**

120 Oregano (*Origanum vulgare*) essential oil (OEO) was obtained from Ferquima Industry and Commerce of
121 Essential Oils (São Paulo, Brazil). This oil was extracted by steam distillation and its density (0.954 g/mL) and
122 composition (main components: carvacrol, thymol, γ-terpinene, p-cymene, and β-caryophyllene) were described in
123 a technical report (CAS number 84012-24-8, batch 224). A stock solution of 50% OEO was prepared in
124 dimethylsulfoxide (DMSO, Sigma-Aldrich; v/v). The maximum DMSO concentration in assays was 0.01%.

125

126 **Drug-likeness prediction of OEO main components**

127 The carvacrol, thymol, γ-terpinene, p-cymene, and β-caryophyllene structure *in silico* study was carried out to
128 evaluate theoretical drug-likeness parameters related to oral bioavailability and absorption, distribution, metabolism,
129 excretion, and toxicity (ADMET) properties. The predictions were calculated in admetSAR software
130 (<http://lmm.d.ecust.edu.cn/admetSar2/>) according to Lipinski's rule of five (Ro5) [22] followed by the additional rule
131 proposed by Veber [23]. ADMET properties predictions were also calculated according to (Cheng et al., 2012).

132

133 **Antipromastigote assay**

134 *Leishmania amazonensis* promastigote forms (10⁶ cells/mL) were treated with a serial concentration of OEO
135 (3.12, 6.25, 12.5, 25, 50, and 100 µg/mL) for 24 h at 25 °C, and the parasites were counted on a Neubauer chamber.
136 Promastigote forms maintained in the culture medium were used as a control, DMSO 0.01% was used as a vehicle,
137 and amphotericin B (AMB) 1µM was used as a positive control. The 50% inhibitory concentration of the parasites
138 (IC₅₀) curve was calculated by logarithmic non-linear regression to the dose-response curve from the data obtained.

139

140 **Viability of peritoneal macrophages**

141 Peritoneal macrophages (5×10^5 cells/mL) were recovered from the peritoneal cavity of BALB/c mice with cold
142 PBS supplemented with 3% of FBS and then cultured in 24-well plates with 200 μ L of RPMI 1640 medium (10%
143 FBS) for 2 h (37 °C, 5% CO₂). Adherent cells were incubated with OEO (3.12, 6.25, 12.5, 25, 50, and 100 μ g/mL)
144 cultured for 24 h under the same conditions. After this period, MTT (3-(4,5-dimethylthiazol-2-yl)-2,5-
145 diphenyltetrazolium bromide) (Sigma-Aldrich) (5 mg/mL) was added for 4 h. Cells under the same conditions
146 without treatment were used as control; DMSO 0.01% was the vehicle, and the positive control was H₂O₂ 0.4%.
147 The plates were read using a spectrophotometer (Thermo Scientific, Multiskan GO) at 550 nm. The results were
148 expressed as a percentage of viability compared to the control group.

149

150 **Analysis of hemolytic activity**

151 Sheep blood was collected with heparin (Ethics Committee for Animal Experimentation of State University of
152 Londrina: 82862016.60) and the toxicity on erythrocytes evaluated as previously described (Bortoleti et al., 2018).
153 PBS was used with negative control and Triton X 1% as a positive control for hemolysis.

154

155 **Scanning electron microscopy (SEM) of promastigote forms**

156 The morphological changes of *L. amazonensis* promastigote forms were evaluated by SEM as previously
157 described (Tomiotto-Pellissier et al., 2018).

158 The length of the promastigote forms (flagellum, cell body, and total length) was analyzed in the Image-Pro
159 Plus software, based on the 10,000x magnification images obtained. At least three images of each condition were
160 subjected to measurement.

161

162 **Determination of the parasites cell size**

163 *L. amazonensis* promastigote forms (10^6 cells/ml) were treated with OEO-IC_{50p} and incubated for 24 h at
164 24°C, harvested, and washed with PBS. Subsequently, the parasites were analyzed using a BD Accuri C6 flow
165 cytometer (BD Biosciences, San Jose, CA), as previously described (Tomiotto-Pellissier et al., 2018). Parasites
166 maintained in the culture medium were used as a control and DMSO 0.01% was used as vehicle control.

167

168 **Mechanism of action determination on *L. amazonensis* promastigote forms**

169 To determine the mechanism of action of OEO, 10^6 promastigote forms were treated for 24 h with IC_{50p} and
170 $2 \times$ IC_{50p}, and different parameters were assessed as previously described by our group (Concato et al., 2020;
171 Tomiotto-Pellissier et al., 2018). Briefly, the inner mitochondrial membrane potential was investigated by incubation
172 with tetramethylrhodamine ethyl ester (TMRE) (Sigma-Aldrich); the reactive oxygen species (ROS) were analyzed
173 by a permeant probe diacetate 2',7'-dichlorofluorescein diacetate (H₂DCFDA) (Sigma-Aldrich), and the lipid
174 droplets detection was performed by labeling with Nile red (Sigma-Aldrich). To verify the death type, we performed
175 autophagic vacuoles quantification by monodansylcadaverine (MDC) (Sigma-Aldrich), phosphatidylserine
176 exposition by annexin-V, and verification of cell membrane integrity by propidium iodide labeling.

177 As positive controls were used carbonyl cyanide m-chlorophenylhydrazone, H₂O₂, camptothecin, and
178 digitonin (all from Sigma) respectively for TMRE, DCF, annexin-V, and propidium iodide analysis, and 24h-
179 parasites cultivated with PBS for Nile Red and MDC.

180

181 **Anti-amastigote assay**

182 Peritoneal macrophages (5x10⁵ cells/mL) were obtained as previously described. The adherent cells were
183 infected with *L. amazonensis* promastigotes (2.5 x10⁶ cells/mL) for 2 h. After, the non-internalized parasites were
184 washed with PBS and the infected cells treated with OEO (3.12, 6.25, 12.5, 25, 50 and 100 µg/mL), RPMI 1640
185 medium (control), DMSO 0.01% (vehicle) or AMB 1 µM (positive control) for 24 h (37 °C, 5% CO₂). After this,
186 the cells were stained with Giemsa (Laborclin), analyzed as (Bortoleti et al., 2018). The supernatant was stored for
187 further analysis.

188

189 **Determination of nitrite as estimative of nitric oxide (NO) levels**

190 NO was determined through the Griess method. Briefly, supernatant aliquots (60 µL) of the anti-amastigote
191 assay were centrifuged at 2370×g for 2 min and a 50 µL of the supernatant was added with 50 µL of Griess reagent
192 (1% sulfanilamide and 0.1% of N-(1-Naphthyl) ethylenediamine in orthophosphoric acid (H₃PO₄) 5%). After 10
193 min incubation at room temperature, the samples were placed in 96-well microplates. A calibration curve was made
194 using dilutions of NaNO₂, and the absorbance was determined at 550 nm on a microplate reader (Thermo Scientific,
195 Multiskan GO).

196

197 **Reactive oxygen species (ROS) generation by macrophages**

198 The ROS generation of peritoneal macrophages (5×10⁵ cells/mL - infected and treated under the same conditions
199 described in the anti-amastigote assay) was evaluated as described in (Tomiotto-Pellissier et al., 2018).

200

201 **Cytokines measurement**

202 The supernatants obtained in the anti-amastigote assay were used to determine the levels of TNF-α, IL-6 e IL-
203 10 using eBioscience commercial kits capture enzyme-linked immune sorbent assay (ELISA) (San Diego, CA,
204 USA), according to the manufacturer's instructions.

205

206 **In vivo analysis of OEO treatment**

207 BALB/c mice (n = 4 per group) were anesthetized and infected at the base of the tail, subcutaneously, with 10⁵
208 promastigote forms of *L. amazonensis*. After the appearance of the lesion in all mice, 100 days post-inoculation
209 (p.i.), the daily treatment started on groups as follows: vehicle group- applications of base gel, OEO group - gel
210 containing OEO 0.2% at the lesion site (0.5 g - applied in circular movements), and positive control group -
211 intraperitoneal injection (i.p.) of glucantime 20 mg/Kg/day. Non-treated animals were used as control. After five
212 weeks of treatment, the animals were anesthetized and euthanized following the approved protocols.

213 The lesions were measured weekly after the onset of treatment using a digital caliper (Starrett 799). Data were
214 expressed as (final lesion size) - (initial lesion size) for each animal.

215

216 **Statistical analysis**

217 Data were expressed as a mean \pm SEM. Data were analyzed using the GraphPad Prism statistical software
218 (GraphPad Software, Inc., USA, 500.288). Significant differences between the groups were determined through
219 one-way ANOVA, followed by Tukey's test for multiple comparisons. Differences were considered as statistically
220 significant upon $p < 0.05$. At least three independent experiments were performed, each with duplicate datasets.

221

222 **Molecular modeling analysis**

223 The crystal structure of *Leishmania mexicana* arginase (ARG; PDB 4IU5) was used for docking calculations
224 (D'Antonio et al., 2013). The carvacrol two-dimensional structure was obtained from the virtual repository
225 PubChem (<https://pubchem.ncbi.nlm.nih.gov/>) and converted to ligand.mol2.

226 Before of test, ions, co-crystallized ligands, and water molecules not relevant to the system structure were
227 removed from the target. The protein binding site for molecular docking was centered between Mg^{2+} at x: 15.141,
228 y: -15.1248, z: -5.40 for calculations as previously described (Camargo et al., 2020).

229 The types of ligand-residue interactions, as well as the figures, were generated using the Discovery Studio
230 Visualizer (Dassault Systèmes BIOVIA, Discovery Studio Modeling Environment, Release 2017, San Diego:
231 Dassault Systèmes, 2016).

232 The protein and ligand preparation was performed using AutoDockTools (ADT) v. 1.5.6 (Morris et al., 2009).
233 The polar hydrogens were added to the protein and atoms charges of protein and ligand are assigned by Kolman and
234 Gasteiger methods, respectively. The dimensions of the grid box were determined as $40 \times 50 \times 38 \text{ \AA}^3$ (Camargo et al.,
235 2020). Molecular docking calculations were carried out considering the Lamarckian Genetic algorithm implemented
236 in Autodock. Scoring functions with 10 iterative runs were performed.

237

238 **RESULTS**

239 **Drug-likeness assessment of main components of OEO**

240 Knowing that the main components of the tested OEO are carvacrol (71%), thymol (3%), γ -terpinene (4.5%),
241 p-cymene (3.5%), and β -caryophyllene (4%) (Figure 1), our first aim was to verify the drug-likeness theoretical
242 potential of those compounds through an *in silico* study. The studied compounds did not present any violation of
243 Lipinski's and Veber's rules, suggesting a good chance of drug-likeness and oral bioavailability (Table 1).

244 Moreover, ADMET properties were analyzed through admetSAR and reported in Table S1. The results predict
245 that the main components of OEO present potential human intestinal absorption, caco2 cell, and blood-brain barrier
246 permeability, human oral bioavailability, non-inhibitor of OATP2B1, MATE, OCT2, BASEP as well as most of the
247 CYPs, besides to no inhibition promiscuity with the main five CYP isoforms (CYP1A2, CYP2C19, CYP2C9,
248 CYP2D6, and CYP3A4). Also, the compounds were predicted as non-carcinogenic, non-mutagenic, non-inductor

249 of micronucleus, and non-hepatotoxic. Regarding the acute oral toxicity, all compounds presented degree III, that
250 is, slightly toxic and slightly irritating predictions (Table S1).

251 On the other side, the compounds also were predicted as OATP1B1 and OATP1B3 inhibitors, with possible
252 effects in eye corrosion and/or irritation. Also, carvacrol and thymol are possible CYP1A2 inhibitors. Besides that,
253 the prediction shows subcellular localization of carvacrol and thymol in the mitochondria, while γ -terpinene, p-
254 cymene, and β -caryophyllene in the lysosomes (Table S1).

255 Table S2 shows the predictive values of the *in silico* study. We found good values for water solubility and low
256 oral toxicity. Concerning the binding to plasma proteins, γ -terpinene, p-cymene and β -caryophyllene presented low
257 predicted values (<85%), while carvacrol and thymol presented higher predictions (\geq 90%).

258

259 **OEO exerts leishmanicidal effect against promastigote forms of *L. amazonensis* in vitro without cytotoxic** 260 **effect for macrophages**

261 The direct effect of OEO on promastigote forms of *L. amazonensis* was evaluated by the parasite
262 proliferation. OEO treatment was able to reduce the viability of the parasites at all concentrations evaluated (3.12-
263 100 μ g/mL) when compared to control and DMSO ($p \leq 0.0001$) (Figure 2A), exhibiting an IC_{50p} value of 16 (± 0.06)
264 μ g/mL (Table 2). This concentration and twice the value ($2 \times IC_{50p}$) were used for the following experiments in
265 promastigote forms.

266 Besides that, the cytotoxic effect of OEO on erythrocytes and macrophages (the main host cells for
267 *Leishmania* sp.) was evaluated. We found that the concentrations up to 100 μ g/mL were not able to reduce the
268 viability of sheep erythrocytes and BALB/c mice peritoneal macrophages (Figure 2B, C). Also, the cytotoxic value
269 for 50% of the cells (CC_{50}) was established as 188.2 (± 10) μ g/mL. The selectivity index of OEO in promastigote
270 forms was calculated from the IC_{50} and CC_{50} values, obtaining a value of 11.7 (Table 2).

271

272 **OEO induces morphological changes and reduction in the cell volume of promastigote forms**

273 The morphological changes of OEO-treated *L. amazonensis* promastigote forms were analyzed by SEM.
274 Untreated (Figure 3A,B) and vehicle-treated (0.01% DMSO) parasites (Figure 3C) showed an elongated body,
275 smooth cell membrane, and all its extracellular structure preserved. Treatment with OEO- IC_{50p} for 24 h induced
276 roughness on the surface of the parasite, reduced cell body size, presence of two flagella, and leakage of cytoplasmic
277 content (Figure 3D-I).

278 Moreover, from the analysis of the images, we could verify that the IC_{50p} OEO-treatment reduced the
279 flagellum, cell body, and total cell size after 24 h concerning the control (Figure 3J). Confirming these data, the flow
280 cytometry analysis showed a reduction in the cell size in both, IC_{50p} and $2 \times IC_{50p}$ concentrations, without differences
281 between the used doses (Figure 3K).

282

283 **OEO treatment promotes death process in *L. amazonensis* promastigote forms by a combination of** 284 **autophagic, apoptotic, and necrotic events**

285 Our next objective was to identify the mechanisms involved in the death of the parasites. The results showed
286 that OEO treatment in both concentrations was able to induce ROS production in parasites and cause mitochondrial
287 depolarization when compared to the control (Figure 4A,B). We also found that treatment with OEO induced the
288 accumulation of neutral lipid droplets and the formation of the autophagic vacuoles when compared to the respective
289 controls. In both parameters, $2 \times IC_{50p}$ showed greater fluorescence than IC_{50p} (Figure 4C,D).

290 Besides, the cell death mechanism triggered by the OEO-treatment involved apoptosis-like death by
291 marking the externalization of phosphatidylserine, as well as the involvement of plasma membrane damage due to
292 the staining of the treated parasites with PI, which diffuses across permeable membranes and binds to nucleic acids
293 (Figure 4E-F).

294

295 **OEO treatment induces *L. amazonensis* amastigotes elimination from infected macrophages**

296 As OEO showed no toxicity in murine peritoneal macrophages at concentrations of 3.12 - 100 $\mu\text{g/mL}$, we
297 investigated its action against intramacrophagic amastigotes. We observed that concentrations of 6.25, 12.5, 25, 50,
298 and 100 $\mu\text{g/mL}$ of OEO significantly reduced the total amount of amastigotes and the percentage of infected
299 macrophages (Figure 5A, B), while the concentrations of 12.5, 25, 50 and 100 $\mu\text{g/mL}$ also induced a reduction in
300 the number of amastigotes by macrophages when compared to the control (Figure 5C). Thus, concentrations of 12.5
301 - 100 $\mu\text{g/mL}$ were chosen for the next set of experiments.

302 Based on the results, we obtained a value of 17.2 $\mu\text{g/mL}$ for the IC_{50} of intracellular amastigote forms, from
303 which a selectivity index of 10.9 times was calculated to peritoneal macrophages (Table 4).

304

305 **OEO treatment down-modulates ROS, NO, and TNF- α levels in *L. amazonensis*-infected macrophages**

306 Knowing that OEO induces the elimination of intramacrophagic amastigote forms, we studied the
307 immunomodulation caused by the treatment. Our data demonstrated that OEO treatment (12.5 - 100 $\mu\text{g/mL}$)
308 significantly reduced the production of ROS and NO when compared to the control (Figure 6A,B). Likewise, the
309 highest concentrations, 50 and 100 $\mu\text{g/mL}$, also significantly reduced TNF- α levels when compared to the control
310 (Figure 6C). The levels of IL-6 and IL-10 did not differ from the control or each other (data not shown).

311

312 ***In vivo* activity of OEO treatment**

313 Once the *in vitro* effect of OEO was showed, its potential action in *L. amazonensis*-infected mice was
314 investigated. For this, BALB/c mice were infected and topically treated for five weeks with 0.2% OEO-based gel
315 after the lesion appearance. The lesion size measurements showed that the control, vehicle, and positive control
316 (glucantime) groups increased the lesion size over time, demonstrated by positive values. The OEO treatment was
317 able to reduce the lesion sizes (negative values), presenting smaller lesions than all other groups (Figure 7). It is
318 important to note that the OEO-treatment was more effective in the tissue repair process than the standard drug
319 (glucantime), in which, despite the treated lesions showed smaller size when compared to the negative control and
320 vehicle, was not able to reduce the size of the injuries, but only delayed its development.

321

322 **Molecular docking of OEO**

323 To gain insights into the OEO mechanism of action against *Leishmania amazonensis*, we predicted the
324 interaction of carvacrol, the main constituent of OEO, with *Leishmania* arginase (ARG) by molecular docking. We
325 found ARG-carvacrol interactions in the active site of the enzyme, with common hydrogen-bonding interactions
326 with Asp141 residue, besides π - π stacking and π -alkyl interaction with His139 (Figure 8). Also, it was provided the
327 binding energy of ARG-carvacrol complex of -6.1 ± 0.33 kcal/mol, suggesting a spontaneous binding between the
328 molecules. Thus, ARG enzyme was identified as one of the probable antileishmanial target of OEO.

329

330 **DISCUSSION**

331 Available drugs for leishmaniasis treatment are unsatisfactory due to their high toxicity and low efficacy
332 (Alvar et al., 2006). Thus, the search for new drugs is necessary and urgent. In this context, our study provided new
333 perspectives on OEO action in experimental leishmaniasis.

334 Drug discovery is a long and expensive process. In the past decade, from hundreds of drug candidates, only
335 a few reached the market due to the high failure rate at the clinical trial stage, mainly due to the lack of efficacy and
336 high toxicity (Cheng et al., 2012). Thus, to predict the influence of the chemical structure on the oral absorption of
337 a compound by analyzing pharmacokinetic parameters is important for the development of a candidate with good
338 chances to become a drug [22]. Also, the absorption, distribution, metabolism, excretion, and toxicity (ADMET)
339 properties of drug candidates play key roles in drug discovery. Thus, *in silico* studies have become a useful, effective,
340 and low-cost tool for studying drug candidates (Cheng et al., 2012).

341 We performed *in silico* studies to assess theoretical drug-likeness parameters and we found that none of
342 Lipinski's and Veber's rules (Lipinski, 2004; Veber et al., 2002) were violated for the main compounds of OEO:
343 carvacrol, thymol, γ -terpinene, p-cymene, and β -caryophyllene. In the same way, the OEO components proved
344 favorable predictions to most of the ADMET properties calculated.

345 It is noteworthy that we found good values for water solubility, since more than 80% of the drugs on the
346 market have a logS value greater than -4 ("logS Calculation - Osiris Property Explorer," n.d.), and low oral toxicity,
347 being classified as degree III ($LD_{50} > 500$) (Yang et al., 2018). Concerning the binding to plasma proteins, γ -
348 terpinene, p-cymene, and β -caryophyllene presented low predicted values, while carvacrol and thymol presented
349 higher predictions. As a general rule, minimally protein-bound drugs (<85%) penetrate tissues better than those that
350 are highly protein-bound ($\geq 90\%$), but clearance of such drugs is also higher (Johnson-Davis and Dasgupta, 2016).

351 Such results are important because poor ADMET properties are among the main reasons for the failure of
352 drug candidates during clinical trials (Cheng et al., 2012). The DNDi also postulated that an ideal leishmaniasis
353 treatment should occur orally, safely, effectively, at a low cost, and over a short period. A possible lead drug to be
354 used should be *in vitro* non-cytotoxic, with a selectivity index ≥ 10 (Ioset et al., 2009). Therefore, taking together
355 the results obtained for OEO components *in silico* and the SI obtained *in vitro* are indicative that OEO is a good
356 drug candidate for leishmaniasis.

357 Aiming to understand the effect of OEO, we investigated the *in vitro* mechanisms of action on *L.*
358 *amazonensis* promastigote forms. We demonstrated the ability of OEO to increase ROS levels, molecules with

359 different biological functions which in excess in the cells can trigger organelles damage and consequently, cellular
360 death (Menna-Barreto, 2019).

361 In this sense, we also found mitochondrial membrane damage upon the reduction of membrane potential.
362 Mitochondrial integrity is essential for parasite survival since trypanosomatids have unique mitochondrion. Also,
363 both ROS and mitochondrial dysfunction are inducers of lipid droplet accumulation, a hallmark of cellular stress
364 (Menna-Barreto, 2019). Thus, we can infer that OEO treatment induced ROS production, causing mitochondrial
365 damage and accumulation of lipid droplets, which culminates in cell stress and promastigote death.

366 Confirming this data, we found autophagic vacuole formation, phosphatidylserine exposure, and cellular
367 membrane damage on the treated parasites. Autophagy is a biological pathway associated with cell recycling through
368 the removal of damaged cell components, which can be activated in the presence of cell stress, forming autophagic
369 vacuoles. When continuous induction of autophagy occurs, it can progress to the cell death of the parasites (Menna-
370 Barreto, 2019).

371 Apoptosis-like cell death has been related to the leishmanicidal mechanism of different compounds. Besides,
372 the promastigote membrane permeabilization, indicated by marking with propidium iodide, can be indicative of
373 both: late-apoptotic and necrotic cell death (Bortoleti et al., 2018; Tomiotto-Pellissier et al., 2018).

374 These results were confirmed by morphological changes in the promastigotes. Roughness on the surface of
375 the parasite, reduction in cell body size, presence of two flagella, and leakage of cytoplasmic content are evident
376 signals of cell death. Similar morphological alterations were found by our group in OEO-treated *S. aureus*
377 (Scandorieiro et al., 2016). Our data are still in accord with the prediction of the *in silico* study, which showed that
378 the main components of OEO accumulate in mitochondria or lysosomes, two organelles intrinsically related to
379 autophagy and apoptosis (Menna-Barreto, 2019).

380 Knowing the mechanisms involved with promastigote forms death, we investigated the OEO action on
381 amastigote-infected macrophages and lesions development in *L. amazonensis*-infected mice. First, we found that
382 the concentrations used were not toxic to macrophages, the main host cells of *Leishmania* sp. [4]. Then, we found
383 that treatment with OEO was able to reduce the rate of macrophage infection *in vitro* and the lesion size *in vivo*.
384 Amastigotes are resistant forms of these parasites, which have complex defense mechanisms that allow their survival
385 in the hostile intramacrophagic environment (Alvar et al., 2006), therefore a potential drug for leishmaniasis must
386 have the ability to act in these evolutionary forms.

387 The immune response against *L. amazonensis* is complex and naturally involves a pro-inflammatory
388 response with TNF- α production that activates macrophages to ROS and NO synthesis, and consequent parasite
389 elimination [4]. However, the excessive pro-inflammatory response is also responsible for tissue damage and disease
390 persistence (Rossi and Fasel, n.d.). OEO was able to reduce *in vitro* the levels of ROS, NO, and TNF- α with
391 concomitant parasite elimination showing a parasite clearance without trigger inflammation. Also, OEO was able
392 to induce tissue repair in the *in vivo* context.

393 Our data corroborate with (Chuang et al., 2018) who found that the ethanolic extract of *Origanum vulgare*
394 suppressed the inflammatory response, reducing the TNF- α production in an inflammation model induced by

395 *Propionibacterium acnes*. Likewise, (Mahran et al., 2019) also identified a reduction in TNF- α levels in a gamma-
396 ray stress induction model after the treatment with carvacrol and thymol.

397 We further predicted that the main OEO component, carvacrol, is capable of binding in the active site of
398 ARG suggesting that it could inhibit this target. In *Leishmania* parasites, ARG is involved in regulating several
399 biological functions, such as replication, growth, differentiation, infectivity, and persistence (Pessenda and Santana
400 da Silva, 2020). Thus, this result provided new evidence that OEO can directly act on intramacrophagic amastigotes,
401 inducing parasite elimination.

402

403 **CONCLUSION**

404 Taking together, our data demonstrated that OEO is a good candidate for the development of a drug
405 according to the *in silico* study, in addition to having an *in vitro* effect both on promastigote forms, triggering death
406 by autophagy, and in amastigote forms, without trigger a pro-inflammatory response. The anti-*Leishmania* effect
407 was further confirmed *in vivo* by the reduction of lesions. Computational docking analysis also suggested the
408 arginase-carvacrol interactions by spontaneous binding proposing a probable antileishmanial target of OEO. These
409 results suggest OEO as a promising agent for further studies and the design of new prototypes of antileishmanial
410 drugs.

411

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416

417 **CONFLICT OF INTERESTS**

418 The authors declared that there is no conflict of interest.

419

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423

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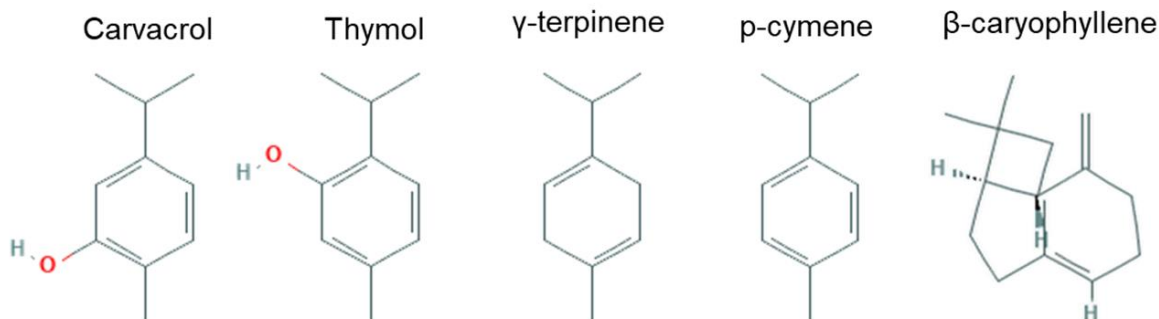
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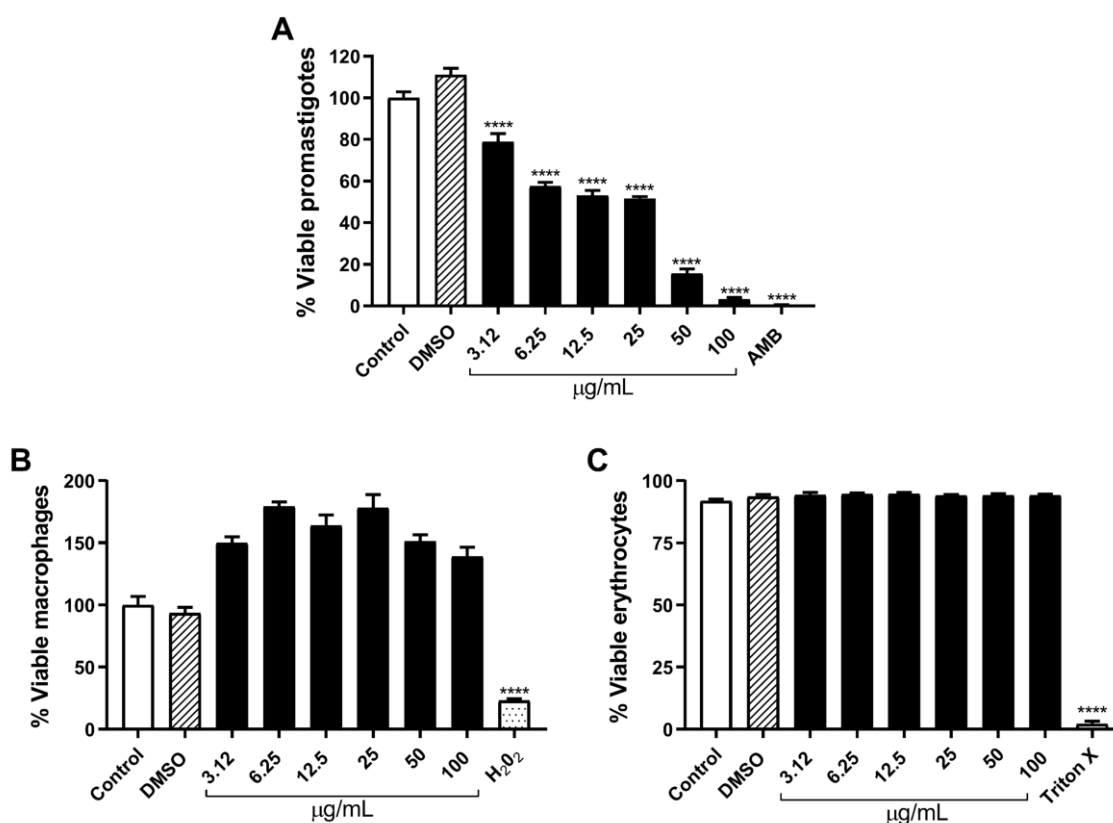
509 **FIGURE LIST**



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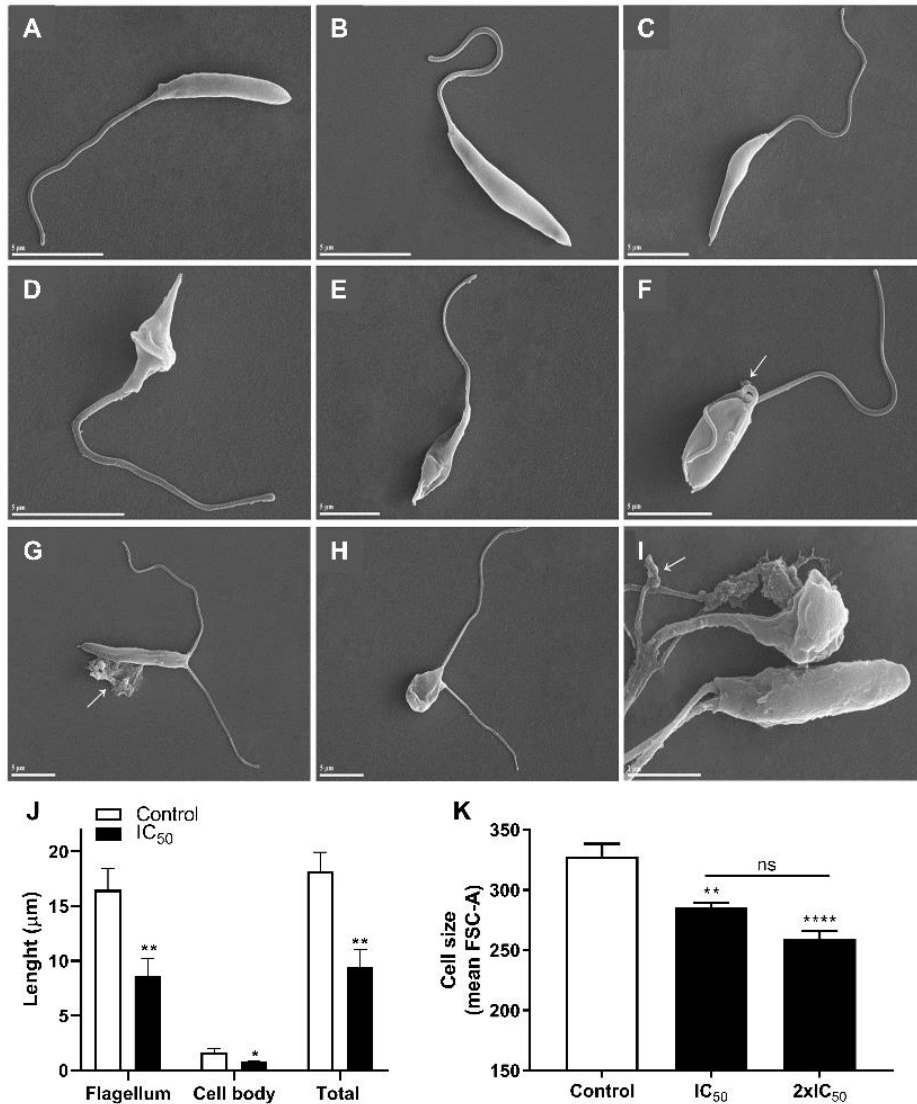
511 **Figure 1 – Molecular structure of the main components of OEO.** OEO composition with 71% carvacrol, 3%
512 thymol, 4.5% γ -terpinene, 3.5% p-cymene, and 4% β -caryophyllene.

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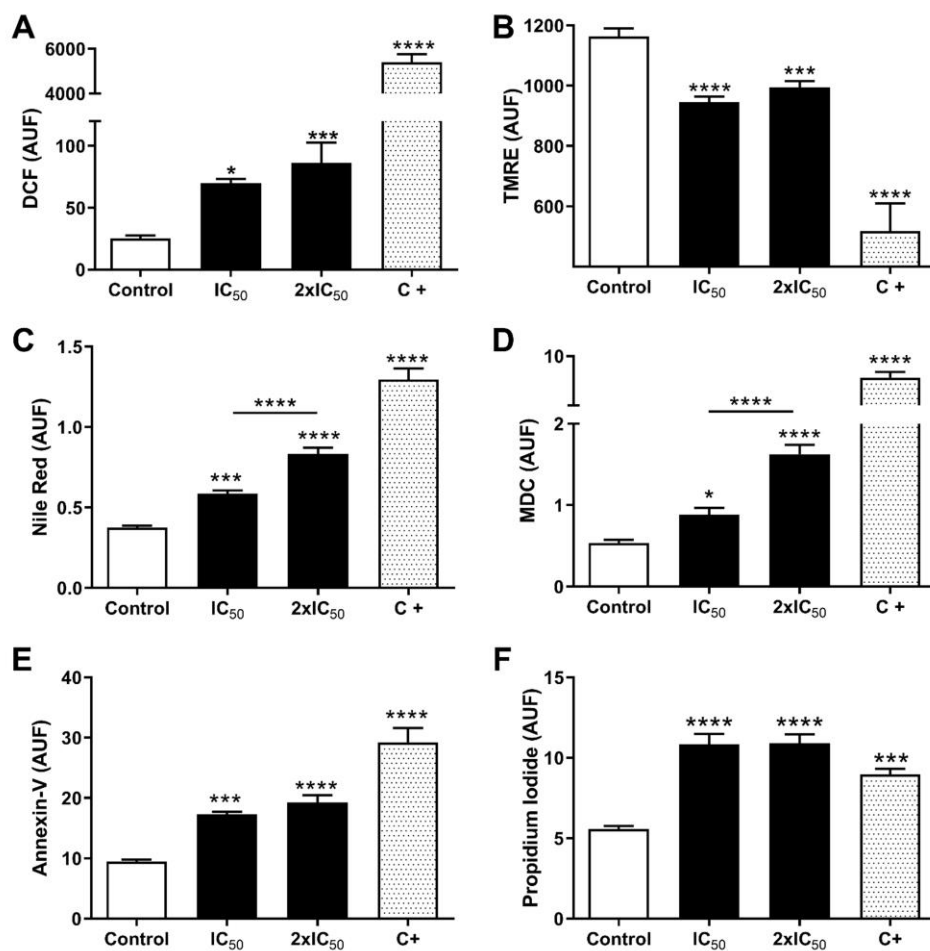
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515 **Figure 2 – Cytotoxic effect of OEO on *L. amazonensis* promastigotes and peritoneal BALB/c macrophages.**
516 (A) *L. amazonensis* promastigote forms were treated with OEO (3.12–100 μ g/mL) and the parasite viability was
517 assessed after 24 h. As control was used non-treated parasites, as vehicle control was used DMSO 0.01% and as a
518 positive control was used amphotericin B (AMB) 1 μ M. (B) Viability of BALB/c peritoneal macrophages treated
519 with OEO (3.12–100 μ g/mL) for 24 h by MTT assay. As control was used non-treated macrophages, as vehicle
520 control was used DMSO 0.01% and as a positive control was used H₂O₂ 0.2%. (C) Viability of erythrocytes treated
521 with OEO (3.12–100 μ g/mL), PBS was used as non-hemolytic control and Triton X 1% was used as a positive
522 control for hemolysis. The values represent the mean \pm SEM of three independent experiments performed in
523 duplicate. ****Significant reduction compared to the control group ($p \leq 0.0001$).



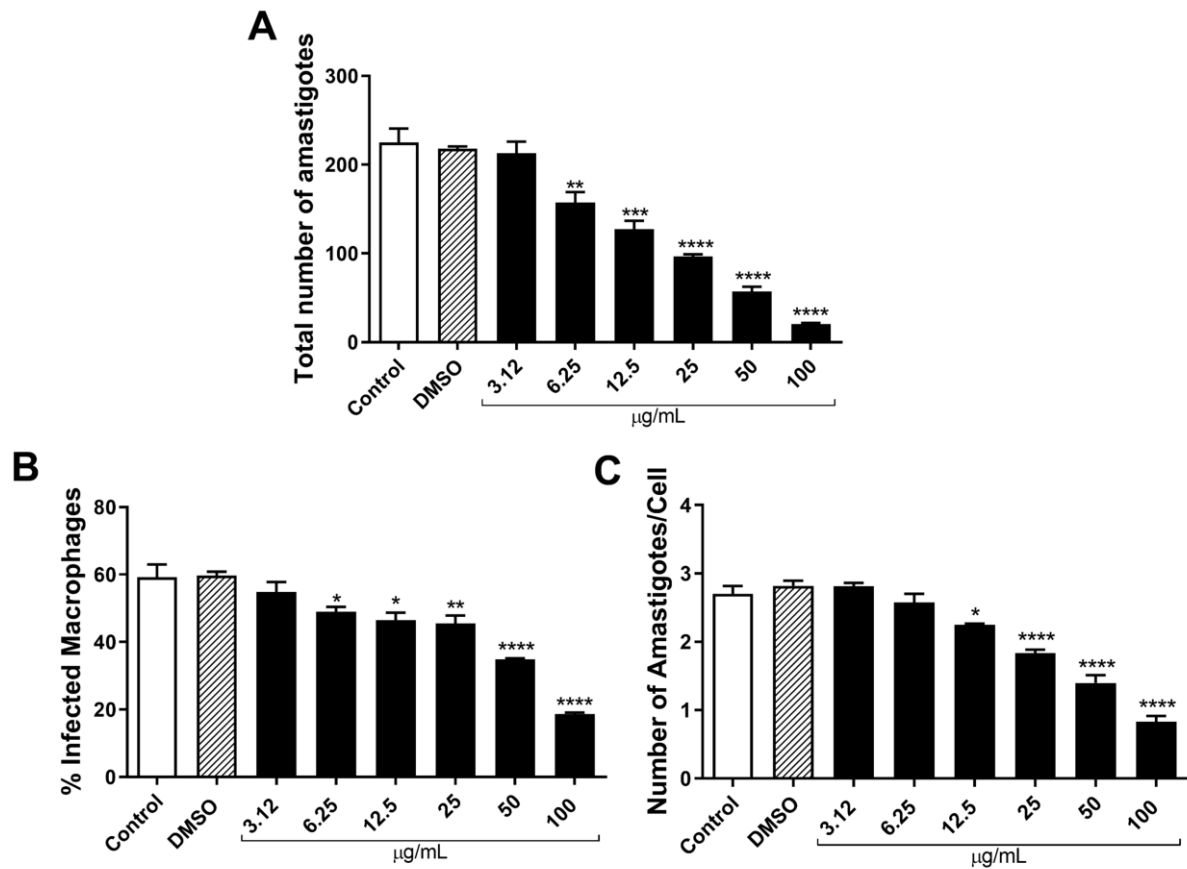
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Figure 3 - Morphological changes in promastigote forms of *L. amazonensis*. The parasites were treated with IC_{50p} (16 µg/mL) of OEO for 24 h at 25 °C and analyzed by SEM. (A-B) Untreated parasites. (C) Parasites were treated with 0.01% DMSO (vehicle). (D-I) Parasites were treated with OEO IC₅₀. Arrows indicate leakage of cellular content. Scale bar = 5 µm (A-H), 2 µm (I). (J) Length of the flagellum, cell body, and total size of promastigotes were measured from SEM images using the Image Pro-plus software. (K) Flow cytometry analysis where the FSC-A was considered a function of cell size. Data represent the mean ± SEM of three independent experiments performed in duplicate. *Significant difference in relation to control (p<0.05), ** (p≤0.01), *** (p≤0.001), **** (p≤0.0001).



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Figure 4 - OEO-induced death in *L. amazonensis* promastigote forms submitted to a 24 h treatment with IC_{50p} and 2xIC_{50p}. The following methods were used for the respective assessments: (A) DCF fluorescence for reactive species of oxygen measurement, (B) TMRE assay for fluorometric analysis of the mitochondrial membrane potential, (C) Nile Red labeling for neutral lipid droplets accumulation labeling, and (D) monodansylcadaverine for autophagic vacuoles formation, (E) annexin-V labeling for phosphatidylserine exposure detection and (F) propidium iodide for membrane permeabilization. AUF – arbitrary units of fluorescence. Data represent the mean ± SEM of three independent experiments performed in duplicate. *Significant difference in relation to control (p<0.05), ***(p≤0.001), *****(p≤0.0001).



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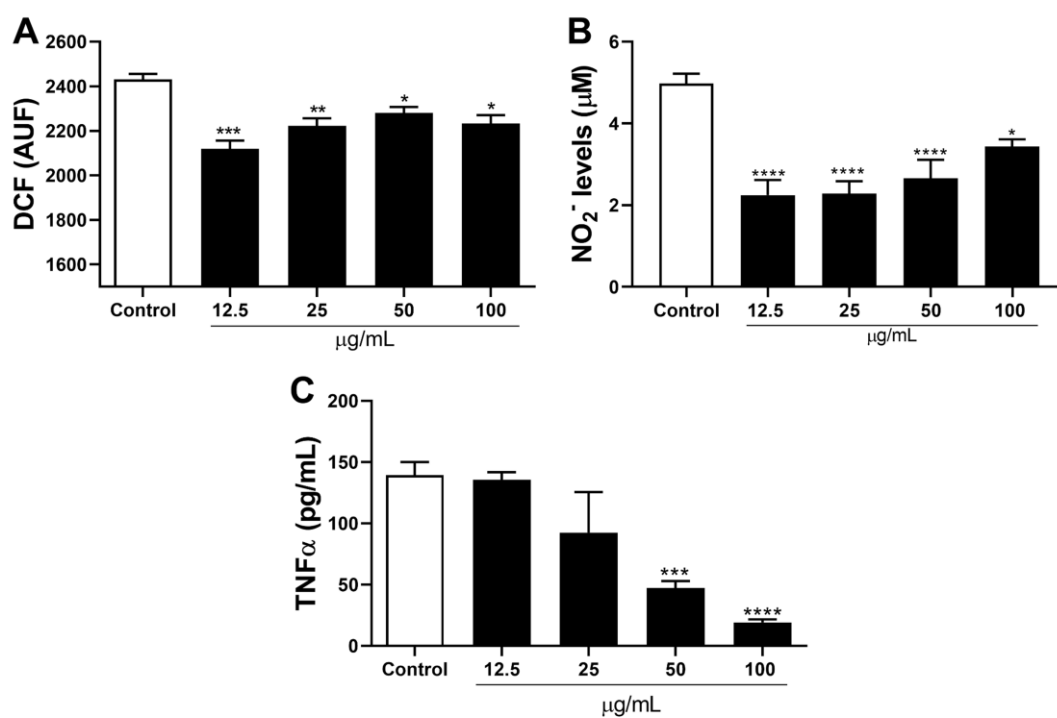
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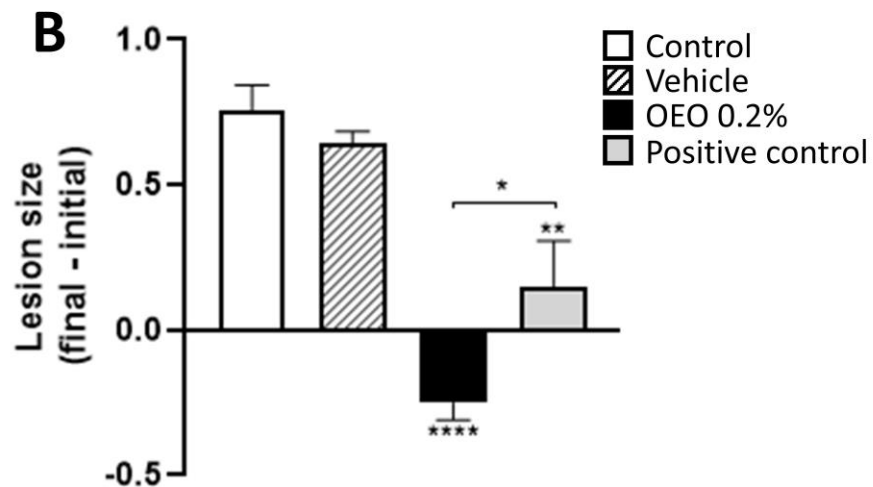
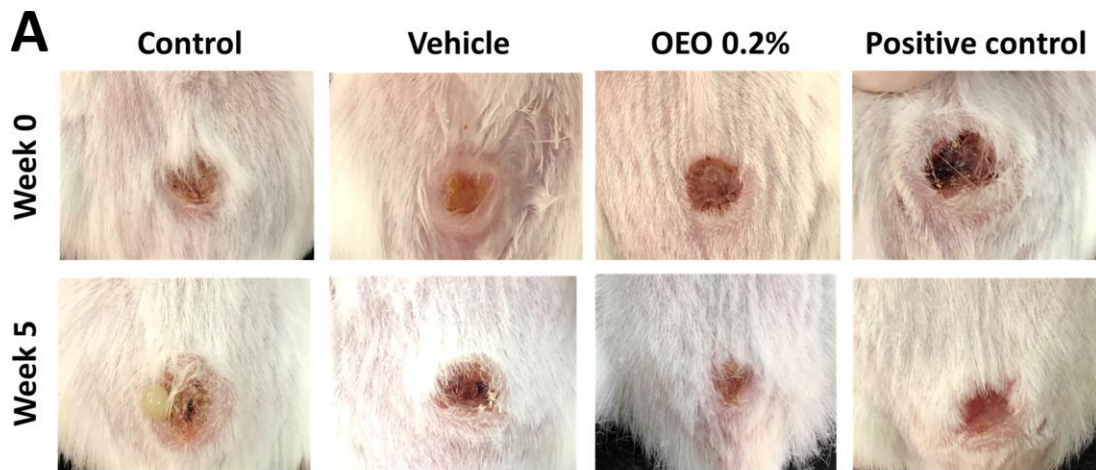
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Figure 5 - OEO-induced elimination of *L. amazonensis* intramacrophagic amastigote forms. *L. amazonensis*-infected peritoneal BALB/c macrophages were submitted to OEO treatment (3.12-100 µg/mL) or DMSO 0.01% (vehicle) and were assessed as the (A) total number of amastigote forms, (B) percentage of infected macrophages, and (C) number of amastigotes per macrophage. The culture medium was used as a control. The values represent the mean ± SEM of three independent experiments performed in duplicate. *Significant difference compared to the control ($p < 0.05$), **($p \leq 0.01$), ***($p \leq 0.001$), ****($p \leq 0.0001$).

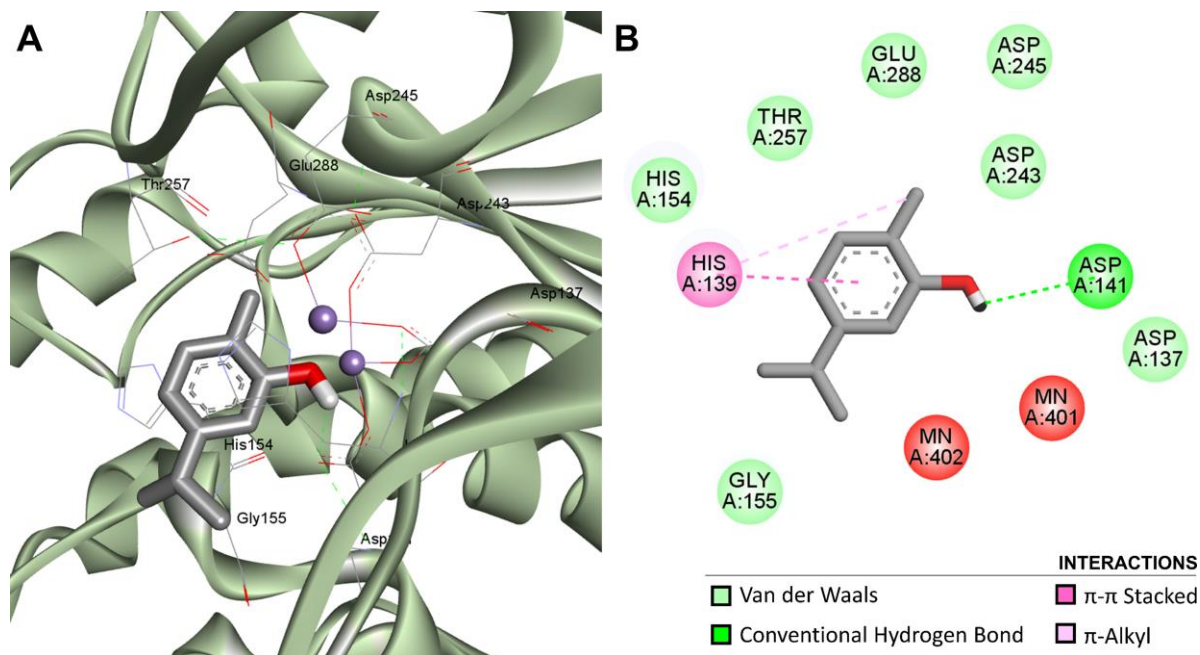


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 553 **Figure 6 - Intramacrophagic *L. amazonensis* death by OEO is not dependent on ROS, NO, or TNF- α .** The
 554 following methods were used for the respective assessments: (A) DCF fluorescence for reactive species of oxygen
 555 measurement, (B) Griess method for nitrite levels, and (C) ELISA for TNF- α measurement in the supernatant of
 556 culture cells. AUF – arbitrary units of fluorescence. Data represent the mean \pm SEM of three independent
 557 experiments performed in duplicate. *Significant difference in relation to control ($p < 0.05$), **($p \leq 0.01$),
 558 ***($p \leq 0.001$), ****($p \leq 0.0001$).
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Figure 7 – *In vivo* activity of OEO treatment. BALB/c mice were infected in the base of the tail with *L. amazonensis* promastigotes for 100 days. After, mice were treated daily topically for five weeks or untreated (control). (A) Representative photographs of lesions before (week 0) and after (week 5) the treatments period. (B) Lesion size measurements (the initial values were subtracted from the final values). The infected groups were defined as control (untreated mice), vehicle (treated topically with the base gel), positive control (glucantime i.p., 20 mg/kg/day), or OEO (treated topically with 0.2% OEO containing gel). Data represent the mean \pm SEM of lesion size for each group (n = 4). **Significant difference in relation to control and vehicle ($p \leq 0.01$), ***($p \leq 0.001$). *Significant difference between the treatments ($p < 0.05$).



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Figure 8 - Interaction diagrams of docking pose results of carvacrol in the active site of ARG (PDB ID: 4IU5). (A) Putative binding profile of carvacrol (gray stick model) toward ARG active site according to the best pose. Purple balls represent the Mn^{2+} . (B) 2D representation of residues interactions. The interactions are represented by dashed lines.

6.2.3. Capítulo de livro

Diterpenes from plants in the breast cancer treatment: new perspectives and recent advances

Capítulo desenvolvido para o livro *Studies in Natural Products Chemistry (Bioactive Natural Products)* da editora *Elsevier* (status: under review).

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All authors equally contributed to the manuscript.

DITERPENES FROM PLANTS IN THE BREAST CANCER TREATMENT: NEW PERSPECTIVES AND RECENT ADVANCES

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Abstract

A variety of plants have been a good source of therapeutic agents, mainly due to its secondary metabolites, which originate from the conversion of light energy into chemical compounds. Among these metabolites are the diterpenes, a group of structurally diverse molecules, widely distributed in nature. Diterpenes present a broad spectrum of biological activities and have been extensively studied by its antitumoral effects, with promising results. Breast cancer is one of the most common cancers worldwide and, varying from mild and potentially curable disease to incurable manifestations using the therapy available. Therefore, the search for new drugs is urgent and necessary. We will cover in this chapter a literature update and a review regarding anti-breast cancer effects of diterpenes from plants, approaching the works published in the main databases in the period 2015-2020. The structure, mechanism of action, molecular targets, and both *in silico*, *in vitro*, and *in vivo* effects are presented.

Key-words: Anti-cancer; Anti-tumor; Natural Compounds; Cell Lineage; Anti-Proliferative; Metastasis; Nanotechnology; Apoptosis.

1. Introduction

Plants are complex organisms that have developed mechanisms that allow them to adapt to various stresses without harm to its cellular and developmental physiological processes, such as protection against predators (due to their bitterness), attracting pollinators, establishing symbiosis, and providing structural components for lignification of cell walls of vascular tissues. The literature shows that secondary metabolites are a central adaptation mechanism in plants (1–3).

Secondary metabolites originate from the conversion of light energy into chemical compounds, although they do not play a significant role in primary life, they are vital for plants allowing their ability to respond not only to environmental stresses but also to protect against attack by pathogens, insects, and herbivores (4). Another particularity is that these metabolites may vary according to the family, genus, or species of the plant, thus, they can be used as taxonomic markers in the botanical classification (5).

Secondary metabolites from plants can provide a range of compounds with biological activity that justifies the growing interest in the development of new drugs (6). Worldwide, approximately 25% of the drugs used are derived from plants, however, there are still over 350,000 species that have not been studied (2). Therefore, this is an area still with great possibilities of expansion and discoveries for biomedical research.

The synthesis of secondary metabolites can be carried out by different biosynthetic pathways: via malonate acetate, mevalonic acid, methylerythritol phosphate, and shikimic acid, by which the three main groups of secondary metabolites formed are phenolic compounds (biosynthesized from the shikimate pathway, containing one or more aromatic hydroxylated rings), alkaloids (nitrogenous substances, in which most are biosynthesized from amino acids) and terpenes (polymeric isoprene derivatives, biosynthesized from acetate via mevalonic acid) (7). Other groups that deserve mention are fatty acid derivatives and aromatic polyketides.

Terpenes are a large and diverse group with a wide structural variety. Having more than 60,000 known compounds (8), these structures are synthesized from isoprenoid building blocks (C5), following the “isoprene rule”. The classification occurs according to the amount of isoprene in the molecule formation, such as hemiterpenes (C5), monoterpenes (C10), sesquiterpenes (C15), diterpenes (C20), sesterterpenes (C25), triterpenes (C30), and tetraterpenes (C40) (**Figure 1**) (9). The biosynthesis of terpenes in plants can occur by two pathways: via mevalonate (present in the cytosol), where it uses three units of acetyl-CoA, and via methylerythritol 4-phosphate (present in

plastids) which synthesizes isopentenyl diphosphate and dimethylallyl diphosphate in plastids (10,11).

The class of diterpenes is considered structurally diverse, widely distributed in nature, originating from the condensation of four isoprene units derived from the mevalonate or phosphate deoxyulose phosphate pathways. The latter, recently discovered, gives rise to diterpene compounds in plants. The diterpenes can be classified into different categories, the main ones being abietane, aconane, beyrane, cassiane, clerodane, gibberellane, kaurane, labdane, lathyrane, primarane, taxane, and trachylobane (**Figure 1**). In nature, they can be found in a polyoxygenated form with keto and hydroxyl groups, these are often esterified by small aliphatic or aromatic acids (12,13). Diterpenes are one of the groups most studied for their broad biological activities, such as antiprotozoal (14–16), antitumor (17–19), antioxidant (20–22), antimutagenic properties (23,24), antimicrobial (25,26). Diterpenes also have been extensively studied by their antitumoral effects, with promising results.

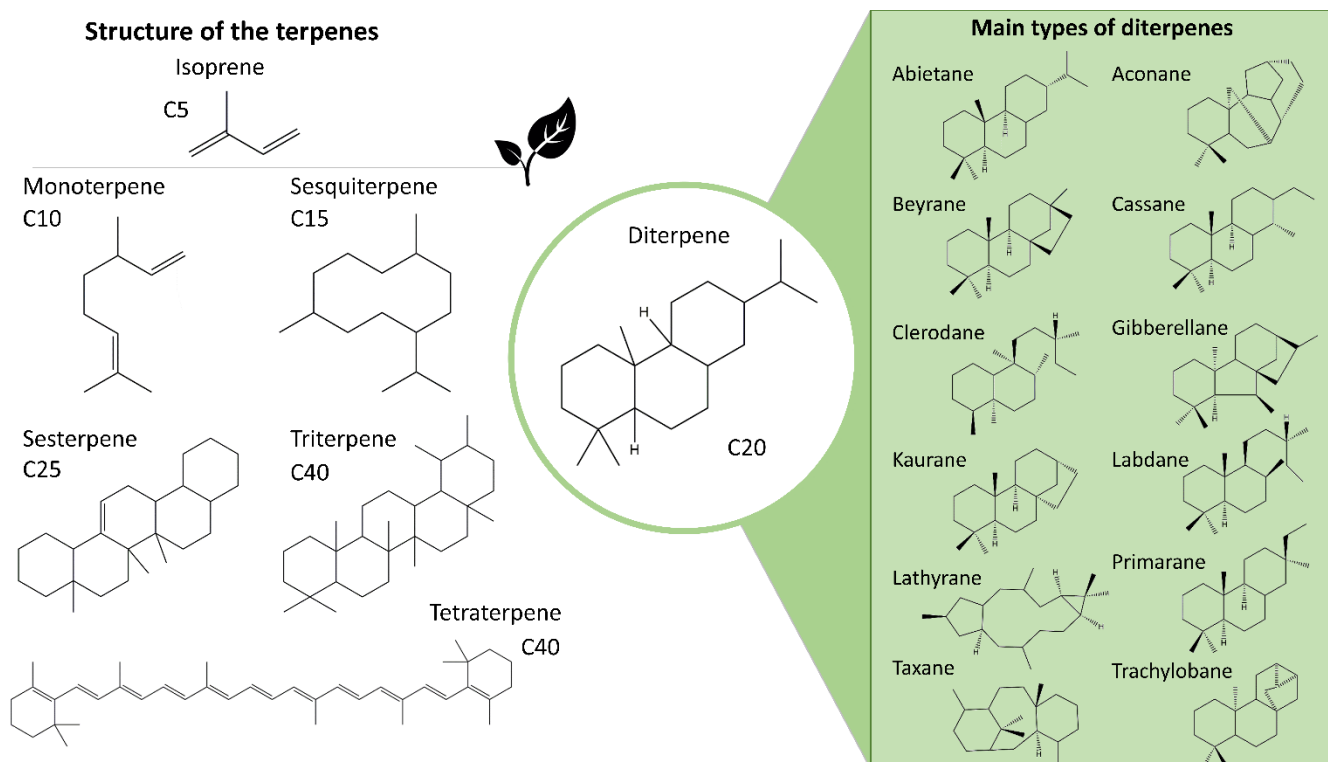


Figure 1 – Basic structure of terpenes. Terpenes are synthesized from isoprenoid building blocks (C5) and the molecules formed are classified according to the amount of isoprene in monoterpenes (C10), sesquiterpenes (C15), diterpenes (C20), sesterterpenes (C25), triterpenes (C30), and tetraterpenes (C40). The main types of diterpenes are abietane, aconane, beyrane, cassiane, clerodane, gibberellane, kaurane, labdane, lathyrane, primarane, taxane, and trachylobane.

The main example of a natural chemotherapeutic drug widely used for clinical treatment of ovarian cancer is Taxol ($C_{47}H_{51}NO_{14}$) (27). This tricyclic diterpenoid is

produced naturally in the bark and needles of *Taxus brevifolia*. It has a unique mechanism of action, promoting the assembly of tubulin in microtubules preventing the dissociation of microtubules, in addition to blocking the progression of the cell cycle, preventing mitosis by inhibiting the growth of cancer cells (28).

Breast cancer is one of the most common cancers worldwide and, depending on the time it is diagnosed, it can be early, which is considered potentially curable, and metastatic, which is considered incurable with available therapies (29). There are four main subtypes of breast cancer, that make up the majority of cases, being luminal A, which has expression of estrogen and progesterone receptors and has a slow growth rate; luminal B, which is also positive for hormone receptors, however, has a rapid growth rate; HER2 is the subtype that has a negative phenotype for hormone receptors, but has a high expression of HER2 receptors; and the fourth subtype is the triple-negative that has no expression of any of these three receptors, has an accelerated growth rate and is related to a worse prognosis. Thus, compounds that inhibit hormone-induced proliferation-inducing signaling may be effective against luminal A and B, but not against HER2 and triple-negative subtypes (30).

According to the biannual Advanced Breast Cancer Conference (ABC), systemic therapy is the first therapeutic choice in metastatic breast cancer, to promotes prolonged survival, maintaining the quality of life, mitigating symptoms, and still inhibit or reverse the development of cancer (30). Thus, new natural products that present cytotoxic activity applicable in cancer therapy are increasingly being researched (31). The main cell lines used to study breast cancer are MCF-7 (luminal A subtype); BT-474 (luminal B subtype); SKBR-3 and MDA-MD-435 (HER2 over-expression subtype); MDA-MB-231 and MCF-10 (basal subtype); and MCF-7/ADR (with multidrug resistance) (32), which will be discussed in this chapter.

According to the literature, the pharmaceutical oncology products research has 49% of medicines derived from or inspired by natural sources, such as microorganisms and marine organisms, and mainly plants (33). Thus, we will cover in this chapter updates the literature and reviews the anti-breast cancer effects of diterpenes, focusing on its chemical properties, approaching the works published in the main databases in the period 2015-2020.

2. Identification of new Diterpenes with anti-breast cancer activity

Knowing that the class of diterpenes provides compounds with important biological actions, many efforts have been concentrated on the discovery and biological characterization of new molecules of this class. This topic will address the new

diterpenes characterized by their anti-breast cancer action, the main findings are summarized in Table 1, and the detailed works are described below.

In the root bark of *Chrozophora oblongifolia*, was isolated new ent-trachylobane-3b-hydroperoxide (5) and four known compounds, ricinine (1), trimethoxy ellagic acid (2), dimethoxy ellagic acid (3), and aleurotic acid (4). The structures of these secondary metabolites were elucidated by using different spectroscopic techniques, ¹H NMR, ¹³C NMR, HSQC, HMBC, 1H-1H COSY, NOESY, HR-ESI-MS, EI-MS, and comparison with published data. Only the new compound 5 showed cytotoxicity against human breast cancer cells (MCF-7) with IC₅₀ of 24.53 μM, and human hepatocyte-derived carcinoma cells (Huh-7) with IC₅₀: 34.13 μM (34).

In the leaves of *Taiwania cryptomerioides* was isolated two uncommon C37 heterodimeric diterpenoids, taicrypnacids A (1) and B (2), and a known labdane-type diterpenoid (3). To define the structures, comprehensive spectroscopic analysis, chemical conversion, X-ray crystallography, and ECD data were realized. The new compounds 1 and 2 causes cytotoxicity in human breast cancer (MCF-7) (IC₅₀: 7.4 and 9.9 μM), osteosarcoma (U-2 OS) (IC₅₀: 8.4 and 9.7 μM), and human colon carcinoma (HCT-116) (IC₅₀: 7.5 and 7.8 μM) cell lines. According to the authors, the presence of the hydroxycarbonyl group can play a crucial role in cytotoxicity, unaffected by the configurational changes in C-7 and C-14' in 1 and 2. Also, compound 1 induces an increase in intracellular Ca²⁺ concentration, ROS level, changes in the endoplasmic reticulum (ER) and mitochondria, causing cell death by activating apoptosis and autophagy. Also, compound 1 showed lower cytotoxicity to noncancerous cell line MCF-10A (35).

C18-, C19- or C20-diterpenoid alkaloids were isolated from the rhizoma of *Aconitum japonicum*, in the total of 199 natural alkaloids and their derivatives isolated, 128 were non-toxic (IC₅₀ > 20 or 40 μM) and 50 alkaloids present moderate antiproliferative effects (IC₅₀ 10-40 μM) against five human tumor cell lines (A549, MDA-MB-231, MCF-7, KB, and MDR KB subline KB-VIN). The aconitine-type C19-diterpenoid alkaloids have been highlighted included 21 natural alkaloids, where 3 natural diterpenoid alkaloids exhibited cytotoxic activity against five human tumor cell lines A549, MDA-MB-231, MCF-7, KB, and MDR KB subline KB-VIN (Lipomesaconitine - 15 IC₅₀=17.2 ~ 21.5 μM; Lipoaconitine-16 IC₅₀= 13.7 ~ 20.3 μM; Lipomesaconitine - 17 IC₅₀: 6.0 ~ 7.3 μM) and 18 of them were inactive (IC₅₀> 20 or 40 μM), based on the results, the fatty acid ester at C-8 and the anisoyl group at C-14 found in 17 may be important to the cytotoxic activity of aconitine-type C19-diterpenoid alkaloids. (36).

Barbatin H (1), a new neo-clerodane diterpenoid, plus fifteen known analogs: scutebata P (2), scutebarbatine F (3), 6-O-nicotinoylscutebarbatine G (4), scutebata G

(5), scutebata E (6), scutebata D (7), barbatin C (8), scutebarbatine A (9), scutebarbatine G(10), scutebarbatine B (11), 6,7-di-O-acetoxybarbatin A (12), scutebata C (13), scutebata A (14), scutebarbatine X (15), and scutebata B (16) were isolated from *Scutellaria barbata*, a plant known as 'Ban zhi lian' in China. Their isolation and structures were elucidated by silica gel column, RPC18 CC, Sephadex LH-20, and preparative HPLC, characterized by NMR and HR-MS data compared to the literature. Most compounds present cytotoxicity against LoVo (colon cancer) cells. Scutebata A (14) showed significant cytotoxic activities against four tested tumor cells LoVo (IC₅₀: 4.57 μM), MCF-7 (IC₅₀: 7.68 μM), SMMC-7721 (hepatoma cancer) (IC₅₀: 5.31 μM) and HCT-116 (colon cancer) (IC₅₀: 6.23 μM). The authors suggest that the results presented by the compound, may be related to the presence of two benzoyloxy groups in C-6 and C-7, which may be closely associated with the strongest aromaticity in the benzene ring (37).

From the *Selaginella moellendorffii*, a traditional Chinese herb was isolated a new modified abietane diterpenoid (3S,4S,5R,10S)-18(4→3)-abeo-3,4,12,18-tetrahydroxy-8,11,13-abietatrien-7-one (1), and two novel dimers, selaginedorffones A (2) and B (3), featuring a new cyclohexene moiety. Their structural elucidation was realized by a combination of NMR spectroscopic analysis and ECD calculations. Selaginedorffone B (3) present cytotoxicity activity only against MCF-7 cells (IC₅₀ 9.0 μM), but it was inactive against SMMC-7721, SW480, HL-60, and A-549 cell lines. Also, compounds 1 and 2 were inactive against all five cancer cell lines used (38).

In the stems and twigs *Ceriops tagal*, a Chinese mangrove, four new diterpenes named tagalons A-D (1-4), comprising an isopimarane (1), two 16-nor-pimaranes (2-3), and a dolabranes (4), plus together with four known dolabranes containing a 4,18-epoxygroup, viz. tagalene I (5), 4-epitagalene I (6), tagalsin A (7), and tagalsin B (8). Their structure and the absolute configuration were elucidated by HR-ESIMS, NMR spectroscopic data and single-crystal X-ray diffraction analyses. The compound tagalene I (5) displayed potent cytotoxic effects against four human breast cancer cell lines MDA-MB-453 (IC₅₀ 8.97 μM), MDA-MB-231 (IC₅₀ 8.97 μM), SK-BR-3 (IC₅₀ 4.62 μM), and MT-1 (IC₅₀ 3.93 μM), while tagalons C (3) and D (4) exhibited cytotoxicity only against MT-1 cell line (IC₅₀ 3.75 and 8.07 μM) (39).

Taxus wallichiana var. *mairei* is a plant found southern area of the Yangtze River in China, and it was isolated a new, taxiwallinine (1), and 25 known taxane diterpenoids: taxinine J (2), 1-dehydroxy-baccatin VI (3), baccatin VI (4), 7-epi-10-deacetyltaxol (5), 10-deacetyltaxol B (6), 2α, 5α, 10β- triacetoxy-14β-(2-methyl)-butyryloxy-4 (20), 11-taxadiene (7), 2α, 5α, 10β, 14β-tetra-acetoxy-4 (20), 11-taxadiene (8), 2-deacetoxytaxinine J (9), taxinine E (10), 2α, 5α, 10β-triacetoxy-14β-propionyloxy-4 (20),

11- taxadiene (11), taxezopidine H (12), taxacin (13), 2 α , 5 α , 7 β , 9 α , 10 β , 13 α -hexaacetoxy-4 (20), 11-taxadiene (14), taxuspinanane K (15), taxayuntin C (16), 7, 9, 10, 13-tetra-O-deacetylabeobacatin VI (17), baccatin V (18), baccatin IV (19), 10-deacetyl taxuyun-nanine (20), taxuspine W (21), taxin B (22), 5-dehydrotaxin B (23), 2 α , 7 β , 13 α - triacetoxy, 5 α , 9 α -dihydroxy-2 (3 \rightarrow 20)-abeotaxa-4 (20), 11-dien-10-one (24), 1 β -hydroxybaccatin I (25) and wallifoliol (26). For the first time, the compounds 10, 12, 14, 15, 17, 21, 22, 23 were isolated from this plant. The structural elucidation was realized by HR-ESI-MS and NMR analyses. The taxiwallinine (1) and taxol has presented cytotoxicity against MCF-7 cell line (IC₅₀ 20.898 μ g/mL and 0.008 μ g/mL) (40).

In aerial parts of *Siegesbeckia pubescens*, widely distributed in China, five new diterpenoid glycosides were isolated: Siegesides A (1), B (2), C (3), D (4), E (5), along with the known compound darutoside (6). The structural elucidation was accomplished by extensive HR-ESI-MS and NMR analysis. Siegesides A (1) showed cytotoxicity activity against MB-MDA-231 breast cancer cells (IC₅₀ 0.27 μ M) and the other compounds showed weak or no effects (IC₅₀ >25 μ M). The presence of a 15S–16 vicinal diol was an effective group for the teaching of inhibitory activities. Also, the authors showed that the number, location, and orientation of hydroxyl groups may be essential for the inhibitory effects of diterpenoids against the invasion of MB-MDA-231 cells. (41).

In the aerial parts of *Salvia leriifolia* was isolated two new (1, 2) and two known (3, 4) labdane diterpenoids, together with three other knowns compounds. The structures, confirmation, and configuration were established by 1D and 2D NMR, HR-ESI-MS, single-crystal X-ray analysis, and electronic circular dichroism spectroscopy. Compound 4 showed cytotoxicity against MCF-7 (IC₅₀ 25 μ M), MDA-MB231 (IC₅₀ 50 μ M), and DU-145 (IC₅₀ 50 μ M) cells. Other compounds were considerably less effective in the tumoral cell lines (IC₅₀ > 10 μ M) (42).

Two strobane diterpenoids, strobols A (1) and B (2), 15 new pimarane diterpenoids (3–6 and 8–18), and the known compounds kirenol (19), darutigenol (20), and ent-2 β ,15,16,19-tetrahydroxypimar-8(14)-ene (7) were isolated in the aerial parts of *Siegesbeckia pubescens* (Makino). The structures and configurations were established based on the interpretation of HR-ESI-MS, NMR analysis, X-ray crystallographic data, and ECD calculations. Compounds 3, 5, and 11 inhibited the EGF-induced invasion of MB-MDA-231 cells (IC₅₀ 4.26, 3.45, and 9.70 μ M, respectively), while other compounds have not affected MB-MDA-231 cells (IC₅₀>10 μ M) (43).

Five fractions of the plant extract of *Plectranthus amboinicus* (Lour.) Spreng inhibited the viability of breast cancer MCF-7 cells, however, hexane fractions demonstrated high cytotoxic activity (average IC₅₀ 8.85 μ g/ml). HPLC-based

metabolomic approach revealed that an abietane diterpene namely 7-acetoxy-6-hydroxyroyleanone was the main compound that contributed to the cytotoxic activity (44).

For the first time in the twigs of *Podocarpus nagi* (Podocarpaceae) was isolated two new abietane-type diterpenoids, named 1 β ,16-dihydroxylambertic acid (1) and 3 β ,16-dihydroxylambertic acid (2), along with two new ent-pimarane-type diterpenoids, name dent-2 β ,15,16,18-tetrahydroxypimar-8(14)-ene (3) and ent-15-oxo-2 β ,16,18-trihydroxypimar-8(14)-ene (4). Their structures were elucidated based on spectroscopic analyses, including 1D- and 2D-NMR, IR, CD, and HR-ESI-MS. Compounds 3 and 4 at 10 μ M inhibited the proliferation of human cervical cancer HeLa cells, human lung cancer A549 cells, and human breast cancer MCF-7 cells. Also, compounds 2 and 3 inhibit NO production in LPS-stimulated RAW264.7 cells (45).

In the roots of *Euphorbia fischeriana* was isolated 23 diterpenoids (1–23), classified into five subtypes, namely, ent-rosane (1–9), ent-abietane (10–16), ent-kaurane (17), ingenane (18–22), and lathyrane (23). The chemical structures of eight new compounds, namely, euphorin A (4), B (5), C (7), D (9), E (13), F (14), G (15), and H (16) were elucidated based on extensive 1D and 2D NMR spectroscopic analyses, as well as single-crystal X-ray structure analysis. The compounds (2, 6, 7, 10, 11, 13, 16, 19, 20, 22, and 23) showed inhibitory activity on the formation of mammospheres in human breast cancer MCF-7 cells at a final concentration 10 μ M. The authors suggested that an aromatic A-ring in the isolated compounds may be beneficial for the activity of ent-rosanes, and the acylation may be important for the activity of ingenanes. However, details about the action of these bioactive diterpenoids are still under investigation (46).

In *Azorella compacta* (Apiaceae), a native Chilean cushion shrub, diterpenoids were fractionated from its resin by high-speed counter-current chromatography (HSCCC). The five major compounds isolated were identified through spectroscopy as diterpenoids azurellanol (7), 13 α ,14 α -dihydroxymulin-11-en-20-oic acid (9), mulinolic acid (10), mulin-11,13-dien-20-oic acid (11), 517-acetoxy-mulinic acid (13), and new diterpenoids characterized as 13 β ,14 β -dihydroxymulin-11-en-20-oic acid (1), 13-epi-azurellanone (2) and 13-epi-7-deacetyl-azurellanol (3), and another four minor diterpenoids as 7-deacetyl-azurellanol (6), 13-epi-azurellanol (4), 7-acetoxy-mulin-9,12-diene (8), and 17-acetoxy-mulin-11,13-dien-20-oic (12). The structures of the new compounds were elucidated by MS, 1D–2D NMR, and molecular modeling techniques. Compounds 4, 6, 7, 8, and 11 showed good cytotoxic activity at 100 μ M against MCF-7 cells. However, compound 7, was considered the most effective (IC₅₀ 25.64 μ M). Among the azurellanes, a necessary feature for its activity is the beta acetate group at position C-7 in the skeleton for the increase of the cytotoxic effect. Besides, the alpha position of the OH group at position C-13 in the same skeleton further increases the cytotoxic effect when comparing

the cytotoxic effect of the epimers 4(OH-17 in beta position) and 7(OH-17 in alpha position (47).

In the tuber of *Icacina trichantha* were isolated seven new 17-norpimarane and (9 β H)-17-norpimarane diterpenoids, icacinlactones A–G (1–7). The structures were elucidated by spectroscopic and HRMS techniques, and the absolute configuration was determined through X-ray crystallographic analysis. The known structures of icacinol, humirianthol, humirianthenolide C, and icacenone exhibited cytotoxicity, however, humirianthenolide C was the most active against MDA-MB-435 (IC₅₀ 0.7 μ M) and MDA-MB-231 cells (IC₅₀ 0.7 μ M) and the new icacinlactone F (6) was moderately active against MDA-MB-435 (IC₅₀ 6.16 μ M), MDA-MB-231 (IC₅₀ 8.94 μ M), and OVCAR3 (IC₅₀ 10.50 μ M) cancer cell lines (48).

In the *Casearia graveolens* twigs was isolated a new clerodane diterpene, caseariagraveolin (1), together with six known compounds casearlucin G (2), casearvestrin C (3), casearvestrin B (4), caseargrewiin L (5), casearvestrin A (6) and caseamembrol B (7). Their structures were elucidated by 1H and 13C NMR, 1H-1H COSY, HMQC, HMBC, 2D NOESY, and mass spectra data. Caseariagraveolin (1) showed cytotoxicity against oral cavity KB (IC₅₀ 2.48 μ M) and breast cancer MCF-7 (IC₅₀ 6.63 μ M) cell lines (49).

Phytochemical investigation of the leaves of *Casearia grewiifolia* was isolated two new clerodane diterpenoids named 1 and 2 and the known compound caseanigrescen D (3). Caseagrewifolin B (2) had inhibitory activity against cancer cell lines as KB (mouth epidermal carcinoma) and HepG-2 (human liver hepatocellular carcinoma) and exhibited a significant selective inhibition in comparison with normal cells (NIH/3T3), but no effect against MCF-7 cells, while caseanigrescen D (3) was cytotoxic against four cancer cell lines KB, HepG-2, LU-1 (human lung adenocarcinoma), and also MCF-7, however, it had no selective inhibition compared with normal cells (50).

Most of the new diterpenes isolated by plants have good cytotoxicity against breast cancer cell lines, with IC₅₀ ranging from 0.23 to 25.64 μ M, indicating a good potency. However, some new diterpenes, despite being unprecedented, were not effective against cancer cell lines.

Is the case of the study performed by He et al. (2020), which has proposed isolation of two new diterpenoid alkaloids: delphatisine D (1), chrysotrichumine A (2), and ten known diterpenoid alkaloids: sharwuphinine A (3), delphatisine A (4), lycoctonine (5), davidisine B (6), delsemine A (7), delavaine A (8), isodelpheline (9), delpheline (10), delcorine (11), del-brusine (12) and 3 β ,6 α -dihydroxysclareolida (13), isolated from *Delphinium chrysotrichum*, a plant native to China. For the first time, a sesquiterpene compound 13 was isolated from this genus. The isolation, structure, and molecular

formula were elucidated by spectroscopic analysis as HRESIM data, ^1H , ^{13}C NMR, DEPT, HSQC spectra, ^1H - ^1H COSY, and HMBC spectrum. However, none of the new compounds present cytotoxicity against human breast cancer cell lines of MCF-7 and MDA-MB-23, and among the known ones, only compound 7 has activity against MDA-MB-23 at 100 μM (51).

In the same way, eighteen new cembrane-type diterpenoids named Boscartins (1–18) and eight known analogs isolated from *Boswellia carterii* didn't show cytotoxicity activity ($\text{IC}_{50} > 10 \mu\text{M}$) against five human tumor cell lines: HCT-8 (human ileocecal adenocarcinoma), Bel-7402 (human hepatoma), BGC-823 (human gastric cancer), A549 (human lung epithelial), and A2780 (human ovarian cancer) (52).

From the plant, *Salvia sahendica* was isolated six compounds: as a new norditerpene (1), and known terpenoids, sclareol (2), oleanolic acid (3), β -sitosterol (4), salvigenin (5) and 3 α -hydroxy-11 α ,12 α -epoxyoleanan-28,13 β -olide (6). The inedited compound showed no toxicity, but sclareol (2) and toxol revealed a moderate cytotoxic activity against the MDA-MB-231 cancer cell line (IC_{50} 81.0 and 0.23 μM) (53).

Similarly, from *Akebia quinata*, one new diterpene glycoside (1) and 13 known compounds (2–14) were isolated. Compounds cyrtophyllones B (2), uncinatone (3), 3-O- α -l-arabinopyranosyl olean-12-en-28-oic acid (6), 3-O-[β -d-glucopyranosyl(1-4)- α -l-arabinopyranosyl]olean-12-en-28-oic acid (8), and 2 α ,3 α ,23-trihydroxyoleane-12-en-28-oic acid (11) showed inhibitory effects on the PTP1B enzyme, and concentration-dependent cytotoxic effects mainly by inducing cell apoptosis on breast cancer cell lines, such as MCF7, MDA-MB-231, and tamoxifen-resistant MCF7 (MCF7/TAMR) (IC_{50} ranging from 0.84 to 7.91 μM), but the new compound did not affect breast cancer cell lines. The presence of a COOH group at C-28 in triterpene compounds (6, 8, and 11) showed a significant increase in inhibitory activity against the PTP1B enzyme. Also, compound 6 showed increased activity due to having one less glucopyranosyl, which is linked to arabinopyranosyl C-3'. The presence of an alcohol group at C-23 may decrease the inhibitory effects on PTP1B. (54).

Finally, eight new diterpenoids were isolated from *Kaempferia pulchra* of Myanmar: kaempulchraols A–H, and five known analogues: 9 α -hydroxyisopimara-8, 15-dien-7-one, 117 β ,9 α -dihydroxypimara-8, 15-diene, 121 α ,11 α -dihydroxypimara-8, 15-diene, 13 sandar-acopimaradien-1 α ,2 α -diol, and (2R)-ent-2-hydroxyisopi-mara-8, 15-diene. None of the isolated compounds present antiproliferative activity against A549, HeLa, PANC-1, PSN-1, and MDA-MB-23 human cancer cell lines, except Kaempulchraol F, that exhibited activity against the human pancreatic PSN-1 cell line (IC_{50} 12.3 μM) (55).

Table 1 – The new identified diterpenes with anti-breast cancer activity.

New diterpen	Plant source	Cell line	IC ₅₀	Ref
Delsemine A	<i>Delphinium chrysotrichum</i>	MDA-MB-23	ND	(51)
Ent-trachylobane-3b-hydroperoxide	<i>Chrozophora oblongifolia</i>	MCF-7	24.53 μ M	(34)
Taicrypnacids A and B	<i>Taiwania cryptomerioides</i>	MCF-7	9.9 μ M	(35)
Lipomesaconitine	<i>Aconitum japonicum</i>	MDA-MB-231 and MCF-7	20 and 19 μ M	(36)
Lipoaconitine			15.5 and 16 μ M	
Lipomesaconitine			6 and 6.7 μ M	
Scutebata A	<i>Scutellaria barbata</i>	LoVo, MCF-7, SMMC-7721, and HCT-116	4.57 μ M, 7.68 μ M, 5.31 μ M, and 6.23 μ M	(37)
Selaginedorffone B	<i>Selaginella moellendorffii</i>	MCF-7	9.0 μ M	(38)
Tagalene I	<i>Ceriops tagal</i>	MDA-MB-453 and MDA-MB-231	8.97 μ M	(39)
Taxiwallinine	<i>Taxus wallichiana var. mairei</i>	MCF-7	20.89 μ g/mL	(40)
Siegesides A	<i>Siegesbeckia pubescens</i>	MB-MDA-231	0.27 μ M	(41)
Pimarane diterpenoid 3	<i>Siegesbeckia pubescens</i>	MB-MDA-231	4.26 μ M	(43)
Pimarane diterpenoid 5			3.45 μ M	
Pimarane diterpenoid 11			9.7 μ M	
15,16,18-tetrahydroypimar-8(14)-ene	<i>Podocarpus nagi</i>	MCF-7	ND	(45)
Ent-15-oxo-2 β ,16,18-trihydroypimar-8(14)-ene			ND	
Euphorin C, E, and H	<i>Euphorbia fischeriana</i>	MCF-7	ND	(46)
Azorellanol	<i>Azorella compacta</i>	MCF-7	25.64 μ M	(47)
Humirianthenolide C	<i>Icacina trichantha</i>	MDA-MB-435 and MDA-MB-231	0.7 μ M	(48)

Icacinlactone F		MDA-MB-435 and MDA-MB-231	6.16 and 8.94 μ M	
Casearia graveolin	<i>Casearia graveolens</i>	MCF-7	6.63 μ M	(49)

IC₅₀: inhibitory concentration for 50% of cells, ND: non-determined.

3. Diterpenes with antiproliferative activity

The chemoprevention of breast cancer is based on chemical compounds capable of inhibiting or reversing the proliferation of cancer cells, thus, new natural products that obtain antiproliferative activity applicable in cancer therapy are increasingly being researched (31).

In the study by Lu et al., (2016) (56), it was observed that when treating two human breast cancer cell lines (AS-B145 and BT-474) with ovatodiolide (Ova) (extracted from *Anisomeles indica* (L.) Kuntze), the compound showed a reduction in the proliferation of these cells, obtaining IC₅₀ values of 6.55 and 4.80 μ M, respectively. In addition, the authors, to enrich and analyze the self-renewal capacity of these strains, used a cluster of mammary tumor cells with stem/progenitor (mammosphere) properties and tested with the same diterpene. They observed that Ova was able to reduce the self-renewal property, to inhibit the expression of stemness genes [including octamer-binding transcription factor 4 (Oct4), Nanog and heat shock protein 27 (Hsp27) and regulate positively the regulatory factor 2 of ubiquitin SMAD (SMURF2). Such data reveal the potential of Ova in the treatment of breast cancer.

In 2017, Win et al., (2017) (57) after isolating 17 bicyclic diterpenoids from the *Curcuma amada* evaluated their antiproliferative activities against the lines MCF-7, HeLa, A549, PANC-1, and PSN-1 (human pancreatic cancer). In the present investigation, compounds 1 and 6 were not effective with any lineage, while compounds 2-4, 7, 8, 12, and 17 were found to be moderately active against the tested cells. Interestingly, (E)-14-hydroxy-15-norlabda-8(17),12-dien-16-al (11) selectively inhibited the proliferation of HeLa, PSN-1 and MCF-7 cells. Structure-activity relationship analyses against PANC-1 and PSN-1 cells suggested that the hydroxymethyl functionality at C-13 might be important to inhibit the proliferation of human pancreatic cancer cell lines, and may also be responsible for the effect on MCF-7 cells.

Triple-negative breast cancers (TNBCs) are considered irresponsible to current therapies because they are more aggressive, have early recurrence and metastases. Thus, Varghese et al., (2018) (58) when evaluating the antiproliferative activity of triptolide, isolated from *Tripterigium wilfordii*, on MDA-MB-231, MDA-MB-468, and MCF-

7, found that the compound was able to inhibit growth, being more evident in MDA-MB-231 after 72 hours of treatment ($IC_{50} > 0.3$ nM). Besides, there was a halt in the cell cycle in the sub-G0/G1 phase and a reduction in G0/G1, an increase in autophagy, and the treatment activated the caspase 3-dependent apoptotic pathway, thus promoting the death of this cell.

In the same year, Awasthee et al., (2018) (59) demonstrated that bharangin, a diterpene isolated from *Pigmacopremna herbacea*, also reduced the proliferation of MCF-7, MDA-MB-231, MDA-MB-453, MDA-MB-468, and T-47D cells. Also, treatment with this diterpene inhibited the migration and invasion of MDA-MB-231, altered the potential of the mitochondrial membrane, reduced proteins associated with cell survival (Bcl-2, Bcl-xL, and survivin), increased levels of Bax (pro-apoptotic) and promoted death by apoptosis, mediated by an increase in caspase-7 and 9. Also, the authors showed that bharangin acts in the MDA-MB-231 line by suppressing the activation of NF- κ B and modulates the expression of lncRNAs (long non-coding RNA), a class of non-coding-protein transcripts, which aberrant expression can act as an oncogene.

In the same way, triptolide was investigated for its antiproliferative effects on tumor lines MCF-7 and MDA-MB-231, as well as for its antitumor action on BALB/c-nu+/nu+ female mice. *In vitro*, the tested compound has been shown to exert potent effects in inhibiting cell viability, mediated through the attenuation and overexpression of ER α in MCF-7 and MDA-MB-231, respectively. In the *in vivo* study, the mice xenografted with MCF-7 and MDA-MB-231 were able to reduce the weight and volume of the tumor, being more evident in MCF-7 (60).

Parvifloron F, a highly oxidized bioactive abietane diterpene, and its synthetic intermediates were studied on MDA-MB-231 and MCF-7 cells, besides A549, nasopharyngeal squamous cell carcinoma (KB) and underlines KB MDR overexpressing P glycoprotein (KB-VIN). Some of the molecules, including parvifloron F, presented good antiproliferative effects. The compounds containing methoxy groups exhibited moderate activity on the cell lines, and that the stereochemistry of molecules did not influence the activity. On the other side, the oxidation level of the abietane ring seemed to be important for the antiproliferative and selective activity of the active compounds (61).

4. Inhibition of tumor angiogenesis and metastasis by diterpenes

Metastasis is an important prognostic factor in breast cancer since around 90% of deaths are not associated with the primary tumor, but with metastasis to distant sites (62). Studies indicate that the cell adhesion to the extracellular matrix (ECM), the structural and functional changes in epithelial cells that permit their migration through the basement membrane, called epithelial-mesenchymal transition, and angiogenesis are

the main mechanisms involved in breast cancer metastasis. The angiogenesis is directly related to the availability of vascular endothelium growth factor (VEGF), which is essential for tumor neovascularization. Therefore, screening for compounds with the ability to control breast cancer metastasis is essential for improving breast cancer treatment (63). Some diterpenoids from plants have been studied about its capacity to inhibit the angiogenesis in breast cancer models, with promising results.

In this sense, Sun et al., (2015) (64) evaluated the anti-metastatic effect of jolkinolide B (JB), a compound isolated from *Euphorbia fischeriana* Steud, on MDA-MB-231, U251 (glioblastoma multiforme), DU145 (prostate cancer), MGC803 (gastric mucinous adenocarcinoma), K562 (erythroleukemic), A549 (lung carcinoma), Bcap37 (endocervical adenocarcinoma), and ACHN (Papillary renal cell carcinoma) cell lines. The authors demonstrated that the higher expression of β 1-integrin (molecules that mediate tumor cell adhesion to ECM) and FAK (focal adhesion kinase) were more expressed in MDA-MB-231, however, after treatment with JB, cell adhesion and invasion were inhibited, in addition to positively regulating the expression of β 1-integrin protein and blocking FAK phosphorylation. Thus, these data contribute to the development of new strategies for the treatment of breast cancer metastasis.

In the same way, Carnosol, a phenolic diterpene extracted from culinary herbs such as oregano and rosemary, presented *in vitro* antimetastatic action, mitigating the migration and invasion of breast cancer cells MDA-MB-231 by downregulating activity and reducing metalloproteinase 9 expressions. Also, carnosol is still able to inhibit the STAT3 signaling pathway, as it induces its proteasomal degradation in a manner dependent on reactive oxygen species. Thus, the reduction in both the activity of metalloproteinase 9 and STAT3 *in vitro* implies that carnosol may have a promising effect in reducing the degradation of the extracellular matrix, inhibiting the metastasis of breast cancers. Besides, carnosol-induced ROS production in breast cancer cells culminates in cell death from apoptosis and autophagy and is therefore a promising compound for the development of new therapeutic approaches in breast cancer (65).

Cembranoids are carbocyclic diterpenes widely present in the plants belonging to the genera *Nicotiana* (44). The cembranoid 1S,2E,4S,6R,7E,11E)-2,7,11-cembratriene-4,6-diol (CDO) isolated from *Nicotiana tabacum* presented significantly reduced VEGFR2 levels in TNBC MDA-MB-231 and BT-474 breast cancer cell lines and showed anti-migratory activity against the TNBC MDA-MB-231. *In vivo*, the intraperitoneal treatment of CDO (40 mg/kg, 3 times/week) significantly reduced the MDA-MB-231 cells breast tumor size in athymic nude mice, with significant downregulation of the vasculogenesis marker CD31 and suppression of VEGFR2-paxillin-FAK pathway. Similar results were found in the swiss albino mice-Matrigel model, which mimic the *in vivo* capillary formation

process, confirming the antiangiogenic potential of CDO. Chemically, CDO consists of 14-membered macrocyclic diterpene skeleton with hydroxyl, methyl, and isopropyl substituents. The docking studies showed that C-4 and C-6 hydroxyl groups are responsible for potent interactions with different amino acids of VEGFR2 (1Y6A and 3CJF). The C-19 and C-20 methyl groups of CDO also promoted favorable interactions with the predicted protein, promoting its inhibition (66).

Oridonin, an active diterpenoid compound isolated from *Rabdosia rubescens*, induced a significant reduction of the migration, invasion, and adhesion of MDA-MB-231 and 4T1 breast cancer cells, and inhibition of tube formation of human umbilical vein endothelial cells in a dose-dependent manner. Oridonin also caused increases in the expression levels of E-cadherin, and decreased expression of N-cadherin, vimentin, and snail, inhibiting the epithelial-mesenchymal transition. Besides that, oridonin exerted its anti-angiogenesis activity through significantly decreasing HIF-1 α , VEGF-A, and VEGF receptor-2 protein expression. Furthermore, oridonin was demonstrated to decrease the micro-vessel density as evidenced by the decreased expression of cluster of differentiation 31, a marker for neovasculature. Thus, oridonin seems to be a good candidate for new studies, since it presented important anti-metastatic features (62).

Zebrafish model was used to study the effect against vein formation of eriocalyxin B (EriB), a natural *ent*-kaurane diterpenoid isolated from *Isodon eriocalyx var. laxiflora*. EriB induced alteration of several angiogenic genes, inhibiting the subintestinal vein formation in zebrafish embryos. In human umbilical vein endothelial cells (UVEC), EriB treatment (50, 100 nM) inhibited the VEGF-induced cell proliferation via suppression of VEGFR-2-mediated downstream signaling pathway, tube formation, cell migration, and cell invasion, together with G1 arrest via the down-regulation of p21-cyclin D1/CDK4-pRb pathway. The *in vivo* C57 mice-Matrigel model showed the further anti-angiogenic effect of EriB, with a reduction of blood vessels formation. Lastly, in BALB/c mouse 4T1 breast tumor model proved that EriB (5 mg/kg/day via intraperitoneal for 21 days) presented anti-tumor and anti-angiogenic activities by reduction on tumor weight, cell proliferation, cell growth, and microvessel density. Also, molecular docking studies showed that EriB could stably bind at the ATP-binding site of VEGFR-2, with seven amino acids actively involved. The findings indicated that the interactions of EriB to VEGFR-2 resulted in the competitive inhibition to ATP against VEGFR-2, leading to the prevention of VEGFR-2 phosphorylation. Therefore, EriB exhibited significant anti-angiogenesis activity through the modulation of the VEGFR-2 mediated signaling pathway both *in silico*, *in vitro*, and *in vivo* (67).

The diterpenoid lactone andrographolide (andro) isolated from *Andrographis paniculata* presented an inhibitory effect against MDA-MB-231, BT-549, MCF-7, MDA-

MB-361, and T47D cell lines. When investigated the mechanism of action of this diterpenoid it was found induction of apoptosis in both MDA-MB-231 and MCF-7 cells by activation of cytochrome c and caspase-dependent apoptotic signaling pathway. Andro treatment *in vitro* also significantly downregulates the COX-2 expression and NF- κ B signaling in MDA-MB-231 cells, and could significantly inhibit the migration, invasion, and tubulogenesis of HUVECs *in vitro*. To confirm its anti-angiogenic activity, human breast cancer xenograft nude mice model was established and the treatment (5 and 10 mg/kg) by intraperitoneal injection every 2 days for 15 days induced an important reduction of the tumor volume and weight, with concomitant reduction of COX-2, microvessel number and expression levels of CD31, when compared with the control group. Therefore, these results may indicate that andro inhibited breast cancer growth and tumor angiogenesis *in vivo* (68).

5. Induction of cell death mechanisms by diterpenes

Regulated cell death is a biological phenomenon resulting from natural homeostatic processes that replace senescent and damaged cells or as a result of diseases (69). On the other hand, the deregulation of the machinery of control and induction of cell death can allow cells with genomic lesions to survive, which initiates the process of carcinogenesis and permit its uncontrolled proliferation. Anticancer therapy is based on the principle of inhibiting cell proliferation and blocking / stimulating signaling pathways that culminate in the death of these aberrant cells (70). Thus, compounds with the ability to selectively induce the death of cancer cells are potential candidates for oncotherapy.

However, the high clinical, morphological, and biological heterogeneity of cancers makes the process of generating a new treatment extremely complex and laborious. Therefore, understanding how new compounds interact with cell signaling and the mechanism by which it induces death in these cells is fundamental (71).

A diterpenoid glycoside compound 6E,0E,14Z-(3S)-17-hydroxygeranylinalool-17-O- β -d-glucopyranosyl-(1 \rightarrow 2)-[α -l-rhamnopyranosyl-(1 \rightarrow 6)]- β -d-glucopyranoside isolated from methanol extract of leaves *Blumea lacera* showed *in vitro* cytotoxic activity against two types of human breast cancer cell lines MCF-7 and MDA-MB-231 and the human stomach adenocarcinoma cell line AGS. Moreover, the compound showed low toxicity in healthy cell lines, such as NIH3T3 and VERO. In this study, the absence of the β -d-glucopyranose on the C-3 oxygen contributed to the significantly cytotoxic activity detected for the compound (72).

Parvifloron D (ParvD), 6,7-dehydroroleanone (DeRoy,1), and 7-acetoxy-6-hydroxyroleanone (Roy, 2) are abietan diterpenoid compounds found mostly in plants

of the genus *Plectranthus*, that have been promising in the *in vitro* treatment of breast cancer cells MCF-7, SkBr3, SUM159, and SUM159 sphere. Also, the *in silico* analysis showed that 1-3 was predicted as isozyme-selective PKC binders (73). PKC family has been intimately associated with proliferation, migration, differentiation, invasion, tumorigenesis, metastasis, and apoptosis of cancer cells (74). Isca et al. 2020 showed that changes in the binding site of PKC isoform change the predicted interaction profiles of the ligands 1-3. Also, minor changes in 1-3 substitution patterns, such as a double substitution only with non-substituted phenyls, or hydroxybenzoate at position four that flips the binding mode, can increase the predicted interactions in certain PKC isoforms. Compound 3 presented the best-calculated interaction profile, given its good interactions of the core structure and amino acid residues in the PKC binding site, as well as experimental inhibition against all cell types, including the most aggressive form. Also, the authors found that substitution groups of the royleanone core (1 and 2), such as hydroxybenzoate on position 4, can provide a better docked binding pose, and they also suggested that further modification in the 1,1-methyl groups can improve its interaction with PKC residues (73). Corroborating with these data, another work showed that compound 1 still can induce apoptosis and arrest the cell cycle in the sub-G1 phase, in addition to reducing the invasive and migratory capacity of MDA-MB-231 cells (75).

Similarly, carnosic acid, a phenolic diterpene found in culinary herbs such as rosemary, has antiproliferative activity in MCF-7 breast cancer cell lines. When used concomitantly with tamoxifen, a standard compound used to treat estrogen receptor-positive breast cancer, carnosic acid was able to induce caspase-3-mediated apoptosis and downregulate the production of anti-apoptotic proteins like Bcl-xl and Bcl-2 and induce pro-apoptotic signals from Bax and Bad. Further, the carnosic acid treatment induced an increase in the activity of the p53 protein, which plays a central role in controlling the replication of aberrant cells, and tumor necrosis factor-related apoptosis-inducing ligand (TRAIL), which can quickly induce apoptosis in different types of cancer. Thus, the pro-apoptotic activity of carnosic acid is related to the induction of the TRAIL / p53 axis and activation of mitochondrial signaling that induces cell death. It is worth highlight that although *in vitro* monotherapy with carnosic acid induces apoptosis and activates death signaling molecules, tamoxifen co-treatment showed more marked and promising results. Besides, concomitant treatment of carnosic acid and tamoxifen inhibited tumor growth *in vivo* of xenographic models of breast cancer with apoptosis induction without generating toxicity (76).

Traditional Chinese medicine is widely used to treat cancer in China. In the herbs used in this type of treatment, oridonin (7,20-epoxy-ent-kauranes) is found in abundance and, therefore, several studies have used it in the development of new therapies for

cancer. About breast cancer, oridonin and oridonine phosphate (from Shifeng biocorporation) were able to inhibit the proliferation of cells of MDA-MB-436 and MDA-MB-231 lineages. Moreover, oridonine phosphate induced apoptosis of breast cancer cells MDA-MB-436, with downregulation of anti-apoptotic proteins such as Bcl-2 and upregulation of caspase-3 and Bax both pro-apoptotic, in addition to occurring caspase-9 activation. The treatment with oridonine phosphate also stimulated autophagy, with an increase in proteins related to the formation of autophagic vacuoles such as beclin-1 and LC-II, however when cells were treated with a 3-methyladenine autophagy inhibitor, the pro-apoptotic effect observed was abolished, indicating that oridonine phosphate induces autophagy-mediated apoptosis in breast cancer cell lines. The best effects found in the compound with the addition of phosphate groups can be explained because molecules attaching to phosphate groups could be easily transported into cells. Additionally, the phosphate group helps to maintain solubility, stability, and a longer half-life time (77).

In the same way, oridonin was also able to induce apoptosis in lung cancer 4T1 cells *in vitro* and inhibit tumor growth of these cells in a xenographic model of breast cancer in nude mice in a dose-dependent manner, significantly inhibiting the cell's invasive capacity by blocking the expression of Notch receptors 1 to 4. When the expression of Notch 1 receptors was hyperstimulated in cells by transfection with plasmids pEGFP-Notch 1, the effect of oridonine was less, indicating that its anti-tumor effect occurs by inhibiting Notch receptors (78).

Similarly, triptolide (from A.G. Scientific, Inc.), another natural compound extracted from herbs commonly used in traditional Chinese medicine, has antitumor activity *in vitro* and induces apoptosis in MCF-7 and MDA-MB-231 breast cancer cells when used concomitantly with a low dose of TNF- α . This effect was due to the induction of caspase-3 activation. Further, the combined triptolide and TNF- α therapy reduced the expression of I κ B, XIAP, and cIAP1/2, which culminates in the activation of caspase-9, impairs the activation of the NF- κ B signaling pathway, leading to apoptosis (79).

Another diterpene compound extracted from herbs (*Salvia corrugata*), 7 β -acetoxy-20-hydroxy-19,20-epoxyroyleanone (AHE), presented an inhibitory effect against MCF10A, SKBR-3, and BT474 cell lines. Also, in SKBR-3 and BT474 cells, AHE induced a complex mechanism that involves cell cycle arrest in the G0/G1 phase, mitochondrial network collapse, induction of reactive oxygen species formation, and activation of ERK1/2 kinase pathway. These events lead to apoptotic death of both breast cancer cell lineages (80).

The use of diterpenic quinones (Cryptotanshinone, dihydrotanshinone I, tanshinone I, tanshinone II A) isolated from Danshen (*Radix Salviae Miltiorrhizae*) has

been studied about its therapeutic potential for the treatment of cancer, with an inhibitory effect *in vitro* on the growth of tumor cell lines, with a more prominent effect on breast cancer cells, such as MCF-7. Besides, treatment with diterpenic quinones induced the activation of caspase 3 and 9 and stimulated the caspase-mediated cleavage of Poly (ADP-ribose) polymerase (PARP), leading to its inactivation. PARP is a protein involved in DNA repair, genomic stability, and apoptosis control, plays an important role in oncogenesis, as it stabilizes the broken DNA strand for homologous recombination repair (HRR)-dependent repair. However, in HRR-defective cancer, classically those with BRCA1 and 2 mutations such as breast and ovarian cancer, PARP activity can prolong cell life and favor cancer development (81). On the other hand, the cleavage of PARP, as observed in the treatment with diterpenic quinones, leads to the separation of the protein into two subunits, one of which remains with the ability to bind to the injured DNA, but without its second subunit, it loses the ability to initiate DNA repair and acts as a pro-apoptotic stimulus (82). Treatment with diterpenic quinones further stimulated dose-dependent induction of p-JNK, caspase-12, and p-eif2 α , and reduced ATF4, indicating the stress of the endoplasmic reticulum, leading to the progressive activation of the Apaf-1/caspase cascade resulting in apoptosis (83).

Yuanhuatine, a daphnane-type diterpene isolated from flowers of *Daphne genkwa*, has a significant inhibitory effect on the *in vitro* growth of breast cancer cells, with a more pronounced effect on MCF-7 cells positive for the alpha estrogen receptor (ER α). Treatment with yuanhuatine reduced the levels of pro-caspase-7 and PARP, increasing the active form of caspase-7, and the treatment also reduced the potential of mitochondrial membrane, induced the generation of reactive oxygen species and superoxide in the mitochondria, and increased Bax and reduced Bcl-2 levels, culminating in apoptosis due to mitochondrial dysfunction (84).

Also, computational molecular docking demonstrated that yuanhuatine forms key hydrophobic interactions in the hydrophobic pocket with some crucial lipophilic residues (Phe 404, Leu 391, Leu 387, Ala 350, Leu 346, Met 421 and Leu 525) as seen in ligand estradiol binding, which could overlap the ATP phosphate-binding region. Further, *in vitro* treatment with yuanhuatine reduces the production of ER α and down-regulated Akt/mTOR and MEK/ERK signaling, confirming that the compound inhibited ER α signaling pathway in MCF-7 cells since Akt/mTOR and MEK/ERK signaling molecules are important downstream molecules of ER α (84).

The natural compound andrographolide belonging to the group of diterpenic lactones, extracted from the plant *Andrographis paniculata* Nees, has been described to have antitumor activity, being three hydroxyls at C-3 (secondary), C-14 (allylic) and C-19 (primary) on the basic structural skeleton reported being responsible for its biological

activities. The *in vitro* treatment with andrographolide in MDA-MB-231 triple-negative breast cancer cell lines inhibited cell growth due to apoptosis triggered by the production of reactive oxygen species and reduced mitochondrial membrane potential. Moreover, the compound was also able to stimulate the production of Bax and reduce Bcl-2, such changes induced the release of cytochrome C in the cytosol, activation of Apaf-1, and consequent activation of caspase-9 and 3, culminating in apoptosis (85).

Similarly, an epoxy clerodane diterpene extracted from *Tinospora cordifolia*, was also able to reduce proliferation by inducing apoptosis in MCF-7 cells *in vitro*, without altering the viability of normal Vero and V79 cells. In MCF-7 cells, treatment induced the production of reactive oxygen species and altered the production of proteins related to cell death, with a reduction in anti-apoptotic (BCL-2, MDM-2 and PCNA), and an increase in pro-apoptotic (CYP1A, GPX, GSK3b, CDKN2A, RB1, BAX, CASP-8, CASP-9, P21, and P53) (61). Similarly, a *cis*-clerodane diterpene, called crispene E, extracted from plants of the same genus *Tinospora*, species *T. crispa*, showed important inhibitory activity of STAT3 dimerization, necessary for its activation and translocation to the nucleus to transcribe several genes related to proliferation and survival of cancer cells. MDA-MB-231 cells treated with crispene E significantly reduced cell viability, in addition to reducing the activation of STAT3 and its target genes bcl-2, cyclin D1, NNMT, and fascin. In this study, the molecular docking suggested that the molecule inhibits STAT3 by interacting with its SH2 domain (86).

Kopetdaghinane, a diterpene extracted from *Euphorbia kopetdaghi* plants, has demonstrated antitumor activity *in vitro* against MCF-7 cells by inducing caspase-6-mediated apoptosis, stimulating increased production of reactive oxygen species and reduced mitochondrial membrane potential, which culminated in the increased pro-apoptotic Bax protein activity and reduced Bcl-2 (87). Reactive oxygen species also seems to be pivotal to the anti-tumoral action of the diterpenoid Crassin, when tested against MDA-MB-231 and 4T1 cell lines. However, surprisingly, the inhibitory effects of Crassin were not related to common mechanisms of cell death (apoptosis, necrosis, necroptosis, and ferroptosis). The authors discuss that the mechanism of action of this compound could be cytostatic instead of cytotoxic (88).

Kahweol is a diterpene found in coffee beans, specifically *Coffea arabica*, and has antitumor activity in human epidermal growth factor receptor-2 (HER2)-overexpressing breast cancer cells in a caspase-3 dependent manner. Kahweol's antiproliferative activity also resulted in reduced Akt phosphorylation and its downstream signaling, mTOR, and cyclin D. Blocking Akt signaling by kahweol results in reduced expression of fatty acid synthase (FASN) and consequently inhibits cell proliferation (89).

Jatrophone, isolated from the root and stem of *Jatropha gossypifolia*, showed antitumor activity in several cells of triple-negative breast cancer lineage (MDA-MB-231, HCC38, MDA-MB-157, and MDA-MB-468) by inhibiting the pathway signaling WNT/ β -catenin, which is involved in the transcription of several genes related to cell proliferation, survival, development, and differentiation. Also, the Wnt/ β -catenin pathway has been associated with the carcinogenesis of several types of cancer. The antitumor activity of the jatrophone occurs by inhibiting the downstream signaling triggered by the Wnt receptor complex, preventing the stabilization of β -catenin and its consequent translocation to the nucleus for transcription of genes such as AXIN2, HMGA2, MYC, PCNA, and CYCLIN D1, an inhibition demonstrated both at the transcriptomic and protein levels, which results in the induction of apoptosis. Thus, jatrophone interferes with triple-negative breast cancer cell proliferation based on the cancer cell line etiology-subtype, the known cellular chemoresistance status, and cell capacity to produce Wnt (90).

Eriocalyxin B (EriB), a diterpenoid with proven anti-angiogenic activity (described in section 4), was also tested against MCF-7 and MDA-MB-231 cell lines with good inhibitory activity, by inducing apoptosis accompanied by the activation of autophagy, which was evidenced by the increased accumulation of autophagosomes, acidic vesicular organelles formation, the microtubule-associated protein 1A/1B-light chain 3B-II (LC3B-II) conversion from LC3B-I and p62 degradation. In these cells, EriB treatment also inhibited the Akt/mTOR/p70S6K signaling pathway. The authors even showed that ROS played an essential role in EriB-induced cell death. Finally, it was shown that in the MDA-MB-231 xenograft breast tumor model, EriB also displayed a significant anti-tumor effect, with a reduction of tumor weight and activation of autophagy and apoptosis. Taken together, the findings demonstrated that EriB presents a potent anti-breast cancer effect both *in vitro* and *in vivo* (91).

6. Diterpenes as a therapeutic option in drug-resistant cell lineages

Multidrug resistance (MDR) is one of the main reasons for the fail of cancer treatments. It is characterized by broad cross-resistance to unrelated drugs that are different both in molecular structure and target specificity. The overexpression of P-glycoprotein (P-gp) is the most studied and common mechanism of MDR (92). P-gp is a plasma membrane-associated transport protein expressed in drug-resistant cancer cells, that recognizes chemotherapeutic drugs as substrates and extrude them out, besides to impair the immunesensitizing functions of the cells, leading to ineffective intracellular drug concentrations, reduced elimination by immune cells, and poor clinical outcomes following chemotherapy (92,93).

Jatrophanes are macrocyclic diterpenes with broad biological effects usually substituted for a variety of acyl groups (94). HU et al., (2018) isolated 15 different jatropane diterpenoids from the fruits of *Euphorbia sororia* and evaluated its effects on multidrug-resistant MCF-7/ADR breast cancer cells with P-gp overexpression. Eight compounds showed good chemoreversal abilities compared to verapamil, a known P-gp inhibitor. From these, Euphosorophane A was the most promising molecule, with low cytotoxicity to normal cells (HEK293 cell line), high therapeutic index, inhibition of P-gp-mediated rhodamine123 efflux, and ability to reverse P-gp-mediated resistance to doxorubicin and other cytotoxic agents associated with MDR as docetaxel and vincristine. The authors further elucidated that the mechanism of action of this diterpene is involved with a competitive inhibition to DOX in the binding site of P-gp, resulting in increasing of intracellular DOX concentration, which culminates in resensitizing MCF-7/ADR to DOX. Also, some correlations of structure-activity relationship were proposed, in which the acyloxyl at C-5 could modulate the reversal effects on DOX resistance. Furthermore, the authors hypothesized that the presence of an aromatic ester group (benzoyl) at C-14 has a positive role in the modulation of the drug accumulation in the MCF-7/ADR cells. Lastly, the presence of the carbonyl at C-14 instead of acetoxy had little influence on the activity (95).

Similar effects were found when myrsinol diterpenes derived from *Euphorbia prolifera*, J196-9-4, and J196-10-1, were tested against MCF-7/ADR cells. Both compounds were non-toxic for cells at low concentrations (<50 μM) and could reverse the resistance to daunorubicin, vincristine, and topotecan by acting as P-gp substrate, inhibiting the P-gp mediated rhodamine 123 efflux, and stimulating the ATP hydrolysis. Thus, J196-9-4 and J196-10-1 also acted as a competitive inhibitor of P-gp and reverses MDR (96,97).

In the same way, XUE et al., (2016) isolated 27 *neo-clerodane* diterpenoids substituted with nicotinoyloxy from *Scutellaria barbata*, which were tested against MCF-7/ADR cells. Among the tested molecules, four exhibited better chemoreversal abilities than verapamil, and the most potent compound, scutebatin A, reduced more than 45 times the IC_{50} value of DOX. Besides that, scutebatin A (10 and 20 μM) significantly increased DOX and rhodamine 123 accumulation in the cells, while decreased the P-gp expression level, and the docking simulations further confirmed scutebatin A is a competitive inhibitor of P-gp, by the interaction of benzoyloxy group at C-6 with residue Phe 974, besides interactions by hydrogen bonds that further stabilized the complex (98).

P-gp inhibition was also verified when MCF-7/ADR cells were treated with five series of 37 new acylate and epoxide derivatives of *Euphorbia* factor L3, a lathyrol diterpene isolated from *Euphorbia lathyris*, designed by modifying the hydroxyl moiety of C-3, C-5,

or C-15. Eight derivatives exhibited greater chemoreversal ability than verapamil against DOX resistance, from which the two most effective were able to effectively reverse the sensitivity to DOX by 34.4 and 28.6 fold, respectively, and exhibited effective efflux inhibition of rhodamine 123. From these experiments, it was proven that benzylation of the OH-3 and OH-5 of lathyrol produced the best inhibitor. Molecular docking indicated the important influence of hydrophobic interactions and hydrogen bonds in the flexible cavity of P-gp for the compound effectiveness (99).

ERBB2 is a member of the epidermal growth factor (EGF) receptor is amplified and overexpressed in 20 % to 30 % of human breast cancers, and it is often associated with aggressive disease, poor prognosis, and Trastuzumab (Tz) resistance (100). The diterpene carnosic acid (CA) isolated from *Salvia somalensis* was tested against MCF7 and MDA-MB-231 (ERBB2⁺), and also against SKBR-3, BT474 ERBB2⁺ cell lines (Tz resistant). Firstly, CA significantly decreased the migratory capability of MDA-MB-231 and the survival of all cells compared to the non-treated control. The combination of Tz and CA in ERBB2⁺ cells strongly enhanced the inhibitory effect by inducing cell cycle arrest in G0/G1 phases, reducing the fraction in G2/M, inhibiting ERBB2 signaling pathway, and cooperatively upregulating CDKN1B/p27KIP1 expression levels, suggesting a cooperative antitumor effect of the two drugs. Also, CA could partially restore Tz sensitivity in Tz-resistant SKBR-3 cells, which results from multiple mechanisms that ultimately cause cell cycle arrest and inhibition of the late stages of the autophagic flux (101).

Taking together, these works show that diterpenes isolates from plants have good potential to revert the drug resistance of breast cancer. However, most of the results are obtained from *in vitro* tests, therefore, further studies are necessary for a complete understanding of the action potential of diterpene compounds for a possible clinical application.

7. Nanotechnology as an enhancer of diterpenes activity

Nanotechnology has increasingly been a useful tool in the management of drugs with low solubility, high toxicity, and difficulty in administration, increasing the targeted and controlled delivery of a drug (102). In this sense, some diterpenes have been nanoencapsulated and studied for its anti-breast cancer potential.

Triptolide (TP) is a diterpenoid triepoxide extracted from the plant *Tripterygium wilfordii* with proven activity against many malignant tumors. However, its poor solubility and high toxicity are limited to its potential clinical application. For this, liposomes containing TP (TPLP) were formulated coated with hyaluronic acid (HA) and tested against MCF-7 and 4T1 breast cancer cell lines. Nanoparticulated TP (TPLP-HA) was

as effective as free TP against tumor lines, however, TPLP-HA presented a lower percentage of cells under late apoptosis and necrosis, which indicates less induction of local inflammation. TPLP-HA were also tested *in vivo* in 4T1 solid tumor-bearing BALB/c mice and the MCF-7 human breast cancer xenograft model in nude mice, both treated with 0.4 mg/kg via intravenous injection once every two days. After 17 days of treatment, it was observed that the TP and TPLP-HA had similar antitumor activity, and the effect on the MCF-7 model was better than 4T1 for both treatments. Also, TPLP-HA displayed lower toxicity than TP, with LD₅₀ of 3.07 and 1.30 mg/kg, respectively, with low effects in the function of the liver and kidney, which indicated the selective tumor targeting of TPLP-HA. The nanoformulated TP also presented higher cellular uptake, which can explain the increase in efficacy while decreasing systemic toxicity (103).

Sclareol (SC) is another diterpene from natural origin with potential antitumor activity, but unfavorable bioavailability due to its poor aqueous solubility. To improve these features, SC was encapsulated in PLGA nanoparticles coated with HA and tested against MCF-7 and MDA-MB-468 breast cancer cell lines. The formulation containing SC coated with HA exerted a noticeable anticancer effect on both the cell lines (10 – 50 µM), promoting a greater decrease in their viability when compared to the uncoated formulation and the free SC. The HA coating of the PLGA nanoparticles favored a better interaction and uptake of particles by cells (104).

SC was also used to improve the cytotoxic effects of DOX, a commercial antitumoral drug from the anthracycline class. For this, OLIVEIRA et al., (2018) developed solid lipid nanoparticles (SLN) containing DOX and SC (SLN-DOX-SC), which were tested against MCF-7 and 4T1 cells. The release profiles revealed that the SLN-DOX-SC showed a controlled release of DOX at pH 7.4 with enhanced drug release at low pH, an important feature for cancer treatment. SLN-DOX-SC resulted in a dose-dependent increase in cell cytotoxicity, which was significantly higher than those observed for the blank SLN. Also, for 4T1 cells, SLN-DOX-SC presented better cytotoxic effects than free DOX, which can be attributed to the combination of DOX and SC in SLN, since SLN-SC (with SC but without DOX) showed no inhibitory effect. The higher cytotoxicity observed for SLN-DOX-SC over free DOX in the 4T1 cell line is a huge achievement of the formulation, as this shows its capability of enhancing drug activity in DOX resistant cell lines (105).

The same research group showed that nanostructured lipid carrier (NLC) co-encapsulating DOX and SC (NLC-DOX-SC) presented a synergic effect on MDA-MB-231 and 4T1 cells in the molar ratio 1:1.9 (DOX: SC), with a significant reduction in the IC₅₀ values when compared to free DOX and SC. Then, the formulation was tested *in vivo*, in a murine breast cancer model (4T1 tumor-bearing mice), and the treatment with NLC-DOX-SC presented a higher capacity to inhibit tumor growth than free DOX and

free SC. Both *in vitro* and *in vivo*, the effects of the association between DOX and SC were similar with or without the encapsulation, however, NLC-DOX-SC provoked a lower body weight loss and hematological alterations compared to free DOX + free SC, suggesting the ability of the NLC to reduce its toxicity (106).

The presented works show potential uses of nanoparticles as alternatives for targeting drugs and reducing toxicity. However, different results can be found when this approach is used for different cell lines. Therefore, studies are still needed to fully clarify the potential use of nanoparticles containing diterpenes for the treatment of breast cancer.

8. Sites of interaction and structure-activity relationship

Some studies involving the activity of diterpenes directly relate structural changes to the activity of the compounds. Still, computational studies allow a predictive identification of targets, sites of interactions, and possible mechanisms of action of diterpenes. The works connecting these features that were presented throughout the chapter are described in Table 2. Also, the specific works on the relationship between structure and activity are described below.

Wada et al (2019) addressed the SAR of a lycotoxine-type C19 diterpenoid alkaloid. From the C-1 and C-14 esterifications 46 new derivatives were obtained (2–29, 2a, 3a, 5a - 7a, 9a, 13a, 13b, 14a, 14b, 16a, 17a, 24a, 32, 34a, 34b, 36a). The synthetic and natural compounds (obtained from the purification of the roots of *Aconitum yesoense* var. *Macroysesense*), were evaluated for antiproliferative activity against five human tumor cell lines: pulmonary adenocarcinoma (A549), breast cancer (MDA-MB-231, MCF-7), cervical carcinoma HeLa derived cell line (KB), and vincristine-resistant KB subline (KB-VIN).

According to the results of the antiproliferative activity, the natural alkaloids Delcosine (1), 14-O-acetyldelcosine (31), and 14-O-acetylbornoniine (33) were inactive ($IC_{50} > 20 \mu M$). However, acylation of the hydroxy group C-1 and/or C-14 of 1 (except with an acetyl group) led to varying degrees of antiproliferative activity. The esterified C-1 compounds, which showed greater potency in all five tumor cell lines tested, were 6 [1-(4-chloro-3-nitrobenzoyl) delcosine], 7 [1-(4-dichloromethylbenzoyl) delcosine], in addition to compounds 13a (pentafluorobenzoate in C-1 and C-14) and 13b (pentafluorobenzoate in C-1, acetate in C-14), with IC_{50} values ranging from 4.3 to 6.0 μM . All potent compounds, except 13a, caused the accumulation of sub-G1 cells within 24 h. Acylation of 1 as a leader appears to be critical for producing antiproliferative activity in this alkaloid class (107).

The determination of the SAR using computational methods was presented by Da Costa et al., (2018) (108). From the isolation of kaurenoic acid (ent-kaur-16-en-19-oic acid) from *Copaifera* ssp, its analogs (ent-7 α -hydroxy-kaur-16-en-19-oic acid, ent-15 β -hydroxy-kaur-16-en-19-oic acid, methyl ent-7 α ,16 β -dihydroxy-kaur-19-oate, and ent-7 α ,11 β -dihydroxy-kaur-16-en-19-oic acid) were obtained from the biotransformation process by the fungus *Aspergillus terreus* and tested against breast cancer cell lines (MCF-7), followed by the multivariate quantitative-structure activity relationship (QSAR) analysis where they presented IC₅₀ of 145.40, 141.00, 505.90, 783.00, and 805.10 respectively. The results showed that the HOMO and HOMO-1 orbital energies and logP are related to the activity against MCF-7 breast cells. That is, the drug-receptor interactions for this class of compounds with carcinogenic activity are predominantly guided by the transferor displacement of charge, suggesting this displacement mainly from the substituent R1, which with the presence of more lipophilic substituents leads to the formation of more complex active compounds, adding to the molecules higher values of HOMO and HOMO-1 as well as logP.

Terpenoids isolated from *Euphorbia kansui* were evaluated according to their anti-proliferative potential in human cancer cells, showing the structure-activity relationship. In this work, twenty-five terpenoids were isolated from *E. kansui*, thirteen diterpenoids of the ingenane type, and eight of the jatrophone type (with two new compounds, kansuinin P and Q), and four triterpenoids. *In vitro* anticancer activity was performed by the MTS assay on five human cancer cell lines (human colorectal cancer Colo205 cells, human melanoma cells breast cancer MDA-MB-435 cells, human leukemia DAUDI cells, human prostate cancer PC3 cells, and human ovary cancer SKOV-3 cells). The results regarding the structure-activity relationship were performed for 12 ingenane-type diterpenoids in colorectal cancer Colo205 cells. The analysis of the structure-activity relationship showed that the presence of the acetoxy (-OAc), and hydroxyl group on position 20 were important for the activity of the compounds in Colo205 cells, while the in position 3 contributed to a more significant biological activity of the compounds both cells more than on position 5 (MDA-MB-435 and Colo205) (109).

Table 2 – Interactions predictions and structure-activity relationship between diterpenes and proteins involved in the breast cancer evolution.

Diterpene (and type)	Source	Interactions predictions/ structure-activity relationship	Ref.
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Taicrypnacids A and B	<i>Taiwania cryptomerioides</i>	Presence of the hydroxycarbonyl group can play a crucial role in cytotoxicity, unaffected by the configurational changes in C-7 and C-14 in the compounds	(35)
Lipomesaconitine Lipoaconitine Lipomesaconitine (diterpenoid alkaloids)	<i>Aconitum japonicum</i>	The fatty acid ester at C-8 and the anisoyl group at C-14 may be important to the cytotoxic activity of aconitine-type C19-diterpenoid alkaloids against human tumor cell lines.	(36)
Barbatin H (neo-clerodane) and its analogs	<i>Scutellaria barbata</i>	Presence of two benzoyloxy groups in C-6 and C-7, associated with the strongest aromaticity in the benzene ring and biological action.	(37)
Siegesides A (diterpenoid glycosides)	<i>Siegesbeckia pubescens</i>	The presence of a 15S-16 vicinal diol was an effective group for the teaching of inhibitory activities, and the number, location, and orientation of hydroxyl groups may be essential for the inhibitory effects of diterpenoids against the invasion of MB-MDA-231 cells.	(41)
ent-2,3,7-trihydroxy-18-nor-rosa-1,3,5,7,15-pentane-6-one (C): ent-rosane. ent-11a,14a-dihydroxy-abieta-7(8),13(15)-dien-16,12a-olide (E), and ent-11a-hydroxy-8a,14a-epoxy-12-oxo-abieta-13(15)-en-16-oic acid methyl ester (H): ent-abietane	<i>Euphorbia fischeriana</i>	Primary consideration of the structure-activity relationship of the isolated compounds suggesting that an aromatic A-ring may be of benefit for the activity of ent-rosanes. However, details about the action of these bioactive diterpenoids are still under investigation.	(46)
Azorellanol (zorellane)	<i>Azorella compacta</i>	In the azorellanes, the beta acetate group at position C-7 in the skeleton increases the cytotoxic effect. The alpha position of the OH group at position C-13 further increases the effect.	(47)
Cyrtophyllones B, Uncinatone, 3-O- α -l-arabinopyranosyl olean-12-en-28-oic acid, 3-O- $[\beta$ -d-glucopyranosyl(1-4)- α -l-arabinopyranosyl]olean-12-en-28-oic acid, and 2 α ,3 α ,23-trihydroxyoleane-12-en-28-oic acid.	<i>Akebia quinata</i>	Presence of a COOH group at C-28 showed a significant increase in inhibitory activity against the PTP1B enzyme. Increased activity due to one less glucopyranosyl, linked to arabinopyranyl C-3'. The presence of an alcohol group at C-23 may decrease the inhibitory effects on PTP1B.	(54)
(E)-14-hydroxy-15-norlabda-8(17),12-dien-16-al (bicyclic diterpenoid)	<i>Curcuma amada</i>	The hydroxymethyl functionality at C-13 might be responsible for the effect.	(57)
Parvifloron F and the synthetic intermediates	genus <i>Plectranthus</i>	The oxidation level of the abietane ring may be important for antiproliferative and selective activity.	(61)
(1S,2E,4S,6R,7E,11E)-2,7,11-cembratriene-4,6-diol (cembranoid)	<i>Nicotiana tabacum</i>	Complete shape fitting at the ATP binding pocket of VEGFR2 kinase domain, with important participation of C-4 and C-6 hydroxyl groups and C-19 and C-20 methyl groups.	(66)

Eriocalyxin B	<i>Isodon eriocalyx</i> var. <i>laxiflora</i>	Stably bind at the ATP-binding site of VEGFR-2, with seven amino acids actively involved.	(67)
6E,0E,14Z-(3S)-17-hydroxygeranylinalool-17-O- β -d-glucopyranosyl-(1 \rightarrow 2)-[α -l-rhamnopyranosyl-(1 \rightarrow 6)]- β -d-glucopyranoside (glycoside)	<i>Blumea lacera</i>	Absence of the β -d-glucopyranose on the C-3 oxygen contributed for the significantly cytotoxic activity	(72)
Parvifloron D, 6,7-dehydroroyleanone and 7-acetoxy-6-hydroxyroyleanone	genus <i>Plectranthus</i>	Double substitution only with non-substituted phenyls, or hydroxybenzoate at position four, can increase the interactions in PKC. Substitution groups of the royleanone core, can provide a better docked binding pose.	(75)
Oridonin (7,20-epoxy-ent-kauranes)	Commercial, found in Chinese herbs	Addition of phosphate group provides better effects.	(77)
Yuanhuatine (daphnane)	<i>Daphne genkwa</i>	Hydrophobic interactions in the hydrophobic pocket with some crucial lipophilic residues (Phe 404, Leu 391, Leu 387, Ala 350, Leu 346, Met 421, and Leu 525).	(84)
Crispene E (clerodane-type)	<i>Tinospora crispa</i>	The inhibition of STAT3 might be by hydrophobic interactions with its SH2 domain.	(86)
Euphosorophane A (jatrophane)	<i>Euphorbia sororia</i>	High binding affinity toward the DOX recognition site of P-gp	(95)
Scutebatin A (neo-clerodane)	<i>Scutellaria barbata</i>	The benzoyloxy group at C-6 interacted with residue Phe 974 of P-gp, with possible roles in the binding pocket. The interaction of scutebatin A with Tye 949 and Gln 721 further stabilized the complex.	(98)
<i>Euphorbia</i> factor L3 (lathyrane)	<i>Euphorbia lathyris</i>	Influence of hydrophobic interactions and hydrogen bonds in the flexible cavity formed by two bundles of six transmembrane helices of P-gp.	(99)
1-(3,5-dichlorobenzoyl) delcosine and 1-(4-dichloromethylbenzoyl) delcosine (lycoctonine-type)	<i>Aconitum yesoense</i> var. <i>macroyesoense</i>	C-1 esterified compounds caused an increase in the number of cells in phases S and G2/M followed by the accumulation of sub-G1	(107)
<i>ent</i> -kaur-16-en-19-oic acid, <i>ent</i> -7 α -hydroxy-kaur-16-en-19-oic acid, <i>ent</i> -15 β -hydroxy-kaur-16-en-19-oic acid, <i>ent</i> -7 α ,11 β -dihydroxy-kaur-16-en-19-oic acid, and methyl <i>ent</i> -7 α ,16 β -dihydroxy-kaur-19-oate (kaurane)	<i>Copaifera</i> <i>ssp</i>	The drug-receptor interactions are predominantly guided by the transfer or displacement of charge, mainly from the R1 substituent, which with the presence of more lipophilic substituents there is the formation of more active compounds.	(108)
Cl-(1-13) (ingenane)	<i>Euphorbia kansui</i>	The presence of the substituent in position 3 contributed to a more significant biological activity.	(109)

We described here the works addressing the structure-activity relationship of plant diterpenes, identifying the structural changes where an increase in antitumor activity was

described when compared to the initial compound, as well as the works where this improvement was not observed. From these studies, it is possible to infer that compounds modified via classical synthesis or from the fungal biotransformation process tend to present a significant improvement in the antitumor activities.

However, some authors do not address the structure-activity relationship but perform an analysis of the molecular interaction with the active site in addition to its therapeutic viability from computational or experimental screening. In these works, it was observed strong interactions of the diterpene molecules with amino acid residues from different breast cancer cell proteins. It is worth mentioning that a large part of the studies did not directly indicate which portion of the molecule or the interaction site was responsible for the biological effects described, which does not diminish the importance and quality of the respective works, showing important results about to the action of diterpenes in the context of breast cancer.

9. Conclusions

In this chapter, it was described the recent discovery diterpenes from plant sources were described by its anti-breast cancer activity, besides the compounds in which the action mechanisms were proved, namely: the antiproliferative action, the inhibition of tumor angiogenesis and metastasis, and induction of cell death. The chapter also covered the diterpenes studied by its effect in drug-resistant breast cancer cell lineages and the use of nanotechnology as an enhancer of its activity. From the structure-activity relationship studies, it was possible to infer that most compounds modified via classical synthesis or from biotransformation process had improved antitumor activities. Thus, we conclude that studies related to diterpenes can be constantly explored since obtaining from natural sources is a vast field and several even more active derivatives can be obtained from structural modification processes, and therefore, diterpenes from plants have an important potential in drug discovery for breast cancer treatment.

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6.3. PARECER DA COMISSÃO DE ÉTICA DE NO USO DE ANIMAIS



COMISSÃO DE ÉTICA NO USO DE ANIMAIS

OF. CIRC. CEUA N° 103/2016

Londrina, 01 de Junho de 2016.

Prezado Pesquisador,

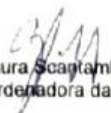
Certificamos que o projeto intitulado "Ação do flavonóide quercetina nos eventos iniciais da infecção experimental de camundongos Balb/c por *Leishmania amazonensis*", protocolo CEUA n° 8286.2016.60, sob a responsabilidade de Wander Rogério Pavanelli, que envolve a produção, manutenção e/ou utilização de animais pertencentes ao filo Chordata, subfilo Vertebrata (exceto o homem), para fins de pesquisa científica (ou ensino), encontra-se de acordo com os preceitos da Lei n° 11.794, de 8 de outubro de 2008, do Decreto n° 6.899, de 15 de julho de 2009, e com as normas editadas pelo Conselho Nacional de Controle da Experimentação Animal (CONCEA), foi **aprovado** pela Comissão de Ética no Uso de Animais da Universidade Estadual de Londrina (CEUA/UJEL), em reunião realizada em **24/05/2016**.

O objetivo do projeto é avaliar os períodos iniciais da infecção experimental por *Leishmania amazonensis* e a ação do flavonóide quercetina como modulador da resposta inflamatória induzida por este parasito em camundongos BALB/c e C57BL/6. Para isso os camundongos serão infectados no coxim plantar de ambas as patas traseiras, via subcutânea, com 1×10^5 formas promastigotas de *L. amazonensis* em fase estacionária de crescimento (5 dias pós-repique). Neste experimento, será avaliada a diferença entre os estágios iniciais da infecção de camundongos BALB/c e C57BL/6 e a ação da quercetina sobre camundongos BALB/c infectados com *L. amazonensis*, nos momentos iniciais da infecção e após o estabelecimento da lesão. Os períodos definidos para as análises serão 6, 24, 72, 144 e 240h pós-infecção e após o aparecimento da lesão (5 semanas pós-infecção). Os grupos experimentais serão: controle sem infecção, que receberá apenas salina via oral (v.o.); controle infectado no coxim plantar de ambas as patas traseiras, via subcutânea, com 1×10^5 formas promastigotas de *L. amazonensis*; e grupos tratados que, além da infecção com 1×10^5 formas promastigotas de *L. amazonensis*, serão tratados por via oral com microcápsulas inertes suspensas em salina (100 mg/kg), quercetina (10 e 100mg/kg, solubilizadas com 10% de tween 80 e 90% de salina) e microcápsulas de quercetina (10 e 100mg/kg, solubilizadas em salina). Em relação ao ovino, o animal será contido em decúbito lateral para coleta do sangue, serão coletados 20 mL de sangue que serão transferidos para frascos de vidro estéreis contendo pérolas de vidro seguido de agitação manual destes frascos durante aproximadamente 15 minutos para o processo de desfibrinação do sangue necessário para o preparo do meio de cultura agar sangue, necessário para o cultivo do protozoário *Leishmania*. Ao longo dos 3 anos de projeto, será utilizado até 160 mL de sangue (20 mL por coleta), GI 1.

Vigência do Projeto	15/05/2016 a 15/04/2019
Espécie/linhagem	Camundongo isogênico / BALB/c e C57BL/6 Ovino / <i>Ovis aries</i>
N° de animais	197 BALB/c, 53 C57BL/6 e 1 Ovino
Peso/Idade	Camundongos: 25g / 6 a 8 semanas Ovino: Indeterminado
Sexo	Machos e Fêmeas
Origem	Camundongos: Biotério Fiocruz/Curitiba Ovino: Fazenda Escola / UJEL
Amostras a serem coletadas	Camundongos: Plasma, patas, linfonodo popliteal, estômago, fígado. Ovino: Sangue

Cumpra orientar que caso pretendam-se quaisquer alterações no protocolo experimental aprovado, deve-se submeter o novo protocolo à apreciação da CEUA/UJEL anteriormente à execução das modificações.

Coloco-me à disposição para quaisquer esclarecimentos que se fizerem necessários. Sem mais para o momento, subscrevo, cordialmente,


Profa. Dra. Glaucia Scantamburlo Alves Fernandes
Coordenadora da CEUA/UJEL

Ilmo. Sr.

Prof. Dr. Wander Rogério Pavanelli

Coordenador do Projeto

Departamento de Ciências Patológicas / Centro de Ciências Biológicas

Com cópia para Diretor da Fazenda Escola/UJEL, Chefe do Departamento de Ciências Patológicas e Diretor(a) do Centro de Ciências Biológicas