

SPECIAL ARTICLE

Autoimmunity, phospholipid-reacting antibodies and malaria immunity

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Several questions regarding the production and functioning of autoantibodies (AAb) during malaria infection remain open. Here we provide an overview of studies conducted in our laboratory that shed some light on the questions of whether antiphospholipid antibodies (aPL) and other AAb associated with autoimmune diseases (AID) can recognize *Plasmodia* antigens and exert anti-parasite activity; and whether anti-parasite phospholipid antibodies, produced in response to malaria, can inhibit phospholipid-induced inflammatory responses and protect against the pathogenesis of severe malaria. Our work showed that sera from patients with AID containing AAb against dsDNA, ssDNA, nuclear antigens (ANA), actin, cardiolipin (aCL) and erythrocyte membrane antigens recognize plasmodial antigens and can, similarly to monoclonal AAb of several specificities including phospholipid, inhibit the growth of *P. falciparum* *in vitro*. However, we did not detect a relationship between the presence of anti-glycosylphosphatidylinositol (GPI) antibodies in the serum and asymptomatic malaria infection, although we did register a relationship between these antibodies and parasitemia levels in infected individuals. Taken together, these results indicate that autoimmune responses mediated by AAb of different specificities, including phospholipid, may have anti-plasmodial activity and protect against malaria, although it is not clear whether anti-parasite phospholipid antibodies can mediate the same effect. The potential effect of anti-parasite phospholipid antibodies in malarious patients that are prone to the development of systemic lupus erythematosus or antiphospholipid syndrome, as well as the (possibly protective?) role of the (pathogenic) aPL on the malaria symptomatology and severity in these individuals, remain open questions. *Lupus* (2014) **23**, 1295–1298.

Key words: Antiphospholipid syndrome; systemic lupus erythematosus; anti-DNA antibodies; malaria

Introduction

Despite the increasing knowledge on immunology during the past decades, several questions regarding autoantibody (AAb) production and function remain open. Some of them concern the relationships between autoimmunity and infectious and parasitic diseases. The concept that AAb production and autoimmunity may follow and complicate infectious and parasitic diseases such as Group A β hemolytical streptococcal infection

and Chagas Disease is now well established.¹ However, evidence has accumulated to indicate that some parasitic diseases, such as malaria, may protect individuals against the development and severity of autoimmune diseases (AID) such as systemic lupus erythematosus (SLE).^{2,3} In addition, it has been proposed that although AAb produced by individuals infected by *Plasmodia* do not seem to participate in the pathogenesis of malaria, they could even exert protective effects against the disease.⁴

Here we give an overview of studies conducted in our laboratory to approach two of the questions concerning autoimmunity and malaria: (a) Do AID-associated AAb, including those directed to negatively charged phospholipids (antiphospholipid antibodies (aPL)), recognize antigens present

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on *Plasmodia* and, could these AAb exert an anti-parasite activity? and, (b) Can similar (anti-parasite phospholipid (PL) antibodies (Ab)), produced in response to malaria, inhibit PL-induced inflammatory responses and protect against the pathogenesis of severe malaria?

Malaria remains the most prevalent parasitic disease worldwide. All five species of *Plasmodia* that naturally infect humans can cause from asymptomatic infection to severe disease, but *Plasmodium falciparum* infection is by far the most prone to cause (and is the most frequently associated with) life-threatening complications. Clinical manifestations of severe falciparum malaria vary according to transmission intensity and include hyperparasitemia, hypoglycemia, cerebral malaria, severe anemia and respiratory distress.

Many of the malaria-associated symptoms and complications result from a severe host innate immune response induced by parasite and host molecules containing pathogen-associated molecular patterns (PAMPs) released in the circulation during schizogony. *P. falciparum* glycosylphosphatidylinositol (GPI) is an abundantly expressed PL that activates leukocytes, triggers the release of pro-inflammatory cytokines and induces the expression of adhesion molecules via toll-like receptors (TLR) -2 and -4.⁵

It has been demonstrated that the administration of *P. falciparum* GPI in mice induces malaria-like symptoms,⁶ and that the immunization with non-toxic moieties of GPI that elicits an anti-GPI Ab response can prevent malaria pathology and death caused by experimental *P. berghei* ANKA and *P. yoelii* 17XL infection in mice.^{7,8} In addition, an association between the presence of anti-GPI Ab, age and prevalence of protected asymptomatic individuals has been shown in areas of high malaria transmission in Kenya and north-west Papua New Guinea.^{9,10} Therefore, although not consensually, it has been considered that anti-GPI Ab could reflect and mediate, at least partially, the anti-disease immunity in malaria, by neutralizing the toxic effect of parasite GPI.⁷

Our group also evaluated the association between anti-GPI Ab response and development of clinical malaria¹¹ by comparing the incidence of these antibodies, in supposedly immune asymptomatic carriers of plasmodial infection living in an area of the Brazilian Amazon (Barcelos) with high transmission rates of malaria, with controls with no infection or previous history of malaria from the same area and Angolan (Lubango) patients with symptomatic disease. Our data indicated that the Ab response against *P. falciparum* GPI was not

associated with *falciparum* asymptomatic infection in individuals chronically exposed to malaria in the Brazilian Amazon. We could not rule out that the high levels of anti-GPI Ab, correlated with degrees of parasitemia, observed in Lubango patients with symptomatic, but non-complicated, malaria could be protecting them from severe disease. However, an alternative and more tempting interpretation is that this Ab response is being boosted by the increasing quantities of *P. falciparum* GPI released in the circulation as the parasitemia level grows (Figure 1).¹¹

Hence, our findings do not support the hypothesis that anti-GPI antibodies mediate clinical (anti-disease) immunity against malaria, but are in accordance with those of Boutlis et al.¹² and Cissoko et al.¹³ These authors also failed to detect a relationship between anti-GPI Ab and asymptomatic infections, but, similarly to us, observed a relationship between these Ab and parasitemia levels in infected individuals from Mali and Papua New Guinea.

On the other hand, it is becoming progressively clear that autoimmune responses generated during malaria infection are not harmful, but might present protective properties against the disease. Evidence exists for AAb against erythrocytes,¹⁴⁻¹⁶ PL,¹⁷⁻²⁰ brain proteins,^{21,22} and nuclear antigens.^{20,23-25}

For the last decade, our team has been collecting evidence that autoimmunity is involved in the protection against malaria.²⁶ Zanini et al.²³ investigated the presence of AAb in the sera of malarious patients, the reactivity of AAb from Brazilian autoimmune patients against plasmodial antigens and the effect of such AAb on the *in vitro* growth of *P. falciparum*. The results showed that the frequency of sera from malarious patients reacting with actin and the cardiolipin PL was higher ($p < 0.001$) than in sera from SLE patients and, conversely, that antinuclear antibodies (ANA) were more frequent in SLE than in malaria sera ($p = 0.004$). In addition, AAb (directed against dsDNA and ssDNA, nuclear Ag, actin, cardiolipin and erythrocyte membrane) from patients with autoimmune processes recognized plasmodial antigens in the absence of malaria, (44% of the SLE – and 81% of the malaria – sera reacted with plasmodial Ag by the immunofluorescent antibody test (IFAT)); and 47.3% of the SLE patients' (and 87.5% of the malaria patients') sera inhibited plasmodial growth *in vitro*. The presence of anti-DNA and ANA was important in such anti-plasmodial activity, but no relationship was found between the *P. falciparum* inhibitory growth of

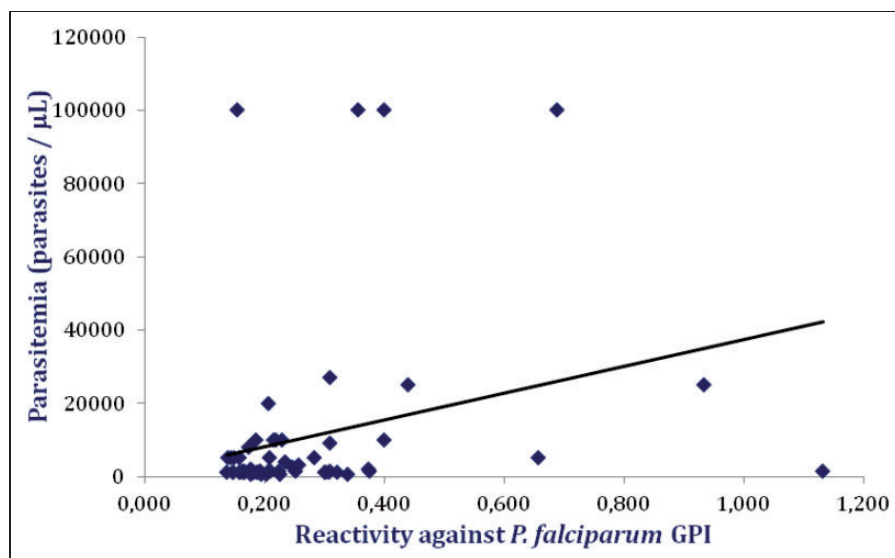


Figure 1 Correlation between the intensity of the immune response directed against the *Plasmodium falciparum* glycosylphosphatidylinositol (GPI) phospholipid (as assessed by ELISA) and the parasitemia (parasites / microliter) in peripheral blood of acute falciparum malaria patients from Lubango (Angola). Spearman's rank correlation $r=0.3198$ and $p=0.0106$.

immunoglobulins from patients with autoimmune disease and the presence of anti-actin or anti-cardiolipin AAb.

We also studied the relationship between autoimmunity and antimalarial responses by determining – in addition to the presence and the pattern of reactivity to plasmodial antigens of sera from 93 French patients with 14 different AID and not previously exposed to malaria – the effect of 21 monoclonal AAb with different specificities on the *in vitro* growth to *P. falciparum*,²⁰ 56% of sera from AID patients and none of healthy individuals reacted with plasmodial antigens. The frequency of sera reacting with at least one of the three strains and two species of *Plasmodia* depended on the specificity of the AAb the sera contained, but the differences were not statistically significant. In addition, they reacted more frequently with the young (less differentiated?) rather than with the mature blood stages of *Plasmodium*. Finally, 85% of the 13 murine monoclonal AAb reacted with plasmodial antigens and 72%, including three out of four anti-cardiolipin AAb, inhibited the *in vitro* growth of *P. falciparum*, although this inhibition did not correlate with the intensity of reactivity against the parasite, as assessed by the IFAT.

Taken together, these results and data from other groups indicate that autoimmune responses may have anti-plasmodial activity, mediated by AAb of different specificities in the presence or not of other serum factors, and protect against malaria. The potential effect of the theoretically

malaria-protective anti-parasite PL in (SLE or anti-PL syndrome-prone) malarious patients as well as the (possibly protective?) role of the (pathogenic) anti-PL AAb in these individuals on the symptomatology and severity of malaria remain open questions.

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Conflict of interest statement

The authors have no conflicts of interest to declare.

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References

- Cunha-Neto E, Teixeira PC, Nogueira LG, Kalil J. Autoimmunity. *Adv Parasitol* 2011; 76: 129–152.
- Greenwood BM. Auto immune disease and parasitic infections in Nigerians. *Lancet* 1968; 1: 380–382.
- Greenwood B, Herrick EM, Voller A. Suppression of auto-immune disease in NZB and (NZB × NZW) F1 hybrid mice by infection with malaria. *Nature* 1970; 226: 266–267.
- Daniel-Ribeiro CT, Zanini G. Autoimmunity and malaria: What are they doing together? *Acta Trop* 2000; 76: 205–221.
- Schofield L, Hackett F. Signal transduction in host cells by a glycosylphosphatidylinositol toxin of malaria parasites. *J Exp Med* 1993; 177: 145–153.
- Schofield L, Vivas L, Hackett F, Gerold P, Schwarz RT, Tachado S. Neutralizing monoclonal antibodies to glycosylphosphatidylinositol, the dominant TNF-alpha-inducing toxin of *Plasmodium falciparum*: Prospects for the immunotherapy of severe malaria. *Ann Trop Med Parasitol* 1993; 87: 617–626.
- Schofield L, Hewitt MC, Evans K, Siomos MA, Seeberger PH. Synthetic GPI as a candidate anti-toxic vaccine in a model of malaria. *Nature* 2002; 418: 785–789.
- Cherif MS, Shuaibu MN, Kodama Y, *et al.* Nanoparticle formulation enhanced protective immunity provoked by PYGPI8p-transamidase related protein (PyTAM) DNA vaccine in *Plasmodium yoelii* malaria model. *Vaccine* 2014; 32: 1998–2006.
- Naik RS, Branch OH, Woods AS, *et al.* Glycosylphosphatidylinositol anchors of *Plasmodium falciparum*: Molecular characterization and naturally elicited antibody response that may provide immunity to malaria pathogenesis. *J Exp Med* 2000; 192: 1563–1576.
- Keenihan SH, Gramzinski R, Ratiwayanto S, *et al.* *Plasmodium falciparum*. Mechanisms of innate and acquired protection against *Plasmodium falciparum* in Javanese transmigrant adults and children newly resident in malaria-endemic Northwest Papua. *Adv Exp Med Biol* 2003; 531: 83–102.
- Gomes LR, Totino PRR, Sanchez MCA, *et al.* Asymptomatic infection in individuals from the municipality of Barcelos (Brazilian Amazon) is not associated with the anti-*Plasmodium falciparum* glycosylphosphatidylinositol antibody response. *Mem Inst Oswaldo Cruz* 2013; 108: 796–800.
- Boutlis CS, Gowda DC, Naik RS, *et al.* Antibodies to *Plasmodium falciparum* glycosylphosphatidylinositols: Inverse association with tolerance of parasitemia in Papua New Guinean children and adults. *Infect Immun* 2002; 70: 5052–5057.
- Cissoko Y, Daou M, Lyke KE, *et al.* Serum antibody levels to glycosylphosphatidylinositols in specimens derived from matched Malian children with severe or uncomplicated *Plasmodium falciparum* malaria and healthy controls. *Am J Trop Med Hyg* 2006; 75: 199–204.
- Schettters TP, Van Run van Breda JH, Hermsen C, Curfs J, Eling WM. Protective and pathological activity in serum of mice developing resistance to *Plasmodium berghei* infection. *Parasite Immunol* 1989; 11: 413–423.
- Hogh B, Petersen E, Crandall I, Gottschau A, Sherman IW. Immune responses to band 3 neoantigens on *Plasmodium falciparum*-infected erythrocytes in subjects living in an area of intense malaria transmission are associated with low parasite density and high hematocrit value. *Infect Immun* 1994; 62: 4362–4366.
- Pantaleo A, Giribaldi G, Mannu F, Arese P, Turrini F. Naturally occurring anti-band 3 antibodies and red blood cell removal under physiological and pathological conditions. *Autoimmun Rev* 2008; 7: 457–462.
- Bate CAW, Taverne J, Bootsma HJ. Antibodies against phosphatidylinositol and inositol monophosphate specifically inhibit tumour necrosis factor induction by malaria exoantigens. *Immunology* 1992; 76: 35–41.
- Facer CA, Agiostratidou G. High levels of anti-phospholipid antibodies in uncomplicated and severe *Plasmodium falciparum* and in *P. vivax* malaria. *Clin Exp Immunol* 1994; 95: 304–309.
- Kwiatkowski MD, Bate CAW, Scragg IG, Beattie P, Udalo I, Knight JC. The malarial fever response pathogenesis, polymorphism and prospects for intervention. *Ann Trop Med Parasitol* 1997; 91: 533–542.
- Brahimi K, Martins YC, Zanini GM, Ferreira-da-Cruz M de F, Daniel-Ribeiro CT. Monoclonal auto-antibodies and sera of auto-immune patients react with *Plasmodium falciparum* and inhibit its *in vitro* growth. *Mem Inst Oswaldo Cruz* 2011; 106(Suppl 1): 44–51.
- Dassé R, Lefranc D, Dubucquoi S, *et al.* Singular, systemic, self-reactive IgG patterns related to age: Relationship with cerebral malaria susceptibility in exposed subjects residing in an endemic area in Abidjan, Côte-d'Ivoire. *Bull Soc Pathol Exot* 2012; 105: 276–283.
- Duarte J, Herbert F, Guiyedi V, *et al.* High levels of immunoglobulin E autoantibody to 14-3-3 epsilon protein correlate with protection against severe *Plasmodium falciparum* malaria. *J Infect Dis* 2012; 206: 1781–1789.
- Zanini GM, De Moura Carvalho LJ, Brahimi K, *et al.* Sera of patients with systemic lupus erythematosus react with plasmodial antigens and can inhibit the *in vitro* growth of *Plasmodium falciparum*. *Autoimmunity* 2009; 42: 545–552.
- Baker VS, Imade GE, Molta NB, *et al.* Cytokine-associated neutrophil extracellular traps and antinuclear antibodies in *Plasmodium falciparum* infected children under six years of age. *Malar J* 2008; 7: 41.
- Mannoor K, Li C, Inafuku M, *et al.* Induction of ssDNA-binding autoantibody secreting B cell immunity during murine malaria infection is a critical part of the protective immune responses. *Immunobiology* 2013; 218: 10–20.
- Daniel-Ribeiro CT. Is there a role for autoimmunity in immune protection against malaria? *Mem Inst Oswaldo Cruz* 2000; 95: 199–207.