

Research Article

In Vitro Antileishmanial Activity of Essential Oil of *Vanillosmopsis arborea* (Asteraceae) Baker

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The search for new immunopharmacological chemical agents to treat various diseases caused by bacteria, fungi, and protozoa, such as leishmaniasis, for example, has led to the exploration of potential products from plant species and their main active ingredients. Antimonial drugs are the current treatment for leishmaniasis. These drugs cause major side effects and frequent discontinuation of treatment. In this study, we evaluated the *in vitro* leishmanicidal activity of essential oil of *Vanillosmopsis arborea* (VAEO) and its major compound α -bisabolol against *Leishmania amazonensis*. The essential oil and α -bisabolol showed activity against promastigotes (IC₅₀ 7.35 and 4.95 μ g/mL resp.) and intracellular amastigotes (IC₅₀ 12.58 and 10.70 μ g/mL, resp.). Neither product showed any cytotoxicity on treated macrophages. The ultrastructural analysis of promastigotes incubated with VAEO or α -bisabolol at 30 μ g/mL, showed morphological changes with the accumulation of vesicles electron dense lipid inclusions. The results give evidence that both VAEO and α -bisabolol have potential as new therapeutic agents against leishmaniasis.

1. Introduction

Natural products obtained from plants have been used in the treatment of infectious diseases, especially in developing countries. It is estimated that about 25% of medicines are derived directly or indirectly from herbal products [1]. Several studies have shown that various plant species have inhibitory action against certain types of parasites such as *Leishmania amazonensis* [2], *Leishmania chagasi* [3], and *Leishmania infantum* [4].

Leishmaniasis is a chronic disease caused by protozoan parasites belonging to the genus *Leishmania*. It is an endemic disease that constitutes a serious public health problem,

threatening about 350 million people in more than 90 countries [5]. *Leishmania* species have a wide distribution among tropical and subtropical countries, including several countries in Latin America, Africa, India, part of Western Asia, and some Central European countries bordering the Mediterranean. The disease presents two clinical forms: cutaneous and visceral and the former can be subdivided into localized cutaneous leishmaniasis, diffuse cutaneous and mucocutaneous [6]. The cutaneous form is the most common, affecting about 1 to 1.5 million people per year [7, 8]. In Brazil, cutaneous leishmaniasis is present in almost all states of the federation, and *L. amazonensis* is the main etiologic agent [7].

Drugs such as sodium stibogluconate (Pentostam), amphotericin B, pentamidine, and antimony N-methylglucamine (Glucantime) are used for leishmaniasis treatment. The pentavalent antimonials have been the mainstay of treatment for human leishmaniasis in recent decades, with cure rates ranging from 60 to 90% [9, 10]. However, treatment with pentavalent antimonial is generally long expensive and toxic and this prolonged course of treatment has caused the development of resistance to these drugs [11, 12].

Thus the development of antileishmanial agents from natural products has become important to find an alternative drug with low toxicity and high efficacy [13].

Essential oils (EOs) are natural compounds and complex mixtures obtained from different parts of plants such as the flowers, leaves, bark, stems, wood, roots, fruits, or seeds and are commonly used in folk medicine to treat various types of diseases [14, 15]. Most of the EOs are comprised of terpenes, especially monoterpenes and sesquiterpenes. Several studies by different research groups have shown biological activity of EOs and their major compounds against protozoal diseases such as leishmaniasis. For example, the essential oil of *Chenopodium ambrosioides* [16], *Cymbopogon citratus* [17] that both contain citral, have been evaluated for their antileishmanial properties. Also the studies of Escobar et al. [18] who tested 19 essential oils, five of which were from different species of *Lippia* and all of them were evaluated for their antileishmanial properties.

Vanillosmopsis arborea Baker is native to the Araripe National Forest, in the Northeast of Brazil in the state of Ceará [19]. There are few studies concerning the traditional use of this plant. However, biological and pharmacological studies have shown that the essential oil of *V. arborea* (VAEO) presents antimicrobial [14], anti-inflammatory [20], and gastroprotective activity [21]. The present study characterized the essential oil of *Vanillosmopsis arborea* chemically and evaluated its leishmanicidal and cytotoxic activity. Its major compound α -bisabolol was also evaluated.

2. Materials and Methods

2.1. Plant Material. *Vanillosmopsis arborea* Baker (Asteraceae) was collected in Crato County, Ceará State, Brazil. The plant material was identified by Dr. Arlene Pessoa, and a voucher specimen was deposited under number 9493 at the Herbarium "Dárdano de Andrade Lima" of the Universidade Regional do Cariri (URCA).

2.2. Essential Oil Extraction. The oil was extracted using a Clevenger-type apparatus. The analysis of the volatile constituents was carried out in a Hewlett-Packard GC/MS, model 5971. The stems were dried (850 g), according to (traditional) folk medicine, and reduced in size, and EO extraction was carried out by hydrodistillation for 2 h using a Clevenger-type apparatus, resulting in an essential oil yield of 0.4% (3.40 mL). After the oil was collected, it was dried using anhydrous sodium sulfate and subsequently stored under low light conditions at 10°C until analysis [22].

2.3. Essential Oil Analysis. Oil analysis was performed using a Shimadzu GC MS-QP2010 series (GC/MS system) under the following conditions: Rtx-5MS capillary column (30 m \times 0.25 mm, 0.25 μ m film thickness); carrier gas—helium at 1.5 mL/min; injector temperature—250°C; detector temperature—290°C; column temperature—60°C–180°C at 5°C/min, then 180°C–280°C at 10°C/min (10 min); scanning speed—0.5 scan/sec from m/z 40 to 350; split ratio—1:200; injected volume—1 μ L of [25 μ L (essential oil)/5 mL CHCl₃] (1:200); solvent cut time—2.5 min; and mass spectrometer—operated at 70 eV of ionization energy. The identification of individual components was based on their mass spectral fragmentation according to the mass spectral library NIST 08, retention indices, and comparison with published data.

2.4. Parasites. Promastigote forms of *Leishmania amazonensis* (MHOM/BR/76/MA-76) were cultured at 26°C in LIBHIT media supplemented with 10% fetal bovine sera (Gibco, USA) and 100 U/mL of penicillin (Gibco, USA). Cultures were used only up to a maximum of ten *in vitro* passages [23].

2.5. Animals. Female BALB/c mice 4–6 weeks old were purchased from Centro de Criação de Animais de Laboratório, Instituto Oswaldo Cruz, Rio de Janeiro, and maintained under pathogen-free conditions. The animals were handled in accordance with Guidelines for Animal Experimentation of the Colégio Brasileiro de Experimentação Animal. The local Ethics Committee on Animal Care and Utilization approved all procedures involving the animals (CEUA FIOCRUZ-LW72/12).

2.6. Cell Cultures. The macrophage J774.G8 line was cultured in RPMI 1640 medium (Sigma, USA) supplemented with 10% fetal bovine sera, penicillin (100 U/mL), and streptomycin (100 μ g/mL) at 37°C and 5% CO₂. Female BALB/c mice were inoculated with 1 mL of sodium thioglycolate 3%, and after 72 hours the peritoneal macrophages were harvested with PBS solution. The harvest was centrifuged at 4000 rpm, and the cells were suspended in RPMI medium supplemented as described before and cultured at 37°C and 5% CO₂.

2.7. Activity against Promastigote Forms. Promastigote forms of *L. amazonensis* (10⁶ parasites/mL) from a 2–4-day-old culture were placed in 96-well plates in the presence of different concentrations of VAEO and α -bisabolol (Sigma-Aldrich, \geq 95% (GC)) (1.9–60 μ g/mL, for both products) to a final volume of 200 μ L per well for 24 hours. Wells without parasites were used as blank, and wells with only parasites were used as control. After the treatment, the viability of parasites was evaluated by the tetrazolium-dye (MTT) colorimetric method modified by Mosmann [24]. MTT (5 mg/mL), a volume equal to 10% of the total, was added to each well. After 2 hours, the plate was centrifuged at 4000 rpm; then 150 μ L supernatant was removed from each well and 100 μ L of DMSO were added to dissolve the formazan. The absorbance

was read on a spectrophotometer at a wavelength of 540 nm. The data were normalized according to the formula:

$$\% \text{ survival} = \frac{\text{DO sample} - \text{DO blank}}{\text{DO control} - \text{DO blank}} \times 100. \quad (1)$$

The results were used to calculate the IC_{50} (50% inhibition of parasite growth). Amphotericin B was used as the reference drug.

2.8. Activity against Intracellular Amastigotes. Peritoneal macrophages were cultured in 24-well plates (10^5 cells/well), with coverslips, at 37°C and 5% CO_2 . The cells were infected with promastigotes forms of *L. amazonensis* using a ratio of 10:1 parasite/cell, and after 2 hours the cells were washed three times with PBS to remove free parasites. The infected cells were treated with different concentrations of VAEO and α -bisabolol (60–1.9 $\mu\text{g}/\text{mL}$, for both products) in triplicate for 24 hours. The coverslips with the infected and treated cells were fixed with Bouin, stained with Giemsa, and examined by light microscopy. The inhibition percentage was calculated using the formula described by Guru et al. [25]: $\text{PI} = 100 - (\text{AN} \times 100)/(\text{INA} \times \text{TI})$, where NA is actual number of amastigotes/100 spleen cell nuclei after treatment; INA is initial number of amastigotes/100 spleen cell nuclei; TI is fold increase in the number of amastigotes in control animals on the corresponding day of the biopsy in treated animals; and PI is percentage inhibition. The IC_{50} was calculated with the GraphPad Prism. Amphotericin B was used as the reference drug.

2.9. Cytotoxicity Assay. Macrophages J774.G8 were cultured in 96-well plates (5×10^5 cells/mL) with different concentrations of VAEO and α -bisabolol (60–1.9 $\mu\text{g}/\text{mL}$, both products) to a final volume of 200 μL per well, at 37°C and 5% CO_2 . Wells without cells were used as blank, and wells with only cells were used as control. After 24 hours, the viability of the cells was evaluated by the tetrazolium-dye (MTT) colorimetric method modified by Mosmann [24]. MTT (5 mg/mL), a volume equal to 10% of the total, was added to each well. After 2 hours, the plate was centrifuged at 4000 rpm; then the supernatant was removed from each well and 100 μL of DMSO was added to dissolve the formazan. The absorbance was read on a spectrophotometer at 540 nm wavelength. The data were normalized following the formula [24]:

$$\% \text{ survival} = \frac{\text{DO sample} - \text{DO blank}}{\text{DO control} - \text{DO blank}} \times 100. \quad (2)$$

The results were used to calculate the cell cytotoxicity by 50% (CC_{50}) with GraphPad Prism 5.

2.10. Transmission Electron Microscopy. Promastigote forms of *L. amazonensis* were treated with VAEO and α -bisabolol at concentrations of 30, 15, 7.5, and 3.75 $\mu\text{g}/\text{mL}$ for 24 hours for both products. The parasites were fixed with 2.5% glutaraldehyde (Sigma, USA) in 0.1 M sodium cacodylate buffer, pH 7.2 overnight. Parasites were washed three times with

0.1 M sodium-cacodylate buffer and postfixed in a solution containing 1% osmium tetroxide, 0.8% ferrocyanide, and 5 mM calcium chloride, washed in 0.1 M sodium-cacodylate buffer, dehydrated in graded acetone, and embedded in epoxy resin. Ultrathin sections were stained with uranyl acetate and lead citrate and examined in a transmission electron microscope JEM-1011 (JEOL, Japan).

2.11. Statistical Analysis. The values were expressed as average \pm S.D. The results were analyzed by Analysis of Variance (ANOVA) followed by the Tukey test. The analyses were performed with the software GraphPad Prism 5.0.4. Differences were considered significant at $P < 0.05$.

3. Results and Discussion

The chemical analysis revealed that α -bisabolol constituted 97.9% (25.5 min.) of VAEO (Figures 1 and 2). Other compounds identified were *o*-methyl eugenol (1.6%, 18.5 min.) and bisabolol oxide (0.5%, 24.9 min.). In another study by Santos et al. [26] the essential oil of *V. arborea* showed α -bisabolol as a major compound, however, at a different concentration (80.43%). The chemical characterization of the essential oil of *Vanillosmopsis pohlii*, too, identified α -bisabolol as a major compound at a concentration of 79.0% [27]. These differences may be due to seasonality and the time of collection of the plant sample [28].

The incubation of VAEO and its major compound, α -bisabolol, efficiently inhibited the growth of *Leishmania amazonensis* promastigotes (Figure 3). The $\text{IC}_{50}/24$ h values were 7.35 and 4.95 $\mu\text{g}/\text{mL}$, respectively (Table 1). The α -bisabolol was more effective than thymol, the major compound of *Lippia sidoides*, which showed an IC_{50} of 22.63 $\mu\text{g}/\text{mL}$ against promastigotes of *L. amazonensis* [15]. In another study, α -bisabolol showed an IC_{50} of 10.99 $\mu\text{g}/\text{mL}$ against promastigotes of *Leishmania infantum*, which demonstrated its antiparasitic potential [29]. Despite having IC_{50} greater than α -bisabolol, VAEO showed significant inhibition against promastigotes of *Leishmania*, especially when compared to other plant species such as *Lippia sidoides* (IC_{50} 44.38 $\mu\text{g}/\text{mL}$), *Cordia verbenacea*, *Cajanus cajan*, *Lantana camara* (IC_{50} 120, 62, and 14 $\mu\text{g}/\text{mL}$, resp.) [30], and *Plectranthus amboinicus*, *Aristolochia cymbifera*, and *Lippia alba* ($\text{IC}_{50} > 500$ $\mu\text{g}/\text{mL}$) [30].

Our results also show that, when the two products were compared, VAEO was less effective against promastigotes and intracellular amastigotes than α -bisabolol (Table 1 and Figure 4). The selectivity index (SI), especially for intracellular amastigotes, showed that α -bisabolol (9.383) was less toxic than VAEO (11.526). These values (9.383 and 11.526) were not statistically different, which confirms the role of the α -bisabolol in the leishmanicidal action of VAEO.

Both products tested showed no cytotoxicity against macrophages with CC_{50} 145 and 100.4 $\mu\text{g}/\text{mL}$, respectively (Table 1), indicating no cytotoxicity, although the essential oils and their compounds have been reported to present cytotoxic effect when incorporated into the cell membrane [31]. The α -bisabolol is a sesquiterpene with low toxicity

TABLE 1: Leishmanicidal and cytotoxic activity of VAEO and its major constituent α -bisabolol.

Compounds	IC ₅₀ $\mu\text{g/mL}$		CC ₅₀ $\mu\text{g/mL}$	SI _{ama} ^a
	Promastigote	Amastigote intracellular	J774.G8	
VAEO	7.35 \pm 0.050	12.58 \pm 0.068	145 \pm 0.023	11.526
α -Bisabolol	4.95 \pm 0.054	10.70 \pm 0.085	100.4 \pm 0.025	9.383
Amphotericin B	3.1 \pm 0.048	7.8 \pm 0.059	1.688 \pm 0.4993	0.216

^aSI_{ama} (selectivity index) = CC₅₀ macrophage/IC₅₀ amastigote.

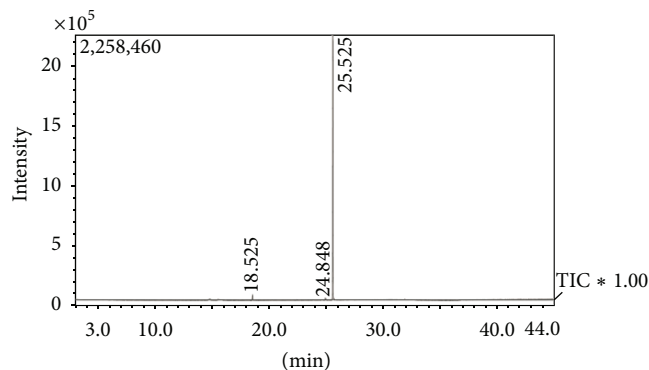


FIGURE 1: Chromatogram of the essential oil of *Vanillosmopsis arborea* Baker.

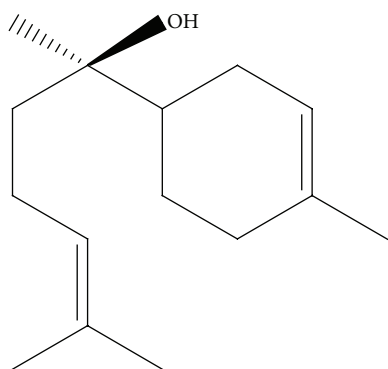


FIGURE 2: Chemical structure of α -bisabolol.

and is used in the cosmetic industry [32, 33] where it presents a pro-apoptotic activity [34]. Its leishmanicidal action against promastigotes and intracellular amastigotes may occur directly or indirectly through the production of cellular mechanisms, such as the production of nitric oxide (NO), which is the major effector molecule that participates in the intracellular killing of *Leishmania* [35]. The hydrophobic nature and the toxic effects of essential oils and their major constituents against various microorganisms are a common feature of plants which have volatile oils. These compounds can preferably incorporate in the cell membranes by inducing a loss of permeability to protons and ions and can induce changes considered vital for the cell; for example, the ergosterol biosynthetic pathway causes extensive lesions on the membranes of the *Candida* [31]. According to the results

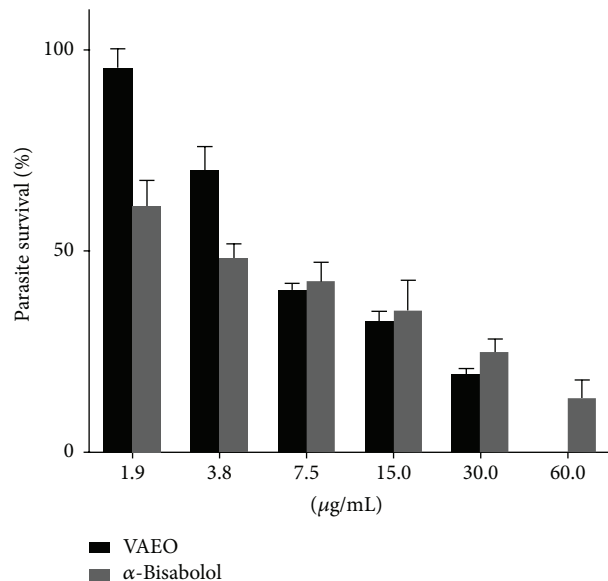


FIGURE 3: Effects of VAEO and α -bisabolol on *Leishmania amazonensis* promastigote forms. Each bar represents the mean \pm standard deviation of three independent experiments in triplicate.

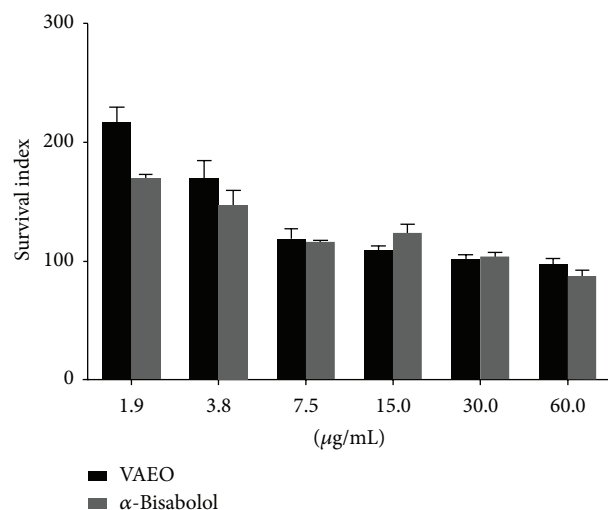


FIGURE 4: Effects of VAEO and α -bisabolol on *Leishmania amazonensis* intracellular amastigotes. Each bar represents the mean \pm standard deviation of three independent experiments in triplicate.

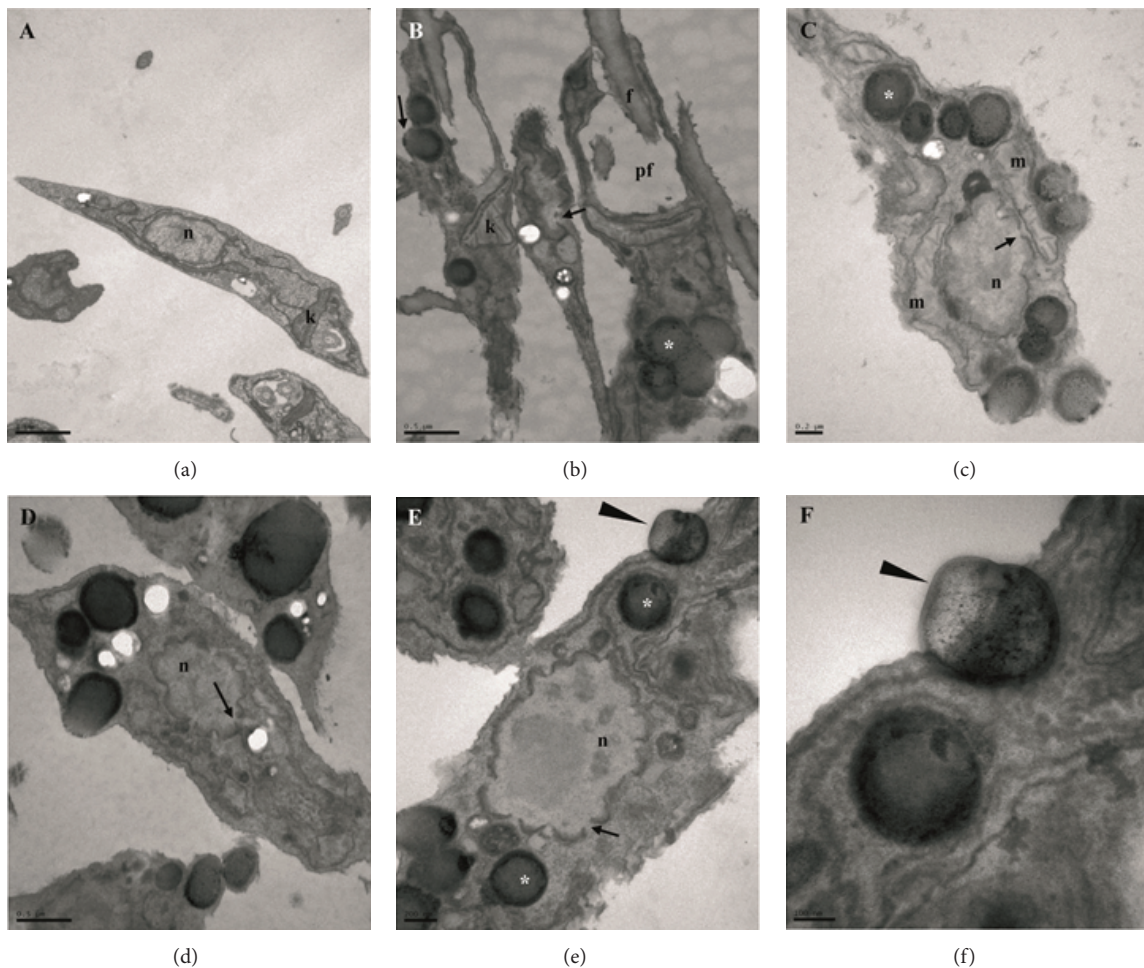


FIGURE 5: Transmission electron microscopy of *Leishmania amazonensis* promastigote with no treatment (a) or incubated with α -bisabolol (b and c) and VAEO (d, e, and f) at concentrations of $30 \mu\text{g/mL}$ for both compounds. Also both treatments show the interaction of lipid droplets with swelling of the plasma membrane. N: nucleus; k: kinetoplast; m: mitochondria; pf: pocket flagellar. Bars $1.0 \mu\text{m}$ (a); $0.5 \mu\text{m}$ (b); $0.2 \mu\text{m}$ (c); $1.0 \mu\text{m}$ (d); $0.5 \mu\text{m}$ (e); 200 nm (f).

shown here, it is possible that α -bisabolol and VAEO have the ability to increase cell permeability to exogenous compounds, since some sesquiterpenes (particularly α -bisabolol) can induce changes in membranes allowing microorganisms to enter the cells and thus augmenting the microbial permeability to antimicrobial agents [36–38].

The ultrastructural analysis of the promastigote control shows the parasite with an elongated cell body and the presence of a well-defined kinetoplast and nucleus (Figure 5).

The promastigotes treated with α -bisabolol for 24 hours, at a concentration of $30 \mu\text{g/mL}$ (Figures 5(b) and 5(c)) presented severe cell damage with the loss of parasite morphology, discontinuity of the nuclear membrane, increased mitochondrial volume and kinetoplast, and presence of vesicles with an electrondense display (white asterisk—Figures 5(b) and 5(c)) with lipid inclusion in the plasma membrane (arrow—Figures 5(b) and 5(c)).

Many terpenes derived from essential oils, such as α -bisabolol, are bioactive, especially against different types of pathogens [39, 40]. Terpenes, hydrocarbons formed from units of isoprene, can easily penetrate the lipid bilayer of

cellular membranes and can thus produce changes in the integrity of cell structures and mitochondrial membranes, for example [40].

The promastigotes treated for 24 h with VAEO at a concentration of $30 \mu\text{g/mL}$ (Figures 5(d), 5(e) and 5(f)) showed increased volumes of flagellar pockets with consequent breakage, increased volumes and changes in mitochondrial kinetoplasts, abnormal condensation of chromatin in the nucleus (small arrow—Figure 5(f)), discontinuity of the nuclear membrane, lipid inclusions in the presence of electrondense vesicles (white asterisk—Figure 5(f)), and visualization of the inclusion of a lipid envelope within the plasma membrane (thin arrow—Figure 5(f)), with the consequent loss of parasite morphology. Similar structural changes have also been described in *Trypanosoma cruzi* [38] and other parasites treated with volatile compounds. Volatile compounds from plants have hydrophobic characteristics with excellent affinity for cell membranes of different microorganisms, including *Leishmania* parasites, which contribute significantly to the toxicity attributed to these compounds [29]. The lipid inclusions observed and the

consequent accumulation of lipid precursors can result from the interference of the oil components, including α -bisabolol, on the lipid biosynthesis pathways, such as ergosterol [37], thus leading to morphological and structural changes affecting the survival of the parasite.

Our results suggest that the essential oil of *Vanillosmopsis arborea* and its major compound, α -bisabolol, show great antileishmanial potential, with the future possibility of a new alternative in the treatment for leishmaniasis or acting as an adjuvant antiprotozoan.

4. Conclusion

The results show that the essential oil *Vanillosmopsis arborea* exhibits leishmanicidal activity *in vitro* against *L. amazonensis* and that this activity is related to the presence of its major compound α -bisabolol. This major compound, α -bisabolol, represents over 97% of the composition of VAEO, and, when tested separately, it also showed similar leishmanicidal activity. The ultrastructural analysis showed that both products induced morphological changes with the presence of cytoplasmic lipid inclusions suggesting an action on the lipid metabolism of the parasite due to increased exocytic activity in the region of the flagellar pocket. Moreover, no cytotoxic effects were observed on the macrophages treated by either product. Further studies with the essential oil of *V. arborea* against cutaneous leishmaniasis will be carried out to demonstrate the potential of natural products derived from this medicinal plant species, thus contributing to advances in allopathic medicine as well as the development of techniques for the conservation of species with potential therapeutic use.

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References

- [1] O. F. Kunle, H. O. Egharevba, and P. O. Ahmadu, "Standardization of herbal medicines—a review," *International Journal of Biodiversity and Conservation*, vol. 4, no. 3, pp. 101–112, 2012.
- [2] T. S. Tiúmana, M. A. Brenzana, T. Ueda-Nakamura, B. P. D. Filho, D. A. G. Corteza, and C. V. Nakamura, "Intramuscular and topical treatment of cutaneous leishmaniasis lesions in mice infected with *Leishmania amazonensis* using coumarin (–) mamea A/BB," *Phytomedicine*, vol. 19, no. 13, pp. 1196–1199, 2012.
- [3] D. S. Corrêa, A. G. Tempone, J. Q. Reimão et al., "Antileishmanial and anti-trypanosomal potential of polygodol isolated from stem barks of *Drimys brasiliensis* Miers (Winteraceae)," *Parasitology Research*, vol. 109, no. 1, pp. 231–236, 2011.
- [4] M. E. Ferreira, A. Rojas de Arias, G. Yaluff et al., "Antileishmanial activity of furoquinolines and coumarins from *Helietta apiculata*," *Phytomedicine*, vol. 17, no. 5, pp. 375–378, 2010.
- [5] J. Alvar, I. D. Velez, C. Bern et al., "Leishmaniasis worldwide and global estimates of its incidence," *PLoS ONE*, vol. 7, no. 5, pp. 1–12, 2012.
- [6] B. L. Herwaldt, "Leishmaniasis," *The Lancet*, vol. 354, no. 9185, pp. 1191–1199, 1999.
- [7] H. W. Murray, J. D. Berman, C. R. Davies, and N. G. Saravia, "Advances in leishmaniasis," *The Lancet*, vol. 366, no. 9496, pp. 1561–1577, 2005.
- [8] M. Ameen, "Cutaneous leishmaniasis: advances in disease pathogenesis, diagnostics and therapeutics," *Clinical and Experimental Dermatology*, vol. 35, no. 7, pp. 699–705, 2010.
- [9] J. Alvar, S. Yactayo, and C. Bern, "Leishmaniasis and poverty," *Trends in Parasitology*, vol. 22, no. 12, pp. 552–557, 2006.
- [10] P. Minodier and P. Parola, "Cutaneous leishmaniasis treatment," *Travel Medicine and Infectious Disease*, vol. 5, no. 3, pp. 150–158, 2007.
- [11] W. Mayrink, A. C. De Carvalho Botelho, P. A. Magalhães et al., "Immunotherapy, immunochemotherapy and chemotherapy for American cutaneous leishmaniasis treatment," *Revista da Sociedade Brasileira de Medicina Tropical*, vol. 39, no. 1, pp. 14–21, 2006.
- [12] R. Reithinger, J.-C. Dujardin, H. Louzir, C. Pirmez, B. Alexander, and S. Brooker, "Cutaneous leishmaniasis," *The Lancet Infectious Diseases*, vol. 7, no. 9, pp. 581–596, 2007.
- [13] A. Dube, N. Singh, A. Saxena, and V. Lakshmi, "Antileishmanial potential of a marine sponge, *Haliclona exigua* (Kirkpatrick) against experimental visceral leishmaniasis," *Parasitology Research*, vol. 101, no. 2, pp. 317–324, 2007.
- [14] J. G. Da Costa, A. R. Campos, S. A. Brito, C. K. B. Pereira, E. O. Souza, and F. F. G. Rodrigues, "Biological screening of araripe basin medicinal plants using *Artemia salina* Leach and pathogenic bacteria," *Pharmacognosy Magazine*, vol. 6, no. 24, pp. 331–334, 2010.
- [15] M. D. G. F. de Medeiros, A. C. da Silva, A. M. D. G. L. Citó et al., "In vitro antileishmanial activity and cytotoxicity of essential oil from *Lippia sidoides* Cham," *Parasitology International*, vol. 60, no. 3, pp. 237–241, 2011.
- [16] L. Monzote, A. M. Montalvo, R. Scull, M. Miranda, and J. Abreu, "Combined effect of the essential oil from *Chenopodium ambrosioides* and antileishmanial drugs on promastigotes of *Leishmania amazonensis*," *Revista do Instituto de Medicina Tropical de Sao Paulo*, vol. 49, no. 4, pp. 257–260, 2007.
- [17] M. R. Santin, A. O. dos Santos, C. V. Nakamura, B. P. Dias Filho, I. C. P. Ferreira, and T. Ueda-Nakamura, "In vitro activity of the essential oil of *Cymbopogon citratus* and its major component (citral) on *Leishmania amazonensis*," *Parasitology Research*, vol. 105, no. 6, pp. 1489–1496, 2009.
- [18] P. Escobar, S. M. Leal, L. V. Herrera, J. R. Martinez, and E. Stashenko, "Chemical composition and antiprotozoal activities of Colombian *Lippia* spp essential oils and their major components," *Memorias do Instituto Oswaldo Cruz*, vol. 105, no. 2, pp. 184–190, 2010.
- [19] M. E. O. Matos, M. P. De Sousa, F. J. A. Matos, and A. A. Craveiro, "Sesquiterpenes from *Vanillosmopsis arborea*," *Journal of Natural Products*, vol. 51, no. 4, pp. 780–782, 1988.
- [20] G. D. O. Leite, L. H. I. Leite, R. D. S. Sampaio et al., "Modulation of topical inflammation and visceral nociception by *Vanillosmopsis arborea* essential oil in mice," *Biomedicine and Preventive Nutrition*, vol. 1, no. 3, pp. 216–222, 2011.
- [21] G. O. Leite, A. R. Penha, C. N. Fernandes, H. H. F. Souza, J. G. M. da Costa, and A. R. Campos, "Gastroprotective mechanism

- of *Vanillosmopsis arborea* bark essential oil," *Fitoterapia*, vol. 80, no. 1, pp. 77–80, 2009.
- [22] F. F. G. Rodrigues, J. G. M. da Costa, and H. D. M. Coutinho, "Synergy effects of the antibiotics gentamicin and the essential oil of *Croton zehntneri*," *Phytomedicine*, vol. 16, no. 11, pp. 1052–1055, 2009.
- [23] S. C. Gonçalves da Costa and P. H. Lagrande, "Development of cell mediated immunity to flagellar antigens and acquired resistance to infection by *Trypanosoma cruzi* in mice," *Memorias do Instituto Oswaldo Cruz*, vol. 76, no. 4, pp. 367–381, 1981.
- [24] T. Mosmann, "Rapid colorimetric assay for cellular growth and survival: application to proliferation and cytotoxicity assays," *Journal of Immunological Methods*, vol. 65, no. 1-2, pp. 55–63, 1983.
- [25] P. Y. Guru, A. K. Agrawal, U. K. Singha, A. Singhal, and C. M. Gupta, "Drug targeting in *Leishmania donovani* infections using tuftsin-bearing liposomes as drug vehicles," *FEBS Letters*, vol. 245, no. 1-2, pp. 204–208, 1989.
- [26] N. K. A. Santos, H. D. M. Coutinho, G. S. B. Viana, F. F. G. Rodrigues, and J. G. M. da Costa, "Chemical characterization and synergistic antibiotic activity of volatile compounds from the essential oil of *Vanillosmopsis arborea*," *Medicinal Chemistry Research*, vol. 20, no. 5, pp. 637–641, 2011.
- [27] I. L. Andrade, J. N. S. Bezerra, M. A. A. Lima et al., "Chemical composition and insecticidal activity of essential oils from *Vanillosmopsis pohlii* baker against *Bemisia argentifolii*," *Journal of Agricultural and Food Chemistry*, vol. 52, no. 19, pp. 5879–5881, 2004.
- [28] E. O. Sousa, A. V. Colares, F. F. G. Rodrigues, A. R. Campos, S. G. Lima, and J. G. M. da Costa, "Effect of collection time on essential oil composition of *Lantana camara* Linn (Verbenaceae) growing in Brazil Northeastern," *Records of Natural Products*, vol. 4, no. 1, pp. 31–37, 2010.
- [29] M. Morales-Yuste, F. Morillas-Márquez, J. Martín-Sánchez, A. Valero-López, and M. C. Navarro-Moll, "Activity of (-)- α -bisabolol against *Leishmania infantum* promastigotes," *Phytomedicine*, vol. 17, no. 3-4, pp. 279–281, 2010.
- [30] F. G. Braga, M. L. M. Bouzada, R. L. Fabri et al., "Antileishmanial and antifungal activity of plants used in traditional medicine in Brazil," *Journal of Ethnopharmacology*, vol. 111, no. 2, pp. 396–402, 2007.
- [31] A. Ahmad, A. Khan, F. Akhtar et al., "Fungicidal activity of thymol and carvacrol by disrupting ergosterol biosynthesis and membrane integrity against *Candida*," *European Journal of Clinical Microbiology and Infectious Diseases*, vol. 30, no. 1, pp. 41–50, 2011.
- [32] M. Wink, "Evolutionary advantage and molecular modes of action of multi-component mixtures used in phytomedicine," *Current Drug Metabolism*, vol. 9, no. 10, pp. 996–1009, 2008.
- [33] L. F. Villegas, A. Marçalo, J. Martin et al., "(+)-epi- α -bisbolol is the wound-healing principle of *Peperomia galioides*: investigation of the in vivo wound-healing activity of related terpenoids," *Journal of Natural Products*, vol. 64, no. 10, pp. 1357–1359, 2001.
- [34] A. Hernández-Ceruelos, E. Madrigal-Bujaidar, and C. de la Cruz, "Inhibitory effect of chamomile essential oil on the sister chromatid exchanges induced by daunorubicin and methyl methanesulfonate in mouse bone marrow," *Toxicology Letters*, vol. 135, no. 1-2, pp. 103–110, 2002.
- [35] E. N. Loria-Cervera, E. I. Sosa-Bibiano, L. E. Villanueva-Lizama et al., "Nitric oxide production by *Peromyscus yucatanicus* (Rodentia) infected with *Leishmania* (*Leishmania*) *mexicana*," *Memórias Do Instituto Oswaldo Cruz*, vol. 108, no. 2, pp. 172–177, 2013.
- [36] Z. Martín-Quintal, M. D. R. García-Miss, M. Mut-Martín, A. Matus-Moo, L. W. Torres-Tapia, and S. R. Peraza-Sánchez, "The leishmanicidal effect of (3S)-16,17-didehydrofalcarinol, an oxylipin isolated from *Tridax procumbens*, is independent of NO production," *Phytotherapy Research*, vol. 24, no. 7, pp. 1004–1008, 2010.
- [37] E. Nibret and M. Wink, "Trypanocidal and antileukaemic effects of the essential oils of *Hagenia abyssinica*, *Leonotis ocyimifolia*, *Moringa stenopetala*, and their main individual constituents," *Phytomedicine*, vol. 17, no. 12, pp. 911–920, 2010.
- [38] B. F. Brehm-Stecher and E. A. Johnson, "Sensitization of *Staphylococcus aureus* and *Escherichia coli* to antibiotics by the sesquiterpenoids nerolidol, farnesol, bisabolol, and apritone," *Antimicrobial Agents and Chemotherapy*, vol. 47, no. 10, pp. 3357–3360, 2003.
- [39] M. D. S. S. Rosa, R. R. Mendonça-Filho, H. R. Bizzo et al., "Antileishmanial activity of a linalool-rich essential oil from *Croton cajucara*," *Antimicrobial Agents and Chemotherapy*, vol. 47, no. 6, pp. 1895–1901, 2003.
- [40] G. F. Santoro, M. G. Cardoso, L. G. L. Guimarães, J. M. Freire, and M. J. Soares, "Anti-proliferative effect of the essential oil of *Cymbopogon citratus* (DC) Stapf (lemongrass) on intracellular amastigotes, bloodstream trypomastigotes and culture epimastigotes of *Trypanosoma cruzi* (Protozoa: Kinetoplastida)," *Parasitology*, vol. 134, no. 11, pp. 1649–1656, 2007.



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