

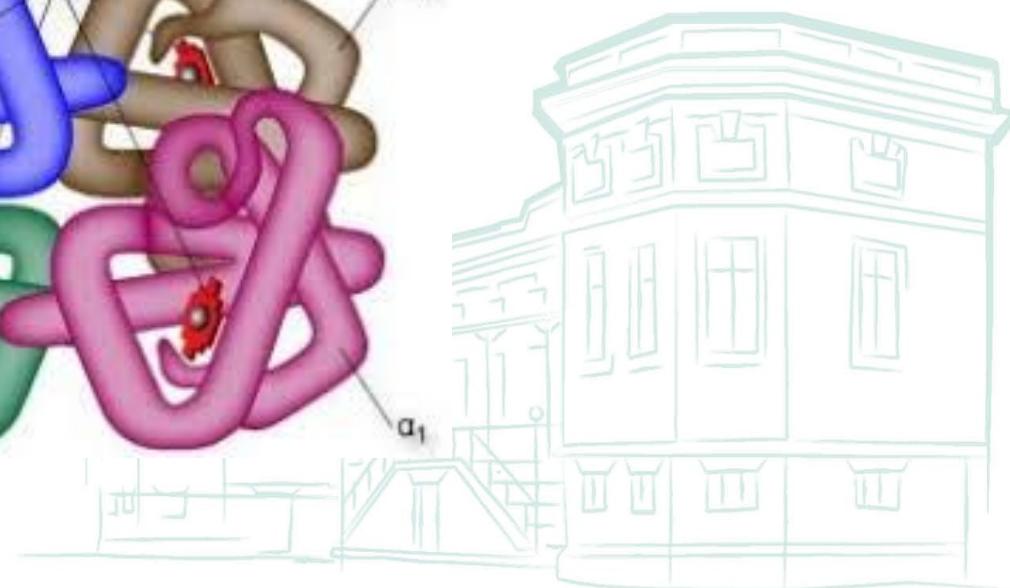
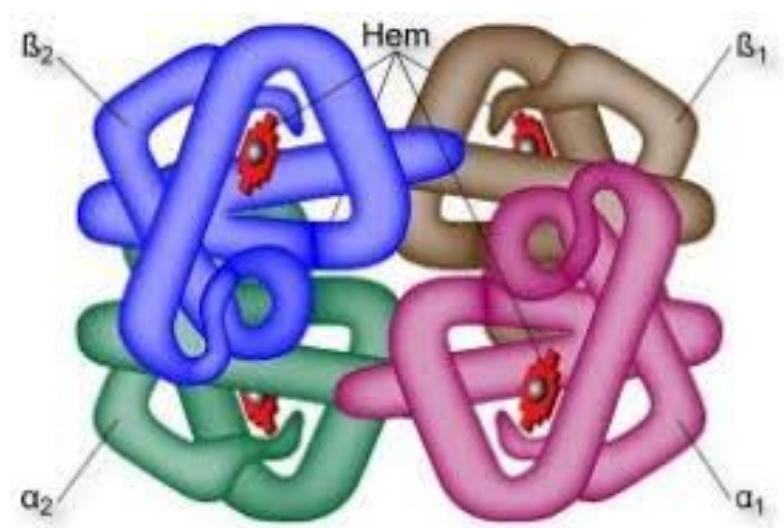
# Iron metabolism in pathogenic *Sporothrix* spp.

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INI/Fiocruz



# Iron importance in living organisms

- Cofactor of several enzymes
- Oxygen carriers
- Electron-transfer systems



# Iron in infectious diseases

- Iron withholding as a defense mechanism

Iron withholding defense system.

#### *Constitutive components*

##### Siderophilins

Transferrin in plasma, lymph, cerebrospinal fluid

Lactoferrin in secretions of lachrymal & mammary glands and of respiratory, gastrointestinal and genital tracts

Ferritin within host cells

#### *Processes induced at times of invasion*

Prompt reduction in plasma iron & in absorption of iron from diet

Increased hepatic synthesis of hepcidin (ferroportin inactivator) to suppress release of iron by macrophages and by duodenal enterocytes. Increased synthesis of macrophage ferritin & decreased synthesis of ferroportin.

Removal of iron from sites of invasion

Release of neutrophils from bone marrow into circulation and then into invasion sites. Release of apolactoferrin from neutrophil granules followed by binding of iron in septic sites. Macrophage scavenging of hololactoferrin. Hepatic release of haptoglobin & hemopexin (to bind extracellular hemoglobin & hemin).

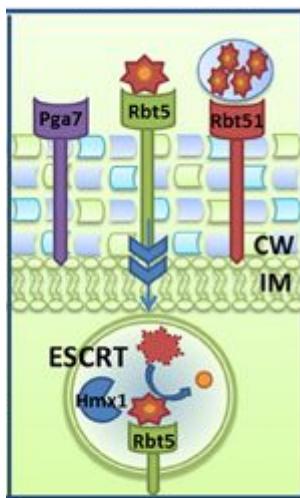
Macrophage suppression of microbial iron metabolism by synthesis of lipocalins (to bind siderophores), of nitric oxide ( to disrupt invader iron metabolism) and of Nramp1 (to withhold growth-essential iron from invaders).

Induction in B lymphocytes of immunoglobulins to microbial cell surface receptors that bind either ferrated siderophores, ferrated siderophilins, hemoglobin, or heme.

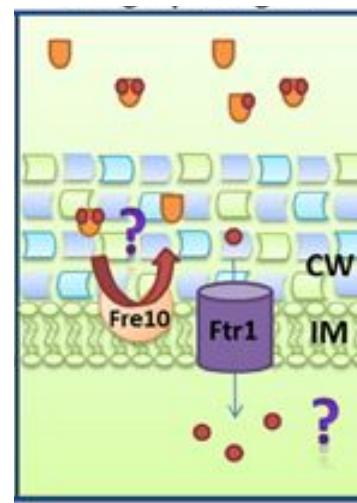
Weinberg, Biochim Biophys Acta. 2009  
Jul;1790(7):600-5.

# Fungal strategies for iron acquisition

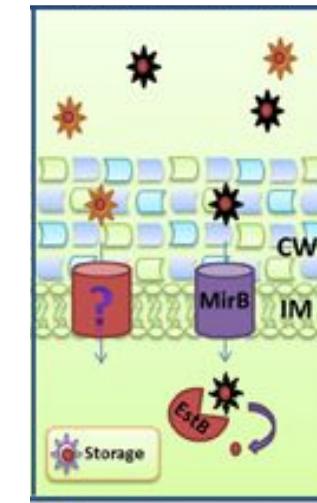
Hemoglobin  
and heme  
uptake



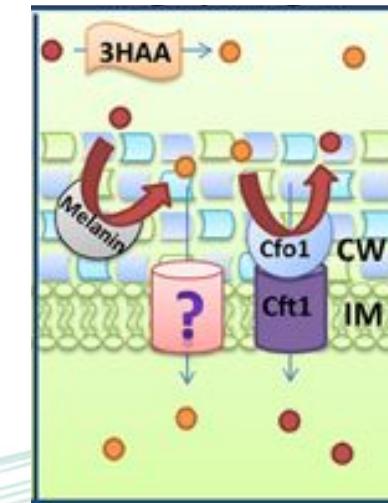
Acquisition  
form transferin



siderophores

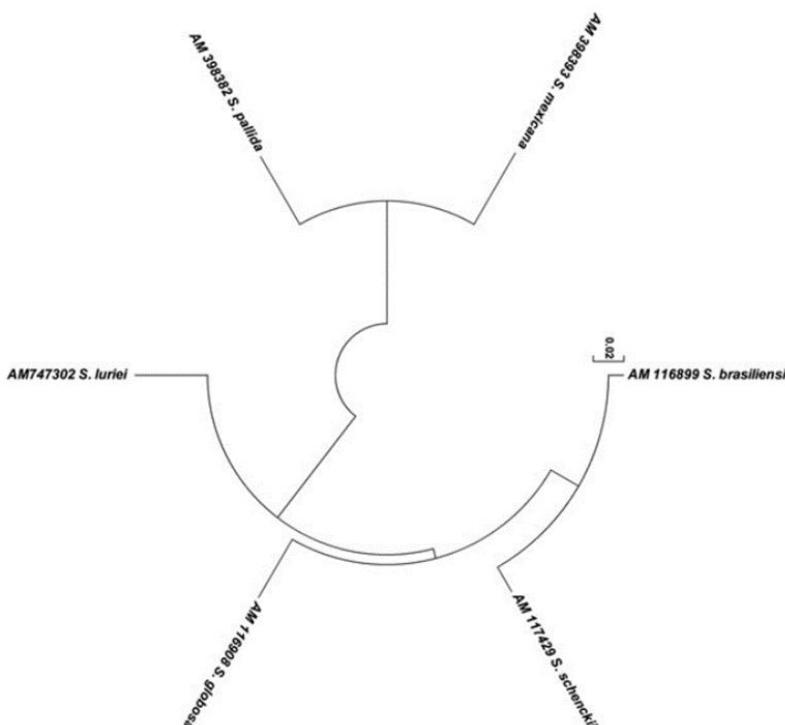
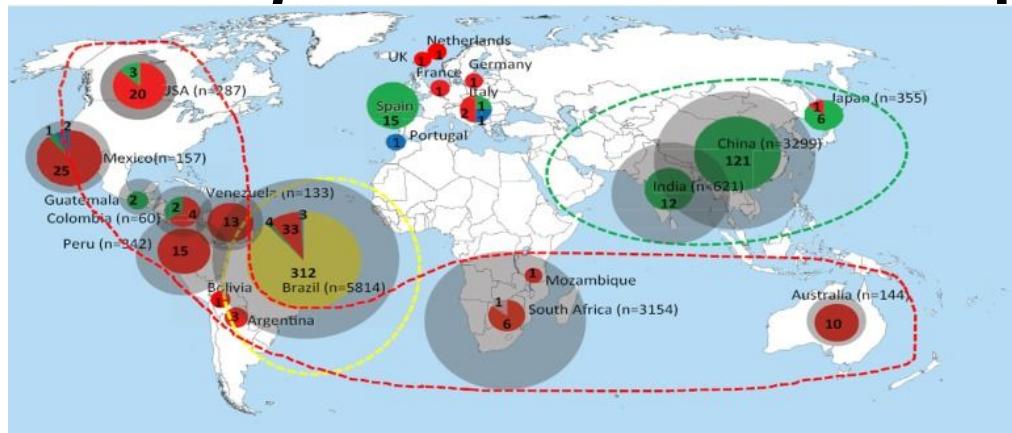


Fe<sup>2+</sup> uptake



Caza & Kronstad. Front Cell Infect Microbiol. 2013 Nov 19;3:80.

# *Sporothrix* and sporotrichosis



# *Sporothrix schenckii* and *Sporothrix brasiliensis* genomes

Teixeira et al. BMC Genomics 2014, 15:943  
<http://www.biomedcentral.com/1471-2164/15/943>



RESEARCH ARTICLE

Open Access

## Comparative genomics of the major fungal agents of human and animal Sporotrichosis: *Sporothrix schenckii* and *Sporothrix brasiliensis*

Marcus M Teixeira<sup>1</sup>, Luiz GP de Almeida<sup>2</sup>, Paula Kubitschek-Barreira<sup>3</sup>, Fernanda L Alves<sup>4,5</sup>, Érika S Kioshima<sup>1,6</sup>, Ana KR Abadio<sup>1</sup>, Larissa Fernandes<sup>7</sup>, Lorena S Derengowski<sup>1</sup>, Karen S Ferreira<sup>8</sup>, Rangel C Souza<sup>2</sup>, Jerônimo C Ruiz<sup>5</sup>, Nathalia C de Andrade<sup>3</sup>, Hugo C Paes<sup>1</sup>, André M Nicola<sup>9,10</sup>, Patrícia Albuquerque<sup>1,10</sup>, Alexandra L Gerber<sup>2</sup>, Vicente P Martins<sup>1</sup>, Luisa DF Peconick<sup>1</sup>, Alan Viggiano Neto<sup>1</sup>, Claudia B Chaucanez<sup>1</sup>, Patrícia A Silva<sup>1</sup>, Oberdan L Cunha<sup>2</sup>, Fabiana FM de Oliveira<sup>1</sup>, Tayná C dos Santos<sup>1</sup>, Amanda LN Barros<sup>1</sup>, Marco A Soares<sup>4</sup>, Luciana M de Oliveira<sup>4,11</sup>, Marjorie M Marini<sup>12</sup>, Héctor Villalobos-Duno<sup>13</sup>, Marcel ML Cunha<sup>3</sup>, Sybren de Hoog<sup>14</sup>, José F da Silveira<sup>12</sup>, Bernard Henrissat<sup>15</sup>, Gustavo A Niño-Vega<sup>13</sup>, Patrícia S Cisalpino<sup>5</sup>, Héctor M Mora-Montes<sup>16</sup>, Sandro R Almeida<sup>17</sup>, Jason E Stajich<sup>18</sup>, Leila M Lopes-Bezerra<sup>3</sup>, Ana TR Vasconcelos<sup>2</sup> and Maria SS Felipe<sup>1,9\*</sup>

# *In silico* analysis of *Sporothrix* spp. genomes

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blastn blastp blastx tblastn tblastx

Enter Query Sequence

Enter accession number(s), gi(s), or FASTA sequence(s) [?](#) Clear Query subrange [?](#)  
From  To

BLASTP programs search protein databases using a protein query. [more...](#) Reset page Bookmark

Enter accession number(s), gi(s), or FASTA sequence(s) [?](#) Clear Query subrange [?](#)  
From  To

Or, upload file [Escolher arquivo](#) Nenhum arquivo selecionado [?](#)

Job Title XP\_755103 L-ornithine N5-oxygenase SidA [Aspergillus...  
Enter a descriptive title for your BLAST search [?](#)

Align two or more sequences [?](#)

Choose Search Set

Database Non-redundant protein sequences (nr) [?](#)  
Organism Sporothrix (taxid:29907)  Exclude [+](#)  
Optional Enter organism common name, binomial, or tax id. Only 20 top taxa will be shown. [?](#)

Exclude Models (XM/XP)  Uncultured/environmental sample sequences  
Optional

Entrez Query Create custom database  
Optional Enter an Entrez query to limit search [?](#)

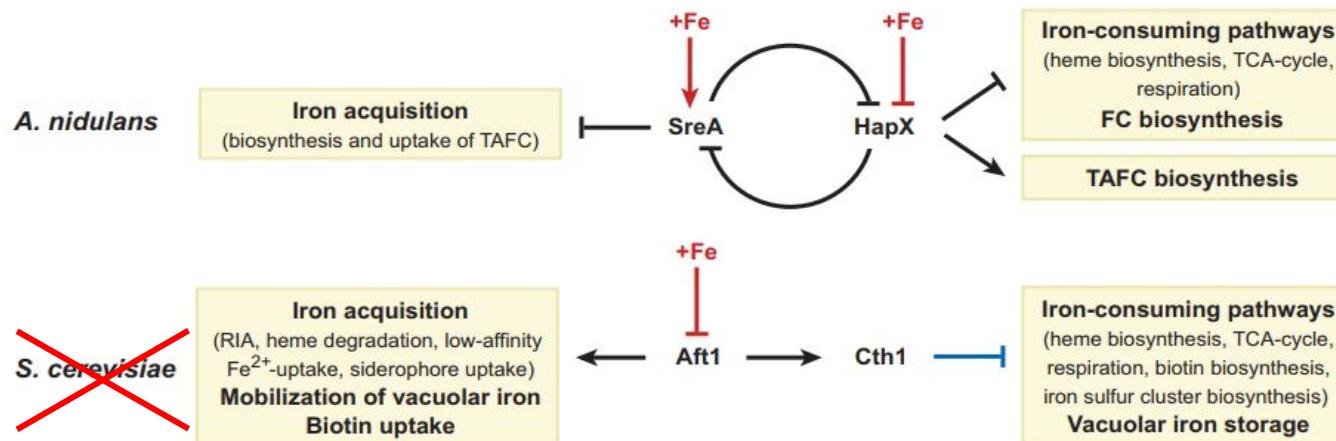
Program Selection

Algorithm Quick BLASTP (Accelerated protein-protein BLAST) [New](#)  
 blastp (protein-protein BLAST)  
 PSI-BLAST (Position-Specific Iterated BLAST)  
 PHI-BLAST (Pattern Hit Initiated BLAST)  
 DELTA-BLAST (Domain Enhanced Lookup Time Accelerated BLAST)  
Choose a BLAST algorithm [?](#)

**BLAST** Search database Non-redundant protein sequences (nr) using Blastp (protein-protein BLAST)  
 Show results in a new window

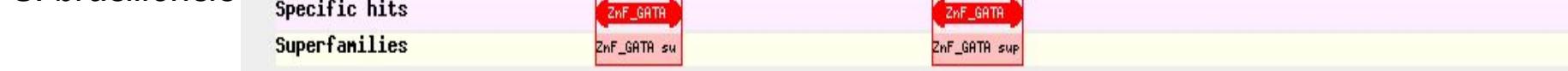
[Algorithm parameters](#)

# Regulation of iron metabolism

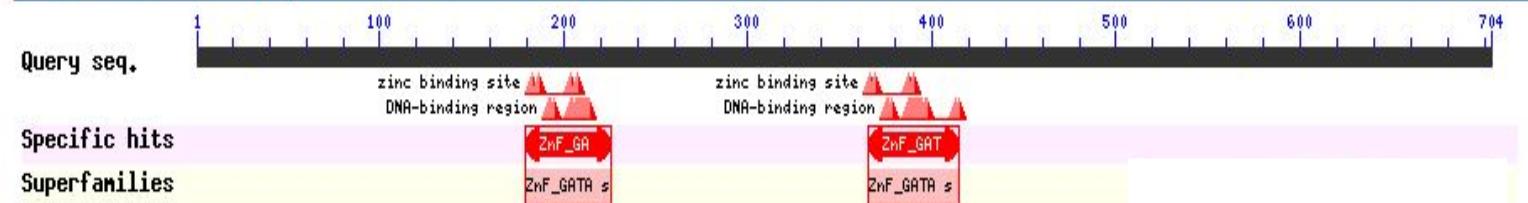


Hass et al. Annu Rev Phytopathol. 2008;  
46:149-187.

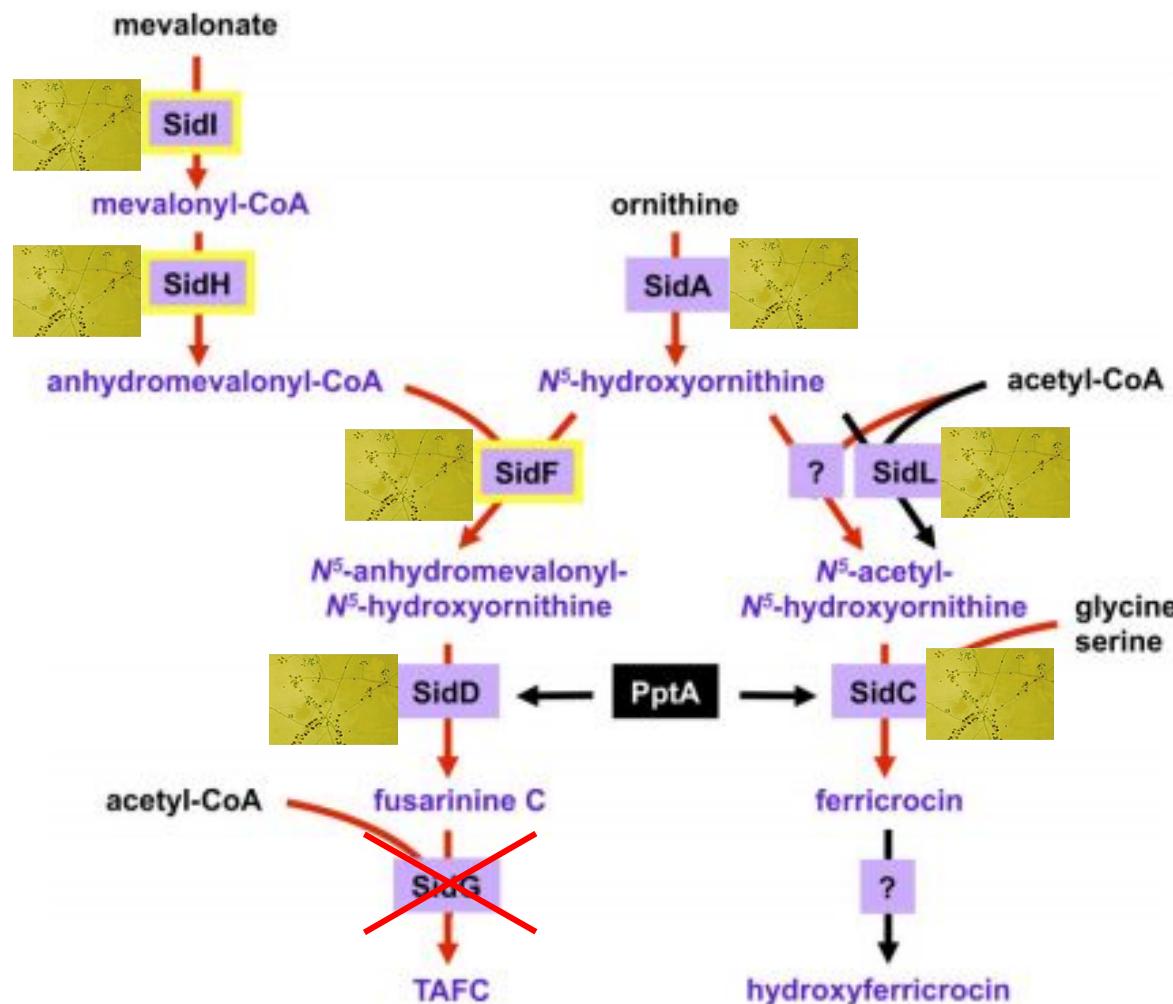
*S. brasiliensis*



*S. schenckii*

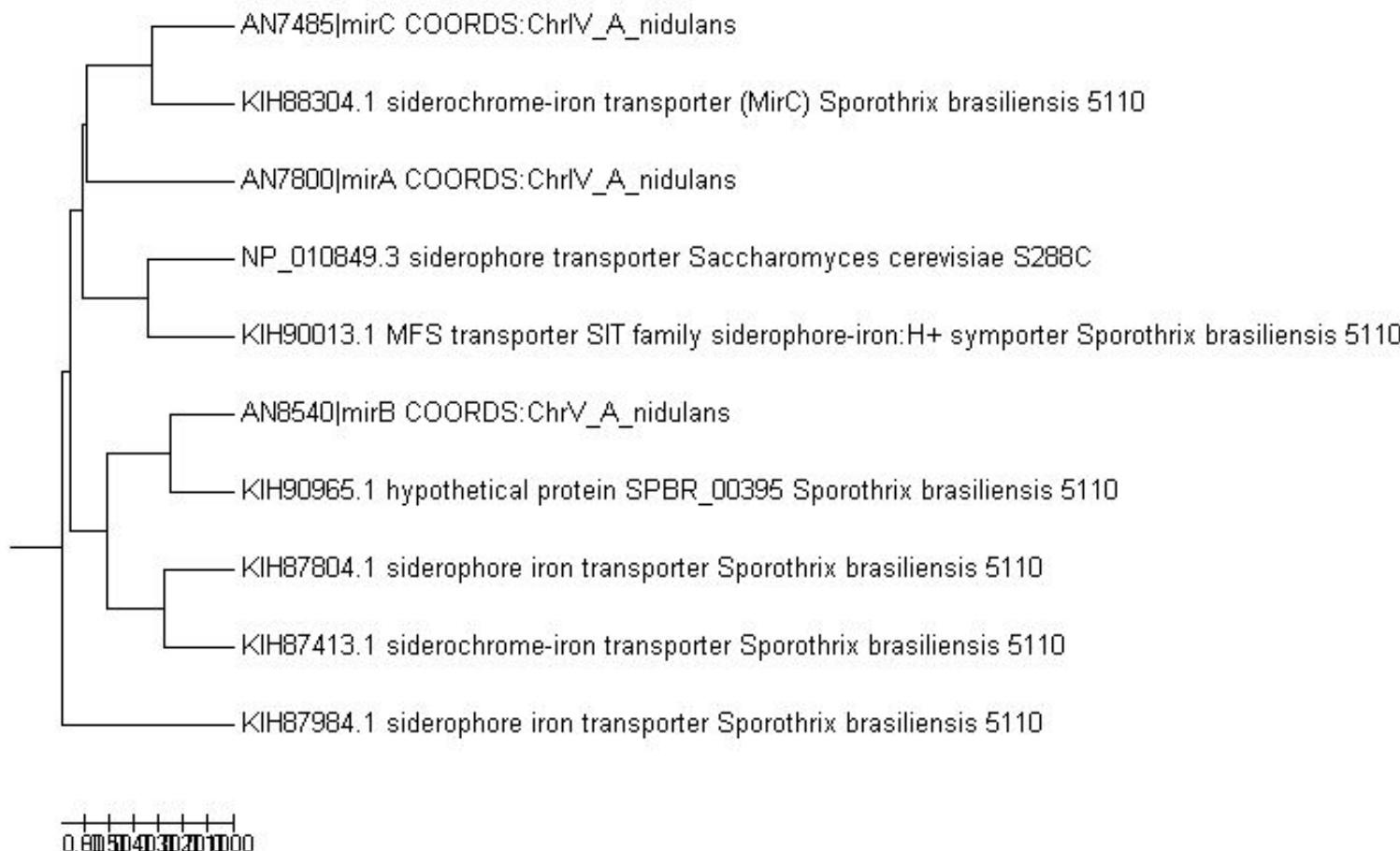


# *Sporothrix* spp. siderophores

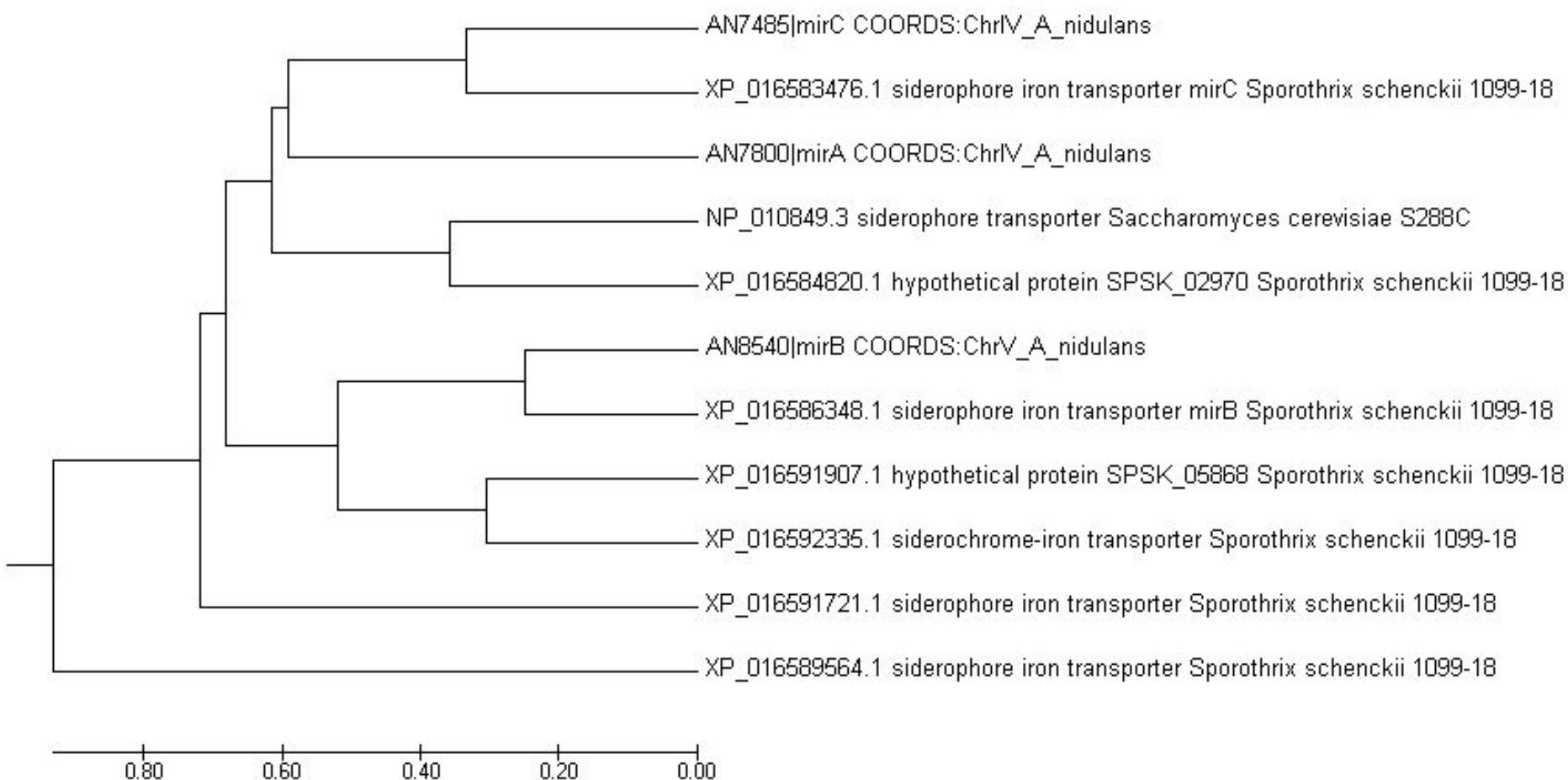


Adapted from Grundlinger et al. Mol Microbiol. 2013; 88(5): 669-675

# *Sporothrix brasiliensis* siderophore transporters

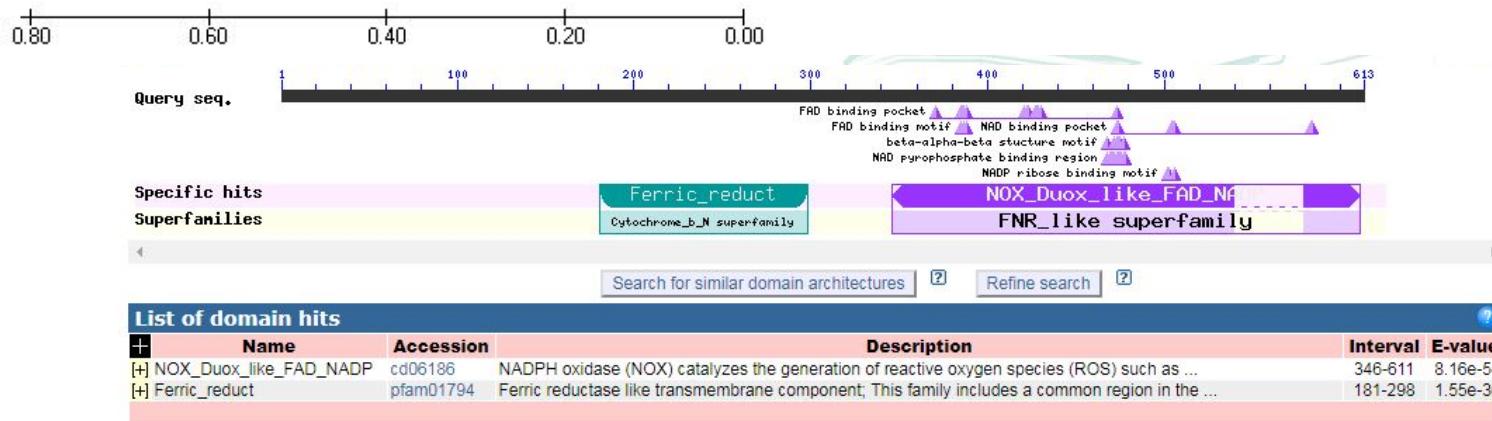
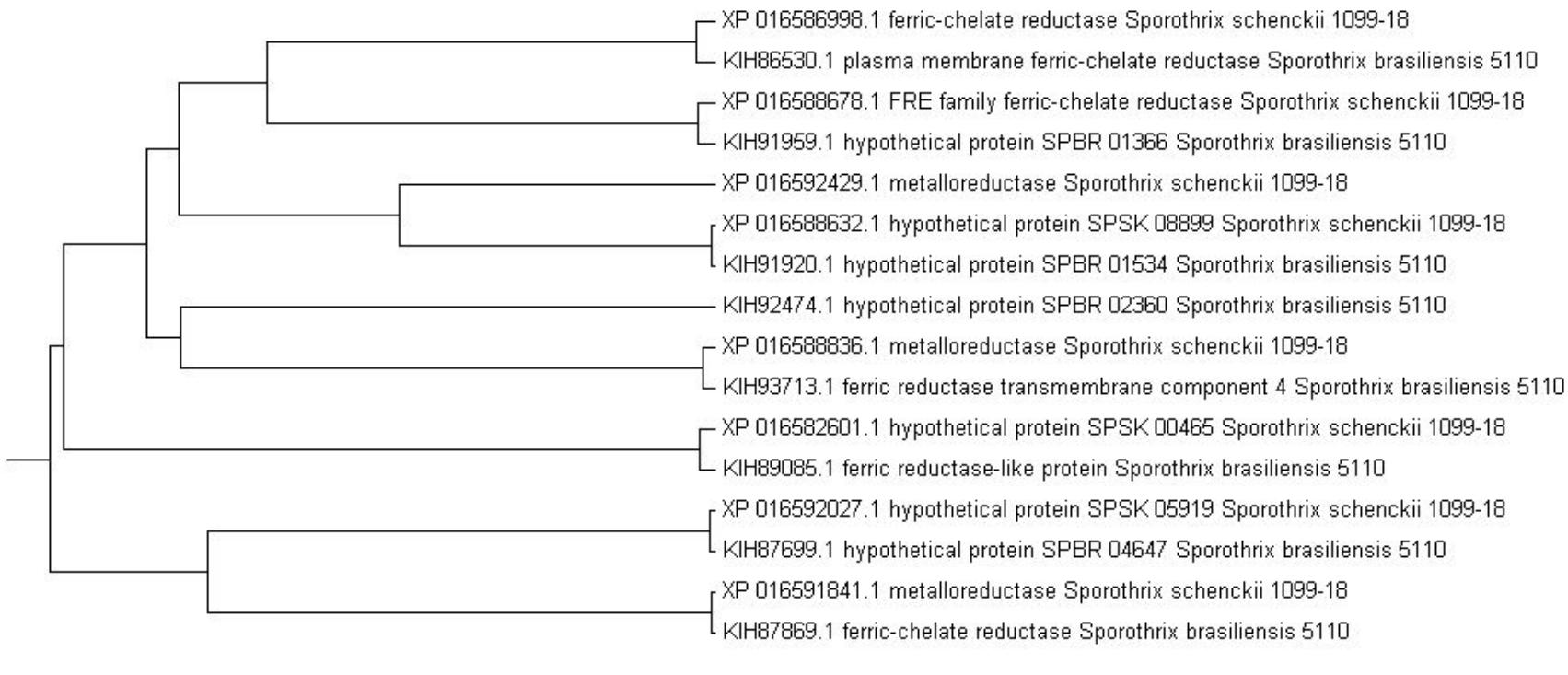


# *Sporothrix schenckii* siderophore transporters

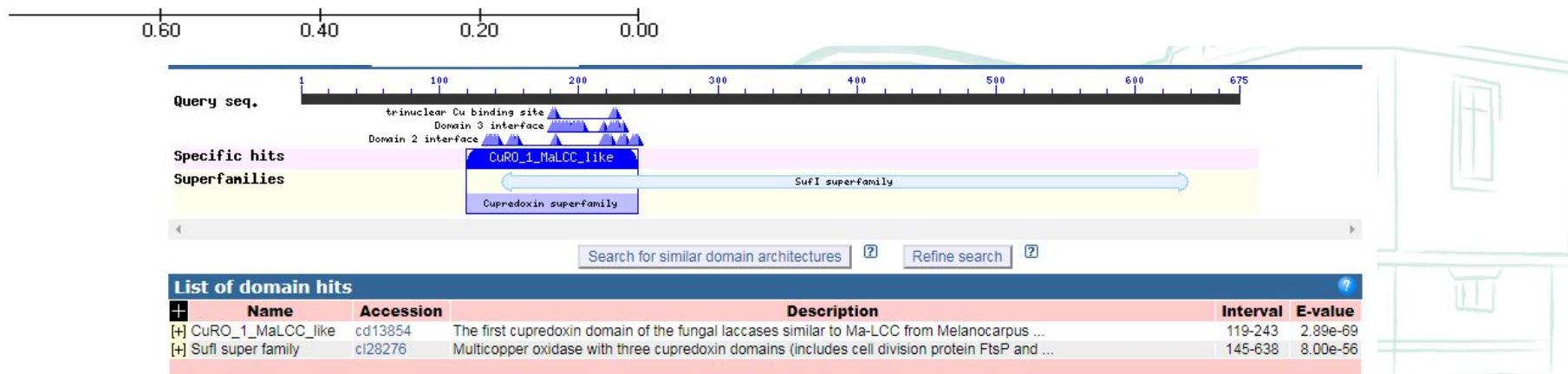
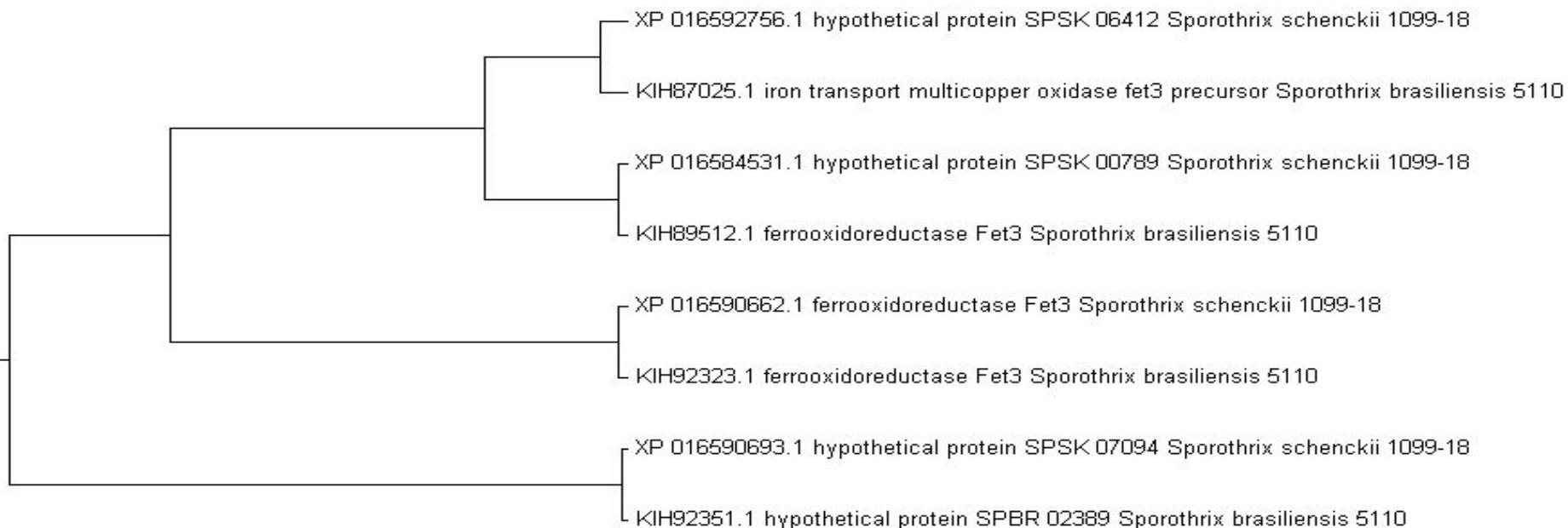




# Reductive iron uptake

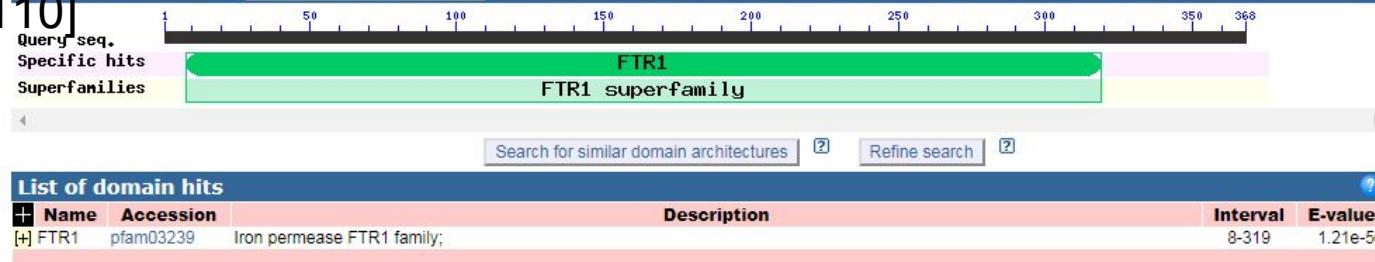


# Ferroxidases

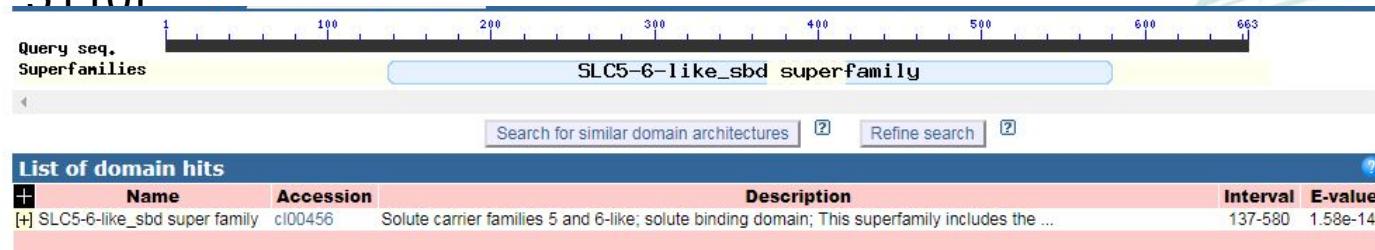


# Iron transporters

KIH92322.1 high-affinity iron transporter [Sporothrix brasiliensis  
5110]



KIH92556.1 metal iron transporter [Sporothrix brasiliensis  
5110]



# Heme uptake

Molecular Microbiology (2004) 53(4), 1209–1220

doi:10.1111/j.1365-2958.2004.04199.x

## A family of *Candida* cell surface haem-binding proteins involved in haemin and haemoglobin-iron utilization

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### Summary

The ability to acquire iron from host tissues is a major virulence factor of pathogenic microorganisms. *Candida albicans* is an important fungal pathogen, responsible for an increasing proportion of systemic infections. *C. albicans*, like many pathogenic bacteria, is able to utilize haemin and haemoglobin as iron sources. However, the molecular basis of this pathway in pathogenic fungi is unknown. Here, we identify a conserved family of plasma membrane-anchored proteins as haem-binding proteins that are involved in haem-iron utilization. We isolated RBT51 as a gene that is sufficient by itself to confer to *S. cerevisiae* the ability to utilize haemoglobin iron. RBT51 is highly homologous to RBT5, which was previously identified as a gene negatively regulated by the transcriptional suppressor CaTup1. Rbt5 and Rbt51 are mannosylated proteins that carry the conserved CFEM domain.

ardson and Warnock, 1997). The ability to cause systemic infection depends on specific virulence factors that enable *C. albicans* to adhere to epithelial cells, penetrate tissues and cause damage (Hube et al., 1997; Lo et al., 1997; Sanglard et al., 1997; Gale et al., 1998). In addition, like all microbial pathogens, *C. albicans* needs to be able to extract nutrients from the host environment in order to proliferate. An important limiting nutrient in host tissues is iron: it has long been known that a number of host proteins play key defence roles against microorganisms by withholding free iron, a defence method that has been called 'nutritional immunity' (Kochan, 1973; Weinberg, 1975); conversely, successful pathogens have by necessity evolved mechanisms to acquire iron from the host tissues (Payne, 1993). Different mechanisms for extracting iron from host proteins have been characterized in bacterial pathogens, including utilization of haem and haemoglobin iron (Ratledge and Dover, 2000). The pathways of haemoglobin iron utilization were extensively characterized in Gram-negative bacteria (reviewed in Genco and Dixon, 2001), and a haem-iron acquisition apparatus was recently described in a Gram-positive pathogen (Dryla et al., 2003; Mazmanian et al., 2003).

Most of our information on fungal iron uptake mechanisms was derived from studies in the related, but non-

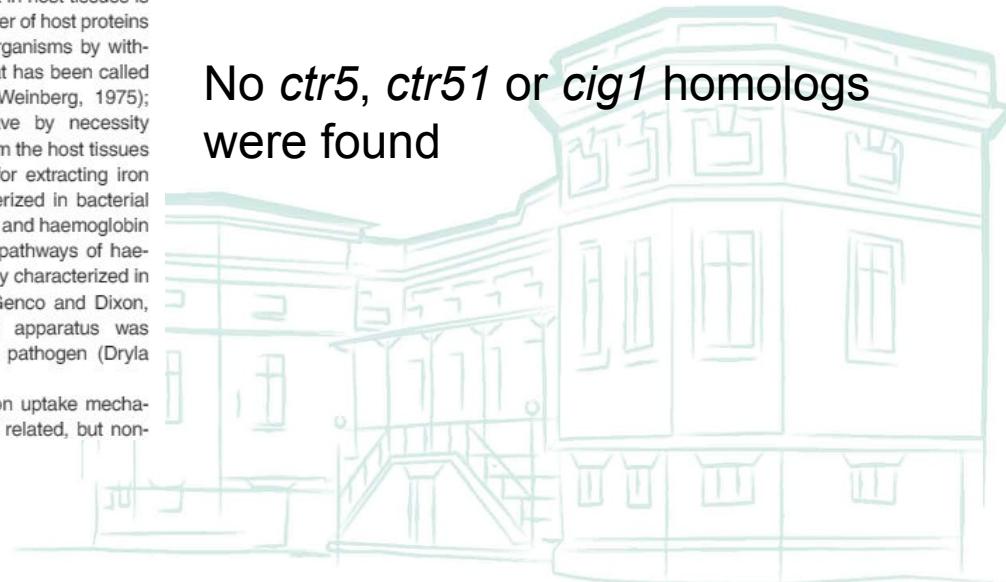
## The Mannoprotein Cig1 Supports Iron Acquisition From Heme and Virulence in the Pathogenic Fungus *Cryptococcus neoformans*

Brigitte Cadieux,<sup>1,2</sup> Tianshun Lian,<sup>1,4</sup> Guanggan Hu,<sup>1</sup> Joyce Wang,<sup>1,5</sup> Carmelo Biondo,<sup>3</sup> Giuseppe Teti,<sup>3</sup> Victor Liu,<sup>1</sup> Michael E. P. Murphy,<sup>2</sup> A. Louise Creagh,<sup>1</sup> and James W. Kronstad<sup>1,2</sup>

<sup>1</sup>Michael Smith Laboratories and <sup>2</sup>Department of Microbiology and Immunology, The University of British Columbia, Vancouver, Canada; and

<sup>3</sup>Department of Pathology and Experimental Microbiology, University of Messina, Italy

No *ctr5*, *ctr51* or *cig1* homologs were found

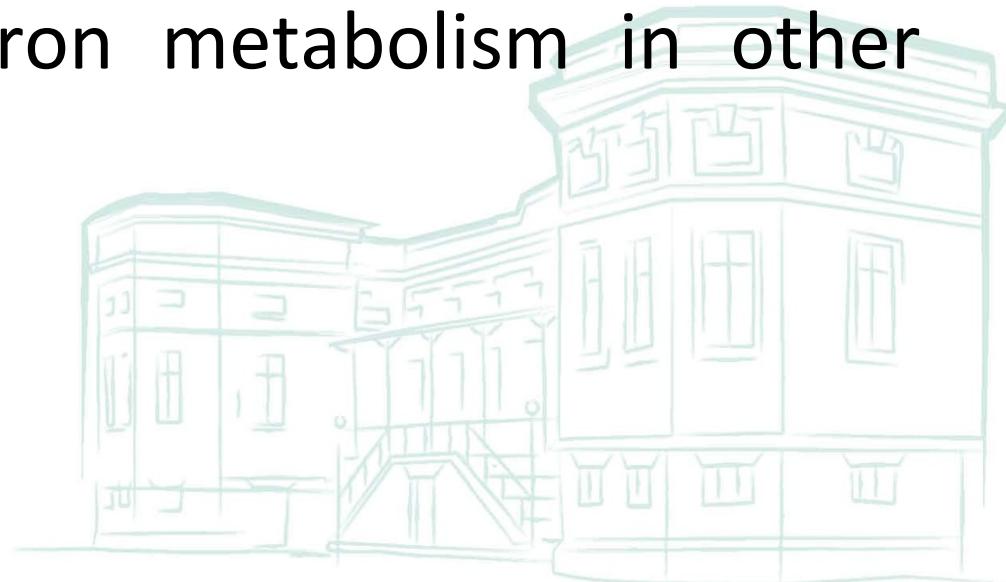


# Conclusions

- Strategies for iron acquisition by *S. brasiliensis* and *S. schenckii* are similar;
- Both species produce siderophores, high and low affinity iron transporters, and reductases;
- Heme utilization by *Sporothrix* spp. remains to be explored.

# Perspectives

- Studies on iron-related gene expression;
- Exploration of heme utilization by *Sporothrix*;
- Characterization of iron metabolism in other *Sporothrix* species.



# Acknowledgements

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  - Dr. Mirelle Garcia Silva Bailão
  - Dr. Lilian Cristiane Baeza