

Systemic Inflammation in Pregnant Women with Latent Tuberculosis Infection

Shilpa Naik¹, Mallika Alexander^{2, 1}, Pavan Kumar³, Vandana Kulkarni^{2, 1}, Prasad Deshpande^{2, 1}, Su Yadana⁴, Cheng-Shiun Leu⁴, Mariana Araújo-Pereira⁵, Bruno B. Andrade⁵, Ramesh Bhosale¹, Subash Babu³, Amita Gupta⁶, Jyoti S. Mathad⁷, Rupak Shivakoti^{8*}

¹B. J. Medical College & Sassoon Hospital, India, ²B. J. Medical College & Sassoon Hospital, India, ³International Centers for Excellence in Research (ICER), India, ⁴Columbia University, United States, ⁵Gonçalo Moniz Institute (IGM), Brazil, ⁶Johns Hopkins Medicine, United States, ⁷Weill Cornell Medicine, Cornell University, United States, ⁸Department of Epidemiology, Columbia University, United States

Submitted to Journal: Frontiers in Immunology

Specialty Section: Microbial Immunology

ISSN: 1664-3224

Article type: Original Research Article

Received on: 26 Jul 2020

Accepted on: 09 Dec 2020

Provisional PDF published on: 09 Dec 2020

Frontiers website link: www.frontiersin.org

Citation:

Naik S, Alexander M, Kumar P, Kulkarni V, Deshpande P, Yadana S, Leu C, Araújo-pereira M, Andrade BB, Bhosale R, Babu S, Gupta A, Mathad JS and Shivakoti R(2020) Systemic Inflammation in Pregnant Women with Latent Tuberculosis Infection. *Front. Immunol.* 11:3619. doi:10.3389/fimmu.2020.587617

Copyright statement:

© 2020 Naik, Alexander, Kumar, Kulkarni, Deshpande, Yadana, Leu, Araújo-pereira, Andrade, Bhosale, Babu, Gupta, Mathad and Shivakoti. This is an open-access article distributed under the terms of the <u>Creative Commons Attribution License (CC BY</u>). The use, distribution and reproduction in other forums is permitted, provided the original author(s) or licensor are credited and that the original publication in this journal is cited, in accordance with accepted academic practice. No use, distribution or reproduction is permitted which does not comply with these terms.

This Provisional PDF corresponds to the article as it appeared upon acceptance, after peer-review. Fully formatted PDF and full text (HTML) versions will be made available soon.

Frontiers in Immunology | www.frontiersin.org





1	Systemic Inflammation in Pregnant Women with
2	Latent Tuberculosis Infection
3	Authors: Shilpa Naik ^{1,2} , Mallika Alexander ¹ , Pavan Kumar ³ , Vandana Kulkarni ¹ , Prasad
4	Deshpande ¹ , Su Yadana ⁴ , Cheng-Shiun Leu ⁴ , Mariana Araújo-Pereira ^{5,6,7} , Bruno B Andrade ^{5, 6, 7, 8,}
5	^{9,10} , Ramesh Bhosale ^{1,2} , Subash Babu ³ , Amita Gupta ^{1,11} , Jyoti S Mathad ¹² , and Rupak Shivakoti ⁴
6	
7	Institutions:
8	¹ Byramjee-Jeejeebhoy Government Medical college-Johns Hopkins University Clinical Research
9	Site, Pune, India
10	² Byramjee Jeejeebhoy Government Medical College, Pune, India
11	³ National Institutes of Health, National Institute for Research in Tuberculosis, International Center
12	for Excellence in Research, Chennai, India
13	⁴ Department of Epidemiology, Columbia University Mailman School of Public Health, New York,
14	USA
15	⁵ Instituto Goncalo Moniz, Fundação Oswaldo Cruz, Salvador, Brazil
16	⁶ Multinational Organization Network Sponsoring Translational and Epidemiological Research,
17	Fundação José Silveira, Salvador, Brazil
18	⁷ Faculdade de Medicina, Universidade Federal da Bahia, Salvador, Brazil
19	⁸ Curso de Medicina, Faculdade de Tecnologia e Ciências, Salvador, Brazil
20	⁹ Universidade Salvador (UNIFACS), Laureate Universities, Salvador, Brazil
21	¹⁰ Escola Bahiana de Medicina e Saúde Pública (EBMSP), Salvador, Brazil

22	¹¹ Department of Medicine, Johns Hopkins University School of Medicine, Baltimore, USA.
23	¹² Department of Medicine, Weill Cornell Medical College, New York, USA
24	Keywords: Latent TB infection, TB disease, inflammation, pregnancy, cytokines
25	
26	
27	
28	
29	
30	
31	
32	
33	
34	
35	
36	
37	
38	
39	
40	
41	
42	
43	
44	
45	
46	

47 Abstract

Background: Recent studies in adults have characterized differences in systemic inflammation
between adults with and without latent tuberculosis infection (LTBI+ vs. LTBI-). Potential
differences in systemic inflammation by LTBI status has not been assess in pregnant women.
Methods: We conducted a cohort study of 155 LTBI+ and 65 LTBI- pregnant women, stratified by

HIV status, attending an antenatal clinic in Pune, India. LTBI status was assessed by interferon gamma release assay. Plasma was used to measure systemic inflammation markers using immunoassays: IFNβ, CRP, AGP, I-FABP, IFNγ, IL-1β, soluble CD14 (sCD14), sCD163, TNF, IL-6, IL-17a and IL-13. Linear regression models were fit to test the association of LTBI status with each inflammation marker. We also conducted an exploratory analysis using logistic regression to test the association of inflammatory markers with TB progression.

59

Results: Study population was a median age of 23 (Interquartile range: 21-27), 28% undernourished
(mid-upper arm circumference (MUAC) <23 cm), 12% were vegetarian, 10% with gestational
diabetes and 32% with HIV. In multivariable models, LTBI+ women had significantly lower levels
of third trimester AGP, IL1β, sCD163, IL-6 and IL-17a. Interestingly, in exploratory analysis,
LTBI+ TB progressors had significantly higher levels of IL1β, IL-6 and IL-13 in multivariable
models compared to LTBI+ non-progressors.

66

67 Conclusions: Our data shows a distinct systemic immune profile in LTBI+ pregnant women
68 compared to LTBI- women. Data from our exploratory analysis suggest that LTBI+ TB progressors
69 do not have this immune profile, suggesting negative association of this profile with TB progression.

Inflammation in LTBI+ pregnant women

70	If other studies confirm these differences by LTBI status and show a causal relationship with TB
71	progression, this immune profile could identify subsets of LTBI+ pregnant women at high risk for
72	TB progression and who can be targeted for preventative therapy.
73	
74	
75	
76	
77	
78	
79	
80	
81	
82	
83	
84	
85	
86	
87	
88	
89	
90	

91 Introduction:

92 Active tuberculosis (TB) disease elicits host responses characterized by an immune profile 93 that is clearly distinct from healthy individuals (O'Garra et al., 2013; Cliff et al., 2015). As the 94 causative agent *Mycobacterium tuberculosis (Mtb)* is actively replicating during TB disease, it causes 95 constant antigen stimulation from the bacterium that shapes the immune response. In contrast, with 96 latent TB infection (LTBI), *Mtb* is not actively replicating in the host and antigen stimulation with 97 *Mtb* antigens is required to generate *Mtb*-specific immune responses (O'Garra et al., 2013). While 98 differences in immunity with *Mtb* antigen stimulation has been extensively studied for active disease 99 or LTBI compared to healthy individuals (Tufariello et al., 2003; Mack et al., 2009; O'Garra et al., 100 2013; Cliff et al., 2015; de Martino et al., 2019), there are limited studies characterizing differences 101 by LTBI status in circulating inflammatory markers, in the absence of antigen stimulation (Cowan et 102 al., 2012; Jensen et al., 2013; LaVergne et al., 2020). This information could potentially explain why 103 an increased risk of certain adverse outcomes (e.g. acute myocardial infarction) have been observed 104 among LTBI+ individuals, or help identify immune profiles associated with TB progression 105 (Andrews et al., 2012; Huaman et al., 2018b). 106 One hypothesis on levels of inflammation by LTBI status is that there is no difference in 107 circulating inflammatory markers between LTBI+ and LTBI- individuals. *Mtb* infection is mainly 108 quiescent during LTBI and can remain in this form for a long time without harm to most individuals

109 (Comstock et al., 1974; Vynnycky and Fine, 2000). However, recent data from studies in adults

110 suggest that there might be differences in systemic inflammation by LTBI status (Cowan et al., 2012;

111 Jensen et al., 2013; Huaman et al., 2016; LaVergne et al., 2020). For example, a study of Indian

adults observed that after adjusting for potential confounders, LTBI+ individuals had significantly

113 higher levels of circulating pro-inflammatory mediators IL-6 and MCP-1 but lower levels of C-

114 reactive protein (CRP), another pro-inflammatory marker, compared to LTBI- individuals (LaVergne115 et al., 2020).

116	While studies have started to assess potential differences in systemic inflammation by LTBI
117	status in non-pregnant adults (Cowan et al., 2012; Jensen et al., 2013; Huaman et al., 2016; LaVergne
118	et al., 2020), there is no data on pregnant women. Pregnant women have a distinct immune profile
119	compared to adults and there are temporal changes in immunity during pregnancy (Mor and
120	Cardenas, 2010). It is not currently known whether there is a difference in systemic inflammation
121	between LTBI+ and LTBI- pregnant women, and how this might change by trimester of pregnancy.
122	Furthermore, LTBI+ women have a higher risk of <i>Mtb</i> progression during pregnancy and post-
123	partum but the reasons are not clear (Mathad and Gupta, 2012; Zenner et al., 2012; Jonsson et al.,
124	2020). The immune profile during pregnancy, including the systemic inflammatory milieu, may
125	inform on potential changes to immunity that increase susceptibility to TB disease during pregnancy.
126	In order to address this research gap in our understanding of systemic immunity in LTBI+ pregnant
127	women, we compared the levels of systemic inflammatory markers, at the second and third trimester,
128	by LTBI status in a cohort of pregnant women from Pune, India, and explored the association of
129	these immune markers with TB progression during pregnancy and post-partum.

130

131 Methods:

132 **Study Design and Population**

133 A cohort study of pregnant women was conducted at Byramjee Jeejeebhoy Government

134 Medical College (BJGMC) in Pune, India from 2016-2019. Adult pregnant women, aged 18-40 years

135 and between 13-34 weeks of gestation (confirmed by early pregnancy ultrasound), receiving

136 antenatal care at BJGMC were enrolled for this study. Pregnant women with active TB at entry were

This is a provisional file, not the final typeset article

137	excluded. We enrolled four cohorts of pregnant women based on their latent tuberculosis infection
138	(LTBI) and HIV status: 1) LTBI+HIV+ (N=35), 2) LTBI+HIV- (N=130), 3) LTBI-HIV+ (N=44) and
139	4) LTBI-HIV- (N=25). The sample size for this cohort was based on the primary objective of the
140	cohort study which was to compare the concentrations of Th1 cytokines after MTB-specific antigen
141	stimulation by stage of pregnancy. LTBI status was determined using Interferon Gamma Release
142	Assay (IGRA Quantiferon TB-Gold) according to manufacturer's instructions. Sampling within each
143	cohort was based on convenience sampling of those that met eligibility criteria.
144	
145	Ethics Statement
146	All clinical investigations were conducted according to the principles expressed in the
147	Declaration of Helsinki. Written informed consent was obtained from all participants. This study was
148	approved by institutional review boards and ethics committees at BJGMC, Johns Hopkins University,
149	Weill Cornell and Columbia University. We followed guidelines for human experimentation from the
150	US Department of Health and Human Services.
151	
152	Data Collection and Laboratory Procedures
153	Sociodemographic information and clinical data were collected from pregnant women at the
154	enrollment visit (13-34 weeks of gestation), at the third trimester visit (for those enrolled in the
155	second trimester), at delivery and approximately every 3 months post-partum. At each follow-up
156	visit, women were administered a World Health Organization (WHO) TB symptom screening
157	questionnaire. Women with a positive WHO symptom screen, unintentional weight loss since last
158	visit or with clinical findings on examination were further investigated with sputum GeneXpert, acid-
159	fast bacilli test, chest X-ray and abdominal ultrasound. Culture in Lowenstein Jensen (LJ) media and
	7

liquid Mycobacteria Growth Indicator Tube (MGIT) were performed for further confirmation inthose with positive findings.

162	Relevant to this analysis, blood was also collected at each visit in heparin tubes and plasma
163	samples were stored in -80°C until further use. We conducted single-plex immunoassays on second
164	and third trimester plasma samples according to the manufacturer's (R&D Systems, Minneapolis,
165	MN) directions for soluble CD163 (sCD163), soluble CD14 (sCD14), intestinal fatty acid-binding
166	protein (I-FABP), C-reactive protein (CRP), alpha 1-acid glycoprotein (AGP) and interferon- β
167	(IFN β). The lower and upper detection limitssensitivity of the assays were as follows: $\frac{1.6-1000.613}{1.6-1000.613}$
168	ng/mL for sCD163, 250-16,000125 pg/mL for sCD14, 15.6-1,0006.21 pg/mL for I-FABP, 0.8-500.02
169	ng/mL for CRP, $3.1-2000.54$ ng/mL for AGP and $50-4,00050$ pg/mL for IFN β . Multiplex
170	immunoassays (Luminex assays from R&D systems) measuring IFN γ , Interleukin (IL)-1 β , IL-6, IL-
171	13, IL-17A and TNF were also performed on these samples. The lower and upper detection
172	limitssensitivity of the assays were as follows: 43.9-10,6900.40 pg/mL for IFNy, 17.5-4,2600.80
173	pg/mL for IL-1β, <u>4.7-1,1501.7</u> pg/mL for IL-6, <u>391-95,06036.6</u> pg/mL for IL-13, <u>12.9-3,1501.8</u>
174	pg/mL for IL-17A, and 7.9-1,9301.2 pg/mL for TNF. These markers were chosen based on their
175	importance to TB, HIV and pregnancy outcomes. For Single-plex immunoassays, SpectraMax plate
176	readers were used with SofMax Pro 6 software. Luminex xMAP technology MAGPIX platform was
177	used for multiplex immunoassays with xPONENT software.

178

179 Statistical Analysis

180 We combined the LTBI+ cohorts (HIV+ and HIV-) and LTBI- cohorts (HIV+ and HIV-) to
181 study the relationship of LTBI status with second or third trimester inflammatory markers among 220

182 women with available inflammatory data. Differences in study population characteristics by LTBI

183 status were determined using Fisher's exact test for categorical variables and Wilcoxon rank-sum test

184	for continuous variables. A p-value less than 0.05 was considered statistically significant and a p-
185	value of less than $0.004 (0.05/12)$ was considered statistically significant after Bonferroni correction
186	for multiple comparisons. We also compared median levels of each inflammatory marker, during the
187	second and third trimester, between LTBI+ and LTBI- pregnant women using the Wilcoxon rank-
188	sum test. Inflammatory markers were log ₂ -transformed for the data to approximate normality.
189	We conducted univariable and multivariable linear regression to determine the change in
190	log ₂ concentrations of each inflammatory marker (outcome variable) by change in LTBI status
191	(exposure variable), with separate cross-sectional analyses for markers measured in second trimester
192	or third trimester. Multivariable models adjusted for age, mid-upper arm circumference (MUAC),
193	HIV status, vegetarian diet and gestational diabetes status. We also tested models that further
194	adjusted for smoking, education or preeclampsia. MUAC at the time of plasma sample collection (i.e.
195	second or third trimester) was used in multivariable models as it is a more reliable indicator of
196	nutritional status during pregnancy compared to body mass index. Sub-set analysis was performed
197	using Wilcoxon rank-sum test to determine whether similar relationships between LTBI status and
198	inflammatory markers were observed for only HIV-negative populations.
199	We also conducted an exploratory analysis, using univariable and multivariable logistic
200	regression analyses, to determine whether third trimester inflammation levels (exposure variable) was
201	associated with TB progression during pregnancy or post-partum (outcome variable). Progressors
202	were defined as those who prospectively developed active TB after sample collection in third
203	trimester and within study follow-up of one-year post-partum. We used STATA software version
204	15.0 for the data analysis.
205	

206 <u>Results</u>:

207

208 Study Population Characteristics

209	Our study population of pregnant Indian women (N=220) had a median age of 23 years
210	(interquartile range (IQR): 21-27) (Table 1). Only 25% had an education of less than secondary
211	education and 34% had an income below India's poverty line (monthly income <10,255 Indian
212	rupees). Around 28% of the women had a mid-upper arm circumference (MUAC) less than 23 cm
213	(an indicator of undernutrition in pregnancy(Ververs et al., 2013)) and 7% had an MUAC>30.5 cm,
214	indicative of overweight (Table 1). Most of the women (88%) did not smoke and 12% were
215	vegetarians. Ten percent had gestational diabetes and 11% had preeclampsia. As this cohort was
216	stratified by HIV status, 32% of the pregnant women were HIV+ (all on antiretroviral therapy). Study
217	population characteristics did not differ by LTBI status except for lower proportion of HIV (p-value
218	<0.001) in LTBI+ women; as mentioned above, this was due to the stratified design of the study.
219	LTBI+ women also had a lower proportion of gestational diabetes (p=0.08) and less post-high school
220	education (p=0.09) but these differences were not statistically significant (Table 1).
221	

222 Levels of Inflammatory markers by LTBI status

We compared the median log₂-transformed levels of third trimester inflammatory markers by LTBI status using Wilcoxon-rank sum tests (**Figure 1**). IL-1β (3.64 vs. 2.25 pg/mL; p=0.0002), TNF (1.76 vs. 1.54 pg/mL; p=0.004), IL-6 (4.08 vs. 1.25 pg/mL; p<0.0001) and IL-17a (2.48 vs. 2.16 pg/mL; p=0.0001) were significantly higher in LTBI- women compared to LTBI+ women (**Figure** 1). IFNγ production upon *Mtb* antigen stimulation is used to define LTBI positivity; of note, IFNγ was lower (3.63 vs. 3.73 pg/mL; p=0.15) in plasma (i.e. unstimulated samples) of LTBI- women compared to LTBI+ women, but this association was not statistically significant (**Figure 1**). Similar

230	results were also observed when using log ₂ concentrations of markers measured in plasma samples
231	from the second trimester (Supplementary Figure 1). LTBI- women had significantly higher levels
232	of second trimester AGP, I-FABP, IL-1 β , TNF, IL-6 and IL-17a compared to LTBI+ women
233	(Supplementary Figure 1). LTBI- women also had lower levels of IFN _γ compared to LTBI+
234	women, although this was not statistically significant (p=0.08) (Supplementary Figure 1).
235	
236	Association of inflammation by LTBI status
237	Next, we assessed the relationship of third trimester inflammation with LTBI status using
238	univariable and multivariable linear regression models. LTBI+ women had significantly lower levels
239	of I-FABP (mean log_2 change: -0.41, 95% confidence intervals (CI): -0.78 to -0.04; p=0.03), IL1 β
240	(mean log ₂ change: -1.03, 95% CI: -1.53 to -0.54; p<0.001), IL-6 (mean log ₂ change: -1.36, 95% CI:
241	-1.93 to -0.80; p<0.001) and IL-17a (mean log ₂ change: -0.34, 95% CI: -0.50 to -0.17; p<0.001)
242	compared to LTBI- women in univariable models (Figure 2). AGP (mean log_2 change: -0.20, 95%
243	CI: -0.42 to 0.02; p<0.08) and sCD163 (mean log ₂ change: -0.18, 95% CI: -0.39 to 0.03; p<0.10) was
244	also lower in LTBI+ women but this relationship was not statistically significant (Figure 2).
245	After adjusting for age, third trimester MUAC, HIV status, vegetarian diet, and gestational
246	diabetes in multivariable models, levels of IL-1 β (mean log ₂ change: -1.15, 95% CI: -1.70 to -0.60;
247	p<0.001), IL-6 (mean log ₂ change: -1.22, 95% CI: -1.87 to -0.58; p<0.001) and IL-17a (mean log ₂
248	change: -0.39, 95% CI: -0.57 to -0.21; p<0.001), but not I-FABP (mean log ₂ change: -0.25, 95% CI: -
249	0.67 to 0.15; p=0.22), remained significantly lower in LTBI+ women compared to LTBI- women
250	(Figure 2). In addition, AGP was also significantly lower in LTBI+ women (mean log_2 change: -
251	0.29, 95% CI: -0.54 to -0.04; p=0.02) (Figure 2). After Bonferroni correction to adjust for multiple
252	comparisons, third trimester IL1 β , IL-6 and IL-17a were significantly lower in LTBI+ women in
253	multivariable models.

254	Further adjusting for smoking, education or preeclampsia in multivariable models did not
255	change the direction or significance of the results. Finally, we also conducted sensitivity analysis to
256	show that when we limited the analysis only to HIV- subjects, the levels of these inflammatory
257	markers were still lower in LTBI+ pregnant women compared to LTBI- women (Supplementary
258	Figure 2), suggesting that HIV was not driving the observed relationships.
259	Results using second trimester inflammatory markers instead of third trimester showed
260	similar associations with LTBI status (Figure 3). In univariable models, LTBI+ pregnant women had
261	significantly lower levels of AGP, I-FABP, IL1 β , TNF, IL-6 and IL-17a compared to LTBI- pregnant
262	women (Figure 3). In multivariable models, we observed similar results observed in univariable
263	models with significantly lower levels of the AGP, I-FABP, IL-1 β , IL-6, and IL-17a, but not TNF in
264	LTBI+ compared to LTBI- women (Figure 3). In addition, sCD163 levels were significantly lower
265	and IFNy was significantly higher in LTBI+ women compared to LTBI- women (Figure 3). After
266	Bonferroni correction to adjust for multiple comparisons, second trimester AGP, IL1β, IL-6 and IL-
267	17a were significantly lower in LTBI+ women in multivariable models.
268	
269	Inflammatory markers during pregnancy and progression of TB
270	We also conducted an exploratory analysis to test whether the systemic immune profile
271	observed in LTBI+ pregnant women was associated with progression to active TB during pregnancy
272	or post-partum. In our study, there were nine women, all LTBI+ at study baseline, who progressed to
273	active TB either during the third trimester of pregnancy (n=1) or post-partum (i.e. within one year of
274	delivery) (n=8). Given that all of the progressors were LTBI+ women, we present data comparing
275	progressors and non-progressors only among LTBI+ women. Interestingly, levels of these markers in
276	LTBI+ progressors, while higher than non-progressor LTBI+ pregnant women, were similar to LTBI-
277	women (data not shown), suggesting that lower levels of these markers might be protective against

This is a provisional file, not the final typeset article

TB progression in LTBI+ pregnant women. There was a significantly increased odds of progression per log_2 increase in third trimester plasma levels of IL-1 β (adjusted odds ratio (aOR): 1.64, 95% CI: 1.05-2.57), IL-6 (aOR: 1.58, 95% CI: 1.05-2.39), and IL-13 (aOR: 2.43, 95% CI: 1.12-5.27) after adjusting for age, MUAC and HIV status (**Figure 4**). There was also an increased odds for IL-17a (aOR: 5.49, 95% CI: 0.84-35.97), but this association was not statistically significant (**Figure 4**). Similar results were observed when we limited the analysis only to post-partum progressors (data not shown).

285

286 **Discussion:**

287 In our study of LTBI+ and LTBI- pregnant women from India, LTBI+ women had lower 288 levels of various pro-inflammatory cytokines such as IL-1β, IL-6 and IL-17a compared to LTBI-289 women. In contract, the levels of IFNy were higher (significant in second trimester) in LTBI+ 290 women. While increased levels of IFNy might be related to the use of this cytokine to define IGRA-291 based LTBI (Pai et al., 2004), the results with the other cytokines were surprising. These findings 292 suggest that LTBI in pregnancy is characterized by a distinct immune profile with higher levels of 293 IFNy but lower levels of other immune markers with known roles in TB disease. Interestingly, 294 LTBI+ women who progressed to active TB during pregnancy and post-partum did not have this 295 profile in our exploratory analysis, suggesting the distinct immune profile in LTBI+ pregnant women 296 might have a protective role against TB progression. Future larger studies will need to confirm these 297 findings and determine whether these markers play a causal role and could be used to identify LTBI+ 298 pregnant women at increased risk for TB progression and a target for preventative therapy.

LTBI+ pregnant women had significantly increased levels of IFNγ in the second trimester
compared to LTBI- women. While the association was not statistically significant, the IFNγ levels
were also higher for LTBI+ women in the third trimester. In our study, we used the IGRA test, which

302	is dependent on IFNy production (Pai et al., 2004), to define LTBI status; thus it might be expected
303	IFN γ is higher in LTBI+ women. On the other hand, it should be noted that we measured IFN γ in
304	plasma samples and it is not obvious that IFNy levels in circulation should also be higher for LTBI+
305	individuals. Our results here do indicate that higher levels of IFN γ are observed in circulation for
306	LTBI+ pregnant women even without <i>Mtb</i> antigen stimulation. Similar results for IFNy have also
307	been observed from plasma samples of non-pregnant LTBI+ adults (Huaman et al., 2016; Huaman et
308	al., 2018a). While the reasons are not clear, it is possible that despite being a latent infection, there
309	could be periodic activity of some component (e.g. mRNA, protein) or low-level replication of Mtb
310	that induces IFN γ production (Huaman et al., 2016). Furthermore, LTBI is thought to be a spectrum
311	of host-pathogen interactions, with ongoing replication and metabolic activity in certain subsets
312	while quiescence in other Mtb subsets (Barry et al., 2009; Huaman et al., 2018b).
313	Our data showed lower levels of immune markers, especially IL-1 β , IL-6, IL-17a and AGP, in
314	both trimesters, in LTBI+ women compared to LTBI- women. Higher levels of IFN γ can partly
315	explain the lower levels of these other markers, as studies of Mtb have shown that IFN γ can have
316	counteractive roles with IL-1 β , IL-6 and IL-17a in certain instances (Nandi and Behar, 2011; Dutta et
317	al., 2012; Eigenbrod et al., 2013). Pregnancy-specific changes in immune profile could also in part
318	help explain these observations (Mor and Cardenas, 2010). For example, during pregnancy there is
319	an increase in neutrophil levels (Sacks et al., 1998; Luppi et al., 2002), which have been linked to
320	lower levels of IL-6 and IL-17 in <i>Mtb</i> infection (Zhang et al., 2009; O'Garra et al., 2013).
321	Interestingly, in our exploratory analyses, LTBI+ TB progressors had a profile more similar
322	to LTBI- women, with higher levels of IL-1 β , IL-6, IL-13 and IL17a and generally lower levels of
323	IFN γ compared to LTBI+ non-progressors. These inflammatory markers have been recognized for
324	their complex role in TB disease where while a deficiency is linked to reduced control of <i>Mtb</i>
325	infection, excessive levels can result in tissue damage and immunopathology (Martinez Cordero et

This is a provisional file, not the final typeset article

326 al., 2008; Tadokera et al., 2011; Martinez et al., 2013; O'Garra et al., 2013; Barber et al., 2014; Zhang 327 et al., 2014; Shen and Chen, 2018) as well as progression to active TB disease in non-pregnant adults 328 (Manabe et al., 2019). Given the small number of progressors in this study, these findings will need 329 to be confirmed in other studies with a larger sample size. If these findings are confirmed, this profile 330 could be used to identify subsets of LTBI+ pregnant women (i.e. those without this profile) at an 331 increased risk of TB progression and would further support the idea of LTBI as a spectrum where 332 subgroups of LTBI+ are protected from progression while others are not (Andrews et al., 2012; 333 Huaman et al., 2018b). In addition, future studies would also need to determine whether this 334 relationship of the systemic immune profile with TB progression is causal as it could partly explain 335 the increased risk of *Mtb* progression during pregnancy and post-partum (Mathad and Gupta, 2012; 336 Zenner et al., 2012; Jonsson et al., 2020).

337 Our study has some limitations. We did not have data on inflammation markers from 338 pregnant women during the first trimester or non-pregnant women. This data would be informative to 339 understand whether the relationship of these markers with LTBI status was also similar in early 340 pregnancy compared to later pregnancy, or in pregnant women compared to non-pregnant women. 341 Regardless, our study did have longitudinal data on inflammatory markers in the second and third 342 trimester of pregnancy, and showed consistent results with LTBI status in both trimesters that was 343 robust to adjustments for multiple comparisons. Another limitation of this study is that we only 344 assessed soluble markers of inflammation. The next steps for this study is to better understand the 345 cellular sources of these differences by assessing potential differences in immune cell phenotype and 346 function by LTBI status. The sample size for the analysis of TB progression was limited; while we 347 were able to detect significant differences in multiple markers, this was an exploratory analysis that 348 will need to be confirmed in larger studies. Future large studies should also address whether the

changes in inflammatory markers due to LTBI status impacts the risk of birth and infant healthoutcomes.

351	In summary, we characterize the systemic immune profile in LTBI+ pregnant women
352	showing higher levels of IFN γ but lower levels of other immune markers compared to LTBI-
353	pregnant women. These findings describe a circulating cytokine and immune milieu indicating a
354	distinct immune profile in LTBI+ women. Exploratory analysis suggests that this profile is negatively
355	associated with TB progression. Future studies should confirm these findings in diverse settings in
356	order to test the potential causal role along with the utility of this profile to identify women at high
357	risk for TB progression and who may benefit from preventative therapy.
358	
359	
360	
361	
362	
363	
364	
365	
366	
367	
368	
369	

370	Acknowledgments: The authors thank the study participants for their time and contributions as well
371	as the study staff who meticulously collected detailed data.

372

373	Financial Support: This work was supported primarily by the United States National Institutes of
374	Health, NIH, Bethesda, MD, USA (R00HD089753 to RS and R01HD081929 to AG). JSM received
375	support from NIAID (K23AI129854). Additional support for this work was the NIH-funded Johns
376	Hopkins Baltimore-Washington-India Clinical Trials Unit for NIAID Networks (U01AI069465 to
377	AG). BBA is a senior investigator from the Conselho Nacional de Desenvolvimento Científico e
378	Tecnológico (CNPq), Brazil. MAP received a research fellowship from the Coordenação de
379	Aperfeiçoamento de Pessoal de Nível Superior (CAPES; finance code 001). The content is solely the
380	responsibility of the authors and does not necessarily represent the official views of the NIH.
381	

381

382 Conflict of Interest: None declared

383

384 Author Contributions: SN contributed to study design, implementation and interpretation. MA 385 contributed to study design and interpretation and led the data collection. PK and SB conducted the 386 laboratory assessments and contributed to interpretation of findings. VK and PD contributed to 387 laboratory data collection and writing of this manuscript. SY and CSL contributed to data analysis. 388 MAP and BBA created the statistical scripts used to plot the analyses and graphs, and helped with the 389 interpretation of findings. RB, AG and JM led the parent study and also contributed to the design, 390 implementation and interpretation of this study. RS led the conceptual design, analysis and wrote the 391 primary version of the manuscript. All authors have approved the final manuscript and agreed to 392 publication.

393 **<u>References</u>**:

- Andrews, J.R., Noubary, F., Walensky, R.P., Cerda, R., Losina, E., and Horsburgh, C.R. (2012). Risk
 of progression to active tuberculosis following reinfection with Mycobacterium tuberculosis.
 Clin Infect Dis 54(6), 784-791. doi: 10.1093/cid/cir951.
- Barber, D.L., Andrade, B.B., McBerry, C., Sereti, I., and Sher, A. (2014). Role of IL-6 in
 Mycobacterium avium--associated immune reconstitution inflammatory syndrome. J
 Immunol 192(2), 676-682. doi: 10.4049/jimmunol.1301004.
- Barry, C.E., 3rd, Boshoff, H.I., Dartois, V., Dick, T., Ehrt, S., Flynn, J., et al. (2009). The spectrum
 of latent tuberculosis: rethinking the biology and intervention strategies. *Nat Rev Microbiol*7(12), 845-855. doi: 10.1038/nrmicro2236.
- Cliff, J.M., Kaufmann, S.H., McShane, H., van Helden, P., and O'Garra, A. (2015). The human
 immune response to tuberculosis and its treatment: a view from the blood. *Immunol Rev*264(1), 88-102. doi: 10.1111/imr.12269.
- 406 Comstock, G.W., Livesay, V.T., and Woolpert, S.F. (1974). The prognosis of a positive tuberculin
 407 reaction in childhood and adolescence. *Am J Epidemiol* 99(2), 131-138. doi:
 408 10.1093/oxfordjournals.aje.a121593.
- Cowan, J., Pandey, S., Filion, L.G., Angel, J.B., Kumar, A., and Cameron, D.W. (2012). Comparison of interferon-gamma-, interleukin (IL)-17- and IL-22-expressing CD4 T cells, IL-22-expressing granulocytes and proinflammatory cytokines during latent and active tuberculosis infection. *Clin Exp Immunol* 167(2), 317-329. doi: 10.1111/j.1365-2249.2011.04520.x.
- de Martino, M., Lodi, L., Galli, L., and Chiappini, E. (2019). Immune Response to Mycobacterium
 tuberculosis: A Narrative Review. *Front Pediatr* 7, 350. doi: 10.3389/fped.2019.00350.
- 415 Dutta, R.K., Kathania, M., Raje, M., and Majumdar, S. (2012). IL-6 inhibits IFN-gamma induced
 416 autophagy in Mycobacterium tuberculosis H37Rv infected macrophages. *Int J Biochem Cell*417 *Biol* 44(6), 942-954. doi: 10.1016/j.biocel.2012.02.021.
- Eigenbrod, T., Bode, K.A., and Dalpke, A.H. (2013). Early inhibition of IL-1beta expression by IFNgamma is mediated by impaired binding of NF-kappaB to the IL-1beta promoter but is
 independent of nitric oxide. *J Immunol* 190(12), 6533-6541. doi: 10.4049/jimmunol.1300324.
- Huaman, M.A., Deepe, G.S., Jr., and Fichtenbaum, C.J. (2016). Elevated Circulating Concentrations
 of Interferon-Gamma in Latent Tuberculosis Infection. *Pathog Immun* 1(2), 291-303. doi:
 10.20411/pai.v1i2.149.
- Huaman, M.A., Henson, D., Rondan, P.L., Ticona, E., Miranda, G., Kryscio, R.J., et al. (2018a).
 Latent tuberculosis infection is associated with increased unstimulated levels of interferongamma in Lima, Peru. *PLoS One* 13(9), e0202191. doi: 10.1371/journal.pone.0202191.
- Huaman, M.A., Ticona, E., Miranda, G., Kryscio, R.J., Mugruza, R., Aranda, E., et al. (2018b). The
 Relationship Between Latent Tuberculosis Infection and Acute Myocardial Infarction. *Clin Infect Dis* 66(6), 886-892. doi: 10.1093/cid/cix910.
- Jensen, A.V., Jensen, L., Faurholt-Jepsen, D., Aabye, M.G., Praygod, G., Kidola, J., et al. (2013).
 The prevalence of latent Mycobacterium tuberculosis infection based on an interferon-gamma release assay: a cross-sectional survey among urban adults in Mwanza, Tanzania. *PLoS One* 8(5), e64008. doi: 10.1371/journal.pone.0064008.
- Jonsson, J., Kuhlmann-Berenzon, S., Berggren, I., and Bruchfeld, J. (2020). Increased risk of active
 tuberculosis during pregnancy and postpartum: a register-based cohort study in Sweden. *Eur Respir J* 55(3). doi: 10.1183/13993003.01886-2019.
- LaVergne, S., Umlauf, A., McCutchan, A., Heaton, R., Benson, C., Kumarasamy, N., et al. (2020).
 Impact of Latent Tuberculosis Infection on Neurocognitive Functioning and Inflammation in
 HIV-Infected and Uninfected South Indians. *J Acquir Immune Defic Syndr*. doi:
 10.1097/QAI.0000000002368.

- Luppi, P., Haluszczak, C., Trucco, M., and Deloia, J.A. (2002). Normal pregnancy is associated with
 peripheral leukocyte activation. *Am J Reprod Immunol* 47(2), 72-81. doi: 10.1034/j.16000897.2002.10041.x.
- Mack, U., Migliori, G.B., Sester, M., Rieder, H.L., Ehlers, S., Goletti, D., et al. (2009). LTBI: latent
 tuberculosis infection or lasting immune responses to M. tuberculosis? A TBNET consensus
 statement. *Eur Respir J* 33(5), 956-973. doi: 10.1183/09031936.00120908.
- Manabe, Y.C., Andrade, B.B., Gupte, N., Leong, S., Kintali, M., Matoga, M., et al. (2019). A
 Parsimonious Host Inflammatory Biomarker Signature Predicts Incident TB and Mortality in
 Advanced HIV. *Clin Infect Dis.* doi: 10.1093/cid/ciz1147.
- 450 Martinez, A.N., Mehra, S., and Kaushal, D. (2013). Role of interleukin 6 in innate immunity to
 451 Mycobacterium tuberculosis infection. *J Infect Dis* 207(8), 1253-1261. doi:
 452 10.1093/infdis/jit037.
- Martinez Cordero, E., Gonzalez, M.M., Aguilar, L.D., Orozco, E.H., and Hernandez Pando, R.
 (2008). Alpha-1-acid glycoprotein, its local production and immunopathological participation in experimental pulmonary tuberculosis. *Tuberculosis (Edinb)* 88(3), 203-211. doi: 10.1016/j.tube.2007.10.004.
- Mathad, J.S., and Gupta, A. (2012). Tuberculosis in pregnant and postpartum women: epidemiology,
 management, and research gaps. *Clin Infect Dis* 55(11), 1532-1549. doi: 10.1093/cid/cis732.
- Mor, G., and Cardenas, I. (2010). The immune system in pregnancy: a unique complexity. *Am J Reprod Immunol* 63(6), 425-433. doi: 10.1111/j.1600-0897.2010.00836.x.
- 461 Nandi, B., and Behar, S.M. (2011). Regulation of neutrophils by interferon-gamma limits lung
 462 inflammation during tuberculosis infection. *J Exp Med* 208(11), 2251-2262. doi:
 463 10.1084/jem.20110919.
- 464 O'Garra, A., Redford, P.S., McNab, F.W., Bloom, C.I., Wilkinson, R.J., and Berry, M.P. (2013). The
 465 immune response in tuberculosis. *Annu Rev Immunol* 31, 475-527. doi: 10.1146/annurev466 immunol-032712-095939.
- Pai, M., Riley, L.W., and Colford, J.M., Jr. (2004). Interferon-gamma assays in the immunodiagnosis
 of tuberculosis: a systematic review. *Lancet Infect Dis* 4(12), 761-776. doi: 10.1016/S14733099(04)01206-X.
- Sacks, G.P., Studena, K., Sargent, K., and Redman, C.W. (1998). Normal pregnancy and
 preeclampsia both produce inflammatory changes in peripheral blood leukocytes akin to those
 of sepsis. *Am J Obstet Gynecol* 179(1), 80-86. doi: 10.1016/s0002-9378(98)70254-6.
- Shen, H., and Chen, Z.W. (2018). The crucial roles of Th17-related cytokines/signal pathways in M.
 tuberculosis infection. *Cell Mol Immunol* 15(3), 216-225. doi: 10.1038/cmi.2017.128.
- Tadokera, R., Meintjes, G., Skolimowska, K.H., Wilkinson, K.A., Matthews, K., Seldon, R., et al.
 (2011). Hypercytokinaemia accompanies HIV-tuberculosis immune reconstitution
- 477 inflammatory syndrome. *Eur Respir J* 37(5), 1248-1259. doi: 10.1183/09031936.00091010.
 478 Tufariello, J.M., Chan, J., and Flynn, J.L. (2003). Latent tuberculosis: mechanisms of host and
 479 bacillus that contribute to persistent infection. *Lancet Infect Dis* 3(9), 578-590. doi:
 - 10.1016/s1473-3099(03)00741-2.

- Ververs, M.T., Antierens, A., Sackl, A., Staderini, N., and Captier, V. (2013). Which anthropometric
 indicators identify a pregnant woman as acutely malnourished and predict adverse birth
 outcomes in the humanitarian context? *PLoS Curr* 5. doi:
- 484 10.1371/currents.dis.54a8b618c1bc031ea140e3f2934599c8.
- 485 Vynnycky, E., and Fine, P.E. (2000). Lifetime risks, incubation period, and serial interval of
 486 tuberculosis. *Am J Epidemiol* 152(3), 247-263. doi: 10.1093/aje/152.3.247.
- Zenner, D., Kruijshaar, M.E., Andrews, N., and Abubakar, I. (2012). Risk of tuberculosis in
 pregnancy: a national, primary care-based cohort and self-controlled case series study. *Am J Respir Crit Care Med* 185(7), 779-784. doi: 10.1164/rccm.201106-1083OC.

490 491 492 493 494 495	 Zhang, G., Zhou, B., Li, S., Yue, J., Yang, H., Wen, Y., et al. (2014). Allele-specific induction of IL- 1beta expression by C/EBPbeta and PU.1 contributes to increased tuberculosis susceptibility. <i>PLoS Pathog</i> 10(10), e1004426. doi: 10.1371/journal.ppat.1004426. Zhang, X., Majlessi, L., Deriaud, E., Leclerc, C., and Lo-Man, R. (2009). Coactivation of Syk kinase and MyD88 adaptor protein pathways by bacteria promotes regulatory properties of neutrophils. <i>Immunity</i> 31(5), 761-771. doi: 10.1016/j.immuni.2009.09.016.
496	
497	
498	
499	
500	
501	
502	
503	
504	
505	
506	
507	
508	
509	
510	
511	
512	
513	

	Overall	LTBI+	LTBI-	P- value
	(N=220)	(N=155)	(N=65)	vulue
Age median (IQR)	23 (21-27)	23 (21-27)	24 (21-27)	0.51
Monthly Income				
≤ Rs. 10,255	75 (34)	51 (33)	24 (38)	0.54
> Rs. 10,255	143 (66)	103 (67)	40 (62)	
Education				
None to primary	54 (25)	40 (26)	14 (22)	
Middle school to high school	139 (63)	101 (65)	38 (58)	0.09
Post-high school	27 (12)	14 (9)	13 (20)	
Mid-upper arm circumference				
< 23 cm	62 (28)	48 (31)	14 (21)	
23 – 30.5 cm	143 (65)	97 (63)	46 (71)	0.37
>30.5 cm	15 (7)	10 (6)	5 (8)	
Smoking status				
Yes	26 (12)	20 (13)	6 (9)	0.50
No	194 (88)	135 (87)	59 (91)	
Preeclampsia				
Yes	25 (11)	18 (12)	7 (11)	0.99
No	195 (89)	137 (88)	58 (89)	
Gestational Diabetes status				
Yes	21 (10)	11 (7)	10 (16)	0.08
No	195 (90)	141 (93)	54 (84)	
HIV				
Yes	70 (32)	31 (20)	39 (60)	< 0.001
No	150 (68)	124 (80)	26 (40)	

Table 1. Characteristics of the study population (N = 220)

515 Legend: Data are presented as number (%) of subjects unless otherwise stated. P-values were

516 calculated using Fisher's exact test for categorical variables and Wilcoxon rank-sum for

517 continuous variables to determine the difference between LTBI+ and LTBI- pregnant women.

525 Figure 1: Levels of third trimester inflammation by LTBI status (N=220)

526 Legend: A) Median and interquartile range (IQR) Log₂ levels of markers, measured in the 3rd

527 trimester is shown for LTBI+ (n=155) and LTBI- (n=65) pregnant women. Wilcoxon rank-sum test

528 was used to calculate p-values. *p < 0.05, **p < 0.01 and ***p < 0.001. B) Relative fold-change is

shown for each marker by LTBI status. Red bars indicate p-value < 0.05.

530

531 Figure 2: Association of LTBI status with third trimester inflammation (N=220)

532 **Legend**: Using linear regression, the mean change in Log₂ concentrations of each inflammation

533 marker and 95% confidence intervals (95% CI) among LTBI+ individuals compared to LTBI-

individuals is shown in the forest plot. Inflammation markers were measured in samples collected at

the third trimester of pregnancy. Multivariate models adjusted for age, mid-upper arm circumference,

536 HIV status, diet and gestational diabetes status. Only immune markers with a p-value <0.2 in the

537 univariate model are shown.

538

539 Figure 3: Association of LTBI status with second trimester inflammation (N=187)

540 Legend: Using linear regression, the mean change in Log₂ concentrations of each inflammation

541 marker and 95% confidence intervals (95% CI) among LTBI+ individuals compared to LTBI-

542 individuals is shown in the forest plot. Inflammation markers were measured in samples collected at

- 543 the second trimester of pregnancy. Multivariate models adjusted for age, mid-upper arm
- 544 circumference, HIV status, diet and gestational diabetes status. Only immune markers with a p-value

545 <0.2 in the univariate model are shown.

546

This is a provisional file, not the final typeset article

547 Figure 4: Association of third trimester inflammation markers with TB progression (N=155; 9
548 progressors)

Legend: Using logistic regression, the odds ratio and 95% confidence intervals (95% CI) of TB progression per log₂ increase in each inflammation marker among LTBI+ pregnant women is shown in the forest plot. Progressors were defined as those who developed TB either during the third trimester of pregnancy (n=1) or up to one year post-partum (n=8). Inflammation markers were measured in samples collected at the third trimester of pregnancy. Multivariable models adjusted for age, mid-upper arm circumference and HIV status. Only immune markers with a p-value <0.2 in the univariate model are shown.



Supplementary Material

1 Supplementary Figure Title and Legends

Supplementary Figure 1: Levels of second trimester inflammation by LTBI status (N=187)

Legend: A) Median and interquartile range (IQR) Log₂ levels of markers, measured in the 2nd trimester is shown for LTBI+ (n=124) and LTBI- (n=58) pregnant women. Wilcoxon rank-sum test was used to calculate p-values. *p < 0.05, **p < 0.01 and ***p < 0.001. B) Relative fold-change is shown for each marker by LTBI status. Red bars indicate p-value < 0.05.

Supplementary Figure 2: Levels of Inflammation by LTBI status in HIV- women in 3rd trimester (N=139)

Legend: A) Median and interquartile range (IQR) Log₂ levels of markers, measured in the 3rd trimester is shown for HIV- pregnant women with (n=124) and without (n=58) LTBI. Wilcoxon rank-sum test was used to calculate p-values. *p < 0.05, **p < 0.01 and ***p < 0.001. B) Relative fold-change is shown for each marker by LTBI status. Red bars indicate p-value < 0.05.



biomarker	model	Mean ch	ange (95% CI)			p-value
AGP	univariate multivariate		↓ ↓		-0.20 (-0.42 to 0.02) -0.29 (-0.54 to -0.04)	0.08 0.02
I-FABP	univariate multivariate				-0.41 (-0.78 to -0.04) -0.25 (-0.67 to 0.15)	0.03 0.22
IL-1β	univariate multivariate				-1.03 (-1.53 to -0.54) -1.15 (-1.70 to -0.60)	<0.001 <0.001
sCD163	univariate multivariate	• •			-0.18 (-0.39 to 0.03) -0.28 (-0.51 to -0.05)	0.10 0.02
TNF	univariate multivariate				-0.19 (-0.49 to 0.10) -0.24 (-0.57 to 0.10)	0.20 0.17
IL-6	univariate multivariate				-1.36 (-1.93 to -0.80) -1.22 (-1.87 to -0.58)	<0.001 <0.001
IL-17a	univariate multivariate	⊢ ∢ −	4		-0.34 (-0.50 to -0.17) -0.39 (-0.57 to -0.21)	<0.001 <0.001
	-	2 -1.5 -1 -0.5	i ı 0 0.5	1 1.5	2	



