



Citation: Müller GA, de Mello CF, Bueno AS, de Alcantara Azevedo WT, Alencar J (2022) Little noticed, but very important: The role of breeding sites formed by bamboos in maintaining the diversity of mosquitoes (Diptera: Culicidae) in the Atlantic Forest biome. PLoS ONE 17(9): e0273774. https://doi.org/10.1371/journal.pone.0273774

Editor: Igor V. Sharakhov, Virginia Polytechnic Institute and State University, UNITED STATES

Received: May 14, 2022

Accepted: August 12, 2022

Published: September 6, 2022

Copyright: © 2022 Müller et al. This is an open access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Data Availability Statement: All relevant data are within the paper and its <u>Supporting Information</u> files. (S1 Table)

Funding: This work was supported by the Research Support Foundation of the state of Rio de Janeiro (FAPERJ; grant number E-26/202.658/ 2018; E-26/010.101076/2018; SEI-260003/ 003471/2022) and the National Council for Scientific and Technological Development (CNPq; grant number 303286/2021-0). The funders had no RESEARCH ARTICLE

Little noticed, but very important: The role of breeding sites formed by bamboos in maintaining the diversity of mosquitoes (Diptera: Culicidae) in the Atlantic Forest biome

Gerson Azulim Müller¹, Cecilia Ferreira de Mello^{2,3}, Anderson S. Bueno⁴, Wellington Thadeu de Alcantara Azevedo^{2,3}, Jeronimo Alencar²*

1 Instituto Federal de Educação, Ciência e Tecnologia Farroupilha, Panambi, RS, Brazil, 2 Laboratório de Diptera, Instituto Oswaldo Cruz (Fiocruz), Rio de Janeiro, RJ, Brazil, 3 Programa de Pós-Graduação em Biologia Animal, Instituto de Biologia (UFRRJ), Universidade Federal Rural do Rio de Janeiro, Seropédica, RJ, Brazil, 4 Instituto Federal de Educação, Ciência e Tecnologia Farroupilha, Júlio de Castilhos, RS, Brazil

* jalencar@ioc.fiocruz.br

Abstract

This study investigated the composition of mosquito species in different kinds of breeding sites in a tropical forest remnant of the Atlantic Forest and identified species of public health concern therein. Collections of immature forms of mosquitoes were carried out monthly at the Poco das Antas Biological Reserve in southeastern Brazil, between June 2014 and June 2015. Samples were collected from four types of breeding sites: bamboos, bromeliads, puddles, and a lake. A total of 1,182 specimens of mosquitoes belonging to 28 species and 13 genera were collected. Three species, Ad. squamipennis, An. neglectus, and Wy. arthrostigma represented 64.8% of the captured specimens. Only three species were found in more than one type of breeding site: Ps. ferox, An. triannulatus, and Tx. trichopygus. Two species of public health concern were found breeding in bamboo (Ae. aegypti and Ae. albopictus) and one in the lake (An. darlingi). Bamboo had the highest species richness, Shannon diversity, abundance of individuals and number of dominant species of all breeding sites. Similar Simpson diversity was obtained for bamboo and bromeliads, with higher values than those obtained for puddles and the lake. The significance of the four breeding sites, especially bamboos, is discussed in the context of controlling populations of sylvatic species of mosquitoes in Atlantic Forest areas.

Introduction

In recent decades, the recurrent emergence of infectious diseases caused by zoonotic agents, especially those transmitted by mosquitoes, has caused severe public health problems, especially in subtropical and tropical countries like Brazil. Epidemic diseases such as chikungunya,

role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Competing interests: NO authors have competing interests Enter: The authors have declared that no competing interests exist.

dengue, yellow fever, malaria, and zika, which cause the illness and deaths of millions of people, are a challenge for control programs by health authorities [1-3].

Entomological surveillance practices and vector control are generally employed together to predict and control urban diseases such as dengue [4]. However, few surveillance and control strategies include natural hedges in sylvatic environments [5]. To extend these programs, it is first necessary to know which species of mosquitoes occur in these areas and understand the role of potential breeding sites in controlling this insect fauna.

The Atlantic Forest extends across the south, southeast, and northeast regions of Brazil. This biome has a forest cover of 32 million hectares, corresponding to 29% of its original area, and is one of the global biodiversity hotspots [6]. Despite its severe degree of forest loss and fragmentation, the Atlantic Forest harbors approximately 100 mosquito species [7–9]. Several of these species are associated with the transmission of pathogens that cause human diseases, like *Anopheles cruzii* [10] and *Haemagogus leucocelaenus* [11].

The number of mosquito species in a given area is associated, among other factors, with the number and types of breeding sites available for these insects [12]. Hence, immature forms of mosquitoes can be found in various aquatic habitats if they contain standing water. These breeding sites, classified as permanent and semi-permanent or transient, are formed by the accumulation of water (e.g., lakes) or natural and artificial containers [13]. The breeding sites formed from water accumulation in plants or plant parts are called phytotelmata. These breeding sites can be formed in bromeliads, bamboo internodes, tree hollows, fruit peels, plant bracts, and even when water accumulates in carnivorous plants [14].

Generally, most mosquito species of the Aedeomyiini, Aedini, and Mansoniini tribes occur in breeding sites formed by water accumulation in the soil. However, Sabethini and Toxor-hynchitini mosquitoes are mainly found in breeding sites formed by natural containers such as phytotelmata. Some species, like *Aedes terrens*, are more generalists, using breeding sites of different kinds [15, 16], while others, such as *Sabethes aurescens*, are more specific [17].

Most studies on mosquito breeding sites in the Atlantic Forest focus on bromeliads (e.g., Chaves et al. [18]) or bamboos (e.g., Müller et al. [19]). However, few have compared different types of breeding sites in the same area. Thus, the present study aimed to compare the composition of the mosquito community found in several types of breeding sites in an tropical forest remnant of the Atlantic Forest and identify areas where species of public health concern occur.

Material and methods

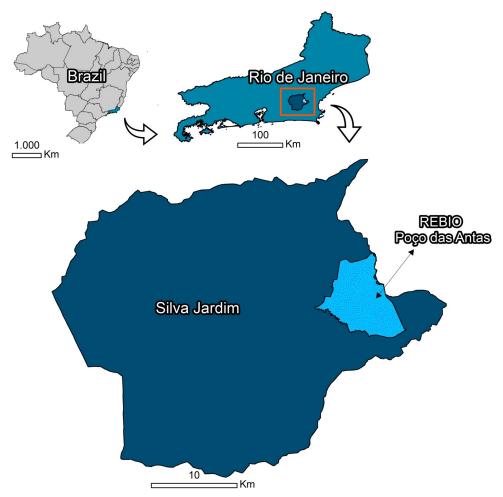
Ethics statement

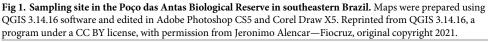
The research project was conducted at the Poço das Antas Biological Reserve with the authorization for collecting zoological material No. 44333–1, issued on 05/06/2014 by the Department of Environment and Agriculture. The permanent license, number 34911–1, for collecting, capturing, and transporting zoological material was granted by SISBIO on 06/14/2012.

Study area

The collections were carried out at the Poço das Antas Biological Reserve (PABioR), located in Silva Jardim, State of Rio de Janeiro, Brazil (22°30'–22°33' S, 42°15'–42°19' W; Fig 1). The region's climate is hot and humid with a rainy season in the summer and high average annual temperatures, without a pronounced winter. According to Takizawa [20] and Cunha [21], the average annual temperature varies from 21.4°C to 24.30°C.

PABioR is a strictly protected area (IUCN category Ia) spanning 5,065 ha, with vegetation cover primarily consisting of Lowland Ombrophilous Dense Forest, which can be divided into





https://doi.org/10.1371/journal.pone.0273774.g001

flooded and non-flooded forests. The relief of the reserve is characterized by low hills [22]. This terrain includes a mosaic of forest fragments with different levels of preservation that are often connected through corridors of native forest.

Collections and laboratory procedures

Immature mosquitoes were collected from four types of breeding sites: bamboos, bromeliads, puddles, and a lake (S1 Fig). The first three types of breeding sites were located within a forest fragment near the PABioR head office, while the lake-the reservoir of an artificial dam-was in an area adjacent to the same forest fragment.

The studied forest fragment is crossed by trails and a small road used to travel through the Biological Reserve. It has dense vegetation, with a forest canopy of up to 6 m high shading an irregular understory. Native and typical plants of the Atlantic Forest were observed in the fragment, belonging to families including Fabaceae, Bignoniaceae, and Myrtaceae. The area surrounding the lake was directly exposed to sunlight and covered by shrubs and herbaceous species.

A total of 13 field trips of two days each were carried out monthly between June 2014 and June 2015. In each field trip, 11 bromeliads, eight bamboos (five internodes per plant), 21 puddles, and a lake were convenience sampled (i.e., non-probabilistic sampling).

Water samples from bromeliads were placed in plastic trays using a manual sucker, and immature mosquitoes were collected from the trays as described by Müller and Marcondes [23]. Bamboo samples were obtained from five internodes per plant, on average, by using an electric drill as described by Marcondes and Mafra [24]. After 30 days, the drilled internodes were cut, and their water content was analyzed in plastic trays to collect immature mosquitoes. Meanwhile, water samples from puddles and the lake were obtained using dippers, as described by Forattini [13], and processed in trays as previously described. Larvae and pupae samples were transported in 250 mL plastic bags (Whirl-Pak®) and labeled with information on the location, date, and type of breeding site where the collection was made.

In the laboratory, immature mosquitoes were sorted and transferred to small individual vessels with water from the breeding site in which they were collected. These vessels were periodically topped up with distilled water. Predatory larvae, including those of the genus *Toxor*-*hynchites*, were fed with *Aedes aegypti* larvae from a colony maintained in the laboratory. The immatures were reared to continue their life cycle, and exuviae and dead larvae were preserved in 70° GL ethanol for later identification. The pupae were transferred to small sludges, in which they stayed until the adult stage.

Specimen identification was carried out by direct observation of morphological traits under an optical microscope. Immatures were fixed on slides with coverslips, while a stereomicroscope was used for adults, which were mounted on paper triangles affixed to entomological pins. The morphological traits observed were consulted in dichotomous keys and descriptions from Lane [25], Consoli and Oliveira [26], Forattini [13] and Stein et al. [27]. In addition, identifications were conducted by comparing the collected specimens with ones kept in the reference collection of the Oswaldo Cruz Institute (Fiocruz). The collected and analyzed specimens were listed in the Entomological Collection of the Instituto Oswaldo Cruz under the title "Coleção Mata Atlântica." Genera and subgenera were abbreviated following Reinert [28].

Data analysis

To assess the effect of the type of breeding site on species diversity, we employed the framework of Hill numbers [29] using individual-based species accumulation curves standardized by the number of individuals captured. Accordingly, three diversity indices were used: (i) species richness, (ii) the exponential of Shannon's entropy index (hereafter, Shannon diversity), and (iii) the inverse of Simpson's concentration index (hereafter, Simpson diversity), which can be interpreted as (i) the number of species, (ii) the number of typical species, and (iii) the number of very abundant species in the community [30]. These three indices share a common set of intuitive mathematical properties and are expressed as the 'effective number of species'– i.e., the number of equally abundant species that would be needed to give the same value of a diversity measure [29]. Four species accumulation curves were generated by extrapolation to 827 individuals based on 1,000 bootstrap replications, one curve for each type of breeding site, with the effective number of species on the *y*-axis and the number of individuals captured on the *x*-axis. If the 95% confidence intervals of the species accumulation curves did not overlap, the difference in the effective number of species was considered significant.

Mosquito species were classified into dominant or subordinate. Accordingly, species with a relative abundance higher than 1/*S*, where *S* is the number of species, are considered dominant and subordinate otherwise. Graphs and analyses were performed in the R software [31] using the *iNEXT* package [32].

Results

A total of 1,182 specimens of mosquitoes were collected, comprising 28 species, 13 genera, six tribes, and two subfamilies. The genus with the highest number of species was *Culex* (n = 6), followed by *Anopheles* (4), *Wyeomyia* and *Aedes* (3), *Limatus* and *Toxorhynchites* (2), and *Aedeomyia, Psorophora, Coquillettidia, Mansonia, Onirion, Sabethes* and *Trichoprosopon* (1). Three species, *Ad. squamipennis, Cx. neglectus* and *Wy. arthrostigma*, represented 64.8% of the total number of captured specimens (Table 1).

Mosquitoes were collected mainly in bamboos, with 827 individuals (69.9%) belonging to 15 species. In the lake, 268 individuals (22.7%) belonging to eight species were collected, while 74 specimens (6.3%) belonging to five species were collected in puddles. Only 13 individuals (1.1%) of three species were collected in bromeliads. Three species were found in more than one breeding site: *Ps. ferox* and *An. triannulatus* in the lake and puddle, and *Tx. trichopygus* in bamboos and bromeliads (Fig 2). Bamboo was the breeding ground with the highest number of dominant species (5), followed by the lake (2), puddle (1), and bromeliads (1) (Fig 3).

Individual-based species accumulation curves for the four types of breeding sites indicated that bamboos had the highest species richness, followed by the lake, puddle, and bromeliads

Subfamily	Tribe	Species	N (%)
Anopheline		Anopheles (Anopheles) maculipes (Theobald, 1923)	2 (0.2)
		Anopheles (Nyssorhynchus) albitarsis Lynch-Arribalzaga, 1878	10 (0.8)
		Anopheles (Nyssorhynchus) darlingi Root, 1926	5 (0.4)
		Anopheles (Nyssorhynchus) triannulatus s.l. (Neiva & Pinto, 1922)	41 (3.5)
	Aedeomyiini	Aedeomyia (Aedeomyia) squamipennis (Lynch Arribalzaga, 1878)	189 (15.9)
	Aedini	Aedes (Ochlerotatus) serratus (Theobald, 1901)	10 (0.8)
		Aedes (Stegomyia) aegypti (Linnaeus, 1762)	2 (0.2)
		Aedes (Stegomyia) albopictus (Skuse, 1894)	5 (0.4)
		Psorophora (Janthinosoma) ferox (von Humboldt, 1819)	66 (5.6)
	Culicini	Culex (Melanoconion) pereyrai Duret, 1967	10 (0.8)
		Culex (Melanoconion) sp.	5 (0.4)
		Culex (Microculex) neglectus Lutz, 1904	369 (31.2)
		Culex (Microculex) pleuristriatus Theobald, 1903	4 (0.4)
		Culex (Microculex) reducens Lane & Whitman, 1951	5 (0.4)
		<i>Culex (Microculex)</i> sp.	2 (0.2)
	Mansoniini	Coquillettidia (Rhynchotaenia) venezuelensis (Theobald, 1912)	2 (0.2)
		Mansonia (Mansonia) titillans (Walker, 1848)	2 (0.2)
	Sabethini	Limatus durhamii Theobald, 1901	3 (0.3)
		Limatus pseudomethysticus (Bonne-Wepster and Bonne, 1920)	2 (0.2)
		Onirion personatum (Lutz, 1904)	62 (5.2)
		Sabethes (Peytonulus) identicus Dyar and Knab, 1907	73 (6.2)
		Trichoprosopon digitatum (Rondani, 1848)	4 (0.4)
		Wyeomyia (Miamyia) oblita (Lutz, 1904)	70 (5.9)
		Wyeomyia (Phoniomyia) sp.	4 (0.4)
		Wyeomyia (Wyeomyia) arthrostigma (Lutz, 1905)	209 (17.7)
		Wyeomyia sp.	6 (0.5)
	Toxorhynchitini	Toxorhynchites (Ankylorhynchus) cf. trichopygus (Wiedemann, 1828)	10 (0.8)
		Toxorhynchites (Lynchiella) bambusicola (Lutz and Neiva, 1913)	10 (0.8)
Total			1182 (100.0)

Table 1. Mosquito taxa collected from June 2014 to June 2015 in breeding sites located at the Poço das Antas Biological Reserve in southeastern Brazil.

https://doi.org/10.1371/journal.pone.0273774.t001

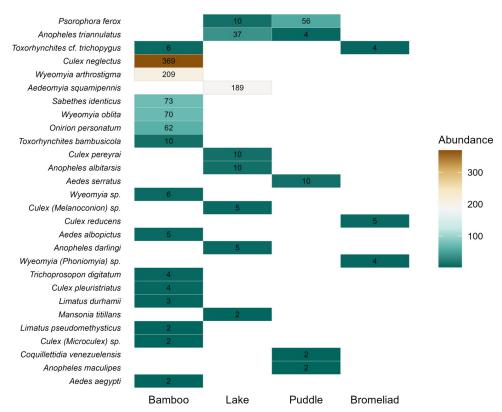


Fig 2. Species-by-site matrix for 28 mosquito species collected in four types of breeding sites in a forest fragment at the Poço das Antas Biological Reserve in southeastern Brazil. Rectangles represent species occurrence in each type of breeding site and are colored according to the number of individuals found therein. Breeding sites are ordered from left to right by the number of species, and species are ordered from top to bottom by the number of sites where the species occur, followed by the number of individuals. Note that only three species occur in more than one type of breeding habitat (*Psorophora ferox, Anopheles triannulatus,* and *Toxorhynchites* cf. trichopygus).

https://doi.org/10.1371/journal.pone.0273774.g002

(Fig 4). Despite significant differences in the species richness, the Shannon and Simpson diversity were broadly similar across the types of breeding sites. The former index indicated a higher number of typical species in the bamboos than in the three other types of breeding sites, which did not differ from each other. The latter index indicated a higher number of very abundant species in bamboos and bromeliads than in the lake and puddle (Fig 5).

Discussion

We found 28 species of mosquitoes belonging to 13 genera, most of which have sylvatic habits, including the species from the tribes Sabethini and Toxorhynchitini and the subgenera of *Culex (Microculex)*. However, taxa with habits associated with human environments, such as those from the tribes Mansoniini and Aedini, were also recorded. This apparent diversity in habit preference between the groups of collected mosquitoes may be associated with the composition of the environment at the PABioR. This strictly protected area is composed of a mosaic of second-growth forest in various stages of development and degrees of conservation, with vegetation covering about half of its area, with the rest consisting of pioneer formations and human-managed fields [33].

The PABioR was created is 1974 to protect populations of the globally endangered Golden Lion Tamarin (*Leontopithecus rosalia*) [34]. Despite this long history, studies on the mosquito fauna in the PABioR are scarce, especially in terms of the collection of immature forms.

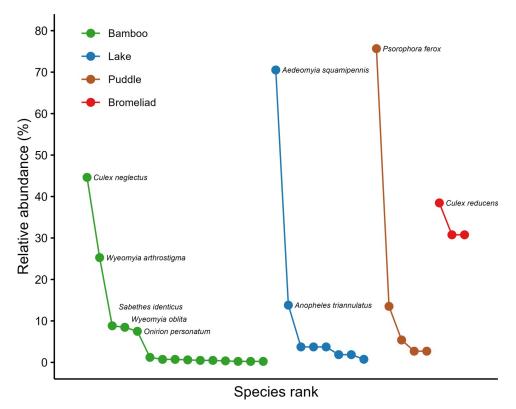


Fig 3. Relative abundance of mosquito species in four types of breeding sites in a forest fragment at the Poço das Antas Biological Reserve in southeastern Brazil. Points represent species, and colors represent the type of breeding habitat type. Dominant species–i.e., those whose relative abundance is higher than 1/S, where S is the number of species in a given breeding habitat–are indicated.

https://doi.org/10.1371/journal.pone.0273774.g003

Alencar et al. [35] used ovitraps to obtain mosquito eggs in the reserve and recorded five species: *Ae. albopictus, Ae. terrens, Hg. leucocelaenus, Hg. capricornii/janthinomys* and *Cx. iridescense*. In another study involving the collection of eggs, Mello et al. [36] recorded a sixth species, *Ps. ferox.* However, none of these studies looked for immature forms of mosquitoes in their natural breeding sites. In a third mosquito survey at the PABioR, Alencar et al. [37] also collected only adult forms, finding 41 species from 12 genera. Of these, 14 species were recorded in the present study. Thus, 13 of the mosquito species recorded herein had not been previously reported in the PABioR, bringing the total number of species to 46.

PABioR is located in an area with recorded circulation of the yellow fever virus, with documented deaths of monkeys from yellow fever in 2018 [38]. In addition, dengue cases have been reported in the region containing this Biological Reserve, with more than one virus serotype appearing to circulate simultaneously [39]. *Haemagogus* species, which are vectors of the yellow fever virus in its sylvatic cycle, were not recorded in the present study since larvae were not collected in tree hollows, the natural breeding sites of these mosquitoes [40]. However, we did find *Ae. aegypti* and *Ae. albopictus* in samples from the internodes of perforated bamboos. *Aedes aegypti* is of high epidemiological importance since it transmits several arboviruses, including dengue and yellow fever viruses [41]. Meanwhile, although *Ae. albopictus* appears to play a role in spreading the dengue virus in rural areas of Asia, its epidemiological importance seems to be secondary in Brazil [42]. Although some specimens of *Ae. albopictus* from a wild area of Rio de Janeiro were found to be naturally infected with yellow fever and zika viruses, their role as vectors has not been established [43].

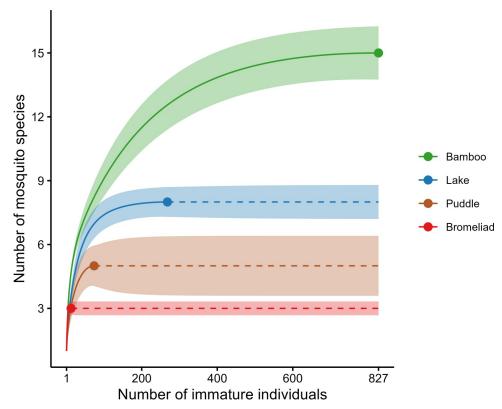


Fig 4. Individual-based species accumulation curves for mosquito species collected in four types of breeding sites in a forest fragment at the Poço das Antas Biological Reserve in southeastern Brazil. The curves are extrapolated to 827 individuals, which corresponds to the maximum number of individuals sampled among the four types of breeding sites. Solid lines, circles and dashed lines represent the interpolated, observed and extrapolated number of species, respectively. Shaded areas represent the 95% confidence intervals.

https://doi.org/10.1371/journal.pone.0273774.g004

Of these two *Aedes* species, *Ae. albopictus* [44] has been recorded as using bamboos for breeding sites since it can breed both in perforated internodes [45] and in bamboo stumps [46]. *Aedes aegypti* seems to prefer only bamboo stumps for oviposition [47], which makes its occurrence in bamboo internodes quite unusual. Indeed, its presence in the samples is probably due to a strong human influence, given that the sampled bamboo grove is located on the edge of a small dirt road crossing part of the PABioR less than 1 km from the entrance of the Biological Reserve, which is in front of a main road. Moreover, Alencar et al. [48] observed populations of *Ae. aegypti* within areas of Atlantic Forest 700 m from the edge of the forest, demonstrating that these mosquitoes can invade forest environments.

With five specimens collected in the lake, *An. darlingi* is another species that merits attention due to its epidemiological importance. It is considered the primary malaria vector in Brazil and is distributed from the Amazon to the Atlantic Forest [49]. *Anopheles darlingi* can be found in natural and artificial breeding sites on the ground, preferably in ponds shaded by riparian vegetation [50]. Furthermore, the larvae of this species are notoriously difficult to collect and thus may not be found along with adult forms [51]. Hence, the reduced number of specimens collected in the present study can be explained by the natural difficulty of capturing immature forms of *An. darlingi* and the scarce riparian vegetation in the sampled lake area, which is widely exposed to solar radiation. It is important to note that prediction models indicate that global warming may displace the transmission of malaria by *An. darlingi* from the Amazon to the Atlantic Forest due to the plastic responses of the mosquito population to rising

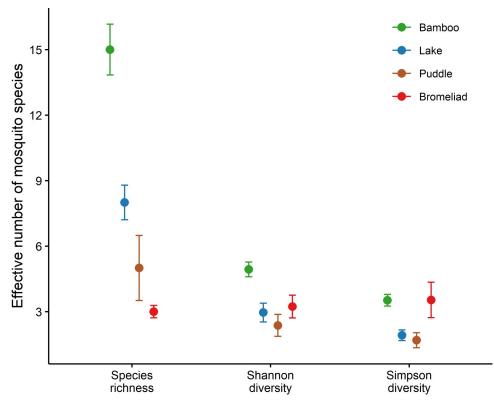


Fig 5. Diversity profile of mosquito assemblages found in four types of breeding sites in a forest fragment at the Poço das Antas Biological Reserve in southeastern Brazil. The graph shows the values of three diversity indices–(i) species richness, (ii) the exponential of Shannon's entropy index (Shannon diversity), and (iii) the inverse of Simpson's concentration index (Simpson diversity)–derived from extrapolation curves for 827 individuals (see Fig 4) for each type of breeding site. Error bars represent the 95% confidence intervals.

https://doi.org/10.1371/journal.pone.0273774.g005

temperatures [52], justifying the constant monitoring of this species in PABioR and other wild areas of Rio de Janeiro.

The present results indicate that bamboos support a considerable part of the mosquito fauna in a wild environment of the Atlantic Forest, at least relative to the other breeding sites analyzed. We made holes in the sides of bamboo internodes using an electric drill, resembling holes made by larvae of Noctuidae (Lepidoptera) [53] and adults of Curculionidae (Coleoptera) [54], which occur in nature while the plant is still young. This procedure may have helped increase the production of mosquito larvae, as several potential breeding sites were artificially created in the same place, a departure from the pattern of natural drilling by these insects. Medeiros-Sousa et al. [55], studying Atlantic Forest fragments in urban parks of São Paulo, observed a lower richness of mosquitoes (three species) in bamboos naturally perforated by sylvatic insects than in natural breeding sites of other types, including bromeliads (six species) and ponds (four species).

We recorded 15 species of mosquitoes in bamboos, five of which were dominant: *Cx. neglectus, Wy. arthrostigma, Sa. identicus, Wy. oblita*, and *On. personatum*. This number of dominant species in bamboo was higher than the observed in the other analyzed breeding sites, resulting in a higher Simpson diversity, which was highest in bamboo and bromeliads. The Shannon diversity was also higher for bamboos than for the other breeding sites due to the larger number of typical species. The species richness observed by a study conducted in an Atlantic Forest fragment in Rio de Janeiro found 19 mosquito *taxa* in internodes of artificially

perforated bamboos, including the five dominant species recorded herein, with *Cx. neglectus* as the most abundant too [56]. In another study carried out in an urban park of São Paulo, an area heavily impacted by human action, only seven *taxa* of mosquitoes inhabiting perforated bamboo internodes were observed, with *Wy. oblita* as the most abundant species [57].

Six specimens of *Toxorhynchites trichopygus* were collected from bamboos and four from bromeliads. This taxon was originally described as two species, *Ankylorhynchus neglectus* Lutz, 1904 and *Ankylorhynchus trichopygus* Dyar, 1928. The first species was described from larvae collected in bromeliads and the second from bamboo internodes. Subsequently, they were discovered to be synonymous and were later reclassified in the genus *Toxorhynchites* [58]. Some species of the genus *Toxorhynchites* follow the pattern of *Tx. trichopygus* regarding oviposition in breeding sites of different types, such as *Tx. amboinensis*, laying its eggs in artificial breeding sites, tree hollows, leaf axils, and bamboos [59].

We recorded another five species in bamboos at low abundance (≤ 10 specimens), *Tx. bamboosicola*, *Tr. digitatum*, *Cx. pleuristriatus*, *Li. durhamii*, and *Li. pseudomethysticus*. Most of these species had already been recorded in bamboos [56, 60], except for *Cx. pleuristriatus*, which had been recorded in bromeliads [61], and *Li. pseudomethysticus*, recorded in artificial breeding sites [62].

The lake was the second-richest type of breeding site of the four analyzed, and the dominant mosquito species there were *Ad. squamipennis* and *An. (Nys.) triannulatus*, and. *Aedeomyia squamipennis* has a Neotropical distribution, with abundant records in all Brazilian biomes [63]. This species is capable of being infected by several parasites participating in the cycle of the Gamboa virus [64], Equine Encephalitis virus [65], and *Plasmodium* spp., which causes avian malaria [66]. *Aedeomyia squamipennis* is directly associated with breeding sites containing aquatic plants [67], matching the sampling points on the lake. *Anopheles (Nyssorhynchus)* is capable of breeding in large permanent breeding sites formed in the ground by artificial and natural causes. In addition, the subgenus *Anopheles (Nyssorhynchus)* is the most common vector species of *Plasmodium* spp.

In contrast to the findings of Jules et al. [68] reported in a study of an area near PABioR, in which seven species of *An*. (*Nyssorhynchus*) were recorded, including *An*. *triannulatus*, we found only three species of this subgenus. As noted for *Ad*. *squamipennis*, the immature forms of *An*. *triannulatus* are commonly associated with breeding sites containing aquatic plants and grasses, as well as preferring water with ample light exposure [13, 69]. In addition to the lake, *An*. *triannulatus* was recorded in the sampled puddle, demonstrating a preference for various breeding sites formed from water accumulation in the soil. The puddle sample area was less than 0.5 m in depth with abundant herbaceous vegetation. A study carried out in the Amazon region also recorded collections of *An*. *triannulatus* in lakes and puddles [70].

The mosquito species collected in the lake had a lower Simpson diversity than in bamboos and bromeliads. The Shannon diversity for the lake did not differ statistically from the values observed in the puddle and bromeliads. Many factors can influence the abundance and composition of the mosquito community in a breeding site like a lake, including the rainfall regime and the presence of competitors, predators, algae, and aquatic plants [71, 72]. The present results show a relatively low number of mosquito species in the lake, with only eight *taxa* captured, including only one species of *Mansonia* (*Ma. titillans*), a genus often dominant in breeding sites of this kind [73]. Meanwhile, Lopes et al. [74] collected 17 mosquito taxa from a lake of an Atlantic Forest fragment under strong anthropic influence. Despite the presence of aquatic plants in the sampled lake, we did not collect any larvae from the roots of these plants, as the present collections were limited to the margins as far as the dippers reached. This methodology could explain the low number of *Mansonia* specimens collected. In addition, in the

study conducted by Lopes et al. [74], the sampling effort was extended beyond the shores of the lake, which increased the chances of collecting a greater number of species.

Other species recorded in the lake were *Cx. pereyrai*, *An. albitarsis*, and *Ps. ferox. Culex pereyrai* can also be found on riverbanks, where the current is slow [75]. *Anopheles albitarsis* is considered a generalist species in terms of breeding sites, which can even use transient breeding sites formed by the accumulation of rainwater, puddles, and permanent breeding sites such as lakes [76, 77]. Meanwhile, *Ps. ferox*, which was also found in puddles in the present study, commonly occurs in transient breeding sites on the ground [78].

The puddle had the third-highest species richness, with five species recorded. In addition, the Shannon and Simpson diversity were the lowest in the puddle. In a study by Medeiros-Sousa et al. [57] an urban park where mosquitoes were collected in different types of breeding sites, puddles were second in terms of species richness, only behind lakes. Some characteristics of this type of breeding site, including the type of vegetation (e.g., presence of grasses) and exposure to sunlight, can affect the mosquito fauna [79].

Psorophora ferox was considered the only dominant species in the sampled puddle. Verna [80] collected immatures in different types of breeding sites in the United States of America and found that *Ps. ferox* was also classified into the dominant species group among the 44 species of mosquitoes collected. This species has its population growth directly affected by the rainfall since, in these periods, transitory breeding sites are formed in the soil, which favors its proliferation [13]. The other species recorded in the puddle were *An. triannulatus*, which was also recorded in the lake, *Ae. serratus*, *Cq. venezuelensis*, and *An. maculipes*. All these species have already been recorded in breeding sites with characteristics similar to the present study [81–83].

Bromeliads had the lowest number of mosquitoes of any breeding site surveyed, with only three *taxa*. However, Simpson diversity was relatively high, and statistically the same as Shannon diversity and species richness, since the number of individuals per species was virtually the same (i.e. high evenness). Bromeliads are considered one of the main breeding sites for sylvatic mosquitoes in the Atlantic Forest, with more than 30 species breeding in the water accumulated in bromeliads' leaf axils [84–86]. Accordingly, Bastos et al. [87] observed a mosquito species richness higher in bromeliads than in bamboos and lakes. The bromeliads sampled in the PABioR were exposed to the sun, with little plant coverage, which implied a low deposition of organic material (e.g., sheets that could fall from the trees and deposit in the bromeliad). This may have influenced the present results since lower amounts of organic matter in the breeding site could have negatively influenced the composition of the insect fauna [88]. In addition, we sampled a small number of bromeliads (11 plants), which may also explain the relatively low mosquito species richness.

Of the three *taxa* recorded in bromeliads, *Cx. reducens* was the most abundants. This species belongs to the subgenus *Cx.* (*Microculex*), which frequently occurs in phytotelmata breeding sites, like *Wy.* (*Phoniomyia*) (e.g., Torreias et al. [89]). Moreover, *Cx. reducens* seems to be habitat generalist, as it is also found in bromeliads in urban and peri-urban areas [90].

Understanding the epidemiological context of the circulation of potentially pathogenic agents to humans that can be transmitted by mosquitoes is related to a greater understanding of the importance of each of the various types of breeding sites in the control of this fauna. The present results reinforce the need to monitor mosquito breeding sites in natural areas of the Atlantic Forest, especially those formed by bamboo internodes and lakes, due to the epidemiological importance of some species found, mainly *Ae. aegypti, Ae. albopictus* and *An. darlingi.* Particularly, we stress the importance of bamboos for maintaining a high number of sylvatic species of mosquitoes in the biome. Despite this, breeding sites formed by bamboos are neglected in vector control strategies, which can thereby pose risks to human populations

living in settlements close to fragments of the Atlantic Forest. Since bamboos proved to be important for the maintenance of the mosquito fauna, we suggest their inclusion in actions to reduce breeding sites for vector control in places close to wild areas of the Atlantic Forest.

Supporting information

S1 Fig. Photograph of the four types of breeding sites sampled at the Poço das Antas Biological Reserve in southeastern Brazil. A and B: lake; C: bromeliad; D: puddle; E and F: bamboo.

(TIF)

S1 Table. Data from the mosquito collections carried out at the Poço das Antas Biological Reserve in southeastern Brazil. (XLS)

Author Contributions

Conceptualization: Gerson Azulim Müller, Jeronimo Alencar.

Formal analysis: Gerson Azulim Müller, Anderson S. Bueno.

Funding acquisition: Jeronimo Alencar.

Investigation: Cecilia Ferreira de Mello, Jeronimo Alencar.

Methodology: Cecilia Ferreira de Mello, Wellington Thadeu de Alcantara Azevedo, Jeronimo Alencar.

Project administration: Jeronimo Alencar.

Supervision: Cecilia Ferreira de Mello, Jeronimo Alencar.

Writing – original draft: Gerson Azulim Müller, Jeronimo Alencar.

Writing - review & editing: Gerson Azulim Müller, Anderson S. Bueno, Jeronimo Alencar.

References

- Brady OJ, Hay SI. The global expansion of dengue: how Aedes aegypti mosquitoes enabled the first pandemic arbovirus. Annu Rev of Entomol. 2019; 65: 191–208. https://doi.org/10.1146/annurev-ento-011019-024918 PMID: 31594415
- Figueiredo PO, Stoffella-Dutra AG, Costa GB, Oliveira JS, Amaral CD, Santos JD, et al. Re-emergence of yellow fever in Brazil during 2016–2019: Challenges, lessons learned, and perspectives. Viruses. 2020; 12(11): 1233. https://doi.org/10.3390/v12111233 PMID: 33143114
- 3. Takken W, Lindsay S. Increased threat of urban malaria from *Anopheles stephensi* mosquitoes, Africa. Emerg Infect Dis. 2019; 25(7): 1431. https://doi.org/10.3201/eid2507.190301 PMID: 31063455
- 4. Valle D, Aguiar R, Pimenta DN, Ferreira V. *Aedes* de A a Z. 1st ed. Rio de Janeiro: SciELO-Editora FIOCRUZ; 2021. 172 p.
- Kitron U, Pener H, Costin C, Orshan L, Greenberg Z, Shalom U. Geographic information system in malaria surveillance: mosquito breeding and imported cases in Israel, 1992. Am J Trop Med Hyg. 1994; 50(5): 550–556. https://doi.org/10.4269/ajtmh.1994.50.550 PMID: 8203702
- Myers N, Mittermeier RA, Mittermeier CG, Fonseca GAB, Kent J. Biodiversity hotspots for conservation priorities. Nature. 2000; 403: 853–858. https://doi.org/10.1038/35002501 PMID: 10706275
- Orlandin E, Piovesan M, Souza VO, Schneeberger AH, Favretto MA, Santos EB. Temporal variation in abundance of mosquitoes (Diptera: Culicidae) in the subtropical Brazilian Atlantic Forest. Ecol Austral. 2021; 31(3): 520–531. https://doi.org/10.25260/EA.21.31.3.0.1702
- Santos EB, Favretto MA, Müller GA. When and what time? On the seasonal and daily patterns of mosquitoes (Diptera: Culicidae) in an Atlantic Forest remnant from Southern Brazil. Austral Entomol. 2020; 59(2): 337–344. https://doi.org/10.1111/aen.12454

- Santos EB, Favretto MA, Navarro-Silva MA. Community structure of mosquitoes (Diptera: Culicidae) in the coast of Southern Brazil. Austral Entomol. 2019; 58(4): 826–835. <u>https://doi.org/10.1111/aen.</u> 12412
- Medeiros-Sousa AR, Christe RO, Duarte AMRC, Mucci LF, Ceretti-Junior W, Marrelli MT. Effects of anthropogenic landscape changes on the abundance and acrodendrophily of *Anopheles (Kerteszia) cruzii*, the main vector of malaria parasites in the Atlantic Forest in Brazil. Malar J. 2019; 18(1): 1–12. https://doi.org/10.1186/s12936-019-2744-8 PMID: 30940142
- Silva SOF, Mello CF, Figueiró R, Docile T, Serdeiro M, Fumian FF, et al. Oviposition behavior of wild yellow fever vector mosquitoes (Diptera: Culicidae) in an Atlantic Forest fragment, Rio de Janeiro state, Brazil. Sci Rep. 2021; 11(1): 1–7. https://doi.org/10.1038/s41598-021-85752-y PMID: 33727688
- 12. Laporta GZ, Sallum MAM. Coexistence mechanisms at multiple scales in mosquito assemblages. BMC Ecology. 2014; 14(1): 1–10. https://doi.org/10.1186/s12898-014-0030-8 PMID: 25384802
- 13. Forattini OP. Culicidologia Médica: Identificação, Biologia, Epidemiologia. São Paulo: EDUSP; 2002. 860 p.
- Mogi M. Phytotelmata: cryptic mosquito habitat. In: Ng FSP, Yong H-S, editors. Mosquitoes and Mosquito-borne Diseases: Biology, Surveillance, Control, Personal and Public Protection Measures. Kuala Lumpur, Malaysia: ASM; 2000. 284 p.
- Mangudo C, Aparicio JP, Rossi GC, Gleiser RM. Tree hole mosquito species composition and relative abundances differ between urban and adjacent forest habitats in northwestern Argentina. Bull Entomol Res. 2018; 108(2): 203–212. https://doi.org/10.1017/S0007485317000700 PMID: 28770688
- Araujo-Oliveira A, Gil-Santana HR, Teixeira CSB, Santos-Mallet JR, Alencar J. Evaluation of the Diversity of Culicidae Vectors of the Sylvatic Yellow Fever Virus in Atlantic Forest Remnants with the Use of Ovitraps and Bamboo Traps. Vector Borne Zoonotic Dis. 2021; 21(11): 875–883. <u>https://doi.org/10.1089/vbz.2021.0033</u> PMID: 34652248
- Müller GA, Navarro-Silva MA, Marcondes CB. Developmental time of immature forms of Sabethes aurescens Lutz (Diptera, Culicidae) from artificially perforated bamboo in the rain forest of southern Brazil. Rev Bras Entomol. 2009; 53(4): 649–652.
- Chaves LSM, Sá ILR, Bergamaschi DP, Sallum MAM. Kerteszia Theobald (Diptera: Culicidae) mosquitoes and bromeliads: a landscape ecology approach regarding two species in the Atlantic rainforest. Acta Trop. 2016; 164: 303–313. https://doi.org/10.1016/j.actatropica.2016.09.023 PMID: 27686960
- Müller GA, Marchi MJ, Marcondes CB. Mosquito immatures in bamboo internodes in eastern Santa Catarina State, South Brazil (Diptera: Culicidae). Biotemas. 2014; 27(1):151–154. <u>https://doi.org/10.5007/2175-7925.2014v27n1p151</u>
- Takizawa FH. Levantamento pedológico e zoneamento ambiental da Reserva Biológica de Poço das Antas. BS monography, Universidade de São Paulo. 1995; 56 p.
- Cunha S. Impactos das obras de engenharia sobre o ambiente biofísico da Bacia do rio São João (Rio de Janeiro–Brasil). Ph. D. Dissertation, Universidade Federal do Rio de Janeiro. 1995; 380 p.
- Saporta LAC, Young CEF. Créditos de carbono e o reflorestamento do entorno da REBIO de Poços das Antas, Brasil. Rev Iberoam Econ Ecol. 2009: 17–32. Available from: <u>http://www.redibec.org/IVO/ rev12_02.pdf</u>.
- Müller GA, Marcondes CB. First report of oviposition of Aedes albopictus (Skuse, 1894) (Diptera: Culicidae) through holes in bamboos in the Americas. Entomol News. 2010; 121(1): 102–103. <u>https://doi.org/10.3157/021.121.0102</u>
- 24. Marcondes CB, Mafra H. Nova técnica para o estudo da fauna de mosquitos (Diptera: Culicidae) em internódios de bambus, com resultados preliminares. Rev Soc Bras Med Trop. 2003; 36:763–764. https://doi.org/10.1590/s0037-86822003000600022 PMID: 15049122
- 25. Lane J. Neotropical Culicidae. São Paulo: University of São Paulo; 1953. 1112 p.
- 26. Consoli RAGB Oliveira RL. Principais mosquitos de importância sanitária no Brasil. Rio de Janeiro: Editora da Fundação Oswaldo Cruz; 1994. 228 p.
- Stein M, Alvarez CN, Alonso AC, Bangher DN, Willener JA, Campos RE. New records of mosquitoes (Diptera: Culicidae) found in phytotelmata in Northern Argentina. Zootaxa. 2018; 4399(1): 87–100. https://doi.org/10.11646/zootaxa.4399.1.5 PMID: 29690331
- Reinert JF. List of abbreviations for currently valid generic-level taxa in family Culicidae (Diptera). Eur Mosq Bull. 2009: 68–76. Available from: https://e-m-b.myspecies.info/sites/e-m-b.org/files/EMB% 2827%2968-76.pdf.
- Chao A, Gotelli NJ, Hsieh TC, Sander EL, Ma KH, Colwell RK, et al. Rarefaction and extrapolation with Hill numbers: a framework for sampling and estimation in species diversity studies. Ecol Monog. 2014; 84: 45–67. https://doi.org/10.1890/13-0133.1

- Chao A, Chun-Huo C, Hsieh TC. Proposing a resolution to debates on diversity partitioning. Ecol. 2012; 93(9): 2037–2051. https://doi.org/10.1890/11-1817.1 PMID: 23094376
- 31. R Core Team. R: A language and environment for statistical computing. 2022.
- 32. Hsieh TC, Ma KH, Chao A. iNEXT: an R package for rarefaction and extrapolation of species diversity (Hill numbers). Methods Ecol Evol. 2016; 7(12): 1451–1456. https://doi.org/10.1111/2041-210X.12613
- Pereira TS, Costa MLMN, Moraes LFD, Luchiari C. Fenologia de espécies arbóreas em floresta Atlântica da Reserva Biológica de Poço das Antas, Rio de Janeiro, Brasil. Iheringia, Ser Bot. 2008; 63(2): 329–339. Available from: https://isb.emnuvens.com.br/iheringia/article/view/153.
- ICMBIO. Rebio de Poço das Antas, Rio de Janeiro. 2021. [cited 2022 May 13]. Available from: https:// www.gov.br/icmbio/pt-br/assuntos/biodiversidade/unidade-de-conservacao/unidades-de-biomas/ mata-atlantica/lista-de-ucs/rebio-de-poco-das-antas.
- Alencar J, Mello CF, Barbosa LS, Gil-Santana HR, Maia DA, Marcondes CB, et al. Diversity of yellow fever mosquito vectors in the Atlantic Forest of Rio de Janeiro, Brazil. Rev Soc Bra Med Trop. 2016; 49: 351–356. https://doi.org/10.1590/0037-8682-0438-2015 PMID: 27384833
- Mello CF, Santos-Mallet JR, Tátila-Ferreira A, Alencar J. Comparing the egg ultrastructure of three Psorophora ferox (Diptera: Culicidae) populations. Braz J Bio. 2018: 505–508. https://doi.org/10.1590/ 1519-6984.171829 PMID: 29091117
- Alencar J, de Mello CF, Rodríguez-Planes L, Silva JS, Gil-Santana HR, Bastos AQ, et al. Ecosystem diversity of mosquitoes (Diptera: Culicidae) in a remnant of Atlantic Forest, Rio de Janeiro state, Brazil. Austral Entomol. 2020; 60(1): 244–256. https://doi.org/10.1111/aen.12508
- Dietz JM, Hankerson SJ, Alexandre BR, Henry MD, Martins AF, Ferraz LP, et al. Yellow fever in Brazil threatens successful recovery of endangered golden lion tamarins. Sci Rep. 2019; 9(1): 1–13. https:// doi.org/10.1038/s41598-019-49199-6 PMID: 31506447
- Simone TS, Nogueira RMR, Araújo ESM, Guimarães FR, Santos FB, Schatzmayr HG, et al. Dengue virus surveillance: the co-circulation of DENV-1, DENV-2 and DENV-3 in the State of Rio de Janeiro, Brazil. Trans R Soc Trop Med Hyg. 2004; 98(9): 553–562. <u>https://doi.org/10.1016/j.trstmh.2003.09.003</u> PMID: 15251405
- Obholz G, Diez F, Blas GS, Rossi G. The austral-most record of the genus *Haemagogus Williston* (Diptera: Culicidae). Rev Soc Bra Med Trop. 2020; 53. <u>https://doi.org/10.1590/0037-8682-0222-2019</u> PMID: 31859948
- Souza-Neto JA, Powell JR, Bonizzoni M. Aedes aegypti vector competence studies: A review. Infec Genet Evol. 2019; 67: 191–209. https://doi.org/10.1016/j.meegid.2018.11.009 PMID: 30465912
- 42. Gratz NG. Critical review of the vector status of *Aedes albopictus*. Med Vet Entomol. 2004; 18(3): 215–227. https://doi.org/10.1111/j.0269-283X.2004.00513.x PMID: 15347388
- 43. Alencar J, Mello CF, Marcondes CB, Guimarães EA, Toma HK, Bastos AQ, et al. Natural Infection and Vertical Transmission of Zika Virus in Sylvatic Mosquitoes Aedes albopictus and Haemagogus leucocelaenus from Rio de Janeiro, Brazil. Trop Med Infect Dis. 2021; 6(2): 99. https://doi.org/10.3390/ tropicalmed6020099 PMID: 34207935
- Singh B, Baruah C, Saikia D, Gurung J. Species composition of mosquito breeding in bamboo stumps in Sikkim, India. J Vector Borne Dis. 2020; 57(1): 96–100. <u>https://doi.org/10.4103/0972-9062.308808</u> PMID: 33818462
- 45. Müller GA, Pacheco FCL, Marcondes CB. Analysis of an alternative method for the study of bromeliadassociated fauna in plants with different foliar organization. A Acad Bras Ciên. 2010; 82:903–906. https://doi.org/10.1590/S0001-37652010000400012 PMID: 21152764
- Mogi M, Armbruster PA, Tuno N. Differences in Responses to Urbanization Between Invasive Mosquitoes, *Aedes japonicus japonicus* (Diptera: Culicidae) and Aedes albopictus, in Their Native Range, Japan. J Med Entomol. 2020; 57(1): 104–112. https://doi.org/10.1093/jme/tjz145 PMID: 31586393
- Cox J, Grillet ME, Ramos OM, Amador M, Barrera R. Habitat segregation of dengue vectors along an urban environmental gradient. The American J Trop Med Hyg. 2007; 76(5): 820–826. <u>https://doi.org/</u> 10.4269/ajtmh.2007.76.820 PMID: 17488898
- Alencar J, Mello CF, Guimarães AÉ, Maia DA, Balbino VQ, Freitas MTS, et al. The first detection of a population of *Aedes aegypti* in the Atlantic Forest in the state of Rio de Janeiro, Brazil. Trop Zool. 2020; 33(2). https://doi.org/10.4081/tz.2020.70
- 49. Emerson KJ, Conn JE, Bergo ES, Randel MA, Sallum MAM. Brazilian Anopheles darlingi Root (Diptera: Culicidae) clusters by major biogeographical region. PLoS One. 2015; 10(7):e0130773. https://doi.org/ 10.1371/journal.pone.0130773 PMID: 26172559
- 50. Barros FSM, Arruda ME, Gurgel HC, Honório NA. Spatial clustering and longitudinal variation of *Anopheles darlingi* (Diptera: Culicidae) larvae in a river of the Amazon: the importance of the forest fringe and

of obstructions to flow in frontier malaria. Bull Entomol Res. 2011; 101(6): 643–658. https://doi.org/10.1017/S0007485311000265 PMID: 21729392

- Roberts DR, Paris JE, Manguin S, Harbach RE, Woodruff R, Rejmankova E, et al. Predictions of malaria vector distribution in Belize based on multispectral satellite data. Am J Trop Med Hyg. 1996; 54 (3): 304–308. https://doi.org/10.4269/ajtmh.1996.54.304 PMID: 8600771
- 52. Chu VM, Sallum MAM, Moore TE, Lainhart W, Schlichting CD, Conn JE. Regional variation in life history traits and plastic responses to temperature of the major malaria vector *Nyssorhynchus darlingi* in Brazil. Sci Rep. 2019; 9(1): 1–11. https://doi.org/10.1038/s41598-019-41651-x PMID: 30926833
- Lozovei AL. Mosquitos dendrícolas (Diptera, Culicidae) em internódios de taquara da Floresta Atlântica, Serra do Mar e do Primeiro Planalto, Paraná, Brasil. Braz Arch Biol Technol. 1998; 41: 501–510. https://doi.org/10.1590/S1516-89131998000400016
- Zequi JAC, Lopes J. Culicideofauna (Diptera) encontrada em entrenós de taquara de uma mata residual na área urbana de Londrina, Paraná, Brasil. Rev Bras Zool. 2001; 18(2): 429–438. <u>https://doi.org/10.1590/S0101-81752001000200014</u>
- 55. Medeiros-Sousa AR, Oliveira-Christe R, Camargo AA, Scinachi CA, Milani GM, Urbinatti PR, et al. Influence of water's physical and chemical parameters on mosquito (Diptera: Culicidae) assemblages in larval habitats in urban parks of São Paulo, Brazil. Acta Trop. 2020; 205: 105394. https://doi.org/10.1016/ j.actatropica.2020.105394 PMID: 32070677
- 56. Bastos AQ, Leite PJ, Mello CF, Maia DA, Machado SL, Gil-Santana HR, et al. Bionomy of Mosquitoes in Bamboo Internodes in an Atlantic Forest Remnant of the State of Rio de Janeiro, Brazil. J Am Mosq Control Assoc. 2021; 37(4): 208–215. https://doi.org/10.2987/21-7044 PMID: 34817616
- Medeiros-Sousa AR, Ceretti-Júnior W, Carvalho GC, Nardi MS, Araujo AB, Vendrami DP, et al. Diversity and abundance of mosquitoes (Diptera:Culicidae) in an urban park: Larval habitats and temporal variation. Acta Trop. 2015; 150: 200–209. https://doi.org/10.1016/j.actatropica.2015.08.002 PMID: 26259817
- Lima AM, Guitton N, Ferreira O. Comentários relativos às espécies da tribo *Toxorhynchitini* (Megarhinini) com a descrição de uma espécie nova de *Lynchiella* (Diptera, Culicidae). Mem Inst Oswaldo Cruz. 1962; 60: 225–51. https://doi.org/10.1590/S0074-02761962000200008 PMID: 13930661
- Donald CL, Siriyasatien P, Kohl A. Toxorhynchites Species: A Review of Current Knowledge. Insects. 2020; 11(11): 747. https://doi.org/10.3390/insects11110747 PMID: 33143104
- Lozovei AL. Microhabitats de mosquitos (Diptera, Culicidae) em internódios de taquara na Mata Atlântica, Paraná, Brasil. Iheringia, Série Zoologia. 2001: 3–13. <u>https://doi.org/10.1590/S0073-</u> 47212001000100001
- Oliveira-Christe R, Medeiros-Sousa AR, Fernandes A, Ceretti-Junior W, Marrelli MT. Distribution of *Culex (Microculex)* (Diptera: Culicidae) in forest cover gradients. Acta Trop. 2020; 202: 105264. <u>https://</u> doi.org/10.1016/j.actatropica.2019.105264 PMID: 31770518
- Alencar J, Serra-Freire NM, Oliveira RFN, Silva JS, Pacheco JB, Guimarães A. Immature mosquitoes of Serra do Mar Park, São Paulo State, Brazil. J Am Mosq Control Assoc. 2010; 26(3): 249–256. https://doi.org/10.2987/09-5896.1 PMID: 21033051
- Pereira AN, Moraes J, Pereira Filho A, Brito G, Rebêlo JM. First record of Aedeomyia squamipennis (Lynch Arribálzaga, 1878) (Diptera: Culicidae) in the state of Maranhão: epidemiological implications and distribution in Brazil. Check List. 2017; 13(2): 1–10. https://doi.org/10.15560/13.2.2084
- Dégallier N, Rosa A, Vasconcelos PFC, Hervé J-P, Sá Filho GC, Rosa JFS, et al. Modifications of arbovirus transmission in relation to construction of dams in Brazilian Amazonia. Ciênc Cult. 1992:124–135. Available from: https://horizon.documentation.ird.fr/exl-doc/pleins_textes/pleins_textes_6/b_fdi_33-34/ 38274.pdf.
- Mitchell CJ, Monath TP, Sabattini MS, Cropp CB, Daffner JF, Calisher CH, et al. Arbovirus investigations in Argentina, 1977–1980. II. Arthropod collections and virus isolations from Argentine mosquitoes. Am J Trop Med Hyg. 1985; 34(5): 945–955. https://doi.org/10.4269/ajtmh.1985.34.945 PMID: 2863989
- 66. Gager AB, Loaiza JR, Dearborn DC, Bermingham E. Do mosquitoes filter the access of *Plasmodium* cytochrome b lineages to an avian host? Mol Ecol. 2008; 17(10): 2552–2561. https://doi.org/10.1111/j. 1365-294X.2008.03764.x PMID: 18422926
- Burkett-Cadena ND, Blosser EM. Aedeomyia squamipennis (Diptera: Culicidae) in Florida, USA, a new state and country record. J Med Entomol. 2017; 54(3): 788–792. <u>https://doi.org/10.1093/jme/tjw226</u> PMID: 28399225
- 68. Jules JR, Albuquerque HG, Suárez-Mutis MC, Oliveira SMP, Gil-Santana HR, Rodrigues W, et al. Species diversity and abundance of *Anopheles* (*Nyssorhynchus*) (Diptera: Culicidae) in Cachoeiras de Macacu Municipality, Rio de Janeiro State: an area of the atlantic forest receptive and vulnerable to malaria. J Med Entomol. 2019; 56(3): 849–858. https://doi.org/10.1093/jme/tjy236 PMID: 30649408

- Stein M, Ludueña-Almeida F, Willener JA, Almirón WR. Classification of immature mosquito species according to characteristics of the larval habitat in the subtropical province of Chaco, Argentina. Mem Inst Oswaldo Cruz. 2011; 106(4):400–407. https://doi.org/10.1590/s0074-02762011000400004 PMID: 21739026
- 70. Nagm L, Luitgards-Moura JF, Neucamp CS, Monteiro-de-Barros FS, Honório NA, Tsouris P, et al. Affinity and diversity indices for anopheline immature forms. Rev Inst Med Trop São Paulo. 2007; 49(5): 309–316. https://doi.org/10.1590/s0036-46652007000500007 PMID: 18026638
- **71.** Overgaard HJ, Tsuda Y, Suwonkerd W, Takagi M. Characteristics of *Anopheles minimus* (Diptera: Culicidae) larval habitats in northern Thailand. Environ Entomol. 2002; 31(1): 134–141. <u>https://doi.org/10.1603/0046-225X-31.1.134</u>
- 72. Obando RG. Influencia de las algas en la densidad larval de Anopheles albimanus Wiedemann (Díptera: Culicidae) en un lago de la zona del Canal de Panamá. Boletín del Museo de Entomología de la Universidad del Valle. 2015; 6(2): 1–7. Available from: https://bibliotecadigital.univalle.edu.co/handle/10893/737.
- 73. Mello CF, Alencar J. Dispersion pattern of Mansonia in the surroundings of the Amazon Jirau Hydroelectric Power Plant. Sci Rep. 2021; 11(1): 1–7. <u>https://doi.org/10.1038/s41598-021-03682-1</u> PMID: 34930973
- Lopes J, Zequi JAC, Nunes V, Oliveira O, Oliveira Neto BP, Rodrigues W. Immature Culicidae (Diptera) collected from the Igapó lake located in the urban area of Londrina, Paraná, Brazil. Braz Arch Biol Technol. 2002; 45(4): 465–471. https://doi.org/10.1590/S1516-8913200200600010
- 75. Silva AM, Swirderski A, Ferreira AC, Massafera R, Fagundes VA, Santos DRD. Primeiro registro da ocorrência de Culex (Melanoconion) pereyrai Duret, 1967 (Diptera: Culicidae) no Paraná, sul do Brasil. Acta Biol Parana. 2021; 50(1–4): 19–25. https://doi.org/10.5380/abp.v50i1-4.83302
- Sanchez-Ribas J, Parra-Henao G, Guimarães AÉ. Impact of dams and irrigation schemes in Anopheline (Diptera: Culicidae) bionomics and malaria epidemiology. Rev Inst Med Trop São Paulo. 2012; 54 (4): 179–191. https://doi.org/10.1590/s0036-46652012000400001 PMID: 22850988
- 77. Almeida NCV, Louzada J, Neves MSAS, Carvalho TM, Castro-Alves J, Silva-do-Nascimento TF, et al. Larval habitats, species composition and distribution of malaria vectors in regions with autochthonous and imported malaria in Roraima state, Brazil. Malar J. 2022; 21(1): 1–16. https://doi.org/10.1186/ s12936-021-04033-1 PMID: 35027049
- 78. Correa FF, Gleiser RM, Leite PJ, Fagundes E, Gil-Santana HR, Mello CF, et al. Mosquito communities in Nova Iguaçu Natural Park, Rio de Janeiro, Brazil. J Am Mosq Control Assoc. 2014; 30(2): 83–90. https://doi.org/10.2987/13-6372.1 PMID: 25102590
- 79. Dida GO, Anyona DN, Abuom PO, Akoko D, Adoka SO, Matano A-S, et al. Spatial distribution and habitat characterization of mosquito species during the dry season along the Mara River and its tributaries, in Kenya and Tanzania. Infec Dis Poverty. 2018; 7(1): 1–16. <u>https://doi.org/10.1186/s40249-017-0385-</u> 0 PMID: 29343279
- Verna TN. Species composition and seasonal distribution of mosquito larvae (Diptera: Culicidae) in southern New Jersey, Burlington County. J Med Entomol. 2015; 52(5): 1165–1169. <u>https://doi.org/10.1093/jme/tjv074</u> PMID: 26336214
- Cardoso JC, Almeida MAB, Santos E, Fonseca DF, Sallum MAM, Noll CA, et al. Yellow fever virus in Haemagogus leucocelaenus and Aedes serratus mosquitoes, southern Brazil, 2008. Emerg Infect Dis. 2010; 16(12): 1918. https://doi.org/10.3201/eid1612.100608 PMID: 21122222
- Alencar J, Ferreira ZM, Lopes CM, Serra-Freire NM, Mello RP, Silva JS, et al. Biodiversity and times of activity of mosquitoes (Diptera: Culicidae) in the biome of the Atlantic Forest in the State of Rio de Janeiro, Brazil. J Med Entomol. 2011; 48(2): 223–231. https://doi.org/10.1603/me09214 PMID: 21485357
- Silva AM. Lista de Anophelinae (Diptera: Culicidae) do Estado do Paraná, sul do Brasil. Acta Biol Parana. 2021; 50(1–4): 117–149. https://doi.org/10.5380/abp.v50i1-4.83536
- Müller GA, Marcondes CB. Bromeliad-associated mosquitoes from Atlantic Forest in Santa Catarina Island, southern Brazil (Diptera, Culicidae), with new records for the State of Santa Catarina. Iheringia, Ser Zool. 2006; 96(3): 315–319. https://doi.org/10.1590/S0073-47212006000300007
- Marques TC, Bourke BP, Laporta GZ, Sallum MAM. Mosquito (Diptera: Culicidae) assemblages associated with *Nidularium* and *Vriesea* bromeliads in Serra do Mar, Atlantic Forest, Brazil. Parasit Vectors. 2012; 5(1): 1–9. https://doi.org/10.1186/1756-3305-5-41 PMID: 22340486
- Cardoso CAA, Lourenço-de-Oliveira R, Codeço CT, Motta MA. Mosquitoes in bromeliads at ground level of the Brazilian Atlantic Forest: the relationship between mosquito fauna, water volume, and plant type. Ann Entomol Soc Am. 2015; 108(4): 449–458. <u>https://doi.org/10.1093/aesa/sav040</u> PMID: 27418695

- Bastos AQ, Mello CF, Silva JS, Gil-Santana HR, Silva SOF, Alencar J. Diversity of Mosquitoes (Diptera: Culicidae) in the Bom Retiro Private Natural Heritage Reserve, Rio de Janeiro State, Brazil. J Med Entomol. 2022; 20(10): 1–8. https://doi.org/10.1093/jme/tjab222 PMID: 35026036
- Brouard O, Cereghino R, Corbara B, Leroy C, Pelozuelo L, Dejean A, et al. Understorey environments influence functional diversity in tank-bromeliad ecosystems. Freshw Biol. 2012; 57(4): 815–823. https:// doi.org/10.1111/j.1365-2427.2012.02749.x
- Torreias SRdS, Ferreira-Keppler RL, Godoy BS, Hamada N. Mosquitoes (Diptera, Culicidae) inhabiting foliar tanks of *Guzmania brasiliensis* Ule (Bromeliaceae) in central Amazonia, Brazil. Rev Bras Entomol. 2010; 54(4): 618–623. https://doi.org/10.1590/S0085-56262010000400013
- 90. Monteiro GRA, Santos RLC, Forattini OP. Aedes albopictus in bromeliads of anthropic environment in São Paulo State, Brazil. Rev Saúde Pública. 2001; 35: 243–248. <u>https://doi.org/10.1590/S0034-89102001000300005</u>