



## Review

Asymptomatic *Leishmania* infection in humans: A systematic review

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## ABSTRACT

**Background:** Leishmaniasis is a highly prevalent neglected tropical disease. It mainly presents as two forms: cutaneous and visceral leishmaniasis, the latter being the most severe form. However, asymptomatic cases of *Leishmania* infection result in an increase in the underreporting and transmission of the protozoan  
**Objectives:** In this study, articles on the incidence of asymptomatic *Leishmania* infection were systematically reviewed.

**Methods:** The publications identified in the Medline/PubMed and Science Direct databases included 4568 articles. Inclusion, exclusion, and eligibility criterion analysis resulted in 83 articles being retained. These studies were mostly performed in Brazil (n = 26) and India (n = 15).

**Results:** Several detection techniques have been used for diagnosis. Among the species found were *L. infantum* and *L. donovani*, which result in visceral leishmaniasis, and *L. amazonensis*, *L. braziliensis*, and *L. panamensis*. The incidence rates varied between the analyzed locations, largely due to sampling and the presence or absence of endemism in the regions. The largest populations analyzed were in two studies performed in India and Nepal. One of these studies evaluated 32,529 people and the incidence rate was 8.3% (n = 2702), while the other study evaluated 21,267 people and the incidence rate was 1.76% (n = 375). Only 14.28% of the studies investigated leishmaniasis in blood donors. Preexisting diseases have also been reported.

**Conclusion:** The findings of this systematic review present the incidence of cases of asymptomatic *Leishmania* infection worldwide, in addition to detailing the studies and offering information for researchers and health authorities to seek alternatives to reduce the number of leishmaniasis cases.

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## Introduction

Leishmaniasis is a parasitic disease caused by more than 20 different *Leishmania* species. It belongs to what are referred to as neglected tropical diseases (NTDs), infecting 700,000–1 million people annually, especially in underdeveloped countries [1]. The disease is transmitted through the bite of sandflies and affects several animal species, including humans, who are a natural reservoir for these protozoa [2]. In addition to vector transmission, other transmission routes include congenital transmission [3], blood transfusion [4], organ transplantation [5], and accidental laboratory exposure [6].

When infecting humans, the disease manifests itself as two main forms: visceral leishmaniasis (VL), or kala-azar, caused by the species *Leishmania (Leishmania) donovani* in the Old World and *Leishmania (Leishmania) infantum* in the New World and also in the vast majority of endemic countries of the Old World, including developed countries characterized by irregular episodes of fever, weight loss, enlarged spleen and liver, and anemia [7], which is fatal in more than 95% of untreated cases [8]; and cutaneous leishmaniasis (CL), which is the most common form, caused by several species belonging to the *Viannia* and *Leishmania* subgenera, causing ulcers and scarring of the skin, in addition to disability [9].

Some infections result in asymptomatic cases and are clinically unnoticeable. Asymptomatic *Leishmania* infection is common in endemic areas and may be more common than symptomatic cases [10]. However, asymptomatic carriers can transmit the infection [11]. Considering the capacity of humans to be a reservoir of *Leishmania*, the objective here is to carry out a systematic review of asymptomatic cases in the world to document the frequency and distribution of these cases and reinforce the importance of monitoring asymptomatic individuals in endemic regions.

## Methods

### Search strategy

A systematic review of asymptomatic *Leishmania* infection in humans was carried out in the Medline/PubMed and Science Direct (Elsevier) databases with the descriptors “Asymptomatic leishmaniasis”. The search was conducted from August to September of 2020. This review was conducted according to the Cochrane Handbook for Systematic Reviews of Interventions [12] described in Fig. 1.

### Inclusion criteria

For the selection of articles, studies had to be on asymptomatic *Leishmania* infection in humans, published in English and in peer-reviewed scientific journals. In the first phase of the search, duplicates were manually removed. The remaining titles and abstracts

were then examined to remove any articles that were unrelated to the inclusion criteria.

### Exclusion criteria

Articles published in a language other than English, bibliographic reviews, articles without access to the full text, studies of molecular markers, pharmacology, diagnostic and treatment techniques, *in vitro* and *ex vivo* studies, book chapters, comments, and letters to the editor were excluded.

### Obtaining and analyzing data

At this stage, the studies included in the systematic review were analyzed to ensure the complete collection of data to be included in Table 1. The analyzed data included the study location, type of detection technique, *Leishmania* species found, total population analyzed, number of positive samples, pre-existing diseases, and age group.

## Results

### Literature revision

The search resulted in 4568 studies: 834 from Medline/PubMed and 3734 from Science Direct. Only 157 articles remained after removing duplicates among the platforms and applying the exclusion criteria. After the eligibility analysis, 84 articles were retained (Fig. 1).

### Studies with asymptomatic Leishmania infection

The reviewed studies reported the global incidence of asymptomatic infection, with the majority of studies conducted in Brazil (n = 27), including the states of Mato Grosso do Sul, São Paulo, Maranhão, Rio de Janeiro, Piauí, Rio Grande do Norte, and Minas Gerais. Other studies were carried out in India (n = 15), Spain (n = 9), Ethiopia (n = 7), Iran (n = 5), Italy (n = 4), Turkey (n = 3), Nepal (n = 2), Israel (n = 2), Croatia, Sri Lanka, Thailand, Austria, France, Tunisia, Morocco, Sudan, the United States, and Argentina (n = 1 each) (Table 1).

### Diagnostic techniques and Leishmania species

The detection techniques used to diagnose the disease were different between the studies. Included parasitological (culture and microscopy), serological tests, enzyme-linked immunosorbent assay (ELISA), fast agglutination screening test (FAST), soluble leishmania antigen (SLA), immunochromatographic test (ICT); immunochromatographic test (rK39), immunofluorescence antibody

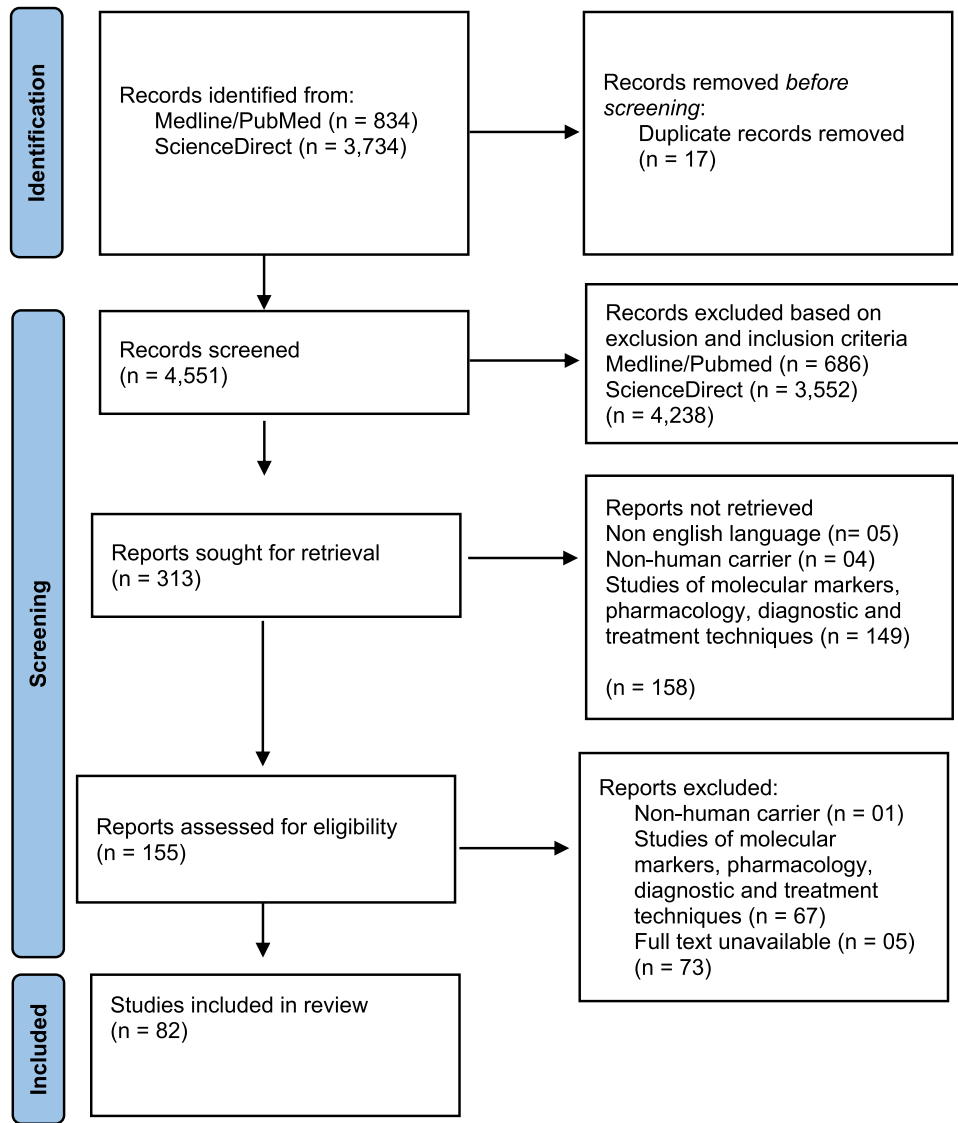


Fig. 1. Flow diagram of selection of articles used for the systematic review.

test (IFAT), western blot (WB), direct agglutination test (DAT), latex agglutination test (KAtex), montenegro skin test (MST), and fucose mannose ligand (FML); molecular test, polymerase chain reaction (PCR), real-time PCR (qPCR) and single nucleotide polymorphism (SNP); cellular test Leishmanin skin test (LST), delayed-type hypersensitivity (DTH), microcapillary culture method (MCM), interferon gama release assay (IGRA); Among the species analyzed were the *L. infantum* and *L. donovani* etiological agents of VL, and *L. amazonensis*, *L. braziliensis*, and *L. panamensis* responsible for CL (Table 1).

Number of people tested and incidence, pre-existing diseases, and age groups

The studies with the highest number of people tested were carried out in India and Nepal, one of which involved 32,529 people and reported an incidence of 8.3% (n=2702), and the other involved 21,267 people and reported an incidence of 1.76% (n=375). The study with the smallest number of tested individuals (n=20) was carried out in Spain, with 100% asymptomatic cases. The locations with the highest positivity were the Balearic Islands (Spain), Pará (Brazil), and Minas Gerais (Brazil), with positivity rates of 100%,

73.2%, and 71.3%, respectively. Only 14.28% of the studies investigated leishmaniasis in blood donors (Table 1).

The vast majority of authors did not report the presence of pre-existing diseases in the participants, but studies carried out in Minas Gerais and Mato Grosso do Sul (Brazil) analyzed liver transplant donor and recipient populations as well as patients with chronic kidney failure. Other analyses carried out in countries such as Thailand, Spain, Iran, Morocco, and Ethiopia determined pre-existing diseases such as HIV, malaria, multiple myeloma, and candidates for kidney transplantation (Table 1).

The age group of the studied population ranged from zero to 94 years of age, and some authors did not describe data regarding the age selected for the study (Table 1).

Discussion

The nature of the articles selected for the systematic review shows that there were only a limited number of studies involving asymptomatic infection in humans, compared to articles involving animals or studies not related to the pathology. Although only 19 countries were the targets of asymptomatic case studies, leishmaniasis is a major public health problem and has a wide geographic distribution, with more than 1 billion people living in endemic areas

**Table 1**  
Characteristics of the studies of asymptomatic *Leishmania* infection and their groups of detection techniques and *Leishmania* species.

Study location	Detection technique	<i>Leishmania</i> species	Total population number (n)	Number of positive (% of asymptomatic)	Preexisting disease	Age groups	Source
Argentina (Salta)	MST	<i>L. panamensis</i> , <i>L. amazonensis</i>	7336	134 (1.8%)	N/I	30–59 years	Sosa-Estani 2000[13]
Austria	ELISA	<i>Leishmania</i> sp.	1048	47 (4.5%)	N/I	18 and 60 years	Poepll 2013[14]
Brazil (Bahia)	ELISA	<i>Leishmania</i> sp.	86	20 (23.2%)	N/I	Average 8.1 years	Badaro 1986[15]
Brazil (Bahia)	ELISA, MST	<i>L. infantum</i>	135	44 (32.59%)	N/I	N/I	D'Oliveira 1997[16]
Brazil (Bahia)	ELISA	<i>Leishmania</i> sp.	700	38 (5.4%)	N/I	Average 42 years	Fukutani 2014[17]
Brazil (Ceará, Piauí and Minas Gerais)	ELISA, WB, PCR	<i>L. infantum</i>	608	37 (6%)	N/I	18–30 years	Ferreira-Silva 2018[18]
Brazil (Federal District)	MST, rK39	<i>L. amazonensis</i> , <i>L. infantum</i>	700	233 (33.28%)	N/I	2–14 years	Carranza-Tamayo 2016[19]
Brazil (Fortaleza)	ELISA	<i>L. infantum</i>	431	57 (13.2%)	N/I	N/I	Monteiro 2016[20]
Brazil (Maranhão)	DTH, ELISA	<i>Leishmania</i> sp.	638 and 572	56%/41% and 58%/98%	N/I	0–5 years	Caldas 2002[21]
Brazil (Maranhão)	ELISA	<i>Leishmania</i> sp.	905	144 (18.3%)	N/I	0–5 years	Gama 2004[22]
Brazil (Maranhão)	ELISA	<i>L. braziliensis</i> , <i>L. infantum</i>	1100	91 (8.2%)	N/I	N/I	Mendes 2007[23]
Brazil (Maranhão)	MST, ELISA	<i>L. infantum</i>	361	35 (9.7%)	N/I	< 16 years	Moura 2012[24]
Brazil (Mato Grosso do Sul)	IFAT, ELISA	<i>Leishmania</i> sp.	220	80 (36.3%)	N/I	4 ≤ > 60 years	De Oliveira 2008[25]
Brazil (Mato Grosso do Sul)	IFAT	<i>L. major</i>	430	67 (15.6%)	N/I	18–68 years	França 2013[26]
Brazil (Mato Grosso do Sul)	IFAT, rK39	<i>Leishmania</i> sp.	50	16 (32%)	N/I	20–77 years	França 2020[27]
Brazil (Minas Gerais)	IFAT, ELISA, rK39, PCR	<i>L. infantum</i>	226	102 (45.1%)	N/I	chronic renal failure	Moreno 2006[28]
Brazil (Minas Gerais)	PCR, ELISA, rK39	<i>L. donovani</i> , <i>L. infantum</i> , <i>L. amazonensis</i>	136	97 (71.3%)	N/I	13–38 years	Moreno 2009[29]
Brazil (Minas Gerais)	IFAT, rK39, rK26, MST	<i>L. amazonensis</i>	246	156 (63.4%)	N/I	14–51 years	Silva 2011[30]
Brazil (Minas Gerais)	PCR	<i>L. amazonensis</i>	67	5 (7.5%)	Liver transplant*	N/I	Clemente 2014[31]
Brazil (Minas Gerais)	SLA, rK39	<i>Leishmania</i> sp.	935	304 (32.5%)	N/I	5–70 years	Marques 2017[32]
Brazil (Minas Gerais)	rK39	<i>L. infantum</i>	1875	6 (0.32%)	N/I	3 months - 10 years	Da Rocha 2018[33]
Brazil (Minas Gerais)	rK39, ELISA, qPCR	<i>L. infantum</i>	179	81 (45.2%)	N/I	2 months - 7 years	Da Cunha 2020[34]
Brazil (Pará)	DTH, IFAT	<i>Leishmania</i> sp.	231	169 (73.2%)	N/I	1–10 years	Silveira 2010[35]
Brazil (Piauí)	PCR	<i>L. infantum</i>	108	8 (7.4%)	N/I	1–89 years	Costa 2002[36]
Brazil (Rio de Janeiro)	MST	<i>L. braziliensis</i>	28	11 (39.28%)	N/I	4–28 years	Bittar 2007[37]
Brazil (Rio Grande do Norte)	FML ELISA, PCR	<i>L. donovani</i>	21	14 (66.66%)	N/I	Average 37 years	Otero 2000[38]
Brazil (Rio Grande do Norte)	ELISA	<i>L. infantum</i>	345	85 (24.6%)	N/I	N/I	De Lima 2012[39]
Brazil (Rio Grande do Sul)	MST	<i>L. amazonensis</i>	151	60 (39.7%)	N/I	N/I	Fagundes 2007[40]
Brazil (São Paulo)	rK39	<i>L. infantum</i>	250	29 (11.6%)	N/I	> 31.5 years	Barão 2007[41]
Croatia	ELISA	<i>L. infantum</i>	2035	231 (11.4%)	N/I	6 months - 88 years	Šiško-Kraljević 2013[42]
Ethiopia	FAST	<i>L. donovani</i>	1390	45 (3.2%)	N/I	0 - > 60 years	Hailu 2002[43]
Ethiopia	DAT, rK39, PCR, KAtex	<i>L. donovani</i>	534	36 (6.7%)	HIV	18 - ≥ 48 years	Griensven 2019[44]
Ethiopia	LST, DAT	<i>L. donovani</i>	650 and 1040	59 (9.08%) and 9 (0.87%)	N/I	> 2 years	Tadesse 2019[45]
Ethiopia (Amhara)	rK39, DAT, DTH	<i>L. donovani</i>	605	61 (10%)	Malaria and HIV	5 < - 15 years	Gadisa 2012[46]
Ethiopia (Amhara)	rK39, DAT	<i>L. major</i>	565	56 (9.9%)	N/I	4–15 years	Custodio 2012[47]
Ethiopia (Aramachio)	rK39	<i>L. donovani</i>	185	14 (7.5%)	N/I	20–30 years	Ayehu 2018[48]
Ethiopia (Omo)	MST, DAT, rK39	<i>Leishmania</i> sp.	1682	30 (1.8%)	N/I	> 18 months	Bekele 2018[49]
France (Monaco)	WB	<i>L. infantum</i>	565	76 (13.4%)	N/I	N/I	Le Fichoux 1999[50]
India (Bihar)	Microscopy	<i>L. donovani</i>	450	6 (1.3%)	N/I	N/I	Sosa 2000
India (Bihar)	rK39, ELISA	<i>L. infantum</i>	150	50 (33.3%)	N/I	N/I	Singh 2002[51]
India (Bihar)	rK39, PCR	<i>L. donovani</i>	997	21 (2.1%)	N/I	17–25 years	Das 2011[52]
India (Bihar)	rK39, DAT	<i>Leishmania</i> sp.	870	287 (32.98%)	N/I	5 - > 15 years	Gidwani 2011[53]
India (Bihar)	rK39, DAT, PCR	<i>L. donovani</i>	355	50 (14%)	N/I	0 - ≥ 60 years	Topno 2010[54]
India (Bihar)	qPCR	<i>Leishmania</i> sp.	210	40 (19.04%)	N/I	6–55 years	Sudarsan 2014[55]
India (Bihar)	rK39	<i>L. donovani</i>	5144	116 (2.2%)	N/I	1 - ≥ 46 years	Das 2016[56]
India (Bihar)	DAT, rK39, qPCR	<i>L. donovani</i>	5794	120 (2.07%)	N/I	≥ 6 years	Das 2020[57]
India (Maida)	rK39	<i>Leishmania</i> sp.	2890	79 (2.7%)	N/I	N/I	Saha 2017[58]

(continued on next page)

Table 1 (continued)

Study location	Detection technique	Leishmania species	Total population number (n)	Number of positive (% of asymptomatic)	Preexisting disease	Age groups	Source
India (Muzaffarpur)	qPCR, DAT, rK39	<i>L. donovani</i>	1469	511 (34.7%)	N/I	≥ 18 years	Sudarsan 2014[59]
India and Nepal	DAT	<i>L. donovani</i>	21,267	375 (1.76%)	N/I	N/I	Ostyn 2011[60]
India and Nepal	DAT	<i>L. donovani</i>	7538	510 (6.7%)	N/I	> 2 years	Picado 2014[61]
India (West Bengal)	qPCR, rK39, ELISA	<i>L. donovani</i>	246	97 (10.9%)	N/I	≤ 18 - ≥ 45 years	Kaushal 2017[62]
Iran	PCR-ELISA	<i>Leishmania</i> sp.	388	25 (24.5%)	N/I	1–35 years	Alborzi 2008[63]
Iran (Ardabil)	DAT, PCR	<i>L. infantum</i>	600	23 (3.8%)	N/I	20–61 years	Asfaram 2017[64]
Iran (Fars)	DAT, PCR	<i>L. infantum</i>	426	68 (15.96%)	N/I	5–10 years	Fakhar 2008[65]
Iran (Fars)	ELISA, PCR	<i>L. infantum</i>	617	17 (2.7%)	N/I	14–83 years	Gigloo 2018[66]
Iran (Fars)	ELISA, PCR	<i>L. infantum</i>	251	19 (7.5%)	HIV	18–45 years	Rezaei 2018[67]
Israel	ELISA	<i>L. donovani</i>	2580	67 (2.59%)	N/I	18–45 years	Adini 2003[68]
Israel (Kafr Yarka)	ELISA	<i>L. donovani</i>	648	67 (10%)	N/I	N/I	Adini 1994
Italy (Bologna)	WB, PCR	<i>L. infantum</i>	119	19 (15.9%)	renal transplant candidates	20–94 years	Comai 2020[69]
Italy (Ferrara)	qPCR	<i>Leishmania</i> sp.	150	22 (14.6%)	CIRDS	31–72 years	Maritati 2018[70]
Italy (Sicily)	ELISA	<i>L. infantum</i>	500	0 (0%)	N/I	N/I	Colomba 2005[71]
Italy (Valsamoggia)	WB, qPCR	<i>L. infantum</i>	240	30 (12.5%)	N/I	22–70 years	Ortali 2020[72]
Morocco	PCR, IFAT	<i>L. infantum</i>	200	10 (5%)	HIV	22–68 years	Echchakery 2018[73]
Nepal	PCR	<i>L. donovani</i>	418	40 (9.6%)	N/I	2 - > 46 years	Ostyn 2015[74]
Nepal (Dharan)	DAT	<i>L. donovani</i>	507	5 (0.9%)	N/I	18–45 years	Timilsina 2016[75]
Spain	IFAT	<i>L. donovani</i>	179	6 (3.3%)	HIV	29–33 years	Ena 2014[76]
Spain	IFAT	<i>L. infantum</i>	625	30 (4.8%)	N/I	11–81 years	Elmahallawy 2015[77]
Spain (Balearic Islands)	rK39, PCR, qPCR	<i>L. infantum</i>	50	29 (58%)	HIV, Myeloma	> 18 years	Molina 2020[78]
Spain (Balearic Islands)	ELISA, PCR, WB	<i>L. infantum</i>	122	36 (29.5%)	N/I	N/I	Riera 2004[79]
Spain (Balearic Islands)	WB	<i>L. infantum</i>	1437	44 (3.1%)	N/I	N/I	Riera 2008[80]
Spain (Balearic Islands)	WB, qPCR	<i>L. infantum</i>	20	20 (100%)	N/I	40–60 years	Jimenez-Marco 2018[81]
Spain (Madrid)	PCR	<i>L. infantum</i>	330	47 (14.24%)	N/I	N/I	Ibarra-Meneses 2016[82]
Spain (Murcia)	ELISA, PCR	<i>L. infantum</i>	657 and 618	13 (2%) and 49 (8%)	N/I	18–65 years	Pé rez-Curtillas 2015[83]
Spain (Province of Grenada)	IFAT, qPCR, PCR-ELISA	<i>L. infantum</i>	1260	129 (10.2%)	N/I	18–65 years	Aliaga 2019[84]
Sri Lanka (Anuradhapura)	rK39, MST	<i>L. donovani</i>	955	31 (3.2%)	N/I	1 - > 65 years	Ranasinghe 2013[85]
Sudan (Gedaref)	DAT, PCR	<i>L. donovani</i>	95	31 (32%)	N/I	≤ 40 and ≥ 40 years	Mohamed 2019[86]
Thailand (Trang)	DAT	<i>Leishmania</i> sp.	724	180 (24.9%)	HIV/AIDS	> 18 years	Manomat 2017[87]
Tunisia	WB	<i>Leishmania</i> sp.	94	38 (40.4%)	N/I	8 months - 75 years	Saghroumi 2012[88]
Turkey	WB, IFAT, ELISA	<i>L. infantum</i>	82	5 (6%)	N/I	1–72 years	Sakru 2007[89]
Turkey (Istanbul)	IFAT	<i>L. infantum</i>	188	12 (6.4%)	N/I	19–64 years	Ates 2012[90]
Turkey (Istanbul)	PCR, MCM, ELISA, IFAT, ICT	<i>L. infantum</i>	343	21 (6.1%)	N/I	18–65 years	Ates 2013[91]
United States (Washington)	ELISA, PCR, IGRA, rK39	<i>L. infantum</i>	200	39 (19.5%)	N/I	24–60 years	Mody 2019[92]

N/I, Uninformed; DAT, Direct Agglutination test; DTH, Delayed-Type Hypersensitivity; ELISA, Enzyme Immunoabsorbent Assay; ELISA and chromatographic test of antibodies against rK39; FAST, Fast Agglutination Screening Test; FML, Fucose Mannose Ligand; IFAT, Antobody Immunofluorescence Test; IGRA, Interferon Gamma Release Assay; ICT, immunochromatographic test; MCM, microcapillary culture method; MST, Montenegro Skin Test; PCR, Polymerase Chain Reaction; qPCR, real time PCR; rK39, Immunochromatographic test rK39; SLA, ELISA based on Soluble *Leishmania* Antigens; SNP, Single nucleotide polymorphism; WB, Western blot; \*Liver transplant donor.

and at risk of contracting the parasite, which is the second most prevalent pathogen of parasitic diseases [1]. The selected studies demonstrated that the occurrence of asymptomatic infection by *Leishmania* is common in endemic regions, especially in developing countries such as Brazil and India.

Asymptomatic infection refers to an individual who has had a confirmed diagnosis of Leishmaniasis, without showing signs or symptoms of the disease, common in endemic regions. Sandflies are the main vectors of *Leishmania* and their evolution with mammals are well documented in the literature. Its transmission can follow an anthroponotic or zoonotic cycle. Domestic dogs are the main reservoirs of *L. infantum*, although other mammals are reservoirs of the parasite. A cohort study in Brazil demonstrated, through PCR tests, infection for asymptomatic leishmaniasis in up to 80% of the analyzed dogs. Despite the amount of clinical research, the nature of asymptomatic infection remains poorly understood [10,93–95].

A variety of diagnostic techniques were used across the different study areas. Most studies used combined immunological methods, while others plot molecular and parasitological tests. Immunological assays, although appropriate for regions with a high infection rate, cannot distinguish whether it is an active infection. Molecular methods, on the other hand, have high sensitivity and specificity, but do not indicate whether the infection is active. Parasitological techniques are time consuming, require trained personnel to visualize the parasite and, in asymptomatic cases, parasite identification is limited. It is difficult to diagnose asymptomatic cases due to the low parasite load and low levels of antibodies. The combination of methods raises the threshold for detecting asymptomatic infections in endemic regions [96].

In Brazil, the leishmaniasis control program involves rapid diagnosis and treatment of human cases [97]. Many asymptomatic infection detection techniques are used for the symptomatic form and have already allowed cross-reactions to be avoided. Owing to the low parasite load, combining tests improves performance and provides greater diagnostic certainty. Once asymptomatic infection in humans is identified, its monitoring is essential for the adoption of adequate therapy upon the onset of the first symptoms.

Four *Leishmania* species that cause CL (*L. braziliensis*, *L. amazonensis*, *L. major*, and *L. panamensis*) and two species that cause CL (*L. infantum* and *L. donovani*) were identified. *L. braziliensis* has a high prevalence in South America and is associated with mucosal forms. In contrast, *L. amazonensis* is responsible for disseminated forms [98].

The main risk factors for leishmaniasis are climate and environmental changes, socioeconomic conditions, malnutrition, and population mobility [99]. These are common factors in developing countries such as Brazil and India, which are the main countries with a high incidence of CL. In the case of asymptomatic infection by *Leishmania*, because it is “silent,” its incidence is underestimated, although epidemiological surveillance of asymptomatic infection allows for subsidization of prevention measures. The determinants of asymptomatic conditions have not yet been elucidated, but factors such as a high albumin concentration, high levels of vitamin A, high birth weight, and high consumption of red meat can favor asymptomatic infection [100].

In the research on asymptomatic infection by *Leishmania*, the presence of comorbidities such as HIV, malaria, multiple myeloma, and chronic renal failure was also described. In the case of infectious diseases, further studies are needed to verify whether the treatment of these diseases could inhibit the activity of *Leishmania*.

Some patients with immunosuppressive diseases such as HIV/AIDS undergoing antiretroviral treatment have asymptomatic *Leishmania* infection even with an increase in the parasite load, which enhances the infectivity of the vector and consequently transmission of the parasite [52,78]. The condition of individuals with asymptomatic *Leishmania* infection in HIV patients needs

further studies to elucidate the immune status that allows this condition.

Asymptomatic *Leishmania* infection has been reported in organ donors [27,31,69] and in blood donors [17,101], thus reinforcing the need for screening in donors in countries that have not yet adopted such screening, especially if they come from endemic areas. To avoid discarding a large number of blood bags from suitable donors with only asymptomatic *Leishmania* infection, leuko-depletion can be adopted, which, in addition to preventing transfusion transmission, improves the quality of blood products [64].

Most of the studies evaluated covered a wide age range, showing that asymptomatic cases can occur from children to the elderly, thus proving that age is not a determining factor for asymptomatic infection. The investigation of asymptomatic *Leishmania* infection is considered a priority to eliminate and reduce the transmissibility of the disease, since transmission remains in these hosts. More studies should be carried out in an attempt to investigate the percentage of individuals who can progress to clinical disease.

The determining factors (such as genetic background, nutritional aspects, and co-infections) for the asymptomatic condition caused by *Leishmania* have not yet been elucidated. Asymptomatic cases are a source of infection and contribute to the endemicity of the disease. Therefore, the detection of asymptomatic infection is important so that treatment can be started in a timely manner if the infection evolves into the symptomatic form, as this increases the chance of a therapeutic response. Studies performed in several endemic countries have highlighted the need to screen blood banks and organ donors, as this allows horizontal transmission through blood and organ donation to be avoided.

## Conclusions

Our review demonstrates the worldwide incidence of asymptomatic *Leishmania* infection cases, describing the relevance of the search for alternatives in order to increase the detection rate and reduce the number of cases.

## Ethics approval

Not applicable.

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## Consent for publication

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## Author contributions

Conceptualization, supervision and critical review: MSCLJ and HCNA; Literature search, data analysis, writing – original draft

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## Data Availability

The authors declare that data supporting the findings of this study are available in the article in the References topic.

## Conflict of interest

The authors declare they have no competing interests.

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## References

- [1] WHO. World Health Organization. Leishmaniasis, <https://www.who.int/news-room/fact-sheets/detail/leishmaniasis> [accessed 11 September 2020].
- [2] Okwor I, Uzonna J. Social and economic burden of human Leishmaniasis. 489–483 *Am J Trop Med Hyg* 2016;94(3). <https://doi.org/10.4269/ajtmh.15-0408>
- [3] Argy N, Lariven S, Rideau A, et al. Congenital Leishmaniasis in a newborn infant whose mother was infected with Leishmaniasis and HIV. *J Pediatr Infect Dis Soc* 2020;9(2):277–80. <https://doi.org/10.1093/jpids/piz055>
- [4] Jimenez-Marco T, Fisa R, Girona-Llobera E, et al. Transfusion-transmitted Leishmaniasis: a practical review. *Transfusion* 2016;56:45–51. <https://doi.org/10.1111/trf.13344>
- [5] Gajurel K, Dhakal R, Deresinski S. Leishmaniasis in solid organ and hematopoietic stem cell transplant recipients. *Clin Transpl* 2017;31(1):1–12. <https://doi.org/10.1111/ctr.12867>
- [6] Xiong YH, Guan YY, Cao JP. Risk assessment on laboratory biosafety of Leishmania. *Chinese J Schistosomiasis Control* 2012;24(3):342–4.
- [7] Gomila H, A, Vanzo C, Garnera A, et al. Visceral Leishmaniasis. Pediatric case report. *Arch Argent Pediatr*. 2017; 115:251–254. <https://doi.org/10.5546/aap.2017.e251>.
- [8] Wilhelm T.J. Visceral Leishmaniasis *Chirurg*. 2019;90(10):833–837. <https://doi.org/10.1007/s00104-019-0994-1>.
- [9] Mokni, M. Cutaneous Leishmaniasis. *Ann Dermatol Venereol*. 2019;143(3):232–246. <https://doi.org/10.1016/j.annder.2019.02.002>.
- [10] Burza S, Croft SL, Boelaert M. Leishmaniasis. *Lancet* 2018;392(10151):951–70. [https://doi.org/10.1016/S0140-6736\(18\)31204-2](https://doi.org/10.1016/S0140-6736(18)31204-2)
- [11] Andrade-Narvaez FJ, Loria-Cervera EN, Sosa-Bibiano EI, et al. Asymptomatic infection with American cutaneous Leishmaniasis: epidemiological and immunological studies. *Mem Inst Oswaldo Cruz* 2016;111(10):599–604. <https://doi.org/10.1590/0074-02760160138>
- [12] Higgins, J.P.T.; Thomas, J.; Chandler, J.; Cumpston, M.; Li, T.; Page, M.J.; Welch, V.A. Eds. *Cochrane Handbook for Systematic Reviews of Interventions* version 6.0 (updated July 2019). Cochrane, 2019. Available online: <http://www.training.cochrane.org/handbook> [accessed 10 August 2020].
- [13] Sosa-Estani S, Segura EL, Salomón OD, et al. Tegumentary leishmaniasis in Northern Argentina: distribution of infection and disease, in three municipalities of Salta, 1990–1992. *Rev Soc Bras Med Trop* 2000;33(6):573–82. <https://doi.org/10.1590/S0037-8682200000600009>
- [14] Poepl W, Herkner S, Tobudic S, et al. Seroprevalence and asymptomatic carriage of Leishmania spp. in Austria, a non-endemic European country. *Parasitology* 2013;19(6):572–7. <https://doi.org/10.1111/j.1469-0691.2012.03960.x>
- [15] Badaro R, Jones TC, Lorenço R, et al. A prospective study of visceral leishmaniasis in an endemic area of Brazil. *J Infect Dis* 1986;154(4):639–49. <https://doi.org/10.1093/infdis/154.4.639>
- [16] D' Oliveira Junior A, Costa SRM, Barbosa AB, et al. Asymptomatic Leishmania chagasi Infection in Relatives and Neighbors of Patients with Visceral Leishmaniasis. *Mem Inst Oswaldo Cruz* 1997;92(1):15–20. <https://doi.org/10.1590/S0074-02761997000100003>
- [17] Fukutani KF, Figueiredo V, Celes FS, et al. Serological survey of Leishmania infection in blood donors in Salvador, Northeastern Brazil. *BMC Infect Dis* 2014;14:1–8. <https://doi.org/10.1186/1471-2334-14-422>
- [18] Ferreira-Silva MM, Teixeira LAS, Tibúrcio MS, et al. Socio-epidemiological characterisation of blood donors with asymptomatic Leishmania infantum infection from three Brazilian endemic regions and analysis of the transfusional transmission risk of visceral Leishmaniasis. *J Br Blood Transf Soc* 2018;28(6):1–7. <https://doi.org/10.1111/tme.12553>
- [19] Carranza-Tamayo CO, Werneck GL, Romero GAS. Are opossums a relevant factor associated with asymptomatic Leishmania infection in the outskirts of the largest Brazilian cities. *Braz J Infect Dis* 2016;20(2):119–26. <https://doi.org/10.1016/j.bjid.2015.11.013>
- [20] Monteiro DCS, Sousa AQ, Lima DM, et al. Leishmania infantum Infection in Blood Donors, Northeastern Brazil. *Emerg Infect Dis* 2016;22(4):739–40. <https://doi.org/10.3201/eid2204.150065>
- [21] Caldas AJM, Costa JML, Silva AAM, et al. Risk factors associated with asymptomatic infection by Leishmania chagasi in north-east Brazil. *Trans R Soc Trop Med Hyg* 2002;96(1):21–8. [https://doi.org/10.1016/S0035-9203\(02\)90227-0](https://doi.org/10.1016/S0035-9203(02)90227-0)
- [22] Gama MEA, Costa JML, Gomes CMC, et al. Subclinical form of the American visceral Leishmaniasis. *Mem Inst Oswaldo Cruz* 2004;99(8):889–93. <https://doi.org/10.1590/S0074-02762004000800018>
- [23] Mendes DG, Lauria-Pires L, Nitz N, et al. Exposure to mixed asymptomatic infections with Trypanosoma cruzi, Leishmania braziliensis and Leishmania chagasi in the human population of the greater Amazon. *Trop Med Int Health* 2007;12(5):629–33. <https://doi.org/10.1111/j.1365-3156.2007.01831.x>
- [24] Moura GS, Santos AM, Aquino DMC, et al. Factors associated with asymptomatic infection in family members and neighbors of patients with visceral Leishmaniasis. *Cad Saúde Pública Rio De Jan* 2012;28(12):2306–14. <https://doi.org/10.1590/S0102-311X2012001400009>
- [25] De Oliveira ALL, Paniago AMM, Sanches MA, et al. Asymptomatic infection in family contacts of patients with human visceral Leishmaniasis in Três Lagoas, Mato Grosso do Sul State, Brazil. *Cad Saúde Pública* 2008;24(12):2827–33. <https://doi.org/10.1590/S0102-311X2008001200011>
- [26] França AO, De Castro VL, Lima Junior MS, et al. Anti-Leishmania antibodies in blood donors from the Midwest region of Brazil. *Transfus Apher Sci* 2013;49(3):627–30. <https://doi.org/10.1016/j.transci.2013.07.009>
- [27] França AO, Da Cunha GMR, Oliveira LP, et al. Presence of anti-Leishmania antibodies in candidates for kidney transplantation. *Int J Infect Dis* 2020;98:470–7. <https://doi.org/10.1016/j.ijid.2020.07.006>
- [28] Moreno EC, Melo MN, Lambertucci JR, et al. Diagnosing human asymptomatic visceral Leishmaniasis in an urban area of the State of Minas Gerais, using serological and molecular biology techniques. *Med Trop* 2006;39(5):421–7. <https://doi.org/10.1590/S0037-86822006000500001>
- [29] Moreno EC, Gonçalves AV, Chaves AV, et al. Inaccuracy of enzyme-linked immunosorbent assay using soluble and recombinant antigens to detect asymptomatic infection by Leishmania infantum. *PLoS Negl Trop Dis* 2009;3(10):536. <https://doi.org/10.1371/journal.pntd.0000536>
- [30] Silva LA, Romero HD, Nascentes GAN, et al. Antileishmania immunological tests for asymptomatic subjects living in a Visceral Leishmaniasis-Endemic Area in Brazil. *Am J Trop Med Hyg* 2011;84(2):261–6. <https://doi.org/10.4269/ajtmh.2011.00-0092>
- [31] Clemente WT, Rabello A, Faria LC, et al. High prevalence of asymptomatic Leishmania spp. infection among liver transplant recipients and donors from an endemic area of Brazil. *Am J Transpl* 2014;14:96–101. <https://doi.org/10.1111/ajt.12521>
- [32] Marques LHS, Da Rocha ICM, Reis IA, et al. Leishmania infantum: illness, transmission profile and risk factors for asymptomatic infection in an endemic metropolis in Brazil. *Parasitology* 2017;144(4):546–56. <https://doi.org/10.1017/S0031182016002134>
- [33] Da Rocha IC, Dos Santos LHM, Coura-Vital W, et al. Effectiveness of the Brazilian Visceral Leishmaniasis Surveillance and Control Programme in reducing the prevalence and incidence of Leishmania infantum infection. *Parasit Vectors* 2018;11(1):586. <https://doi.org/10.1186/s13071-018-3166-0>
- [34] Da Cunha GMR, Carneiro M, Xavier MAP, et al. Prospection of immunological biomarkers for characterization and monitoring of asymptomatic Leishmania (Leishmania) infantum infection. *Parasitology* 2020;147(10):1–34. <https://doi.org/10.1017/S0031182020000852>
- [35] Silveira FT, Laison R, Crescente JA, et al. A prospective study on the dynamics of the clinical and immunological evolution of human Leishmania (L.) infantum chagasi infection in the Brazilian Amazon region. *Trans R Soc Trop Med Hyg* 2010;104(8):529–35. <https://doi.org/10.1016/j.trstmh.2010.05.002>
- [36] Costa CHN, Stewart JM, Gomes RBB, et al. Asymptomatic human carriers of Leishmania chagasi. *Am J Trop Med Hyg* 2002;66(4):334–7. <https://doi.org/10.4269/ajtmh.2002.66.334>
- [37] Bittar RC, Nogueira RS, Vieira-Gonçalves R, et al. T-cell responses associated with resistance to Leishmania infection in individuals from endemic areas for Leishmania (Viannia) braziliensis. *Mem Inst Oswaldo Cruz* 2007;102(5):625–30. <https://doi.org/10.1590/S0074-02762007005000069>
- [38] Otero AC, Da Silvai VO, Luz KG, et al. Short report: occurrence of Leishmania donovani DNA in donated blood from seroreactive Brazilian blood donors. *Am J Trop Med Hyg* 2000;62(1):128–31. <https://doi.org/10.4269/ajtmh.2000.62.128>
- [39] De Lima I, Queiroz JW, Lacerda HG, et al. Leishmania infantum chagasi in Northeastern Brazil: asymptomatic infection at the Urban Perimeter. *Am J Trop Med Hyg* 2012;86(1):99–107. <https://doi.org/10.4269/ajtmh.2012.10-0492>
- [40] Fagundes A, Marzochi MCA, Fernandes O, et al. First encounter of subclinical human Leishmania (Viannia) infection in the State of Rio Grande do Sul, Brazil. *Mem Inst Oswaldo Cruz* 2007;102(8):1003–5. <https://doi.org/10.1590/S0074-02762007000800018>
- [41] Barão SC, Camargo-Neves VLF, Resende MR, et al. Human asymptomatic infection in visceral Leishmaniasis: a seroprevalence study in an urban area of low endemicity. *Am J Trop Med Hyg* 2007;77(6):1051–3. <https://doi.org/10.4269/ajtmh.2007.77.1051>
- [42] Šiško-Kraljević N, Jerončić A, Mohar B, et al. Asymptomatic Leishmania infantum infections in humans living in endemic and non-endemic areas of

- Croatia, 2007 to 2009. *Eur Surveill* 2013;18(29):1–8. <https://doi.org/10.2807/1560-7917.ES2013.18.28.20533>
- [43] Hailu A, Kroon CCM, Schoone GJ, et al. Sero-epidemiological assessment and diagnosis of visceral Leishmaniasis in an endemic locality using Fast Agglutination Screening Test (FAST). *Acta Trop* 2002;83(2):93–101. [https://doi.org/10.1016/s0001-706x\(02\)00063-3](https://doi.org/10.1016/s0001-706x(02)00063-3)
- [44] Griensven JV, Henten SV, Mengesha B, et al. Longitudinal evaluation of asymptomatic Leishmania infection in HIV-infected individuals in North-West Ethiopia: a pilot study. *PLOS Negl Trop Dis* 2019;13(10):1–17. <https://doi.org/10.1371/journal.pntd.0007765>
- [45] Tadese D, Hailu A, Bekele F, et al. An epidemiological study of visceral leishmaniasis in North East Ethiopia using serological and leishmanin skin tests. *PLOS ONE* 2019;14(12):1–10. <https://doi.org/10.1371/journal.pone.0225083>
- [46] Gadisa E, Custodio E, Cañavate C, et al. Usefulness of the rK39-immunochromatographic test, direct agglutination test, and leishmanin skin test for detecting asymptomatic leishmania infection in children in a new Visceral Leishmaniasis focus in Amhara State, Ethiopia. *Am J Trop Med Hyg* 2012;86(5):792–8. <https://doi.org/10.4269/ajtmh.2012.11-0196>
- [47] Custodio E, Gadisa E, Sordo L, et al. Factors associated with leishmania asymptomatic infection: results from a cross-sectional survey in highland Northern Ethiopia. *PLOS Negl Trop Dis* 2012;6(9):1–9. <https://doi.org/10.1371/journal.pntd.0001813>
- [48] Ayehe A, Aschale Y, Lemma W, et al. Seroprevalence of asymptomatic leishmania donovani among laborers and associated risk factors in agricultural camps of West Armachiho District, Northwest Ethiopia: a cross-sectional study. *J Parasitol Res* 2018;1–8. <https://doi.org/10.1155/2018/5751743>
- [49] Bekele F, Belay T, Zeynudid A, et al. Visceral Leishmaniasis in selected communities of Hamar and Banna-Tsamai districts in Lower Omo Valley, South West Ethiopia: Sero-epidemiological and Leishmanin Skin Test Surveys. *PLoS ONE* 2018;13(5):1–13. <https://doi.org/10.1371/journal.pone.0197430>
- [50] Le Fichoux YI, Quaranta JF, Aueuvre JP, et al. Occurrence of leishmania infantum parasitemia in asymptomatic blood donors living in an area of endemicity in Southern France. *J Clin Microbiol* 1999;37(6):1953–7. <https://doi.org/10.1128/jcm.37.6.1953-1957.1999>
- [51] Singh S, Kumari V, Singh N. Predicting kala-azar disease manifestations in symptomatic patients with latent Leishmania donovani infection by detection of antibody against recombinant K39 antigen. *Clin Diagn Lab Immunol* 2002;9(3):568–72. <https://doi.org/10.1128/cdli.9.3.568-572.2002>
- [52] Das VNR, Siddiqui NA, Verma RB, et al. Asymptomatic infection of visceral Leishmaniasis in hyperendemic areas of Vaishali district, Bihar, India: a challenge to kala-azar elimination programmes. *Trans R Soc Trop Med Hyg* 2011;105(11):661–6. <https://doi.org/10.1016/j.trstmh.2011.08.005>
- [53] Gidwani K, Ostyn B, Picado A, et al. Persistence of human Leishmania antibodies in past Kala azar cases in the Indian subcontinent. *Trop Med Int Health* 2011;18(2):346–8. <https://doi.org/10.1128/CVI.00473-10>
- [54] Topno RK, Dasm VNR, Ranjan A, et al. Asymptomatic infection with visceral leishmaniasis in a disease-endemic area in Bihar, India. *Am J Trop Med Hyg* 2010;83(3):502–6. <https://doi.org/10.4269/ajtmh.2010.09-0345>
- [55] Sudarshan M, Sudar S. Parasite load estimation by qPCR differentiates between asymptomatic and symptomatic infection in Indian visceral leishmaniasis. *Parasitology* 2014;80(1):40–2. <https://doi.org/10.1016/j.diagmicrobio.2014.01.031>
- [56] Das VNR, Pandey RN, Siddiqui NA, et al. Longitudinal study of transmission in households with visceral leishmaniasis, asymptomatic infections and PKDL in highly endemic villages in Bihar, India. *PLoS Negl Trop Dis* 2016;10(12):1–12. <https://doi.org/10.1371/journal.pntd.0005196>
- [57] Das VNR, Bimal S, Siddiqui NA, et al. Conversion of asymptomatic infection to symptomatic visceral Leishmaniasis: a study of possible immunological markers. *PLOS Negl Trop Dis* 2020;14(6):1–18. <https://doi.org/10.1371/journal.pntd.0008272>
- [58] Saha P, Ganguly S, Chatterjee M, et al. Asymptomatic Leishmaniasis in kala-azar endemic areas of Malda district, West Bengal, India. *PLoS Negl Trop Dis* 2017;11(2):1–10. <https://doi.org/10.1371/journal.pntd.0005391>
- [59] Sudarshan M, Singh T, Singh AK, et al. Quantitative PCR in epidemiology for early detection of Visceral Leishmaniasis cases in India. *PLoS Negl Trop Dis* 2014;8(12):1–7. <https://doi.org/10.1371/journal.pntd.0003366>
- [60] Ostyn B, Gidwani K, Khanal B, et al. Incidence of symptomatic and asymptomatic leishmania donovani infections in high-endemic foci in India and Nepal: a prospective study. *PLoS Negl Trop Dis* 2011;5(10):1–7. <https://doi.org/10.1371/journal.pntd.0001284>
- [61] Picado A, Ostyn B, Singh SP, et al. Risk factors for visceral leishmaniasis and asymptomatic Leishmania donovani infection in India and Nepal. *PLoS ONE* 2014;9(1):1–8. <https://doi.org/10.1371/journal.pone.0087641>
- [62] Kaushal H, Bhattacharya SK, Verma S, et al. Serological and molecular analysis of leishmania infection in healthy individuals from two districts of West Bengal, India, Endemic for Visceral Leishmaniasis. *Am J Trop Med Hyg* 2017;96(6):1448–55. <https://doi.org/10.4269/ajtmh.16-0592>
- [63] Alborzi A, Pourabbas B, Shahian F, et al. Detection of Leishmania infantum kinetoplast DNA in the whole blood of asymptomatic individuals by PCR-ELISA and comparison with other infection markers in Endemic Areas, Southern Iran. *Am J Trop Med Hyg* 2008;79(6):839–42. <https://doi.org/10.4269/ajtmh.2008.79.839>
- [64] Asfaram S, Fakhar M, Mohebbi M, et al. Asymptomatic human blood donors carriers of Leishmania infantum: potential reservoirs for visceral Leishmaniasis in northwestern Iran. *Transfus Apher Sci* 2017;56(3):474–9. <https://doi.org/10.1016/j.transci.2017.06.001>
- [65] Fakhar M, Motazedian MH, Hatam GR, et al. Asymptomatic human carriers of Leishmania infantum: possible reservoirs for Mediterranean visceral Leishmaniasis in southern Iran. *Ann Trop Med Parasitol* 2008;102(7):577–83. <https://doi.org/10.1179/136485908X337526>
- [66] Gigloo AL, Sarkari B, Rezaei Z, et al. Asymptomatic Leishmania Infected Children: a Seroprevalence and Molecular Survey in a Rural Area of Fars Province, Southern Iran. *J Trop Med* 2018;15:1–6. <https://doi.org/10.1155/2018/8167247>
- [67] Rezaei Z, Sarkari B, Dehghani M, et al. High frequency of subclinical Leishmania infection among HIV-infected patients living in the endemic areas of visceral Leishmaniasis in Fars province, southern Iran. *Parasitol Res* 2018;117(8):2591–5. <https://doi.org/10.1007/s00436-018-5949-9>
- [68] Adini I, Ephros M, Chen J, Jaffe CL. Asymptomatic Visceral Leishmaniasis, Northern Israel. *Emerg Infect Dis* 2003;9(3):397–8. <https://doi.org/10.3201/eid0903.020297>
- [69] Comai G, De Pascali AM, Busutti M, et al. Screening strategies for the diagnosis of asymptomatic Leishmania infection in dialysis patients as a model for kidney transplant candidates. *J Nephrol* 2020;34(1):191–5. <https://doi.org/10.1007/s40620-020-00705-4>
- [70] Maritati M, Trentini A, Michel G, et al. Subclinical Leishmania infection in patients with rheumatic diseases under biological drugs. *Infection* 2018;46(6):401–9. <https://doi.org/10.1007/s15010-018-1189-2>
- [71] Colomba C, Saporito L, Polara FV, et al. Serological screening for Leishmania infantum in asymptomatic blood donors living in an endemic area (Sicily, Italy). *Transfus Apher Sci* 2005;33(3):311–4. <https://doi.org/10.1016/j.transci.2005.07.009>
- [72] Ortalli M, De Pascali AM, Longo S, et al. Asymptomatic Leishmania infantum infection in blood donors living in an endemic area, northeastern Italy. *J Infect* 2020;80(1):116–20. <https://doi.org/10.1016/j.jinf.2019.09.019>
- [73] Echchakery M, Nieto J, Boussaia S, et al. Asymptomatic carriers of Leishmania infantum in patients infected with human immunodeficiency virus (HIV) in Morocco. *Parasitol Res* 2018;117(4):1237–44. <https://doi.org/10.1007/s00436-018-5805-y>
- [74] Ostyn B, Uranw S, Bhattarai NR, et al. Transmission of Leishmania donovani in the Hills of Eastern Nepal, an Outbreak Investigation in Okhaldhunga and Bhojpur Districts. *PLOS Negl Trop Dis* 2015;9(8):1–13. <https://doi.org/10.1371/journal.pntd.0003966>
- [75] Timilsina S, Bhattarai NR, Khanal B, et al. Serological assessment for leishmania donovani infection in blood donors of Sunsari District, Dharan, Nepal. *Indian J Hematol Blood Transfus* 2016;32(1):95–9. <https://doi.org/10.1007/s12288-015-0505-6>
- [76] Ena J, Pasquau F, López-Perezguz MM, et al. Screening for subclinical Leishmania infection in HIV-infected patients living in eastern Spain. *Pathog Glob Health* 2014;108(8):356–61. <https://doi.org/10.1179/204773214Y.0000000164>
- [77] Elmahallawy EK, Cuadros-Moronta E, Liébana-Martos MC, et al. Seroprevalence of Leishmania infection among asymptomatic renal transplant recipients from southern Spain. *Transpl Infect Dis* 2015;17(6):795–9. <https://doi.org/10.1111/tid.12444>
- [78] Molina R, Jiménez M, García-Martínez J, et al. Role of asymptomatic and symptomatic humans as reservoirs of visceral leishmaniasis in a Mediterranean context. *PLoS Negl Trop Dis* 2020;14(4):1–16. <https://doi.org/10.1371/journal.pntd.0008253>
- [79] Riera C, Fisa R, Udina M, et al. Detection of Leishmania infantum cryptic infection in asymptomatic blood donors living in an endemic area (Eivissa, Balearic Islands, Spain) by different diagnostic methods. *Trans R Soc Trop Med Hyg* 2004;98(2):102–10. [https://doi.org/10.1016/s0035-9203\(03\)00015-4](https://doi.org/10.1016/s0035-9203(03)00015-4)
- [80] Riera C, Fisa R, López-Chejade P, et al. Asymptomatic infection by Leishmania infantum in blood donors from the Balearic Islands (Spain). *J Transf* 2008;48(7):1383–9. <https://doi.org/10.1111/j.1537-2995.2008.01708.x>
- [81] Jimenez-Marco T, Riera C, Girona-Llobera E, et al. Strategies for reducing the risk of transfusion-transmitted leishmaniasis in an area endemic for Leishmania infantum: a patient- and donor-targeted approach. *Blood Transfus* 2018;16(2):130–6. <https://doi.org/10.2450/2017.0201-16>
- [82] Ibarra-Meneses AV, Carrillo E, Sanchez C, et al. Interleukin-2 as a marker for detecting asymptomatic individuals in areas where Leishmania infantum is endemic. *Clin Microbiol Infect* 2016;22(8):1–739. <https://doi.org/10.1016/j.cmi.2016.05.021>
- [83] Pérez-Cutillas P, Goyena E, Chitima L, et al. Spatial distribution of human asymptomatic Leishmania infantum infection in southeast Spain: a study of environmental, demographic and social risk factors. *Acta Trop* 2015;146:127–34. <https://doi.org/10.1016/j.actatropica.2015.03.017>
- [84] Aliaga L, Ceballos J, Sampedro A, et al. Asymptomatic Leishmania infection in blood donors from the Southern of Spain. *J Infect Dis* 2019;47(5):739–47. <https://doi.org/10.1007/s15010-019-01297-3>
- [85] Ranasinghe S, Wickremasinghe R, Munasinghe A, et al. Cross-sectional study to assess risk factors for leishmaniasis in an endemic Region in Sri Lanka. *Am J Trop Med Hyg* 2013;89(4):742–9. <https://doi.org/10.4269/ajtmh.12-0640>
- [86] Mohamed NS, Osman HA, Muneer MS, et al. Identifying asymptomatic Leishmania infections in non-endemic villages in Gedaref state, Sudan. *BMC Res* 2019;12:1–7. <https://doi.org/10.21203/rs.2.11134/v2>
- [87] Manomat J, Leelayoova S, Bualert L, et al. Prevalence and risk factors associated with Leishmania infection in Trang Province, southern Thailand. *PLOS Negl Trop Dis* 2017;11(11):1–15. <https://doi.org/10.1371/journal.pntd.0006095>



- [88] Saghrouni F, Khammari I, Kaabia N, et al. Asymptomatic carriage of Leishmania in family members of patients with visceral leishmaniasis in Central Tunisia. *Pathol Biol* 2012;60(5):55–8. <https://doi.org/10.1016/j.patbio.2011.11.001>
- [89] Sakru N, Korkmaz M, Ozbel Y, et al. Investigation of asymptomatic visceral Leishmaniasis cases using western blot in an endemic area in Turkey. *N Microbiol* 2007;30(1):13–8.
- [90] Ates SC, Bagirova M, Allahverdiyev AM, et al. Detection of antileishmanial antibodies in blood sampled from blood bank donors in Istanbul. *Future Microbiol* 2012;7(6):773–9. <https://doi.org/10.2217/fmb.12.46>
- [91] Ates SC, Bagirova M, Allahverdiyev AM, et al. Utility of the microculture method for Leishmania detection in non-invasive samples obtained from a blood bank. *Acta Trop* 2013;128(1):54–60. <https://doi.org/10.1016/j.actatropica.2013.06.009>
- [92] Mody RM, Lakhali-Naouar I, Sherwood JE, et al. Asymptomatic Visceral Leishmania infantum Infection in US Soldiers Deployed to Iraq. *Clin Infect Dis* 2019;68(12):2036–44. <https://doi.org/10.1093/cid/ciy811>
- [93] Courtenay O, Quinnell RJ, Garcez LM, et al. Infectiousness in a cohort of Brazilian dogs: why culling fails to control visceral leishmaniasis in areas of high transmission. *J Infect Dis* 2002;186(9):1314–20. <https://doi.org/10.1086/344312>
- [94] Singh OP, Hasker E, Sacks D, et al. Asymptomatic Leishmania infection: a new challenge for Leishmania control. *Clin Infect Dis* 2014;58(10):1424–9. <https://doi.org/10.1093/cid/ciu102>
- [95] Pace D. Leishmaniasis. *J Infect* 2014;69:01–9. <https://doi.org/10.1016/j.jinf.2014.07.016>
- [96] Neitzke-Abreu HC, Venazzi MS, Bernal MVZ, et al. Detection of DNA from Leishmania (Viannia): accuracy of polymerase chain reaction for the diagnosis of Cutaneous Leishmaniasis. *PLoS One* 2013;5(8):1–8. <https://doi.org/10.1371/journal.pone.0062473>
- [97] Urias EVR, Carvalho SFG, Oliveira CL, et al. Prevalence of adults infected with Leishmania (Leishmania) chagasi among blood donors of the Hemominas Foundation in Montes Claros, Minas Gerais, Brazil. *Rev Bras Hematol Hemoter* 2009;31:348–54. <https://doi.org/10.1590/S1516-84842009005000073>
- [98] P.A.H.O. Pan American Health Organization. Leishmaniasis: Epidemiological Report in the Americas. Number 9, December 2020. Washington, D.C. 2020. <https://iris.paho.org/handle/10665.2/53090>.
- [99] Kammona O, Tsanaktisidou E. Nanotechnology-aided diagnosis, treatment and prevention of leishmaniasis. *Int J Pharm* 2021;605:120761 <https://doi.org/10.1016/j.ijpharm.2021.120761>
- [100] Michel G, Pomares C, Ferrua B, et al. Importance of worldwide asymptomatic carriers of Leishmania infantum (L. chagasi) in human. *Acta Trop* 2011;119(2–3):69–75. <https://doi.org/10.1016/j.actatropica.2011.05.012>
- [101] Silva LP, Werkauser SMR, Silva KG, et al. Asymptomatic Leishmania infection in blood donors from a major blood bank in Northeastern Brazil: a cross-sectional study. *Rev Inst Med Trop* 2020;62:1–7. <https://doi.org/10.1590/S1678-9946202062092>