

**SOUTH AMERICAN  
TREMATODES PARASITES  
OF AMPHIBIANS AND REPTILES**



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We dedicate this book to the memory  
of our unforgettable Professors  
**Lauro Travassos**  
and  
**João Ferreira Teixeira de Freitas**





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# CONTENTS

Summary .....	13
Introduction .....	13
Methodology .....	14
<b>CLASS TREMATODA .....</b>	<b>17</b>
<b>AMPHIBIA SUBCLASS DIGENEA .....</b>	<b>17</b>
Superfamily Allocreadioidea .....	17
Family Allocreadiidae .....	17
Superfamily Gorgoderoidea .....	17
Family Brachycoelidae .....	17
Family Dicrocoeliidae .....	18
Family Gorgoderidae .....	19
Family Mesocoelidae .....	22
Superfamily Hemiuroidea .....	23
Family Derogenidae .....	23
Superfamily Microphalloidea .....	24
Family Pleurogenidae .....	24
Superfamily Paramphistomoidea .....	24
Family Diplodiscidae .....	24
Superfamily Plagiorchioidea .....	27
Family Glypthelminthidae .....	27
Family Haematoloechidae .....	28
Family Macroderoididae .....	30
Family Plagiorchiidae .....	33
Family Telorchiidae .....	36
<b>REPTILIA SUBCLASS ASPIDOGASTREA .....</b>	<b>37</b>
Family Aspidogastridae .....	37
<b>REPTILIA SUBCLASS DIGENEA .....</b>	<b>37</b>
Superfamily Allocreadioidea .....	37
Family Allocreadiidae .....	37
Superfamily Clinostomoidea .....	37
Family Clinostomidae .....	37
Family Liolopidae .....	38
Superfamily Diplostomoidea .....	38
Family Cyathocotylidae .....	38
Family Proterodiplostomidae .....	39
Superfamily Echinostomatoidea .....	43
Family Echinostomatidae .....	43
Family Psilostomidae .....	45

Family Rhytidodidae .....	46
Superfamily Gorgoderoidea.....	46
Family Brachycoeliidae.....	46
Family Braunotrematidae .....	46
Family Dicrocoeliidae .....	46
Family Mesocoeliidae .....	48
Superfamily Microphalloidea .....	49
Family Pachysolidae.....	49
Family Pleurogenidae .....	49
Superfamily Microscaphidioidea.....	50
Family Microscaphidiidae .....	50
Superfamily Opisthorchioidea.....	51
Family Cryptogonimidae .....	51
Superfamily Paramphistomoidea.....	52
Family Cladorchiidae .....	52
Family Diplodiscidae .....	55
Superfamily Plagiorchioidea.....	56
Family Macroderoididae .....	56
Family Opisthogonimidae.....	56
Family Plagiorchiidae .....	60
Family Reniferidae .....	65
Family Styphlotrematidae.....	67
Family Telorchiidae .....	67
Family Urotrematidae .....	70
Syperfamily Pronocephaloidea .....	71
Family Pronocephalidae .....	71
Superfamily Schistosomatoidea.....	73
Family Spirorchiidae.....	73
<b>Host-parasite list .....</b>	<b>77</b>
<b>References .....</b>	<b>105</b>
<b>Index to Trematoda .....</b>	<b>205</b>
<b>Index to Hosts .....</b>	<b>217</b>

# SOUTH AMERICAN TREMATODES PARASITES OF AMPHIBIANS AND REPTILES

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## SUMMARY

This catalog lists 215 species and 207 figures of trematodes parasites of amphibians and reptiles from South America: 71 species have been recorded from amphibians and 150 from reptiles. Six species have been reported in both groups of hosts. Parasitizing amphibians 36 species have been reported from Brazil, 24 species from Argentina, 19 from Uruguay, 11 from Peru and Venezuela each, 06 from Colombia and Paraguay each, 04 from Ecuador, 03 from Chile, and one from Bolivia. No amphibian trematode was reported from the The Falkland Islands, French Guyana, Galapagos Islands, Guyana and Surinam. From reptiles 112 species have been reported from Brazil, 32 from Argentina, 23 from Uruguay, 18 from Venezuela, 10 from Colombia, 06 from Ecuador, 05 from Paraguay, 04 from Galapagos Islands, 03 from Peru and one from Bolivia, French Guyana and Surinam each. Two species have been reported from South American reptiles without specified locality. No trematode parasitizing reptiles was reported from Chile, Falkland Islands and Guyana. This survey is based on bibliographic sources and includes main measurements, figures, hosts, geographical distribution and references.

**Key words:** Trematoda –Aspidogastrea - Digenea - Amphibia – Reptilia - South America

## INTRODUCTION

South America includes: Argentina, Bolivia, Brazil, Chile, Colombia, Ecuador (with Galapagos Islands), French Guyana (overseas region of France), Guyana, Paraguay, Peru, Surinam, Uruguay, Venezuela and the Falkland Islands (British overseas territory).

A catalog of Aspidogastrea and Digenea parasites of fishes from South America has been edited by Kohn, Fernandes & Cohen (2007) and in this opportunity we present the trematodes parasites of Amphibia and Reptilia from the same region. We have attempted to include in one paper all published reports based on original records and some published in journals not easily available to scientists from other regions of the globe. Parasites species referred in thesis and scientific meetings do not constitute formal publications and are, consequently, not considered herein. However, some species cited in thesis or scientific meetings were

included, considering that they have been cited in published check-lists or catalogues. Undetermined species without figures and/or measurements were not included.

Several check-lists and catalogs including the parasites of amphibians and reptiles had been published from these regions such as Viana (1924) and Travassos, Freitas & Kohn (1969) for Brazil; Thatcher (1993) for the Neotropical Region; Caballero & Diaz-Ungria (1958), Heyneman, Brenes & Diaz-Ungria (1960) and Diaz-Ungria (1967; 1973) for Venezuela; Masi-Pallares, Benitez-Usher & Maciel (1976) for Paraguay; Tantaleán, Sarmento & Huiza (1992) for Peru and Lunaschi & Drago (2007; 2010) for Argentina.

One hundred and thirteen species of Amphibia and 256 species of Reptilia were found parasitized by trematodes in South America. Among the amphibians, *Leptodactylus latrans* was the host-species most parasitized with 31 different species of trematodes and among the reptiles, the most parasitized were *Caiman crocodilus* and *Chelonia mydas* parasitized by 18 species each.

Two hundred and fifteen species of trematodes have been reported parasitizing amphibians and reptiles: 71 species of Digenea have been recorded from amphibians and 149 species of Digenea and one species of Aspidogastrea from reptiles. Six species have been reported in both groups of hosts.

In 1819 Rudolphi described the first two species of Digenea from South America, collected by Natterer in Brazil: *Mesocoelium monas* from the amphibian *Siphonops annulatus* and *Rhytidodes gelatinosus* from the reptiles *Caretta caretta* and *Podocnemis expansa*.

In spite of the Galapagos Islands belonging to Ecuador, we judge most convenient to refer the species of this locality separately, considering its geographical and ecological characteristics.

Parasitizing amphibians, 36 species of Digenea have been reported from Brazil, 23 species from Argentina, 19 from Uruguay, 11 from Peru and Venezuela each, 06 from Colombia and Paraguay each, 04 from Ecuador, 03 from Chile, and one from Bolivia. No amphibian trematode was reported from the Falkland Islands, French Guyana, Galapagos Islands, Guyana and Surinam.

Parasitizing reptiles, 112 species of trematodes have been reported from Brazil, 32 from Argentina, 23 from Uruguay, 18 from Venezuela, 10 from Colombia, 06 from Ecuador, 05 from Paraguay, 04 from Galapagos Islands, 03 from Peru and one from Bolivia, French Guyana and Surinam each. Two species had been reported from South American reptiles without specified locality. No trematode parasite of reptiles were reported from Chile, The Falkland Islands and Guyana. The only species of Aspidogastrea from South America, *Lophotaspis vallei* (Stossich, 1899) was found in Brazil in the reptile *Caretta caretta*.

## METHODOLOGY

Parasites species referred in thesis and scientific meetings do not constitute formal publications and are, consequently, not considered herein. However, some species cited in thesis or scientific meetings were included, considering that they have been cited in published check-lists or catalogues. Undetermined species without figures and/or measurements were not included.

The superfamilies, families, genera and parasite species are presented in alphabetical order. For each parasite species listed, the following information is provided:

- The current scientific name, including author(s) and date(s). No attempt has been made to evaluate systematically the validity of published records. The country is followed by updated scientific name of host species, with original host record name in parentheses (when changed), references into brackets, the site and the main measurements.
- The abbreviations correspond to: B: body, E: eggs, FB: forebody, HC: head collar, HD: hindbody, OS: oral sucker, PD: peristomic disc, VS: ventral sucker, mng: measurements not given, wmd: species without morphological data.
- Measurements in micrometers, unless otherwise specified, are referred only to South American material. Measurements of different authors may be presented in some species considering different ranges.
- Species described or referred in South America with different names are stated in remarks.

Classification of the trematodes used in this study are that of Gibson, Jones & Bray (2002), Jones, Bray & Gibson (2005) and Bray, Gibson & Jones (2008). The adopted valid names of host species follow Frost (2011) and Uetz, P. (2012). Sometimes the name of the host do not correspond to the author's reference which is cited in brackets. The amphibians and reptiles are listed in the families and species in alphabetical order, followed by their trematode parasites. Host's names not found in the web.orgs are cited as in the original reference.





# AMPHIBIA

## CLASS TREMATODA

### SUBCLASS DIGENEA Carus, 1863

#### SUPERFAMILY ALLOCREADIOIDEA Looss, 1902

#### FAMILY ALLOCREADIIDAE Looss, 1902

#### CREPTOTREMA Travassos, Artigas & Pereira, 1928

*Creptotrema lynchi* Brooks, 1976 (Fig. 1)

**COLOMBIA:** *Rhinella marina* (= *Bufo marinus*) [Brooks, 1976b].

Site: Intestine.

B: 0.85-1.49 mm x 0.39-0.67 mm; OS: 156-176 x 192-336; VS: 264-396 x 276-372; E: 55-67 x 35-38 (after Brooks, 1976b).

\***MAICURU** Freitas, 1960

*Maicuru solitarium* Freitas, 1960 (Fig. 2)

**BRAZIL:** *Incilius nebulifer* (= *Bufo granulatus*) [Freitas, 1960a].

Site: Intestine.

B: 0.84 mm x 0.51 mm; OS: 120 x 150; VS: 180 x 160; E: 50-55 x 34-36 [after Freitas, 1960a].

\*Remarks: Considered *incertae sedis* by Caira & Bogéa (2005).

#### SUPERFAMILY GORGODEROIDEA Looss, 1899

#### FAMILY BRACHYCOELIIDAE Johnston, 1912

#### BRACHYCOELIUM (Dujardin, 1845) Stiles & Hassall, 1898

*Brachycoelium salamandrae* (Frolich, 1789) Luehe, 1909

**BRAZIL:** *Leptodactylus martinezi*; *Leptodactylus rhodomystax* [Goldberg, Bursey, Caldwell, Vitt & Costa, 2007, wmd].

Site: Intestine.

## **IQUITOS Mañé-Garzón & Gil, 1963**

### ***Iquitos ceii* Mañé-Garzón & Gil, 1963 (Fig. 3)**

**PERU:** *Rana palmipes* [Mañé-Garzón & Gil, 1963].

Site: Intestine.

B: 0.96-1.19 mm x 0.59-1.08 mm; OS: 78-169 in diameter; VS: 104-183 in diameter; E: 20 x 9-11 (after Mañé-Garzón & Gil, 1963).

## **FAMILY DICROCOELIIDAE Odhner, 1910**

### ***INFIDUM* Travassos, 1916**

***Infidum infidum*** (Faria, 1910) Travassos, 1916

**BRAZIL:** *Leptodactylus podicipinus* [Campião, Silva & Ferreira, 2009, wmd].

Site: Gall bladder.

## **FAMILY GORGODERIDAE Looss, 1899**

### ***GORGODERA* Looss, 1899**

***Gorgodera australiensis*** Johnston, 1912 (Fig. 4)

**ARGENTINA:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Suriano, 1965b; 1978].

Site: Urinary bladder.

B: 4.40 mm x 0.90-1.30 mm; OS: 330-500 x 400-530; VS: 550-700 x 670; E: 31-32 x 20-22 (after Suriano, 1978).

### ***GORGODERINA* Looss, 1902**

\****Gorgoderina carioca*** Fernandes, 1958 (Fig. 5)

**BRAZIL:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Fernandes, 1958].

Site: Urinary bladder.

B: 6.00-11.95 mm x 1.29-1.82 mm; OS: 510-740 in diameter; VS: 880-1,120 in diameter; E: 38-42 x 25-29 (after Fernandes, 1958).

\*Remarks: Described as *Gorgoderina* (*Metagorgoderina*) *carioca* by Fernandes (1958).

***Gorgoderina cedroi*** Travassos, 1924 (Fig. 6)

**BRAZIL:** *Hylodes nasus* (= *Elosia nasus*) [Travassos, 1924b; Fernandes, 1958].

Site: Urinary bladder.

B: 4.30-6.00 mm x 1.00 mm; OS: 400-450 in diameter; VS: 700-750 in diameter; E: 35-40 x 21 (after Travassos, 1924b).

\**Gorgoderina chilensis* Dioni, 1947 (Fig. 7)

**CHILE:** *Rhinoderma darwinii* [Dioni, 1947; Puga, 1994; Olmos & Muñoz, 2006].

**URUGUAY:** *Rhinoderma darwinii* [Dioni, 1947].

**PERU:** *Telmatobius brachydactylus* (= *Batrachophrynus brachydactylus*) [Tantaleán & García, 1993].

Site: Urinary bladder.

B: 2.76-3.63 mm x 0.40-0.46 mm; OS: mng; VS: 238-330 in diameter; E: 27 x 17 (after Dioni, 1947).

B: 2.70-4.10 mm x 0.56-0.77 mm; OS: 200-300 in diameter; VS and E: mng (after Tantaleán & García, 1993).

\*Remarks: Referred as *Gorgoderina* (*Neogorgoderina*) *chilensis* by Dioni (1947).

\**Gorgoderina cryptorchis* Travassos, 1924 (Fig. 8)

**ARGENTINA:** *Pseudopaludicola boliviana* [Hamann, Kehr & Gonzalez, 2013a].

**BRAZIL:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Travassos, 1924b; Lent, Freitas & Proença, 1946]; *Rhinella crucifer* (= *Bufo crucifer*) [Travassos, 1924b; Lent, Freitas & Proença, 1946]; *Rhinella marina* (= *Bufo marinus*) [Lent, Freitas & Proença, 1946].

**ECUADOR:** *Atelopus ignescens* [Dyer, 1986].

**PARAGUAY:** *Rhinella dorbignyi* (= *Bufo dorbigny*) [Lent, Freitas & Proença, 1946].

Site: Urinary bladder.

B: 4.00-4.70 mm x 0.90-1.00 mm; OS: 400-500 in diameter; VS: 540-700 in diameter; E: 35-40 x 21 (after Travassos, 1924b).

\*Remarks: Referred as *Gorgoderina* (*Gorgoderina*) *cryptorchis* by Fernandes (1958).

*Gorgoderina darwini* Mañé-Garzón & Gonzáles, 1978 (Fig. 9)

**URUGUAY:** *Melanophryniscus stelzneri* [Mañé-Garzón & Gonzáles, 1978a].

Site: Urinary bladder.

B: 2.85-3.99 mm x 0.31-0.44 mm; OS: 273-417 x 244-345; VS: 2,850-3,990 x 311-444; E: 24-28 x 16-20 (after Mañé-Garzón & Gonzáles, 1978a).

\**Gorgoderina diaster* Lutz, 1926 (Fig. 10)

**COLOMBIA:** *Rhinella marina* (= *Bufo marinus*) [Brooks, 1976b].

**VENEZUELA:** *Pseudis paradoxa* [Lutz, 1926; Fernandes, 1958]; *Rana palmipes* [Lutz, 1926; Fernandes, 1958; Diaz-Hungria, 1973]; *Rana* sp. [Lutz, 1928; Caballero & Diaz-Ungria, 1958].

Sites: Urinary bladder, ureters.

B: 5.39 mm x 0.96 mm; OS: 510 x 490; VS: 660 x 630; E: 29 x 21 (after Fernandes, 1958 based on Lutz's material).

\*Remarks: Referred as *Gorgoderina* (*Gorgoderina*) *diaster* by Pereira & Cuocolo (1940b) and as *Gorgoderina* (*Metagorgoderina*) *diaster* by Fernandes (1958).

*Gorgoderina festoni* Mata-López, 2005

**BRAZIL:** *Rhinella fernandezae* [Santos & Amato, 2010, wmd].

Site: Urinary bladder.

*Gorgoderina megacysta* Mañé-Garzón & Gonzáles, 1978 (Fig. 11)

**URUGUAY:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Mañé-Garzón & Gonzáles, 1978b].

Site: Urinary bladder.

B: 3.97-4.84 mm x 0.64-0.84 mm; OS: 440-592 x 473-592; VS: 287-340 in diameter; E: 16-20 long (after Mañé-Garzón & Gonzáles, 1978b).

\**Gorgoderina parvicava* Travassos, 1922 (Fig. 12)

**ARGENTINA:** *Leptodactylus chaquensis* [Hamann, Kehr, González, 2006; Schaefer, Hamann, Kehr, González & Duré, 2006; Hamann, Kehr & González, 2013a]; *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Suriano, 1965a; 1978; Hamann, Kehr & González, 2013a]; *Rhinella fernandezae* [Hamann, Kehr & González, 2013b].

**BRAZIL:** *Leptodactylus labyrinthicus* (= *Leptodactylus pentadactylus labyrinthicus*) [Dobbin Jr., 1957b]; *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Travassos, 1922b; Travassos, 1944; Travassos, Freitas & Mendonça, 1964; Vicente & Santos, 1976; Faria, 1978]; *Leptodactylus pentadactylus* [Lutz, 1926; Travassos, 1928; Fernandes, 1958]; *Rhinella crucifer* (= *Bufo crucifer*) [Travassos, 1924b]; *Rhinella icterica* (= *Bufo ictericus*) [Luque, Martins & Tavares, 2005]; *Rhinella marina* (= *Bufo aqua*; *Bufo marinus*) [Lent, Freitas & Proença, 1946].

**PARAGUAY:** *Rhinella schneideri* (= *Bufo paracnemis*; *Bufo schneideri*) [Lent, Freitas & Proença, 1946].

**PERU:** *Atelopus bomolochus* [Iannacone, 2003a]; *Leptodactylus rhodonotus* [Floríndez & Morales, 1994]; *Rhinella limensis* (= *Bufo spinulosus limensis*) [Tantaleán, Martínez & Juárez, 1974/1975]; *Telmatobius culeus* [Tantaleán, Martínez & Juárez, 1974/1975]; *Telmatobius jelskii* [Iannacone, 2003b]; *Telmatobius macrostomus* (= *Batrachophrynus macrostomus*) [García & Tantaleán, 1987]; *Telmatobius peruvianus* [Ibáñez & Córdova, 1979; Ibáñez, 1998]; *Telmatobius* sp. [Ibáñez & Córdova, 1979].

**URUGUAY:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Mañé-Garzón & Gonzáles, 1978b].

**VENEZUELA:** *Pseudis paradoxa*; *Rana palmipes* [Lent, Freitas & Proença, 1946].

Site: Urinary bladder.

B: 6.00-11.00 mm x 1.00-2.00 mm; OS: 470-700 in diameter; VS: 310-560 in diameter; E: 39-42 x 28 (after Travassos, 1922b).

B: 11.00-14.00 mm x 1.67-2.00 mm; OS: 770-970 x 770-1,030; VS: 730-930 in diameter; E: 25-33 x 21 (after Fernandes, 1958).

B: 3.02-9.70 mm x 0.62-1.52 mm; OS: 500-900 in diameter; VS: mng; E: 11-27 x 10-20 (after Suriano, 1978).

\*Remarks: Referred as *Gorgoderina permagna* by Lutz (1926) and as *Gorgoderina* (*Gorgorimma*) *parvicava* by Fernandes (1958).

*Gorgoderina parvicava minuta* Tantaleán & García, 1993 (Fig. 13)

**PERU:** *Telmatobius macrostomus* (= *Batrachophrynus macrostomus*) [Tantaleán & García, 1993].

Site: Urinary bladder.

B: 3.19-3.49 mm x 1.10-1.20 mm; OS: 530-550 in diameter; VS: mng; E: 33-41 x 24-26 (after Tantaleán & García, 1993).

\**Gorgoderina pigulevskyi* Fernandes, 1958 (Fig. 14)

**BRAZIL:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Fernandes, 1958].

Site: Urinary bladder.

B: 10.50-14.70 mm x 1.50-2.10 mm; OS: 600-650 in diameter; VS: 660-840 in diameter; E: mng (after Fernandes, 1958).

\*Remarks: Referred as *Gorgoderina* (*Metagorgoderina*) *pigulevskyi* by Fernandes (1958).

\**Gorgoderina rochalimai* Pereira & Cuocolo, 1940 (Fig. 15)

**ARGENTINA:** *Leptodactylus chaquensis* [Hamann, Kehr & González, 2006; Hamann, Kehr & González, 2013a].

**BRAZIL:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Dobbin Jr., 1957a; Fernandes, 1958]; *Rhinella arenarum* (= *Bufo arenarum*; *Bufo arenarius*) [Dobbin Jr., 1957a; Fernandes, 1958]; *Rhinella schneideri* (= *Bufo paracnemis*; *Bufo schneideri*) [Pereira & Cuocolo, 1940b; Fernandes, 1958].

Site: Urinary bladder.

B: 10.30-16.60 mm x 1.70-2.00 mm; OS: 600-770 in diameter; VS: 940-1,200 x 1,030-1,200; E: 33-38 x 21 (after Fernandes, 1958).

\*Remarks: Referred as *Gorgoderina* (*Gorgoderina*) *rochalimai* by Pereira & Cuocolo (1940b) and as *Gorgoderina* (*Metagorgoderina*) *rochalimai* by Fernandes (1958).

*Gorgoderina valdiviensis* Puga, 1979 (Fig. 16)

**CHILE:** *Calyptocephalella gayi* (= *Caudiverbera caudiverbera*) [Puga, 1979; 1982; 1994; Olmos & Muñoz, 2006].

Site: Urinary bladder.

B: 1.20 mm x 0.90 mm; OS: 400 in diameter; VS: 700 in diameter; E: 30 x 20 (after Puga, 1979).

*Gorgoderina* sp. (Fig. 17)

**PERU:** *Telmatobius* sp. [Ibáñez & Córdova, 1979; Tantaleán, Sarmiento & Huiza, 1992].

Site: Urinary bladder.

## FAMILY MESOCOELIDAE Dollfus, 1929

### MESOCOELIUM Odhner, 1910

*Mesocoelium lanfrediae* Gomes, Melo, Giese, Furtado, Gonçalves & Santos, 2013 (Fig. 18)

**BRAZIL:** *Rhinella marina* [Gomes, Melo, Giese, Furtado, Gonçalves & Santos, 2013].

Site: Intestine.

B: 1.37-3.00 mm x 0.79-1.23 mm; OS: 170-270 x 220-280; VS: 160-240 x 150-230; E: 30-34 x 20-20 (after Gomes, Melo, Giese, Furtado, Gonçalves & Santos, 2013).

\**Mesocoelium meggitti* Bhalerao, 1927 (Fig. 19)

**BRAZIL:** *Rhinella icterica* (= *Bufo marinus ictericus*) [Perez, 1964]; *Rhinella marina* (= *Bufo horribilis*; *Bufo marinus marinus*) [Pereira & Cuocolo, 1940a]; *Rhinella* sp. (= *Bufo* sp.) [Pereira & Cuocolo, 1940a].

Site: Intestine.

B: 2.67-3.11 mm x 1.25-1.36 mm; OS: 300-330 in diameter; VS: 200-240 in diameter; E: 33-37 x 25-26 (after Pereira & Cuocolo, 1940a).

\*Remarks: *Mesocoelium travassosi* Pereira & Cuocolo (1940a) was considered synonym of *M. meggitti* by Dronen, Calhoun & Simcik (2012).

\**Mesocoelium monas* (Rudolphi, 1819) Freitas, 1958 (Fig. 20)

**ARGENTINA:** *Rhinella schneideri* [Lunaschi & Drago, 2010].

**BRAZIL:** *Leptodactylus fuscus* (= *Leptodactylus sibilatrix*) [Fábio, 1982]; *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Freitas, 1963; Fábio, 1982]; *Leptodactylus mystaceus*; *Leptodactylus mystacinus* [Fábio, 1982]; *Leptodactylus pentadactylus* [Freitas, 1963]; *Rhinella arenarum* (= *Bufo arenarum*) [Freitas, 1963]; *Rhinella crucifer* (= *Bufo crucifer*) [Travassos, 1921b; 1924a; 1945; Freitas, 1963; Faria, 1978; Rodrigues, Rodrigues & Cristófaró, 1978; 1982]; *Rhinella icterica* (= *Bufo ictericus*; *Bufo marinus ictericus*) [Freitas, 1963; Travassos, Freitas & Mendonça, 1964; Faria, 1978; Rodrigues, Rodrigues & Cristófaró, 1978; 1982; Luque, Martins & Tavares, 2005]; *Rhinella marina* (= *Bufo marinus*; *Bufo marinus bimaculatus*) [Travassos, 1921b; Freitas, 1963; Rodrigues, Rodrigues & Faria, 1990]; *Rhinella schneideri* (= *Bufo paracnemis*; *Bufo schneideri*) [Travassos & Freitas, 1942; Freitas, 1963; Perez, 1964]; *Siphonops annulatus* [Diesing, 1850; Freitas, 1958].

**COLOMBIA:** *Rhinella marina* (= *Bufo marinus*; *Chaunus marinus*) [Ucrós, 1959; Bechara-Escudero & Asprilla-Murillo, 2007; Bechara & Vélez, 2010].

**PARAGUAY:** *Rhinella schneideri* (= *Bufo paracnemis*, *Bufo schneideri*) [Lent, Freitas & Proença, 1946; Masi-Pallarés & Benítez-Usher, 1976].

**PERU:** *Rhinella marina* (= *Bufo marinus*) [Tantaleán, Martínez & Juarez, 1974/1975].

**VENEZUELA:** *Incilius nebulifer* (= *Bufo granulatus*); *Rhinella marina* (= *Bufo marinus*) [Nasir & Diaz, 1971a].

Site: Intestine.

B: 1.26-2.41 mm x 0.69-1.07 mm; OS: 230-330 x 230-360; VS: 170-280 x 180-300; E: 34-44 x 21-25 (after Freitas, 1958).

\*Remarks: Referred as *Mesocoelium incognitum* by Travassos (1921b), Lent, Freitas & Proença (1946), Ucrós (1959), Perez (1964) and Masi-Pallarés & Benítez-Usher (1976); as *Mesocoelium sociale* by Ucrós (1959) and as *Mesocoelium* sp. by Travassos (1945) and Travassos & Freitas (1942).

\**Mesocoelium waltoni* Pereira & Cuocolo, 1940 (Fig. 21)

**BRAZIL:** *Rhinella marina* (= *Bufo marinus*) [Pereira & Cuocolo, 1940a].

B: 1.15-1.75 mm x 0.49-0.70 mm; OS: 200-260 in diameter; VS: 140-200; E: 37-41 x 21-25 (after Pereira & Cuocolo, 1940a).

**PERU:** *Rhinella marina* (= *Bufo marinus*) [Miyazaki, Kifune, Habe & Uyema, 1978].

Site: Intestine.

\*Remarks: This species was considered synonym of *M. monas* by Freitas (1963) and revalidated by Dronen, Calhoun & Simcik (2012).

## SUPERFAMILY HEMIUROIDEA Looss, 1899

### FAMILY DEROGENIDAE Nicoll, 1910

#### *HALIPEGUS* Looss, 1899

*Halipegus dubius* Klein, 1905 (Fig. 22)

**ARGENTINA:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Suriano, 1978].

**BRAZIL:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Kohn & Fernandes, 1988; Paraense, 1992]; *Leptodactylus pentadactylus* [Paraense, 1992].

**URUGUAY:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Cordero, 1942].

Sites: Mouth, pharynx.

B: 3.30-4.00 mm x 1.10-1.50 mm; OS: 357-485 x 457-528; VS: 542-656 x 614-656; E: 42-52 x 17-22 (after Cordero, 1942).

B: 3.29 mm x 1.07 mm; OS: 559 x 598; VS: 754 x 819; E: 44-47 x 16 with filament (after Suriano, 1978).

B: 2.06-4.14 mm x 0.86-1.65 mm; OS: 310-490 x 370-550; VS: 500-740 x 510-750; E: 40-59 x 16-23, with long polar filament (after Kohn & Fernandes, 1988).

Nomen nudum

*Halipegus similis* Lutz, 1928. Considered synonym of *Halipegus dubius* Klein, 1905 by Cordero (1942), is herein considered *nomen nudum*.

**SUPERFAMILY MICROPHALLOIDEA Ward, 1901**

**FAMILY PLEUROGENIDAE Looss, 1899**

**LOXOGENES Stafford, 1905**

*Loxogenes macrocirra* (Caballero & Bravo-Hollis, 1949) Yamaguti, 1958

**ECUADOR:** *Rana palmipes* [Dyer & Altig, 1977; Dyer, 1986, wmd].

Site: Intestine.

**PSEUDOSONSINOTREMA Dollfus, 1951**

*Pseudosonsinotrema chabaudi* (Caballero & Caballero, 1969) Sullivan, 1974 (Fig. 23)

**COLOMBIA:** *Rhinella marina* [Bechara & Vélez, 2010].

Site: Intestine.

B: 0.73-0.87 mm x 0.51-0.63 mm; OS: 78-126 x 141-165; VS: 110-157 x 118-141; E: 18-26 x 10-16 (after Bechara & Vélez, 2010).

*Pseudosonsinotrema megalorchis* Flowers, Law & Carvajal-Endara, 2011 (Fig. 24)

**ECUADOR:** *Gastrotheca pseustes* [Flowers, Law & Carvajal-Endara, 2011].

Site: Intestine

B: 0.41-0.61 mm x 0.31-0.45 mm; OS: 69-111 x 102-134; VS: 92-121 x 98-126; E: 20-31 x 11-18 (after Flowers, Law & Carvajal-Endara, 2011).

**SUPERFAMILY PARAMPHISTOMOIDEA Fiscoeder, 1901**

**FAMILY DIPLODISCIDAE Cohn, 1904**

**CATADISCUS Cohn, 1904**

*Catadiscus cohni* Travassos, 1926 (Fig. 25)

**BRAZIL:** *Rhinella icterica* [Santos, Amato & Borges-Martins, 2013]; *Rhinella marina* (= *Bufo marinus*) [Travassos, 1926b; 1934; Freitas & Lent, 1939b].

Site: Intestine.

B: 1.21-2.21 mm x 0.80-1.17 mm; OS: 100-150 (without diverticles); 220-310 (with diverticles) x 220-270 (level of diverticles); VS: 700-770 x 570-600; E: 80-88 x 42-46 (after Freitas & Lent, 1939b).

*Catadiscus corderoi* Mañé-Garzón, 1958 (Fig. 26)

**ARGENTINA:** *Leptodactylus latrans* [Lunaschi & Drago, 2010].



**URUGUAY:** *Pseudis minuta* (= *Pseudis meridionalis*) [Mañé-Garzón, 1958].

Site: Intestine.

B: 3.11 mm x 0.94 mm; OS: 310 in diameter; VS: 700 x 390; E: 57-60 x 20-30 (after Mañé-Garzón, 1958).

*Catadiscus eldoradiensis* Artigas & Pérez, 1964 (Fig. 27)

**BRAZIL:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Artigas & Pérez, 1964].

Site: Intestine.

B: 2.16-3.00 mm x 0.80-1.04 mm; OS: 100-180 x 160-250 (without diverticles); VS: 600-750 x 560-870; E: 60-82 x 30-32 (after Artigas & Pérez, 1964).

*Catadiscus freitaslenti* Ruiz, 1943 (Fig. 28)

**PARAGUAY:** *Leptodactylus fuscus* (= *Leptodactylus typhonius*) [Masi-Pallarés & Maciel, 1974; Masi-Pallarés, Benítez-Usher & Maciel, 1976]; *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Lent, Freitas & Proença, 1946; Masi-Pallarés, Benítez-Usher & Maciel, 1976]; *Rhinella schneideri* (= *Bufo schneideri*, *Bufo paracnemis*) [Lent, Freitas & Proença, 1946; Masi-Pallarés, Benítez-Usher & Maciel, 1976].

Site: Intestine.

B: 2.82-3.40 mm x 1.22-1.53 mm; OS: 170-200 long (without diverticles), 320-350 x 210-290 (with diverticles); VS: 790-810 x 630-760; E: 82-92 x 45-53 (after Lent, Freitas & Proença, 1946).

*Catadiscus hylae* Incorvaia, 1983 (Fig. 29)

**ARGENTINA:** *Hypsiboas pulchellus* (= *Hyla pulchella*) [Incorvaia, 1983].

Site: Intestine.

B: 1.38-3.15 mm x 0.57-1.56 mm; OS: 80-140 x 110-220 (without diverticles); VS: 500-900 x 400-900; E: 54-87 x 18-39 (after Incorvaia, 1983).

*Catadiscus inopinatus* Freitas, 1941 (Fig. 30)

**ARGENTINA:** *Leptodactylus bufonius* [González & Hamann, 2006; Hamann, Kehr & González, 2013a]; *Leptodactylus chaquensis* [Hamann, Kehr & González, 2006; Hamann, Kehr & González, 2013a]; *Leptodactylus latinasus* [Hamann, González & Kehr, 2006; Hamann, Kehr & González, 2013a]; *Leptodactylus latrans* [Hamann, Kehr & González, 2013a]; *Physalaemus santafecinus* [Hamann, Kehr & González, 2013a]; *Rhinella fernandezae* [Hamann, Kehr & González, 2013b]; *Scinax nasicus* [Hamann, Kehr, González, Duré & Schaefer, 2009; Hamann, Kehr & González, 2013a].

**BRAZIL:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Freitas, 1941b].

**PARAGUAY:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Lent, Freitas & Proença, 1946; Masi-Pallarés & Maciel, 1974; Masi-Pallarés, Benítez-Usher & Maciel 1976].

Site: Intestine.

B: 2.51-3.95 mm x 1.21-1.71 mm; OS: 96-113 (without diverticles); 261-278 (with diverticles) x 240-320; VS: 650-880 x 530-780; E: 84-101 x 50-55 (after Freitas, 1941b).

*Catadiscus marinholutzi* Freitas & Lent, 1939 (Fig. 31)

**ARGENTINA:** *Rhinella fernandezae* [Hamann, Kehr & González, 2013b].

**BRAZIL:** *Leptodactylus latrans* (= *Leptodactylus caliginosus*, *Leptodactylus ocellatus*) [Freitas & Lent, 1939b; Travassos & Freitas, 1941a,b; Goldberg, Bursey, Caldwell & Shepard, 2009].

Site: Intestine.

B: 1.61-2.49 mm x 1.05-1.29 mm; OS: 80-100 (without diverticles); 200-250 (with diverticles) x 180-220; VS: 700-900 x 530-980; E: 113-126 x 59-71 (after Freitas & Lent, 1939b).

*Catadiscus mirandai* Freitas, 1943 (Fig. 32)

**BRAZIL:** *Pipa carvalhoi* (= *Hemipipa carvalhoi*) [Freitas, 1943; Travassos, 1944a].

Site: Intestine.

B: 2.91 mm x 0.84 mm; OS: 116 (without diverticles); 249 (with diverticles) x 230; VS: 550-460; E: 113-122 x 61-78 (after Freitas, 1943).

*Catadiscus propinquus* Freitas & Dobbin Jr., 1956 (Fig. 33)

**ARGENTINA:** *Leptodactylus chaquensis* [Hamann, Kehr & González, 2006]; *Lysapsus limellum* (= *Lysapsus limellus*) [Hamann & Kehr, 1997; Kehr, Manly & Hamann, 2000; Hamann, 2004].

**BRAZIL:** *Leptodactylus podicipinus* [Campião, Silva & Ferreira, 2009; Campião, Delatorre, Rodrigues, Silva & Ferreira, 2012]; *Pseudis platensis* [Campião, Silva & Ferreira, 2010]; *Rana palmipes* [Freitas & Dobbin Jr., 1956; Dobbin Jr., 1957a].

Site: Intestine.

B: 1.12-1.67 mm x 0.67-1.10 mm; OS: 90-160 (without diverticles); 180-230 (with diverticles) x 150-220; VS: 430-580 in diameter; E: 118-155 x 59-88 (after Freitas & Dobbin Jr., 1956).

\**Catadiscus pygmaeus* (Lutz, 1928) Freitas & Lent, 1939 (Fig. 34)

**VENEZUELA:** *Pseudis paradoxa* [Lutz, 1928; Travassos, 1934; Freitas & Lent, 1939b; Caballero & Diaz-Ungria, 1958].

Site: Rectum.

B: 1.04 mm x 0.60 mm; OS: 120 x 176 (without diverticles); VS: 350 x 320; E: 80-88 x 56 (after Freitas & Lent, 1939).

\*Remarks: Referred as *Diplodiscus pygmaeus* by Lutz (1928) and Travassos (1934).

*Catadiscus uruguayensis* Freitas & Lent, 1939 (Fig. 35)

**ARGENTINA:** *Hypsiboas pulchellus* (= *Hyla pulchella*) [Ostrowski de Núñez, 1978/1979]; *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Suriano, 1970; 1978; Ostrowski de Núñez, 1978/1979]; *Phyllomedusa azurea* [Lunaschi & Drago, 2010]; *Pseudis minuta* (= *Lysapsus mantidactylus*) [Ostrowski de Núñez, 1978/1979].

**BRAZIL:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Freitas, 1960b]; *Lisapsus limellum* [Freitas, 1960b; Travassos & Freitas, 1964].

**URUGUAY:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Freitas & Lent, 1939b].

Sites: Intestine, stomach.

B: 0.96-2.16 mm x 0.45-0.71 mm; OS: 100-130 (without diverticles); 180-230 (with diverticles) x 130-220; VS: 400-610 x 310-560; E: 105-113 x 53-55 (after Freitas & Lent, 1939b).

B: 0.37-1.70 mm x 0.17-0.70 mm; OS: 42-124 x 52-116 (without diverticles); VS: 105-581 x 162-464; E: 84-105 x 35-63 (after Ostrowski de Núñez, (1978/1979).

## **SUPERFAMILY PLAGIORCHIOIDEA Lühe, 1901**

### **FAMILY GLYPTHELMINTHIDAE Cheng, 1959**

#### **GLYPTHELMINS Stafford, 1905**

*Glypthelmins bilialis* Suriano, 1968 (Fig. 36)

**ARGENTINA:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Suriano, 1968; 1978].

Site: Gall bladder.

B: 1.29-2.62 mm x 0.37-0.81 mm; OS: 130-270 x 130-290; VS: 130-220 x 150-230; E: 19-28 x 14 (after Suriano, 1968).

*Glypthelmins festina* Cordero, 1944 (Fig. 37)

**URUGUAY:** *Rhinella arenarum* (= *Bufo arenarum*) [Cordero, 1944].

Site: Gall bladder.

B: 1.74-3.25 mm x 0.69-0.96 mm; OS and VS: 210 in diameter; E: 40 x 23 (after Cordero, 1944).

\**Glypthelmins parva* Travassos, 1924 (Fig. 38)

**BRAZIL:** *Leptodactylus latrans* (= *Cystignathus ocellatus*; *Leptodactylus ocellatus*) [Travassos, 1924a].

**PERU:** *Dendropsophus leucophyllatus* (= *Hyla leucophyllata*); *Scinax pedromedinai*; *Trachycephalus coriaceus* (= *Phrynohyas coriacea*) [Burseley, Goldberg & Parmelee, 2001].

Site: Intestine.

B: 1.30 mm x 0.46 mm; OS: 140 in diameter; VS: 130 in diameter; E: 28 in length (after Travassos, 1924a).

\*Remarks: Bursey, Goldberg & Parmelee (2001) referred by mistake that Freitas (1941a) reported this species in Uruguay.

\**Glypthelmins sanmartini* (Mañé-Garzón & Holcman-Spector, 1974) (Fig. 39)

**URUGUAY:** *Chthonerpeton indistinctum* [Mañé-Garzón & Holcman-Spector, 1974].

Site: Intestine.

B: 3.84-4.30 mm x 1.22-1.30 mm; OS: 410-430 in diameter; VS: 280 in diameter; E: 28-30 x 15 (after Mañé-Garzón & Holcman-Spector, 1974).

\*Remarks: Referred as *Margeana sanmartini* by Mañé-Garzón & Holcman-Spector (1974). Razo-Mendivil, León-Règagnon & Ponce de León (2006) suggested the possible inclusion of this species in genus *Rauschiella*.

## FAMILY HAEMATOLOECHIDAE Freitas & Lent, 1939

### HAEMATOLOECHUS Looss, 1899

*Haematoloechus arequipensis* Ibañez & Córdova, 1979 (Fig. 40)

PERU: *Telmatobius peruvianus* [Ibañez & Córdova, 1979].

Site: Lungs.

B: 7.02-8.38 mm x 1.82-2.47 mm; OS: 400-520 x 465-578; VS: 400-546 x 450-600; E: 35-40 x 18-21 (after Ibañez & Córdova, 1979).

*Haematoloechus freitasi* Mañé-Garzón & Gil, 1959 (Fig. 41)

BRAZIL: *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Fróes & Lima, 1974].

URUGUAY: *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Mañé-Garzón & Gil, 1959].

Site: Lungs.

B: 13.28-13.94 mm x 1.69-3.51 mm; OS: 309-590 x 332-557; VS: 357-787 x 656-746; E: 21-23 x 11-15 (after Mañé-Garzón & Gil, 1959).

\**Haematoloechus fuelleborni* (Travassos & Darriba, 1930) Ingles, 1933 (Fig. 42)

BRAZIL: *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Faria, 1978]; *Rhinella icterica* (= *Chaunus ictericus*) [Hoppe, Pedrassani, Hoffmann-Inocente, Tebaldi, Storti, Zanuzzo, Avancini, & Nascimento, 2008]; *Rhinella marina* (= *Bufo marinus*) [Travassos & Darriba, 1930; Dobbin Jr., 1957b].

Site: Lungs.

B: 10.00 mm x 2.30 mm; OS: 670 in diameter; VS: 310 x 370; E: 32-39 x 21 (after Travassos & Darriba, 1930).

\*Remarks: Referred as *Pneumonoeces fuelleborni* by Travassos & Darriba (1930).

*Haematoloechus legrandi* Mañé-Garzón & Gil, 1959

URUGUAY: *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Mañé-Garzón & Gil, 1959].

Site: Lungs.

B: 6.23 mm x 1.38 mm; OS: 426 x 394; VS: 231 x 259; E: 30 x 12 (Mañé-Garzón & Gil, 1959).

*Haematoloechus longipectus* Stafford, 1902 (Fig. 43)

ARGENTINA: *Ceratophrys cranwelli* [Hamann & Pérez, 1999]; *Leptodactylus chaquensis* [Hamann & Pérez, 1999; Hamann, Kehr & González, 2006; Schaefer, Hamann, Kehr,

González & Duré, 2006; Hamann, González & Kehr, 2012; Hamann, Kehr & González, 2013]; *Leptodactylus latinasus* [Hamann, González & Kehr, 2006; Hamann, González & Kehr, 2012]; *Leptodactylus latrans* [Hamann, González & Kehr, 2012]; *Pseudopaludicola boliviana* [Hamann, González & Kehr, 2012]; *Rhinella fernandezae* (= *Bufo fernandezae*) [Hamann & Pérez, 1999; Hamann, González & Kehr, 2012, 2013b].

Site: Lungs.

B: 1.12-8.55 mm x 0.57-2.80 mm; OS: 180-490 x 198-480; VS: 168-350 x 178-350; E: 30-43 x 18-25 (after Hamann & Pérez, 1999).

\**Haematoloechus lutzi* Freitas & Lent, 1939 (Fig. 44)

**VENEZUELA:** *Rana palmipes* [Cordero & Vogelsang, 1939; Freitas & Lent, 1939a; Caballero, Vogelsang & Zerecero, 1953; Dobbin Jr., 1957b]; *Rana* sp. [Walton, 1951; Caballero & Diaz-Ungria, 1958].

Site: Lungs.

B: 4.05-4.49 mm x 1.00-1.31 mm; OS: 380-410 x 430-450; VS: 400 x 360-400; E: 34 x 17 (after Freitas & Lent, 1939a).

\*Remarks: Referred as *Pneumonoeces tejerae* by Cordero & Vogelsang (1939) and as *Haematoloechus* (*Haematoloechus*) *lutzi* by Freitas & Lent (1939a).

\**Haematoloechus ozorioi* Freitas & Lent, 1939 (Fig. 45)

**ARGENTINA:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Suriano, 1978].

**URUGUAY:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Freitas & Lent, 1939a; Dobbin Jr., 1957b].

Site: Lungs.

B: 7.03 mm x 2.14 mm; OS: 350 x 330; VS: 410 x 460; E: 21 x 17 (after Freitas & Lent, 1939a).

B: 7.10-9.43 mm x 1.66-1.88 mm; OS: 280-350 in diameter; VS: 360-520 x 410-540; E: 19-22 x 11-14 (after Suriano, 1978).

\*Remarks: Referred as *Pneumonoeces ozorioi* by Suriano (1978).

*Haematoloechus pukinensis* Ibañez & Córdova, 1979 (Fig. 46)

**PERU:** *Telmatobius peruvianus* [Ibañez & Córdova, 1979].

Site: Lungs.

B: 3.50-4.22 mm x 0.70-0.88 mm; OS: 250-300 x 258-300; VS: 218-290 x 260-300; E: 34-38 x 18-20 (after Ibañez & Córdova, 1979).

## **NEOHAEMATOLOECHUS** Odening, 1960

\**Neohaematoloechus iturbei* (Cordero & Vogelsang, 1939) Odening, 1960 (Fig. 47)

**BRAZIL:** *Rana palmipes* [Cordero & Vogelsang, 1939; Freitas & Dobbin Jr., 1956; Dobbin Jr., 1957b].

**COLOMBIA:** *Rana palmipes* [Uribe-Piedrahita, 1948].

**ECUADOR:** *Rana palmipes* [Dyer, 1986].

**VENEZUELA:** *Rana palmipes* [Cordero & Vogelsang, 1939].

Site: Lungs.

B: 9.03-15.39 mm x 1.63-2.49 mm; OS: 365-432 x 365-465; VS: absent; E: 25 x 13-17 (after Dobbin Jr., 1957b).

\*Remarks: Referred as *Pneumonoeces iturbei* by Cordero & Vogelsang (1939), as *Haematoloechus medioplexus* Stafford, 1902 by Uribe-Piedrahita (1948) and as *Haematoloechus iturbei* (Cordero & Vogelsang, 1939) Walton, 1949 by Freitas & Dobbin Jr. (1956) and Dobbin Jr. (1957b).

\**Neohaematoloechus neivai* (Travassos & Artigas, 1927) Odening, 1960 (Fig. 48)

**BRAZIL:** *Leptodactylus labyrinthicus* (= *Leptodactylus pentadactylus labyrinthicus*) [Fahel, 1952; Dobbin Jr., 1957b]; *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Travassos & Artigas, 1927; Dobbin Jr., 1957b; Vicente & Santos, 1976; Rodrigues, 1986; Fábio & Pinheiro, 2001]; *Leptodactylus pentadactylus* [Fahel, 1952]; *Pseudis paradoxa* [Travassos & Freitas, 1941c; Dobbin Jr., 1957b].

**VENEZUELA:** *Pseudis paradoxa* [Lutz, 1928; Travassos & Darriba, 1930; Caballero & Diaz-Ungria, 1958]; *Rana palmipes* [Lutz, 1928; Freitas & Lent, 1939a; Caballero & Diaz-Ungria, 1958].

Site: Lungs.

B: 3.00-7.00 mm x 1.30-2.50 mm; OS: 320-550 in diameter; VS: absent; E: 53-61 x 30 (after Travassos & Darriba, 1930).

\*Remarks: Referred as *Pneumonesces neivai* by Travassos & Artigas (1927), Travassos & Darriba (1930), as *Haematoloechus neivai* Travassos & Artigas, 1927 by Travassos & Freitas (1941c), Fahel (1952), Dobbin Jr. (1957b) and as *Pneumonoeces planorbinus* and *Pneumonoeces pseudis* by Lutz (1928).

## FAMILY MACRODEROIDIDAE McMullen, 1937

### RAUSCHIELLA Babero, 1951

\**Rauschiella chaquensis* (Mañé-Garzón & Holcman-Spector, 1967) Razo-Mendivil, León-Règagnon & Ponce de León, 2006 (Fig. 49)

**ARGENTINA:** *Leptodactylus laticeps* [Mañé-Garzón & Holcman-Spector, 1967b].

Site: Intestine.

B: 2.18-2.41 mm x 1.18-1.26 mm; OS: 290 x 290-330; VS: 160-230 x 240-260; E: 21-30 x 12 (after Mañé-Garzón & Holcman-Spector, 1967b).

\*Remarks: Referred as *Margeana chaquensis* by Mañé-Garzón & Holcman-Spector (1967b).

\**Rauschiella lenti* (Freitas, 1941) Razo-Mendivil, León-Règagnon & Ponce de León, 2006 (Fig. 50)

**URUGUAY:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Freitas, 1941a; Artigas & Zerpa, 1961; Mañé-Garzón & Holcman-Spector, 1974].

Sites: Intestine, stomach.

B: 2.38-5.69 mm x 0.67-1.17 mm; OS: 300-500 in diameter; VS: 150-250 in diameter; E: 29-36 x 15-17 (after Freitas, 1941a).

\*Remarks: Referred as *Plagiorchis lenti* by Freitas (1941a), Artigas & Zerpa (1961) and by Mañé-Garzón & Holcman-Spector (1974).

\**Rauschiella linguatula* (Rudolphi, 1819) Razo-Mendivil, Leon-Regagnon & Ponce de Leon, 2006 (Fig. 51)

**ARGENTINA:** *Leptodactylus latrans* [Lunaschi & Drago, 2010].

**BOLIVIA:** *Leptodactylus bolivianus* (after Yamaguti, 1971).

**BRAZIL:** *Anaxyrus terrestris* (= *Bufo musicus*); *Ceratophrys cornuta* [Travassos, 1924a]; *Incilius nebulifer* (= *Bufo granulatus*) [Freitas, 1960b]; *Leptodactylus labyrinthicus* (= *Leptodactylus pentadactylus labyrinthicus*) [Dobbin Jr., 1957a]; *Leptodactylus latrans* (= *Cystignathus ocellatus*, *Leptodactylus ocellatus*) [Travassos, 1924a; 1926a; Pereira & Cuocolo, 1941; Dobbin Jr., 1957a; Stumpf, 1981/1982; Rodrigues, 1986; Rodrigues, Rodrigues & Faria, 1990]; *Leptodactylus pentadactylus* [Fahel, 1952]; *Rhinella crucifer* (= *Bufo crucifer*) [Freitas, 1960b]; *Rhinella icterica* (= *Bufo marinus ictericus*) [Rodrigues, Rodrigues & Cristóforo, 1978; 1982]; *Rhinella marina* (= *Bufo aqua*, *Bufo marinus*, *Bufo marinus bimaculatus*) [Travassos, 1924a; Dobbin Jr., 1957a; Ruiz & Leão, 1942c]; *Rhinella schneideri* (= *Bufo paracnemis*, *Bufo schneideri*) [Freitas, 1960b].

**URUGUAY:** *Chthonerpeton indistinctum* [Cordero, 1944]; *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Mañé-Garzón & Holcman-Spector, 1974].

**VENEZUELA:** *Rhinella marina* (= *Bufo marinus*) [Nasir & Diaz, 1970].

Sites: Gall bladder, pancreas, intestine, liver, biliary ducts.

B: 3.00-3.20 mm x 1.40-1.70 mm; OS: 250-370 in diameter; VS: 250-270 in diameter; E: 33-38-15-20 (after Travassos, 1926a).

B: 4.00 x 1.20 mm; OS: 290 in diameter; VS: 240 in diameter; E: 28-32 x 14 (after Dobbin Jr., 1957a).

B: 5.74-8.14 mm x 1.25-1.77 mm; OS: 500-590 in diameter; VS: 230-290 in diameter; E: 29-32 x 11-14 (after Mañé-Garzón & Holcman-Spector, 1974).

\*Remarks: Referred as *Glyphelmims linguatula* by Travassos (1924a), Fahel (1952), Dobbin Jr. (1957a), Freitas (1960b) and Rodrigues, Rodrigues & Faria (1990); as *Glyphelmims elegans* by Travassos (1926a); as *Choledocystus eucharis* by Pereira & Cuocolo (1941); as *Choledocystus vesicalis* by Ruiz & Leão (1942c); as *Choledocystus elegans* by Ruiz (1949), Dobbin Jr. (1957a), Rodrigues, Rodrigues & Cristóforo (1978; 1982), Stumpf (1981/1982), Rodrigues (1986) and Lunaschi & Drago (2010); as *Glyphelmims vesicalis* by Ruiz & Leão (1942) and by Nasir & Diaz (1970); as *Margeana linguatula* by Mañé-Garzón & Holcman-Spector (1974) and as *Margeana sera* by Cordero (1944) and by Mañé-Garzón & Holcman-Spector (1974).

\**Rauschiella palmipedis* (Lutz, 1928) Sullivan, 1977 (Fig. 52)

**ARGENTINA:** *Leptodactylus chaquensis* [Hamann, Kehr & González, 2006; Schaefer, Hamann, Kehr, González & Duré, 2006; Hamann, Kehr & González, 2009; Hamann, Kehr

& González, 2013a]; *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Savazzini, 1930; Suriano, 1978; Hamann, Kehr & González, 2013a]; *Rhinella fernandezae* [Hamann, Kehr & González, 2013a].

**BRAZIL:** *Incilius nebulifer* (= *Bufo granulatus*) [Freitas, 1960b; Travassos & Freitas, 1964]; *Leptodactylus latrans* (= *Leptodactylus caliginosus*, *Leptodactylus ocellatus*) [Freitas, 1941a; Travassos & Freitas, 1941a; Dobbin Jr., 1957a; Freitas, 1960b; Travassos & Freitas, 1964; Vicente & Santos, 1976; Rodrigues, Rodrigues & Cristófaró, 1978; 1982; Rodrigues, 1986; Rodrigues, Rodrigues & Faria, 1990]; *Leptodactylus pentadactylus* (= *Leptodactylus pentadactylus labyrinthicus*) [Fahel, 1952; Dobbin Jr., 1957a]; *Pseudis paradoxa* [Travassos & Freitas, 1941a,b,c]; *Pseudis platensis* [Campiãó, Silva & Ferreira, 2010]; *Rana palmipes* [Freitas, 1941a; Dobbin Jr., 1957a]; *Rhinella icterica* (= *Chaunus ictericus*) [Hoppe, Pedrassani, Hoffmann-Inocente, Tebaldi, Storti, Zanuzzo, Avancini, & Nascimento, 2008]; *Rhinella marina* (= *Bufo aqua*, *Bufo marinus*) [Freitas, 1941a; Travassos & Freitas, 1964; Rodrigues, Rodrigues & Faria, 1990].

**PARAGUAY:** *Leptodactylus fuscus* (= *Leptodactylus typhonius*) [Masi-Pallarés & Maciel, 1974]; *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Lent, Freitas & Proença, 1946]; *Leptodactylus pentadactylus* [Fahel, 1952]; *Rhinella schneideri* (= *Bufo paracnemis*, *Bufo schneideri*) [Freitas, 1960b; Masi-Pallarés & Maciel, 1974].

**URUGUAY:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Freitas, 1941a].

**VENEZUELA:** *Leptodactylus bolivianus* [Sullivan, 1977a]; *Rana palmipes* [Lutz, 1928; Caballero, Vogelsang & Zerecero, 1953; Caballero & Diaz-Ungria, 1958]; *Rhinella marina* (= *Bufo marinus*) [Freitas, 1941a; Travassos & Freitas, 1964; Sullivan, 1977a; Rodrigues, Rodrigues & Faria, 1990].

Sites: Intestine, stomach, lung.

B: 3.35-8.88 mm x 0.94-2.08 mm; OS: 430-700 x 410-730; VS: 220-400 in diameter; E: 25-34 x 13-17 (after Freitas, 1941a).

B: 2.22-5.32 mm x 0.55-0.66 mm; OS: 260-450 in diameter; VS: 150-260 in diameter; E: 28-33 x 16 (after Suriano, 1978).

\*Remarks: Referred as *Haplometra palmipedis* by Lutz (1928); as *Glypthelmins palmipedis* by Travassos (1930), Savazzini (1930), Freitas (1941a) Travassos & Freitas (1941a,b,c), Lent, Freitas & Proença, 1946, Fahel (1952), Caballero, Vogelsang & Zerecero (1953), Dobbin Jr. (1957a), Caballero & Diaz-Ungria (1958), Freitas (1960b), Travassos & Freitas (1964), Diaz-Ungria (1967), Rodrigues (1968), Masi-Pallarés, Benítez-Usher & Maciel (1976), Vicente & Santos (1976), Suriano (1978), Rodrigues, Rodrigues & Cristófaró (1978, 1982), Rodrigues, Rodrigues & Faria (1990), Hamann, Kehr & González (2006), Schaefer, Hamann, Kehr, González & Duré (2006), Hoppe, Pedrassani, Hoffmann-Inocente, Tebaldi, Storti, Zanuzzo, Avancini, & Nascimento (2008), **Hamann, Kehr & González (2009)**, Hamann, Kehr & González (2012); and as *Metorchis leptodactylus* by Savazzini (1930).

\**Rauschiella proxima* (Freitas, 1941) Sullivan, 1977 (Fig. 53)

**URUGUAY:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Freitas, 1941a; Mañé-Garzón & Holcman-Spector, 1974].

Sites: Intestine, stomach.



B: 2.68-4.22mm x 0.74-1.00 mm; OS: 280-360 x 310-450; VS: 170-250 x 170-220; E: 42 x 13 (after Freitas, 1941a).

\*Remarks: Referred as *Glyphelminis proximus* by Freitas (1941a) and as *Margeana proximus* by Mañé-Garzón & Holcman-Spector (1974).

\****Rauschiella repandum*** (Rudolphi, 1819) Babero, 1951 (Fig. 54)

**ARGENTINA:** *Leptodactylus bufonius* [González & Hamann, 2006; Hamann, Kehr & González, 2012]; *Leptodactylus chaquensis* [Hamann, Kehr & González, 2006; Schaefer, Hamann, Kehr, González & Duré, 2006; **Hamann, Kehr & González, 2009**; Hamann, Kehr & González, 2012]; *Leptodactylus latinasus* [Hamann, González & Kehr, 2006; Hamann, Kehr & González, 2012]; *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Hamann, Kehr & González, 2012]; *Odontophrynus americanus* [Hamann, Kehr & González, 2013a]; *Physalaemus santafecinus* [Hamann, Kehr & González, 2012].

**BRAZIL:** *Leptodactylus latrans* (= *Cystignatus ocellatus*, *Leptodactylus ocellatus*) [Travassos, 1924a; Travassos & Freitas, 1941a; Pinto & Noronha, 1972; Faria, 1978; Rodrigues, Rodrigues & Cristóforo, 1978; 1982].

Site: Intestine.

B: 8.00 mm x 1.40 mm; OS: 360 in diameter; VS: 270 in diameter; E: 32 x 16 (after Travassos, 1924a).

\*Remarks: Referred as *Glyphelminis repandum* by Travassos (1924a), Travassos & Freitas (1941a), Pinto & Noronha (1972), Faria (1978), Rodrigues, Rodrigues & Cristóforo (1978, 1982), Hamann, González & Kehr (2006), González & Hamann (2006), Schaefer, Hamann, Kehr, González & Duré (2006), **Hamann, Kehr & González (2009)** and by Hamann, Kehr & González (2012).

\****Rauschiella robusta*** (Brooks, 1976) Razo-Mendivil, León-Règagnon & Ponce de León, 2006 (Fig. 55)

**COLOMBIA:** *Rhinella marina* (= *Bufo marinus*) [Brooks, 1976b].

Site: Intestine.

B: 1.96-2.30 mm x 0.66-0.74 mm; OS: 264 x 288-300; VS: 156 x 145; E: 23-26 x 12 (after Brooks, 1976b).

\*Remarks: Referred as *Glyphelminis robustus* by Brooks (1976b).

## FAMILY PLAGIORCHIIDAE Lühe, 1901

### CHOLEDOCYSTUS Pereira & Cuocolo, 1941

\****Choledocystus hepaticus*** (Lutz, 1928) Sullivan, 1977 (Fig. 56)

**VENEZUELA:** *Hypsiboas crepitans* (= *Hyla crepitans*) [Sullivan, 1977b]; *Incilius nebulifer* (= *Bufo granulosus*) [Nasir & Diaz, 1970]; *Rhinella marina* (= *Bufo marinus*) [Lutz, 1928; Caballero & Diaz-Ungria, 1958; Nasir & Diaz, 1970; Sullivan, 1977b].

Sites: Intestine, liver, biliary ducts, gall bladder.

B: 1.11-6.21 mm x 0.38-2.47 mm; OS: 120-380 x 130-370; VS: 130-440 x 140-460; E: 17-24 x 8-14 (after Sullivan, 1977b).

\*Remarks: Referred as *Plagiorchis hepaticus* by Lutz (1928); as *Choledocystus intermedius* by Lutz (1928) and Caballero & Diaz-Ungria (1958); as *Glyphelmins hepaticus* and as *Glyphelmins linguatula* by Nasir & Diaz (1970).

\**Choledocystus incurvatum* (Nasir, 1966) Sullivan, 1976 (Fig. 57)

**VENEZUELA:** *Pseudis paradoxa* [Nasir, 1966].

Sites: Intestine, lungs.

B: 1.73-2.20 mm x 0.54-0.61; OS: 193-206 in diameter; VS: 131-165 in diameter; E: 25-33 x 14-16 (after Nasir, 1966).

\*Remarks: Referred as *Glyphelmins incurvatum* and as *Glyphelmins ramitesticularis* by Nasir (1966) which was considered synonym of *Choledocystus incurvatum* by Sullivan (1976).

\**Choledocystus pseudium* (Mañé-Garzón & Holcman-Spector, 1967) Sullivan, 1976 (Fig. 58)

**URUGUAY:** *Pseudis minuta* (= *Pseudis mantidactylus*) [Mañé-Garzón & Holcman-Spector, 1967a; 1974].

Site: Intestine.

B: 2.30-3.90 mm x 0.90-1.64 mm; OS: 300-450 x 280-360; VS: 200-360 in diameter; E: 35-41 x 14 (after Mañé-Garzón & Holcman-Spector, 1967a).

\*Remarks: Referred as *Margeana pseudium* by Mañé-Garzón & Holcman-Spector (1967a) and as *Glyphelmins pseudium* by Travassos, Freitas & Kohn (1969).

\**Choledocystus simulans* (Freitas, 1941) Razo-Mendivil, León-Règagnon & Ponce de León, 2006 (Fig. 59)

**URUGUAY:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Freitas, 1941a; Mañé-Garzón & Holcman-Spector, 1974].

Sites: Intestine, stomach, gall bladder.

B: 2.51 mm x 0.97 mm; OS: 250 in diameter; VS: 220 in diameter; E: 29 x 13 (after Freitas, 1941a).

\*Remarks: Referred as *Glyphelmins simulans* by Freitas (1941a).

\**Choledocystus vitellinophilum* (Dobbin Jr., 1958) Sullivan, 1973 (Fig. 60)

**ARGENTINA:** *Lysapsus limellum* (= *Lysapsus limellus*) [Hamann & Kehr, 1997; 1999; Kehr, Manly & Hamann, 2000; Hamann, 2006].

**BRAZIL:** *Hypsiboas raniceps* (= *Hyla raniceps*) [Dobbin Jr., 1958]; *Lisapsus limellum* [Freitas, 1960b; Travassos & Freitas, 1964].

Site: Intestine.

B: 2.48-5.12 mm x 0.96-2.51 mm; OS: 270-460 x 240-450; VS: 180-360 x 190-330; E: 30-31 x 16-18 (after Dobbin Jr., 1958).

\*Remarks: Referred as *Glyphelmims vitellinophilum* by Dobbin Jr. (1958), Freitas (1960b), Travassos & Freitas (1964), Hamann & Kehr (1997, 1999), Kehr, Manly & Hamann (2000) and Hamann (2006).

### **PLAGIORCHIS** Lühe, 1899

*Plagiorchis rangeli* Artigas & Zerpa, 1961 (Fig. 61)

**BRAZIL:** *Leptodactylus latrans* (= *Leptodactylus ocellatus*) [Artigas & Zerpa, 1961].

Site: Intestine.

B: 2.33 mm x 0.80 mm; OS: 180 in diameter; VS: mng; E: 35 x 14 (after Artigas & Zerpa, 1961).

### **RUDOLPHITREMA** Travassos, 1926

*Rudolphitrema chilensis* Puga, 1986 (Fig. 62)

**CHILE:** *Alsodes roseus* (= *Eusophus roseus*) [Puga, 1986; 1994; Puga & Torres, 1999; Olmos & Muñoz, 2006].

Site: Intestine.

B: 2.03-4.06 mm x 0.63-1.20 mm; OS: 130-300 x 200-280; VS: 170-320 x 180-310; E: 31-47 x 20-27 (after Puga, 1986).

*Rudolphitrema physalaemi* Mañé-Garzón & Ponce de León, 1976 (Fig. 63)

**URUGUAY:** *Physalaemus gracilis* [Mañé-Garzón & Ponce de León, 1976].

Site: Intestine.

B: 1.03-1.37 mm x 0.51-0.97 mm; OS: 145-185 in diameter; VS: 108-126 x 114-153; E: 43-50 x 28-34 (after Mañé-Garzón & Ponce de León, 1976).

\**Rudolphitrema rudolphii* (Travassos, 1924) Travassos, 1926 (Fig. 64)

**BRAZIL:** *Rhinella crucifer* (= *Bufo crucifer*) [Travassos, 1924a; 1926a; 1930]; *Rhinella icterica* [Santos, Amato & Borges-Martins, 2013].

**PERU:** *Atelopus ignescens* (= *Atelopus laevis*) [Ibañez, 1980].

Site: Intestine.

B: 2.00-2.20 mm x 1.00 mm; OS: 200 in diameter; VS: 140 in diameter; E: 42-45 x 28 (after Travassos, 1924a).

\*Remarks: Referred as *Rudolphiella rudolphi* by Travassos (1924a, 1930).

### **TRAVTREMA** Pereira, 1929

\**Travtrema stenocotyle* (Cohn, 1902) Goodman, 1951

**ARGENTINA:** *Scinax nasicus* [Hamann, Kehr & González, 2010, wmd].

**BRAZIL:** *Leptodactylus podicipinus* [Campião, Silva & Ferreira, 2009, wmd].

Site: Intestine.

\*Remarks: Referred as *Travtrema* aff. *stenocotyle* by Hamann, Kehr & González (2010).

## FAMILY TELORCHIIDAE Looss, 1899

### OPISTHIOGLYPHE Looss, 1899

\**Opisthioglyphe amplicavus* (Travassos, 1924) Travassos, 1930 (Fig. 65)

**BRAZIL:** *Hylodes nasus* (= *Elosia nasus*) [Travassos, 1924a; 1930].

Site: Intestine.

B: 2.00-2.25 mm x 0.90-1.00mm; OS: 230-250 in diameter; VS: 410-450 in diameter; E: 60-64 x 32-40 (after Travassos, 1930).

\*Remarks: Referred as *Dolichosaccus amplicava* by Travassos (1924a).

## SPECIES REFERRED BY MISTAKE IN SOUTH AMERICA

*Cephalogonimus americanus* Stafford, 1902

This species was redescribed by Lent & Freitas (1941) from *Rana pipiens* from North America and by mistake it was referred as from Brazil by Yamaguti (1971).

*Haematolechus travdarribus* (Skrjabin & Antipin, 1962) Yamaguti, 1971

This species was described by Travassos and Darriba (1930) as *Pneumonoeces schulzei* Wundsch, 1911 from *Rana* sp. from the Zoological Museum of Hamburg; Yamaguti (1971) considered it synonym of *Haematolechus travdarribus* and by mistake referred it as from Brazil.

# REPTILIA

## CLASS TREMATODA

### SUBCLASS ASPIDOGASTREA Faust & Tang, 1936

#### FAMILY ASPIDOGASTRIDAE Poche, 1907

##### *LOPHOTASPIS* Looss, 1901

*Lophotaspis vallei* (Stossich, 1899) Looss, 1901 (Fig. 66)

BRAZIL: *Caretta caretta* (= *Thalassochelys caretta*) [Araujo, 1941].

Sites: Oesophagus, stomach.

B: 10 mm x 1.81 mm; VD: 7 mm x 1.60 mm with 77 alveoli; E: 13 mm x 0.05 mm (after Araujo, 1941).

### SUB CLASS DIGENEA Carus, 1863

#### SUPERFAMILY ALLOCREADIOIDEA Looss, 1902

#### FAMILY ALLOCREADIIDAE Looss, 1902

##### *LEUROSOMA* Ozaki, 1932

*Leurosoma rudolfbarthi* Kohn & Fernandes, 1976 (Fig. 67)

BRAZIL: *Chironius fuscus* [Kohn & Fernandes, 1976].

Site: Ureter.

B: 4.19 mm x 0.92 mm; OS: 210 x 220; VS: 150 x 140; E: 40-50 x 30-34 (after Kohn & Fernandes, 1976).

#### SUPERFAMILY CLINOSTOMOIDEA Lühe, 1901

#### FAMILY CLINOSTOMIDAE Lühe, 1901

##### *ODHNERIOTREMA* Travassos, 1928

\**Odhneriotrema microcephala* (Travassos, 1922) Travassos, 1928 (Fig. 68)

BRAZIL: *Caiman crocodilus* (*Caiman sclerops*) [Travassos, 1922a; 1928a].

Site: Oesophagus.

B: 12.00-25.00 mm x 3.00-5.00 mm; OS: 700-1,000 in diameter; VS: 1,800 in diameter; E: 134 x 63 (after Travassos, 1928).

\*Remarks: Referred as *Nephrocephalus microcephalus* by Travassos (1922a).

## FAMILY LIOLOPIDAE Odhner, 1912

### *HELICOTREMA* Odhner, 1912

\**Helicotrema asymmetricum* (Travassos, 1922) Viana, 1924 (Fig. 69)

**BRAZIL:** *Iguana iguana* (= *Iguana tuberculata*) [Travassos, 1922a; 1928a].

Site: Intestine.

B: 28.00 mm x 1.00-1.50 mm; OS: 260 in diameter; VS: 300 in diameter; E: 163 x 92 (after Travassos, 1928).

\*Remarks: Referred as *Helicometra asymmetrica* by Travassos (1922a).

*Helicotrema magniovatum* Odhner, 1912 (Fig. 70)

**BRAZIL:** *Iguana iguana* [Odhner, 1912; Ávila & Silva, 2013].

Site: Intestine.

B: 20 mm x mng; OS: 230-270 in diameter; VS: 260-300 in diameter; E: 155-167 x 90 (after Odhner, 1912).

\**Helicotrema spirale* (Diesing, 1850) Odhner, 1912 (Fig. 71)

**BRAZIL:** *Chelonoidis denticulata* (= *Geochelone denticulata*; *Testudo tabulata*); *Iguana iguana* (= *Hypstophus tuberculatus*; *Iguana tuberculata*); *Peltocephalus dumerilianus* (= *Podocnemis dumeriliana*; *Podocnemis tracaxa*) [Odhner, 1912].

Site: Intestine.

B: 21.00-25.00 mm long; OS: 54 in diameter; VS: 114-137 in diameter; E: 114 x 73 (after Odhner, 1912).

\*Remarks: Referred as *Monostomum spirale* by Diesing (1850).

## SUPERFAMILY DIPLOSTOMOIDEA Poirier, 1886

### FAMILY CYATHOCOTYLIDAE Poche, 1926

#### *CYATHOCOTYLE* Mühling, 1896

*Cyathocotyle brasiliensis* Ruiz & Leão, 1943 (Fig. 72)

**BRAZIL:** *Caiman crocodilus* (= *Caiman sclerops*) [Ruiz & Leão, 1943a]; *Caiman yacare* (= *Caiman crocodilus yacare*) [Catto & Amato, 1994a].

Site: Intestine.

B: 2.40-2.47 mm x 1.76-1.83 mm; OS: 226-232 in diameter; VS: 169-240 in diameter; E: 117-137 x 67-70 (after Ruiz & Leão, 1943).

## FAMILY PROTERODIPLOSTOMIDAE Dubois, 1936

### CHELONIODIPLOSTOMUM Sudarikov, 1960

\**Cheloniodiplostomum testudinis* (Dubois, 1936) Sudarikov, 1960 (Fig. 73)

**ARGENTINA:** *Phrynops hilarii* [Lombardero & Moriena, 1977].

**BRAZIL:** *Testudo* sp. [Dubois, 1936; 1938]

Site: Intestine.

B: 1.84-2.07 mm x 0.52-0.66; OS: 53-62 x 48-75; VS: mng; E: no eggs (after Dubois, 1938)

\*Remarks: Referred as *Herpetodiplostomum testudinis* by Dubois (1936; 1938) and by Ruiz & Rangel (1954).

### CROCODILICOLA Poche, 1926

*Crocodilicola pseudostoma* (Willemoes-Suhm, 1870) Poche, 1926 (Fig. 74)

**BRAZIL:** *Caiman* sp. (= *Crocodilus* sp.) [Viana, 1924; Dubois, 1938].

Site: Intestine.

B: 2.91-3.50 mm x 0.54-0.70 mm; OS: 310-570 x 310-550; VS: 50-101 x 58-117; E: 85-104 x 48-62 (after Dubois, 1938).

### CYSTODIPLOSTOMUM Dubois, 1936

*Cystodiplostomum hollyi* Dubois, 1936 (Fig. 75)

**BRAZIL:** *Caiman crocodilus* (= *Caiman sclerops*) [Dubois, 1936; 1938; Ruiz & Rangel, 1954]; *Caiman latirostris* [Dubois, 1948]; *Caiman yacare* (= *Caiman crocodilus yacare*) [Catto & Amato, 1994b].

**VENEZUELA:** *Caiman crocodilus* [Nasir & Diaz, 1971b].

Site: Intestine.

B: 1.05-1.84 mm x 0.60-0.97 mm (anterior portion); 0.95-2.58 mm x 0.53-0.66 mm (posterior portion); OS: 50-90 in diameter; VS: 160-228 in diameter; E: 62-104 x 43-62 (after Ruiz & Rangel, 1954).

### HERPETODIPLOSTOMUM Dubois, 1936

\**Herpetodiplostomum caimancola* (Dollfus, 1935) Dubois, 1936 (Fig. 76)

**BRAZIL:** *Caiman crocodilus* [Dubois, 1936; 1938]; *Caiman latirostris* [Dollfus, 1935]; *Caiman yacare* (= *Caiman crocodilus yacare*) [Catto & Amato, 1994b]; *Melanosuchus niger* [Dubois, 1936].

**VENEZUELA:** *Caiman crocodilus* [Nasir & Diaz, 1971b].

Site: Intestine.

B: 1.16-2.67 mm x mng; OS: 50-80 x 32-65; VS: 80-142 x 80-144; E: 84-130 x 53-88 (after Dubois, 1938).

B: 1.76-2.82 mm x 0.51-0.83 mm; OS: 36-66 in diameter; VS: 90-177 in diameter; E: 96-120 x 63-96 (after Nasir & Diaz, 1971b).

\*Remarks: Referred as *Crocodicicola caimancola* by Dollfus (1935) and as *Prohemistomum babai* by Nasir & Diaz (1971). These species were considered synonym of *Herpetodiplostomum caimancola* by Dubois (1979).

## **HETERODIPLOSTOMUM Dubois, 1936**

*Heterodiplostomum helicopsis* Mañé-Garzón & Alonso, 1976 (Fig. 77)

**URUGUAY:** *Helicops infrataeniatus* [Mañé-Garzón & Alonso, 1976].

Site: Intestine.

B: 3.94-4.64 mm x 0.52-0.73 mm; 2.06-2.92 mm (anterior portion); 1.73-2.04 mm (posterior portion); OS: 36-57 in diameter; VS: 112-176 in diameter; E: 128 x 42 (after Mañé-Garzón & Alonso, 1976).

*Heterodiplostomum lanceolatum* Dubois, 1936 (Fig. 78)

**ARGENTINA:** *Bothrops alternata* (= *Bothrops alternatus*) [Poumarau, 1968; Lunaschi & Sutton, 1985]; *Helicops infrataeniatus* [Lunaschi & Sutton, 1985]; *Helicops leopardinus* (= *Helicops leopardina*); *Hydrodynastes gigas* (= *Cyclagras gigas*) [Poumarau, 1968; Lunaschi & Sutton, 1985; Lunaschi & Drago, 2010].

**BRAZIL:** *Coluber* sp. [Dubois, 1936; 1953; 1988]; *Mastigodryas bifossatus* [Dubois, 1988]; *Xenodon guentheri* [Ruiz & Rangel, 1954].

**PARAGUAY:** *Hydrodynastes gigas*; *Liophis poecilogyrus reticulatus* [Dubois, 1988]; *Mastigodryas bifossatus* [Dubois, 1986].

Site: Intestine.

B: 3.97-4.66 mm x 1.34-1.42 mm (anterior portion); 4.08-5.05 mm x 0.79-0.92 mm (posterior portion); OS: 80-111 in diameter; VS: 296-320 in diameter; E: 135-178 x 74-104 (after Ruiz & Rangel, 1954).

## **MASSOPROSTATUM Caballero, 1948**

*Massoprostatum longum* Caballero, 1948 (Fig. 79)

**COLOMBIA:** *Caiman crocodilus fuscus* [Álvarez, Lenis & Vélez, 2005].

Site: Intestine.

B: 1.40-2.06 mm x mng; OS: 39-49 in diameter; VS: 23-55 x 39-61; E: 87-100 x 59-79 (after Álvarez, Lenis & Vélez, 2005).



## MESODIPLOSTOMUM Dubois, 1936

*Mesodiplostomum gladiolum* Dubois, 1936 (Fig. 80)

**BRAZIL:** *Caiman* sp.; *Melanosuchus niger* [Dubois, 1938; 1953].

Site: Intestine.

B: 2.60-4.00 mm x 0.27-0.50 mm; OS: 31-48 x 35-55; VS: 76-105 x 76-120; E: 72-97 x 43-58 (after Dubois, 1938).

## OPHIODIPLOSTOMUM Dubois, 1936

Syn: *Petalodiplostomum* Dubois, 1936

\**Ophioidiplostomum ancyloides* (Dubois, 1936) Dubois, 1979 (Fig. 81)

**BRAZIL:** *Coluber* sp. [Dubois, 1936; 1938; Ruiz & Rangel, 1954].

Site: Intestine.

B: 3.30-4.14 mm x 0.90-0.96 mm; OS: 40-60 x 50-70; VS: 165-180 x 200-215; E: 108-126 x 60-75 (after Dubois, 1938).

\*Remarks: Referred as *Petalodiplostomum ancyloides* by Dubois (1936; 1938). The genus *Petalodiplostomum* Dubois, 1936 was considered synonym of *Ophioidiplostomum* by Dubois (1979) and accepted by Niewiadomska (2002).

*Ophioidiplostomum spectabile* Dubois, 1936 (Fig. 82)

**ARGENTINA:** *Philodryas* sp. [Lunaschi & Drago, 2010].

**BRAZIL:** *Coluber* sp. [after Travassos, Freitas & Kohn, 1969]; *Crotalus durissus terrificus*; *Eunectes deschauenseei* [Pinto, Mati & Melo, 2012]; *Liophis poecilogyrus* (= *Leimadophis poecilogyrus*) [Ruiz & Rangel, 1954; Pinto, Mati & Melo, 2012]; *Liophis miliaris* (= *Liophis miliaris miliaris*) [Ruiz & Rangel, 1954; Pinto, Mati & Melo, 2012]; *Liophis reginae*; *Liophis typhlus* [Pinto, Mati & Melo, 2012]; *Mastigodryas bifossatus* (= *Dryadophis bifossatus*, *Drymobius bifossatus*) [Dubois, 1936; 1938; Pinto, Mati & Melo, 2012]; *Xenodon merremi* [Noronha, Sá, Knoff, Muniz-Pereira & Pinto, 2009].

Site: Intestine.

B: 1.05-2.11 mm 0.79-1.55 mm (anterior portion); 1.45-2.82 mm x 0.58-1.18 mm (posterior portion); OS: 37-133 in diameter; VS: 61-123 in diameter; E: 86-154 x 55-123 (after Ruiz & Rangel, 1954).

\*Remarks: Referred as *Petalodiplostomum aristoterisi* by Ruiz & Rangel (1954).

## PARADIPLOSTOMUM La Rue, 1926

\**Paradiplostomum abbreviatum* (Brandes, 1888) La Rue, 1926 (Fig. 83)

**ARGENTINA:** *Caiman latirostris* [Lunaschi & Sutton, 1990].

**BRAZIL:** *Caiman crocodilus* (= *Caiman sclerops*, *Crocodylus* sp.) [Brandes, 1888; Dubois, 1936; 1938]; *Caiman yacare* (= *Caiman crocodilus yacare*) [Catto & Amato, 1994b].

Site: Intestine.

B: 1.06-1.27 mm x 0.41-0.50 mm; OS: 48-57 x 57-70; VS: 65-89 x 75-94; E: 91-108 x 55-65 (after Dubois, 1938).

\*Remarks: Referred as *Diplostomum abbreviatum* by Viana (1924).

### **PROLECITHODIPILOSTOMUM Dubois, 1936**

\**Prolecithodiplostomum constrictum* Dubois, 1936 (Fig. 84)

**BRAZIL:** *Caiman crocodilus* (= *Caiman sclerops*) [Dubois, 1936; 1938; 1953]; *Caiman* sp. [Ruiz & Rangel, 1954]; *Caiman yacare* (= *Caiman crocodilus yacare*) [Catto & Amato, 1994b].

**COLOMBIA:** *Caiman crocodilus fuscus* [Álvarez, Lenis & Vélez, 2005].

Site: Intestine.

B: 1.41 mm x 0.84-0.90 mm (anterior portion); 1.47-1.62 mm x 0.43-0.63 (posterior portion); OS: 43 x 67; VS: 380-422 x 309-352; E: 90-99 x 55-62 (after Dubois, 1938).

\*Remarks: Referred as *Prolecithodiplostomum cavum* by Dubois (1936) (after Dubois, 1979), Ruiz & Rangel (1954) and by Álvarez, Lenis & Vélez (2005).

### **PROTERODIPILOSTOMUM Dubois, 1936**

*Proterodiplostomum breve* Catto & Amato, 1994 (Fig. 85)

**BRAZIL:** *Caiman yacare* (= *Caiman crocodilus yacare*) [Catto & Amato, 1994b].

Site: Intestine.

B: 2.69-4.22 mm in total length x 366-658 mm at posterior part of body; OS: 102-146 x 109-175; VS: 109-164 in diameter; E: 80-102 x 44-65 (after Catto & Amato, 1994b).

*Proterodiplostomum globulare* Catto & Amato, 1994 (Fig. 86)

**BRAZIL:** *Caiman yacare* (= *Caiman crocodilus yacare*) [Catto & Amato, 1994b].

Site: Intestine.

B: 2.56-4.75 mm in total length x 237-713 mm at posterior part of body; OS: 51-73 x 43-73; VS: 73-109 x 80-124; E: 98-131 x 44-66 (after Catto & Amato, 1994b).

\**Proterodiplostomum longum* (Brandes, 1888) Dubois, 1936 (Fig. 87)

**BRAZIL:** *Caiman* sp. (= *Crocodylus* sp.) [Brandes, 1888]; *Melanosuchus niger*; *Paleosuchus* sp. (= *Crocodylus coroa*) [Dubois, 1938; 1953; Ruiz & Rangel, 1954].

**PARAGUAY:** *Caiman yacare* (= *Caiman crocodilus yacare*) [Dubois, 1988].

**VENEZUELA:** *Caiman crocodilus* [Diaz & Nasir, 1969; Nasir & Diaz, 1971b; Diaz-Ungria, 1973].

Site: Intestine.

B: 3.75-9.30 mm x 0.12-0.72 mm; OS: 55-96 x 53-95; VS: 96-144 x 100-160; E: 77-96 x 48-65 (after Dubois, 1938).

\*Remarks: Referred as *Diplostomum longum* Brandes, 1888 by Viana (1924).

\**Proterodiplostomum medusae* (Dubois, 1936) Caballero, Hidalgo & Grocott, 1937 (Fig. 88)

**BRAZIL:** *Caiman crocodilus* (= *Caiman sclerops*) [Dubois, 1936; 1938]; *Caiman* sp. [Ruiz & Rangel, 1954]; *Caiman yacare* (= *Caiman crocodilus yacare*) [Catto & Amato, 1994b].

**VENEZUELA:** *Caiman crocodilus crocodilus* [Nasir & Rodrigues, 1967; Diaz-Ungria, 1973].  
Site: Intestine.

B: 0.84-0.98 mm x 0.56-0.59 mm (anterior portion); 1.55-2.18 mm x 0.42 mm (posterior portion); OS: 61-64 in diameter; VS: 61-74 x 92; E: 111-123 x 67-80 (after Ruiz & Rangel, 1954).

B: 1.68-2.34 mm in length; OS: 34-50 x 53-70; VS: 62-71 x 68-82; E: 96-115 x 57-65 (after Dubois, 1938).

\*Remarks: Referred as *Pseudoneodiplostomum brasiliensis* by Ruiz & Rangel (1954); as *Proterodiplostomum intermedium* by Nasir & Rodrigues (1967) and by Diaz-Ungria (1973) and as *Proterodiplostomum brasiliensis* (Ruiz & Rangel, 1954) Sudarikov, 1960 by Travassos, Freitas & Kohn (1969).

*Proterodiplostomum tumidulum* Dubois, 1936 (Fig. 89)

**BRAZIL:** *Caiman crocodilus* (= *Caiman sclerops*) [Dubois, 1936; 1938]; *Caiman yacare* (= *Caiman crocodilus yacare*) [Catto & Amato, 1994b].

Site: Intestine.

B: 1.80-2.50 mm (total length) x 0.37-0.54 mm (anterior portion); OS: 62-86 x 62-85; VS: 62-77 x 65-89; E: 86-96 x 55-64 (after Dubois, 1938).

## SUPERFAMILY ECHINOSTOMATOIDEA Looss, 1899

### FAMILY ECHINOSTOMATIDAE Looss, 1899

#### CABALLEROTREMA Prudhoe, 1960

*Caballerotrema* sp. (Fig. 90)

**BRAZIL:** *Melanosuchus niger* [Ostrowski de Núñez, 2003].

Site: Intestine (probably).

B: 4.10-6.80 mm x 0.42-0.66 mm; OS: 132-182 in diameter; HC: with 29 spines; VS: 295-408 x 245-364; E: 60-82 x 41-57 (after Ostrowski de Núñez, 2003).

#### ECHINOSTOMA Rudolphi, 1809

*Echinostoma* sp. (Fig. 91)

**BRAZIL:** *Melanosuchus niger* [Ostrowski de Núñez, 2003].

Site: Intestine (probably).

B: 6.50-14.30 mm x 0.45-1.33 mm; OS: 176-283 x 157-283; HC: with 47 spines; VS: 438-1,280 x 320-880; E: 88-113 x 54-63 (after Ostrowski de Núñez, 2003).

## PRIONOSOMA Dietz, 1909

*Prionosoma phrynopsis* Mañé-Garzón & Gil, 1961 (Fig. 92)

**URUGUAY:** *Phrynops hillarii* (= *Phrynops geoffroyana hillarii*) [Mañé-Garzón & Gil, 1961a].  
Site: Intestine.

B: 8.09-14.86 mm x 1.19-1.20 mm; OS: 180-240 in diameter with 47 spines; VS: 530-590 in diameter; E: 72-93 x 48-55 (after Mañé-Garzón & Gil, 1961a).

## PRIONOSOMOIDES Freitas & Dobbin Jr., 1967

*Prionosomoides scalaris* Freitas & Dobbin Jr., 1967 (Fig. 93)

**ARGENTINA:** *Phrynops hilarii* [Lombardero & Moriena, 1977].

**BRAZIL:** *Phrynops geoffroanus* (= *Phrynops geoffroanus geoffroanus*) [Freitas & Dobbin Jr., 1967].

Site: Intestine.

B: 13.33-19.32 mm x 1.01-1.60 mm; PD: with 46 spines; OS: 230-280 x 280-330; VS: 670-930 in diameter; E: 133-153 x 67-107 (after Freitas & Dobbin Jr., 1967).

## PULCHROSOMOIDES Freitas & Lent, 1937

*Pulchrosomoides elegans* Freitas & Lent, 1937 (Fig. 94)

**BRAZIL:** *Iguana iguana* (= *Iguana tuberculata*) [Freitas & Lent, 1937; Travassos, 1951]; *Mabuya macrorhyncha* [Vrcibradic, Rocha, Bursey & Vicente, 2002]; *Tupinambis teguixin* [Ávila & Silva, 2010].

Site: Stomach.

B: 11.40-13.50 mm x 2.63-3.40 mm; OS: 789-1,050 x 973-1,184; VS: 1,525-2,262; E: 136-157 x 72-86 (after Freitas & Lent, 1937).

## STEPHANOPRORA Odhner, 1902

*Stephanoprora campomica* Nasir & Diaz, 1971 (Fig. 95)

**VENEZUELA:** *Caiman crocodilus* [Nasir & Diaz, 1971b].

Site: Intestine.

B: 4.64-5.76 mm x 0.38-0.45 mm; OS: 84-120 in diameter; HC: with 22 spines; VS: 188-309 in diameter; E: 60-87 x 40-59 (after Nasir & Diaz, 1971b).

\**Stephanoprora jacaretinga* (Freitas & Lent, 1938) Yamaguti, 1958 (Fig. 96)

**BRAZIL:** *Caiman crocodilus* (= *Caiman sclerops*) [Freitas & Lent, 1938a]; *Caiman yacare* (= *Caiman crocodilus yacare*) [Catto & Amato, 1994a].

Site: Intestine.

B: 3.02-5.60 mm x 0.58-0.68 mm; OS: 128-144 x 120-144; HC: with 22 spines; VS: 357-368 x 386-429; E: 89-94 x 57-65 (after Freitas & Lent, 1938a).

\*Remarks: Referred as *Echinostoma jacaretinga* by Freitas & Lent (1938a). According to Ostrowski de Núñez (2003) the specimens studied by Catto & Amato (1994a) can not belong to *S. jacaretinga*, and this material should be restudied.

***Stephanoprora nattereri*** Ostrowski de Núñez, 2003 (Fig. 97)

**BRAZIL:** *Melanosuchus niger* [Ostrowski de Núñez, 2003].

Site: Intestine (probably).

B: 8.80-13.00 mm x 0.38-0.58 mm; OS: 126-176 x 157-220; HC: with 22 spines; VS: 480-768 x 448-720; E: 71-84 x 44-59 (after Ostrowski de Núñez, 2003).

***Stephanoprora* sp.** (Fig. 98)

**BRAZIL:** *Caiman crocodilus* [Ostrowski de Núñez, 2003].

Site: Intestine (probably).

B: 2.99-3.31 mm x 0.29-0.43 mm; OS: 79-107 x 95-117; HC: with 22 spines and 2 corner spines; VS: 167-227 x 195-252; E: 60-76 x 44-50 (after Ostrowski de Núñez, 2003).

## FAMILY PSILOSTOMIDAE Looss, 1900

### \**COTYLOTRETUS* Odhner, 1902

***Cotylotretus rugosus*** Odhner, 1902 (Fig. 99)

**BRAZIL:** *Spilotes pullatus* (= *Coluber pullatus*) [Odhner, 1902].

Site: Intestine.

B: 9.50-19.00 mm x 1.75-4.00 mm; PD: without spines; OS: 310-390 in diameter; VS: 1,600 x 1,45-2,350; E: 120 x 60 (after Odhner, 1902).

\*Remarks: The genus *Cotylotretus* Odhner, 1902 was considered as a *genus inquirenda* by Kostadinova (2005).

### *SPHAERIDIOTREMA* Odhner, 1913

***Sphaeridiotrema echinosauense*** O'Brien, Sidner & Etges, 1979 (Fig. 100)

**ECUADOR:** *Echinosaura horrida horrida* [O'Brien, Sidner & Etges, 1979].

Site: Intestine.

B: 930-1,180 x 570-750; OS: 112-203 x 130-229; VS: 280-350 x 240-430; E: 40-70 x 26-36 (after O'Brien, Sidner & Etges, 1979).

**FAMILY RHYTIDODIDAE Odhner, 1926**

**RHYTIDODES Looss, 1901**

\**Rhytidodes gelatinosus* (Rudolphi, 1819) Looss, 1901 (Fig. 101)

**BRAZIL:** *Caretta caretta* (= *Thalassochelis caretta*) [Diesing, 1850]; *Podocnemis expansa* [Rudolphi, 1819].

Site: Intestine.

B: 12.50-13.20 mm x 3.00-3.50 mm; OS: 580 x 480; VS: 450 x 412; E: 62 x 36 (after Braun, 1901).

\*Remarks: Referred as *Distoma gelatinosum* by Rudolphi (1819).

**SUPERFAMILY GORGODEROIDEA Looss, 1899**

**FAMILY BRACHYCOELIIDAE Looss, 1899**

**BRACHYCOELIUM (Dujardin, 1845) Stiles & Hassall, 1898**

\**Brachycoelium salamandrae* (Frolich, 1789) Lühe, 1909 (Fig. 102)

**BRAZIL:** *Anolis scypheus* (= *Anolis nitens*) [Freitas, 1961]; *Leposoma osvaldoi* [Goldberg, Bursey, Caldwell, Vitt & Costa, 2007].

Site: Intestine.

B: 1.10-1.36 mm x 0.87-0.92 mm; OS: 180-200 x 220; VS: 120-140 x 140-150; E: 47-55 x 31-34 (after Freitas, 1961).

\*Remarks: Referred as *Brachycoelium mesocoeliiformis* by Freitas (1961).

**FAMILY BRAUNOTREMATIDAE Yamaguti, 1958**

**BRAUNOTREMA Price, 1930**

\**Braunotrema pulvinatum* (Braun, 1899) Price, 1930 (Fig. 103)

**BRAZIL:** *Podocnemis expansa* [Braun, 1899; Lent & Freitas, 1938].

Site: Intestine.

B: 2.81-4.47 mm x 0.81-1.26 mm; OS: 543-601 x 486-572; VS: 443-543 x 515-715; E: 40-52 x 28-32 (after Lent & Freitas, 1938).

\*Remarks: Referred as *Distomum pulvinatum* by Braun (1899; 1901) and as *Thaumatocotyle pulvinatum* by Viana (1924).

**FAMILY DICROCOELIIDAE Odhner, 1910**

**INFIDUM Travassos, 1916**

\**Infidum infidum* (Faria, 1910) Travassos, 1916 (Fig. 104)

**ARGENTINA:** *Bothropoides diporus* (= *Bothrops neuwiedi diporus*) [Martínez, Troiano, Binda, Selles, Jara & Fescina, 1996]; *Eunectes notaeus*; *Hydrodynastes gigas* (= *Cyclagras gigas*) [Poumarau, 1968; Martínez, Troiano, Binda, Selles, Jara & Fescina, 1996]; *Philodryas psammophidea* (= *Philodryas psammophideus*) [Poumarau, 1968].

**BOLIVIA:** *Hydrodynastes gigas* (= *Cyclagras gigas*) [Travassos, 1944b].

**BRAZIL:** *Bothrops moojeni* [Barrella & Silva, 2003]; *Eunectes murinus* [Faria, 1910; Travassos, Pinto & Muniz, 1928; Travassos, 1944b]; *Hydrodynastes gigas* (= *Cyclagras gigas*) [Faria, 1910; Travassos, 1944b].

Site: Gall bladder.

B: 2.80-5.10 mm x 1.50-2.60 mm; OS: 366-550 in diameter; VS: 351-612 in diameter; E: 26-34 x 13-16 (after Travassos, 1944b).

\*Remarks: Referred as *Dicrocoelium infidum* by Faria (1910).

*Infidum luckeri* McIntosh, 1939 (Fig. 105)

**GALAPAGOS ISLANDS:** *Philodryas hoodensis* (= *Leimadophis chamissonis*, *Oreophis (Driomicus) hoodensis*) [McIntosh, 1939].

Site: Gall bladder.

B: 3.31 mm x 1.63 mm; OS: 410 in diameter; VS: 410 x 440; E: 26 x 16 (after Travassos, 1944b).

\**Infidum similis* Travassos, 1916 (Fig. 106)

**ARGENTINA:** *Philodryas olfersii* [Poumarau, 1968]; *Philodryas* sp. [Lunaschi & Drago, 2010].

**BRAZIL:** *Bothropoides jararaca* (= *Bothrops jararaca*) [Travassos, 1944b]; *Drymarchon corais* (= *Drymarchon corais corais*) [Travassos, 1944b]; *Liophis miliaris* (= *Liophis miliaris miliaris*) [Travassos, 1944b]; *Liophis poecilogyrus* (= *Leimadophis poecilogyrus*) [Ruiz & Leão, 1943b; Travassos, 1944b]; *Mastigodryas bifossatus* (= *Dryadophis bifossatus*; *Drymobius bifossatus*; *Eudryas bifossatus*) [Travassos, 1916; 1944b; Fábio & Rolas, 1974].

Sites: Gall bladder, bile ducts, liver, pancreas.

B: 2.90-5.90 mm x 1.60-2.70 mm; OS: 320-500 in diameter; VS: 320-750 in diameter; E: 30-34 x 15-17 (after Travassos, 1944b).

\*Remarks: Referred as *Infidum intermedium* by Ruiz & Leão (1943b).

### PARADISTOMUM Kossack, 1910

\**Paradistomum boae* (MacCallum, 1921) Travassos, 1924 (Fig. 107)

**BRAZIL:** *Boa constrictor* (= *Constrictor constrictor*) [MacCallum, 1921; Viana, 1924; Travassos, 1944b].

Site: Oesophagus.

B: 2.00 mm x 1.40 mm; OS and VS: mng; E: 20 in length (after Travassos, 1944b).

\*Remarks: Referred as *Zoogonoides boae* MacCallum, 1921 by Viana (1924).

\**Paradistomum parvissimum* (Travassos, 1918) Travassos, 1919 (Fig. 108)

**BRAZIL:** *Ameiva ameiva* [Rodrigues, 1968]; *Bothropoides jararaca* (= *Bothrops jararaca*) [Travassos, 1944b]; *Chironius carinatus* [Travassos, 1944b]; *Hemidactylus mabouia* [Rodrigues, 1968; Rodrigues, 1970; 1986]; *Iguana iguana* [Ávila & Silva, 2010]; *Kentropyx calcarata* [Ávila & Silva, 2013]; *Liolaemus lutzae* [Rodrigues, 1992]; *Mabuya agilis*; *Mabuya macrorhyncha* [Vrcibradic, Rocha, Bursey & Vicente, 2002]; *Mastigodryas bifossatus* (= *Dryadophis bifossatus*) [Fábio & Rolas, 1974]; *Philodryas patagoniensis* (= *Philodryas schottii*) [Travassos, 1944b]; *Plica plica* [Ávila & Silva, 2013]; *Tropidurus torquatus* (= *Tropidurus torquatus torquatus*) [Travassos, 1918; Travassos, 1944a; Travassos, Freitas, Mendonça & Rodrigues, 1962; Vicente, 1978; Rodrigues, Rodrigues & Faria, 1990]; *Tupinambis teguixin* [Travassos, 1918]; *Uranoscodon superciliosus* [Ávila & Silva, 2010].

Sites: Gall bladder, liver, bile ducts.

B: 1.50- 3.70 mm x 0.50-1.30 mm; OS: 186-321 in diameter; VS: 244-390 in diameter; E: 38-45 x 22-30 (after Travassos, 1944b).

\*Remarks: Referred as *Eurytrema parvum* by Travassos (1918), as *Paradistomum lutzi* by Travassos (1919a) and as *Paradistomum magnum* by Travassos (1919a).

*Paradistomum rabusculum* Kossack, 1910 (Fig. 109)

**BRAZIL:** *Gymnodactylus geckoides* [Travassos, 1944b].

Site: Liver.

B: 1.50-1.70 mm x 0.72-0.76 mm; OS: 290 x 310 in diameter; VS: 270-280 in diameter; E: 49 x 28 (after Travassos, 1944b).

## FAMILY MESOCOELIIDAE Dollfus, 1929

### MESOCOELIUM Odhner, 1910

*Mesocoelium monas* (Rudolphi, 1819) Freitas, 1958 (Fig. 110)

**ARGENTINA:** *Sibynomorphus turgidus*; *Sibynomorphus ventrimaculatus* [Poumarau, 1968]; *Sibynomorphus* sp. [Lunaschi & Drago, 2010]; *Tomodon ocellatus* [Poumarau, 1968; Led & Boero, 1973].

**BRAZIL:** *Amphisbaena ridleyi* [Ramalho, Silva, Schwartz & Péres, 2009]; *Amphisbaena* sp. [Rudolphi, 1819; Freitas, 1963]; *Anolis fuscoauratus* [Ávila & Silva, 2013]; *Cercosaura eigenmanni* (= *Prionodactylus eigenmanni*) [Bursey & Goldberg, 2004]; *Diploglossus lessonae*; *Lepostemon microcephalum*; *Liophis poecilogyrus* (= *Leimadophis poecilogyrus*) [Freitas, 1963]; *Lygodactylus klugei* [Anjos, Bezerra, Passos, Zanchi & Galdino, 2011]; *Plica plica* [Goldberg, Bursey & Vitt, 2009, Ávila & Silva, 2013]; *Thecadactylus solimoensis* [Ávila & Silva, 2013]; *Trachylepis atlantica* (= *Mabuya maculata*) [Freitas, 1963; Goldberg, Bursey & Vitt, 2009; Ramalho, Silva, Schwartz & Péres, 2009]; *Tropidurus torquatus* [Rodrigues, Rodrigues & Faria, 1990]; *Uranoscodon superciliosus* [Bursey, Goldberg & Vitt, 2005; Ávila & Silva, 2013].



**ECUADOR:** *Alopoglossus angulatus* [Goldberg, Bursey & Vitt, 2007].

Site: Intestine.

B: 1.26-2.41 mm x 0.690-1.07 mm; OS: 230-330 x 230-360; VS: 170-280 x 180-300; E: 34-44 x 21-25 (after Freitas, 1963).

*Mesocoelium sibynomorphi* Ruiz & Leão, 1943 (Fig. 111)

**ARGENTINA:** *Uranoscodon superciliosus* [Poumarau, 1968].

**BRAZIL:** *Mastigodryas bifossatus* (= *Dryadophis bifossatus*) [Fortes & Hoffman, 1987/1988]; *Sibynomorphus mikanii* (= *Sibynomorphus mikanii mikanii*) [Ruiz & Leão, 1943].

Site: Intestine.

B: 3.70-4.70 mm x 1.40-1.70 mm; OS: 325-381 in transversal diameter; VS: 4282-339 in diameter; E: 36-39 x 22-26 (after Ruiz & Leão, 1943).

## SUPERFAMILY MICROPHALLOIDEA Ward, 1901

### FAMILY PACHYPSOLIDAE Yamaguti, 1958

#### PACHYPSOLUS Looss, 1901

\**Pachypsolus sclerops* (Travassos, 1922) Travassos, 1928 (Fig. 112)

**BRAZIL:** *Caiman crocodilus* (= *Caiman sclerops*) [Travassos, 1922a; 1928a]; *Caiman yacare* (= *Caiman crocodilus yacare*) [Catto & Amato, 1994a]; *Paleosuchus palpebrosus* [Gomes & Pinto, 1978].

Sites: Cloaca, intestine.

B: 2.00-3.00 mm x 0.80-1.10 mm; OS: 500 in diameter; VS: 600 in diameter; E: 42 x 17 (after Travassos, 1928).

\*Remarks: Referred as *Gastris sclerops* by Travassos (1922a).

### FAMILY PLEUROGENIDAE Looss, 1899

#### ALIPTREMA Ruiz & Leão, 1955

*Aliptrema riberoi* Ruiz & Leão, 1955 (Fig. 113)

**BRAZIL:** *Liophis miliaris* (= *Liophis miliaris miliaris*) [Ruiz & Leão, 1955].

Sites: Buccal cavity, oesophagus.

B: 1.98-2.98 mm x 1.06-1.24 mm; OS: 560-650 in diameter; VS: 320-350 in diameter; E: 27-30 x 13-16 (after Ruiz & Leão, 1955).

**SUPERFAMILY MICROSCAPHIDIOIDEA Looss, 1900**

**FAMILY MICROSCAPHIDIIDAE Looss, 1900**

**NEOCTANGIUM Ruiz, 1943**

*Neoctangium travassosi* Ruiz, 1943 (Fig. 114)

**BRAZIL:** Marine turtle [Ruiz, 1943].

Site: Intestine.

B: 7.98-11.17 mm x 2.12-2.92 mm; OS: 310-420 x 370-550; VS: absent; E: 81-95 x 56-57 (after Ruiz, 1943b).

**NEODEUTEROBARIS Brooks, 1976**

*Neodeuterobaris pritchardae* Brooks, 1976 (Fig. 115)

**COLOMBIA:** *Podocnemis lewyana* [Brooks, 1976a; Lenis & Vélez, 2011].

Site: Stomach.

B: 3.08-4.18 mm x 1.79-2.02 mm; OS: 280-480 x 360 x 480; VS: absent; E: 138-149 x 58-92 (after Brooks, 1976a).

**OCTANGIODES Price, 1937**

*Octangioides tlacotalpensis* Caballero, 1942

**ECUADOR:** *Rhinoclemmys nasuta* [Dyer & Carr, 1990].

Site: Intestine.

B: mng; OS and VS: mng; E: 93-110 x 50-70 (after Dyer & Carr, 1990).

**PODOCNEMITREMA Alho & Vicente, 1964**

*Podocnemitrema papillosus* Alho & Vicente, 1964 (Fig. 116)

**BRAZIL:** *Podocnemis expansa* [Alho & Vicente, 1964].

Site: Stomach.

B: 11.25 mm x 6.25 mm; OS: 1.85 x 2.00 mm; VS: absent; E: 132-140 x 79-82 (after Alho & Vicente, 1964).

**POLYANGIUM Looss, 1902**

*Polyangium linguatula* (Looss, 1899) Looss, 1902 (Fig. 117)

**BRAZIL:** *Chelonia mydas* (= *Chelone mydas*) [Travassos, 1934; Freitas & Lent, 1938b].

Site: Intestine.

B: 6.71-9.34 mm x 1.66-1.97 mm; OS: 170-200 x 200-240; VS: absent; E: 76-84 x 46-51 (after Freitas & Lent, 1938b).

## SUPERFAMILY OPISTHORCHIOIDEA Looss, 1899

### FAMILY CRYPTOGONIMIDAE Ward, 1917

#### ACANTHOSTOMUM Looss, 1899

*Acanthostomum scyphocephalum* (Braun, 1899) Hughes, Higginbotham & Clary, 1954 (Fig. 118)

**BRAZIL:** *Caiman crocodilus* (= *Caiman sclerops*); *Chelus fimbriatus* (= *Chelys fimbriata*, *Testudo matamata*) [Braun, 1901; Ostrowski de Núñez, 1986].

**URUGUAY:** *Phrynops hilarii* (= *Phrynops geoffroyana hillarii*) [Mañé-Garzón & Gil, 1961a].

**VENEZUELA:** *Caiman crocodilus*; *Drymachon corais* [Nasir, 1974].

Site: Intestine.

B: 2.30-3.00 mm long; OS: 200-250 in diameter with 22-30 spines; VS: 80-110 in diameter; E: 20-28 x 11 (after Braun, 1901).

#### CAIMANICOLA Freitas & Lent, 1938

\* *Caimanicola brauni* (Mañé-Garzón & Gil, 1961) Brooks, 1980 (Fig. 119)

**ARGENTINA:** *Phrynops hilarii* [Ostrowski de Núñez, 1987 (experimental)]

**URUGUAY:** *Phrynops hilarii* (= *Phrynops geoffroyana hillarii*) [Mañé-Garzón & Gil, 1961a].

Site: Intestine.

B: 4.13-5.22 mm x 0.45-0.47 mm; OS: 310-390 x 330-430 with 23-24 spines; VS: 144-205 in diameter; E: 21 x 10 (after Mañé-Garzón & Gil, 1961a).

\*Remarks: Referred as *Acanthostomum brauni* by Mañé-Garzón & Gil (1961a) and Ostrowski de Núñez (1987).

\* *Caimanicola marajoara* Freitas & Lent, 1938 (Fig. 120)

**BRAZIL:** *Caiman crocodilus* (= *Caiman sclerops*) [Freitas & Lent, 1938a; Ostrowski de Núñez, 1984a,b]; *Caiman yacare* (= *Caiman crocodilus yacare*) [Catto & Amato, 1993a].

**COLOMBIA:** *Caiman crocodilus* (= *Caiman sclerops*) [Carter & Etges, 1972]; *Paleosuchus* sp. [Ostrowski de Núñez, 1984a,b].

**VENEZUELA:** *Crocodylus intermedius* [Ostrowski de Núñez, 1984a,b (experimental)]

Site: Intestine.

B: 1.18-1.24 mm x 0.50-0.55 mm; OS: 270-280 in diameter, with 20 spines; VS: 100-150 x 150; E: 24 x 11 (after Freitas & Lent, 1938a).

\*Remarks: Referred as *Acanthostomum marajoarum* by Carter & Etges (1972) and by Ostrowski de Núñez (1984a,b) and as *Acanthostomum* (*Acanthostomum*) *scyphocephalum* (Braun, 1899) by Nasir (1974).

## PROCTOCAECUM Baugh, 1957

*Proctocaecum dorsale* Catto & Amato, 1993 (Fig. 121)

**BRAZIL:** *Caiman yacare* (= *Caiman crocodilus yacare*) [Catto & Amato, 1993a].

Site: Intestine.

B: 2.04-6.03 mm x 0.016-0.043 mm; OS: 102-211 x 124-248 surrounded by 23 spines; VS: 94-153 x 94-175; E: 20-25 x 9-13 (after Catto & Amato, 1993a).

## TIMONIELLA Rebecq, 1960

\**Timoniella incognita* Brooks, 1980 (Fig. 122)

**VENEZUELA:** *Caiman crocodilus crocodilus*; *Drymarchon corais* [Nasir, 1974; Brooks, 1980].

Site: Intestine.

B: 2.4 mm in length; OS: 41 x 15 with 23 spines; VS: mng; E: 23 x 12 (after Brooks, 1980).

\*Remarks: Referred as *Acanthostomum (Acanthostomum) scyphocephalum* (Braun, 1899) Hughes, Higginbotham & Clary, 1941 (*in partim*) by Nasir (1974).

\**Timoniella ostrowskiae* Brooks & Holcman, 1993 (Fig. 123)

**URUGUAY:** *Phrynops hillarii* (= *Phrynops geoffroyana hillarii*) [Mañé-Garzón & Gil, 1961a].

**VENEZUELA:** *Caiman crocodilus crocodilus*; *Drymarchon corais* [Nasir, 1974].

Site: Intestine.

B: 2.30-3.00 mm in length; OS: 200-250 in diameter, with 22-30 spines; VS: 80-110 in diameter; E: 20-28 x 11 (after Braun, 1901).

\*Remarks: Referred as *Acanthostomum scyphocephalum* (Braun, 1899) Hughes, Higginbotham & Clary, 1941 by Mañé-Garzón & Gil (1961a) and by Nasir (1974).

## SUPERFAMILY PARAMPHISTOMOIDEA Fiscoeder, 1901

### FAMILY CLADORCHIIDAE Fiscoeder, 1901

#### HALLTREMA Lent & Freitas, 1939

*Halltrema avitellina* Lent & Freitas, 1939 (Fig. 124)

**BRAZIL:** *Chelonoidis denticulata* (= *Testudo denticulata*) [Alho, 1965]; *Podocnemis expansa* [Lent & Freitas, 1939; Freitas & Lent, 1942].

**VENEZUELA:** *Podocnemis* sp. [Cordero & Vogelsang, 1940; Caballero & Diaz-Ungria, 1958; Diaz-Ungria, 1967; 1973].

Sites: Stomach, intestine.

B: 7.52-11.49 mm x 3.58-4.42 mm; OS: 63-79 x 58-76; VS: 1.31-1.63 mm x 1.37-1.63 mm; E: 128-136 x 64-68 (after Lent & Freitas, 1939).

\**Halltrema heteroxenus* (Cordero & Vogelsang, 1940) Jones, 2005 (Fig. 125)

**ECUADOR:** *Rhinoclemmys nasuta* [Dyer & Carr, 1990].

**VENEZUELA:** *Podocnemis* sp. [Cordero & Vogelsang, 1940].

Site: Stomach.

B: 5.70 mm x 2.95 mm; OS: mng; VS: 1.30 mm in diameter; E: no eggs (after Cordero & Vogelsang, 1940)

\*Remarks: Referred as *Cladorchis heteroxenus* by Cordero & Vogelsang (1940) and as *Pseudallassostoma heteroxenus* by Dyer & Carr (1990). The genus *Pseudallassostoma* Yamaguti, 1958 was considered synonym of *Halltrema* Lent & Freitas, 1939 by Jones (2005).

### **NEMATOPHILA Travassos, 1934**

\**Nematophila argentinum* (Cordero & Vogelsang, 1940) Lenis & Vélez, 2011 (Fig. 126)

**ARGENTINA:** *Phrynops* sp. (cited as probably *Hydraspis* sp.) [Cordero & Vogelsang, 1940].

**COLOMBIA:** *Podocnemis lewyana*; *Trachemys callirostris callirostris* [Lenis & Vélez, 2011].

Site: Intestine.

B: 10.14-15.62 mm x 4.06-5.38 mm; OS: absent; VS: 11.83-2.49 mm x 1.91-2.75 mm; E: 124-169 x 82-119 (after Lenis & Vélez, 2011).

\*Remarks: Referred as *Paramphistomum argentinum* by Cordero & Vogelsang (1940).

\**Nematophila grandis* (Diesing, 1839) Travassos, 1934 (Fig. 127)

**ARGENTINA:** *Phrynops hilarii* [Lombardero & Moriena, 1977].

**BRAZIL:** *Chelus fimbriatus* (= *Chelys fimbriata*) [Travassos, 1934]; *Hydraspis schoppii* [Travassos, 1934; Alho, 1964]; *Kinixys erosa* [after Travassos, Freitas & Kohn, 1969]; *Kinosternon scorpioides* (= *Kinosternon scorpioides scorpioides*) [Alho, 1964]; *Mesoclemmys gibba* (= *Hydraspis gibba*; *Phrynops gibbus*); *Mesoclemmys nasuta* (= *Batrachemys nasuta*; *Rhynemis nasuta*) [Travassos, 1934]; *Peltocephalus dumerilianus* (= *Podocnemis dumeriliana*; *Podocnemis tracaxa*) [Diesing, 1850; Travassos, 1934; Alho, 1964]; *Phrynops geoffroanus* (= *Hydraspis geoffroyana*); [Travassos, 1934]; *Podocnemis erythrocephala* [Diesing, 1850]; *Podocnemis expansa* [Diesing, 1839; Travassos, 1934; Lent & Freitas, 1939]; *Rhinoclemmys punctularia* (= *Geoemyda punctularia punctularia*; *Rhinoclemmys punctularia punctularia*) [Alho, 1964].

**ECUADOR:** *Rhinoclemmys areolata*; *Rhinoclemmys nasuta* [Dyer & Carr, 1990].

**FRENCH GUYANA:** *Rhinoclemmys punctularia* [Dyer & Carr, 1990].

**PARAGUAY:** *Hydromedusa tectifera* [Masi-Pallarés & Benítez-Usher, 1973; Masi-Pallarés, Benítez-Usher & Vergara, 1973; Masi-Pallarés, Benítez-Usher & Maciel, 1976].

**PERU:** *Podocnemis unifilis* [Salízar & Sánchez, 2004; Sánchez, Tantaleán, Vela & Méndez, 2006].

**VENEZUELA:** *Kinosternon scorpioide* [Diaz-Ungria, 1978; 1979]; *Podocnemis expansa* [Heyneman et al. 1960; Diaz-Ungria, 1978; 1979]; *Podocnemis unifilis*; *Podocnemis vogli* [Heyneman, Brenes & Diaz-Ungria, 1960]; *Podocnemis* sp. [Caballero & Diaz-Ungria, 1958].

Sites: Intestine, stomach.

B: 11.25-22.81 mm x 5.81-10.00 mm; OS: absent; VS: 2.06-3.25 mm x 2.00-3.94 mm; E: 143 x 71 (after Lent & Freitas, 1939).

\*Remarks: Referred as *Amphistoma grande* by Diesing (1839) and as *Nematophila grande* by Lent & Freitas (1939) and Caballero & Diaz-Ungria (1958).

\**Nematophila venezuelensis* (Cordero & Vogelsang, 1940) Lenis & Vélez, 2011 (Fig. 128)

**COLOMBIA:** *Podocnemis lewyana* [Lenis & Vélez, 2011].

**VENEZUELA:** *Phrynops* sp. (= *Hydraspis* sp.) [Cordero & Vogelsang, 1940].

B: 9.42-22.11 mm x 4.82-8.56 mm; OS: absent; VO: 1.29-2.01 mm x 1.41-2.11 mm; E: 109-149 x 78-94 (after Lenis & Vélez, 2011).

\*Remarks: Referred as *Allassostoma venezuelensis* by Cordero & Vogelsang (1940).

### **ORIXIMINATREMA Knoff, Brooks, Mullins & Gomes, 2012**

*Oriximinatrema noronhae* Knoff, Brooks, Mullins & Gomes, 2012 (Fig. 129)

**BRAZIL:** *Podocnemis expansa* [Knoff, Brooks, Mullins & Gomes, 2012].

Sites: Stomach, intestine.

B: 1.10-2.75 mm x 0.48-1.05 mm; OS: absent; VS: 290-640 x 380-760; E: 60-110 x 50-70 (after Knoff, Brooks, Mullins & Gomes, 2012).

### **PSEUDOCLEPTODISCUS Caballero, 1961**

*Pseudocleptodiscus margaritae* Caballero, 1961

**ECUADOR:** *Rhinoclemmys nasuta* [Dyer & Carr, 1990, wmd].

Site: Intestine.

### **PSEUDONEMATOPHILA Lenis & Vélez, 2011**

\**Pseudonematophila ovalis* (Cordero & Vogelsang, 1940) Lenis & Vélez, 2011 (Fig. 130)

**COLOMBIA:** *Podocnemis lewyana* [Lenis & Vélez, 2011].

**VENEZUELA:** *Podocnemis* sp. [Cordero & Vogelsang, 1940].

Site: Stomach.

B: 8.23-17.15 mm x 4.58-8.88 mm; OS: mng; VS: 1.39-2.41 mm x 1.37-2.05 mm; E: 111-123 x 79-89 (after Lenis & Vélez, 2011).

\*Remarks: Referred as *Nematophila ovalis* by Cordero & Vogelsang (1940).

## FAMILY DIPLODISCIDAE Cohn, 1904

### CATADISCUS Cohn, 1904

\**Catadiscus dolichocotyle* (Cohn, 1903) Cohn, 1904 (Fig. 131)

**ARGENTINA:** *Philodryas* sp. [Lunaschi & Drago, 2010].

**BRAZIL:** *Chironius fuscus* (= *Herpetodryas fuscus*) [Freitas & Lent, 1939b].

**URUGUAY:** *Liophis miliaris* (= *Liophis miliaris miliaris*) [Mañé-Garzón & Gortari, 1965].

Site: Intestine.

B: 0.90-1.00 mm x 0.42 mm; OS: 160 x 130; VS: 370 x 300; E: 73 x 36 (after Cohn, 1903).

\*Remarks: Referred as *Amphistomum dolichocotyle* Cohn (1903).

*Catadiscus freitaslenti* Ruiz, 1943 (Fig. 132)

**ARGENTINA:** *Bothropoides diporus* (= *Bothrops neuwiedii diporus*); *Bothropoides neuwiedi* (= *Bothrops neuwiedii meridionalis*); *Bothrops alternatus* [Poumarau, 1968]; *Erythrolampus aesculapii*; *Liophis almadensis* (= *Leimadophis almada*); *Lygophis flavifrenatus* (= *Liophis flavifrenatus*); *Xenodon dorbignyi* (= *Lystrophis dorbignyi*); *Xenodon merremi* (= *Waglerophis merremi*) [Poumarau, 1968].

**BRAZIL:** *Liophis miliaris* (= *Liophis miliaris miliaris*) [Ruiz, 1943a].

Site: Intestine.

B: 3.11-3.15 mm x 0.96-1.06 mm; OS: 205-217 x 282-294 (without diverticles); VS: 791-876 x 777-791; E: 70-84 x 33-53 (after Ruiz, 1943a).

*Catadiscus longicoecalis* Poumarau, 1965 (Fig. 133)

**ARGENTINA:** *Bothropoides diporus* (= *Bothrops neuwiedii diporus*); *Bothropoides neuwiedi* (= *Bothrops neuwiedii meridionalis*) [Poumarau, 1965; 1968]; *Philodryas olfersii*; *Xenodon dorbignyi* (= *Lystrophis dorbignyi*); [Poumarau, 1968].

Site: Intestine.

B: 2.84 mm x 1.18 mm; OS: 240 x 296 (without diverticles); VS: 904 x 760; E: 91-104 x 52-65 (after Poumarau, 1965).

*Catadiscus rochai* Correa & Artigas, 1978/1979 (Fig. 134)

**BRAZIL:** *Lygophis typhlus* (= *Dromicus typhlus*) [Correa & Artigas, 1978/1979].

Site: Intestine.

B: 1.85 mm x 0.75 mm; OS: 270 in diameter (with diverticles); VS: 490 x 600; E: 115 x 52 (after Correa & Artigas, 1978/1979).

*Catadiscus uruguayensis* Freitas & Lent, 1939 (Fig. 135)

**ARGENTINA:** *Liophis poeciloyurus* [Lunaschi & Drago, 2002].

Site: Intestine.

B: 1.49-1.64 mm x 0.71-0.82 mm; OS: mng; VS: 509-566 x 394-434; E: 67-93 x 36-55 (after Lunaschi & Drago, 2002).

## SUPERFAMILY PLAGIORCHIOIDEA Lühe, 1901

### FAMILY MACRODEROIDIDAE McMullen, 1937

#### RAUSCHIELLA Babero, 1951

*Rauschiella linguatula* (Rudolphi, 1819) Razo-Mendivil, León-Règagnon & Perez-Ponce de León, 2006

**BRAZIL:** *Mastigodryas bifossatus* (= *Dryadophis bifossatus*) [after Noronha, Sá, Knoff, Muniz-Pereira & 2009, wmd]  
Site: intestine.

### FAMILY OPISTHOGONIMIDAE Travassos, 1928

#### LIOPHISTREMA Artigas, Ruiz & Leão, 1942

*Liophistrema buccalis* Holcman-Spector & Mañé-Garzón, 1973 (Fig. 136)

**URUGUAY:** *Thamnodynastes strigatus* [Holcman-Spector & Mañé-Garzón, 1973].  
Site: Mouth.

B: 1.11-1.34 mm x 0.38-0.54 mm; OS: 250-300 x 200-290; VS: 70-100 x 80-110; E: 15-17 x 7-10 (after Holcman-Spector & Mañé-Garzón, 1973).

\**Liophistrema pulmonale* (Artigas, Ruiz & Leão, 1942) Tkrach, 2008 (Fig. 137)

**BRAZIL:** *Liophis miliaris* (= *Liophis miliaris miliaris*) [Artigas, Ruiz & Leão, 1942].

**URUGUAY:** *Liophis miliaris* (= *Liophis miliaris miliaris*) [Mañé-Garzón & Gortari, 1965].  
Site: Lungs.

B: 9.31-17.29 mm x 1.33-2.66; OS: 931-1,729 in diameter; VS: mng; E: 25-30 x 14-19 (after Artigas, Ruiz & Leão, 1942).

\*Remarks: Referred as *Liophistrema pulmonalis* by Artigas, Ruiz & Leão (1942) and emended to *L. pulmonale* by Tkach (2008).

#### OPISTHOGONIMUS Lühe, 1900

\**Opisthogonimus afranioi* Preira, 1929 (Fig. 138)

**BRAZIL:** *Bothropoides newiedi* (= *Bothrops newiedii*); *Chironius carinatus*; *Philodryas patagoniensis* (= *Philodryas schotti*) [Pereira, 1929a].

**URUGUAY:** *Philodryas patagoniensis* (= *Philodryas schotti*) [Mañé-Garzón & Gortari, 1965].  
Site: Oesophagus.



B: 4.25-7.70 mm x 1.50-2.00 mm; OS: 625-1,000 in diameter; VS: 420-650 in diameter; E: 30 x 15 (after Pereira, 1929a).

\*Remarks: Referred as *Westella afranioi* by Freitas (1956) and Travassos, Freitas & Kohn (1969).

***Opisthognimus artigasi*** Ruiz & Leão, 1942 (Fig. 139)

**ARGENTINA:** *Bothropoides jararaca* (= *Bothrops jararaca*); *Lygophisttyphlus* (= *Leimadophis typhlus*); *Mastigodryas bifossatus* (= *Drymobius bifossatus*); *Mastigodryas bifossatus triseriatus*; *Philodryas psammophidea* (= *Philodryas psammophideus*); *Thamnodynastes* sp. (cited as *Thamnodynastes pallidus*); *Xenodon dorbignyi* (= *Lystrophis dorbignyi*) [Poumarau, 1968].

**BRAZIL:** *Bothrops moojeni* [Barrella & Silva, 2003]; *Thamnodynastes pallidus* (= *Dryophylax pallidus*) [Ruiz & Leão, 1942a].

Sites: Mouth, oesophagus, stomach.

B: 3.50-6.25 mm x 1.00-1.84 mm; OS: 470-850 x 497-884; VS: 552-644 x 515-644; E: 27 x 15-17 (after Ruiz & Leão, 1942a).

***Opisthognimus fariai*** Leão & Ruiz, 1943 (Fig. 140)

**BRAZIL:** *Liophis miliaris* (= *Liophis miliaris miliaris*) [Leão & Ruiz, 1943; Pinto, Mati & Melo, 2012]; *Xenodon merremi* (= *Waglerophis merremii*) [Ruiz & Leão, 1942a].

Sites: Mouth, oesophagus.

B: 3.04-5.98 mm x 0.74-1.23 mm; OS: 370-530 x 400-530; VS: 330-480 x 310-440; E: 29-30 x 14-15 (after Leão & Ruiz, 1943).

***Opisthognimus fonsecai*** Ruiz & Leão, 1942 (Fig. 141)

**ARGENTINA:** *Liophis miliaris semiaureus* [Boero, Led & Brandetti, 1972]; *Xenodon merremi* (= *Waglerophis merremii*) [Lunaschi & Drago, 2001].

**BRAZIL:** *Bothropoides jararaca* (= *Bothrops jararaca*); *Bothropoides neuwiedi* (= *Bothrops neuwiedii*) [Mati & Melo, 2012]; *Bothrops alternatus* [Pinto, Mati & Melo, 2012]; *Bothrops moojeni* [Barrella & Silva, 2003; Silva, 2005]; *Chironius bicarinatus*; *Chironius foveatus*; *Clelia occipitolutea*; *Erythrolamprus aesculapii* [Pinto, Mati & Melo, 2012]; *Liophis miliaris* (= *Liophis miliaris miliaris*) [Pinto, Mati & Melo, 2012]; *Liophis poecilogyrus*; *Liophis typhlus*; *Mastigodryas bifossatus*; *Philodryas patagoniensis* [Pinto, Mati & Melo, 2012]; *Xenodon merremi* (= *Waglerophis merremii*) [Ruiz & Leão, 1942a; Pinto, Mati & Melo, 2012].

Sites: Mouth, oesophagus, intestine.

B: 3.31-4.14 mm x 1.53-1.66 mm; OS: 474-552 x 460-552; VS: mng; E: 30 x 15 (after Ruiz & Leão, 1942a).

B: 3.11-4.52 mm x 0.83-1.26 mm; OS: 374-503 x 360-484; VS: 317-420 x 288-415; E: 19-24 x 12-14 (after Lunaschi & Drago, 2001).

***Opisthognimus interrogativus*** (Nicoll, 1914) Pereira, 1929 (Fig. 142)

**ARGENTINA:** *Xenodon merremi* (= *Waglerophis merremii*) [Lunaschi & Drago, 2001].

**BRAZIL:** *Bothropoides jararaca* (= *Bothrops jararaca*); *Philodryas patagoniensis* (= *Philodryas schottii*); *Xenodon merremi* (= *Ophis merremii*; *Waglerophis merremii*) [Pereira, 1929a].

Sites: Mouth, oesophagus.

B: 3.30-4.70 mm x 1.60-1.80 mm; OS: 700-750 in diameter; VS: 500-570 in diameter; E: 30 x 15 (after Pereira, 1929a).

B: 2.10-2.29 mm x 0.74-0.84 mm; OS: 355-365 x 260-374; VS: 250-269 x 250-317; E: 22-24 x 12-14 (after Lunaschi & Drago, 2001).

\**Opisthognomus megabothrium* Pereira, 1928 (Fig. 143)

**ARGENTINA:** *Liophis jaegeri* [Lunaschi & Sutton, 1985]; *Xenodon merremi* (= *Waglerophis merremii*) [Lunaschi & Drago, 2001].

**BRAZIL:** *Liophis miliaris*; *Xenodon merremi* (= *Liophis merremii*; *Ophis merremii*; *Waglerophis merremii*) [Pereira, 1928; 1929a].

**URUGUAY:** *Liophis miliaris* (= *Liophis miliaris miliaris*) [Mañé-Garzón & Gortari, 1965].

Sites: Mouth, oesophagus, intestine.

B: 2.80-4.10 mm x 0.90-1.20 mm; OS: 450-570 in diameter; VS: 720-800 in diameter; E: 27-34 x 15-27 (after Pereira, 1929a).

B: 1.60-3.43 mm x 0.65-1.05 mm; OS: 227-389 x 202-389; VS: 291-617 x 272-579; E: 23-28 x 14-15 (after Lunaschi & Drago, 2001).

\*Remarks: Referred as *Opisthognomus* (*Opisthognomus*) *megabothrium* by Lunaschi & Sutton (1985).

*Opisthognomus misionesensis* Lunaschi & Drago, 2001 (Fig. 144)

**ARGENTINA:** *Xenodon merremi* (= *Waglerophis merremii*) [Lunaschi & Drago, 2001].

Site: Intestine.

B: 3.10-3.62 mm x 0.94-1.40 mm; OS: 365-461 x 347-461; VS: 345-461 x 316-422; E: 21-28 x 11-16 (after Lunaschi & Drago, 2001).

*Opisthognomus pereirai* Ruiz & Leão, 1942 (Fig. 145)

**BRAZIL:** *Chironius carinatus* [Ruiz & Leão, 1942a].

Site: Oesophagus.

B: 9.50-12.00 mm x 2.25-2.75 mm; OS: 1.10 mm x 1.10-1.20 mm; VS: 920-1,010 in diameter; E: 26-34 x 15 (after Ruiz & Leão, 1942a).

\**Opisthognomus lecithonotus* Lühe, 1900 (Fig. 146)

**ARGENTINA:** *Bothropoides diporus* (= *Bothrops neuwiedii diporus*) [after Lunaschi & Drago, 2007]; *Bothropoides jararaca* (= *Bothrops jararaca*) [Martínez, Troiano, Binda, Selles, Jara & Fescina, 1996]; *Bothropoides neuwiedi* (= *Bothrops neuwiedii meridionalis*) [Poumarau, 1968]; *Bothrops alternata* [Poumarau, 1968; Martínez, Troiano, Binda, Selles, Jara & Fescina, 1996]; *Bothrops moojeni* [Martínez, Troiano, Binda, Selles, Jara & Fescina, 1996];

*Clelia clelia* (= *Cloelia cloelia*, *Pseudoboa cloelia*) (referred as *Boiruna maculata* by Lunaschi & Drago (2007); [Poumarau, 1968; Boero & Led, 1971]; *Helicops carinicaudus* (= *Helicops carinicauda*) [Martínez, Troiano, Binda, Selles, Jara & Fescina, 1996]; *Helicops infrataeniatus* [after Lunaschi & Drago, 2007]; *Helicops leopardinus* (= *Helicops leopardina*) [Poumarau, 1968]; *Hydrodynastes gigas* (= *Cyclagras gigas*) [Poumarau, 1968; Martínez, Troiano, Binda, Selles, Jara & Fescina, 1996]; *Lygophis anomalus* (= *Liphis anomalus* sic) [Poumarau, 1968]; *Lygophis flavifrenatus* [Poumarau, 1968]; *Lygophis typhlus* (= *Leimadophis typhlus*) [Poumarau, 1968]; *Mastigodryas bifossatus* (= *Drymobius bifossatus*) [Poumarau, 1968]; *Mastigodryas bifossatus triseriatus* [after Lunaschi & Drago, 2007]; *Micrurus pyrrhocryptus*; *Philodryas patagoniensis* (= *Philodryas schottii*) [Poumarau, 1968]; *Thamnodynastes pallidus* [Poumarau, 1968]; *Thamnodynastes strigatus* [Martínez, Troiano, Binda, Selles, Jara & Fescina, 1996]; *Thamnodynastes* sp. [Cordero & Vogelsang, 1928]; *Xenodon dorbignyi* (= *Lystrophis dorbignyi*) [Poumarau, 1968]; *Xenodon merremi* (= *Ophis merremii*; *Waglerophis merremi*) [Cordero & Vogelsang, 1928; Poumarau, 1968].

**BRAZIL:** *Bothrops atrox*; *Bothrops jararacussu* [Pereira, 1929a; Silva, 2004]; *Bothrops moojeni* [Silva, 2004]; *Bothrops* sp. [Pereira, 1929a; Travassos & Freitas, 1941c]; *Hydrodynastes gigas* (= *Cyclagras gigas*) [Pereira, 1929a]; *Mastigodryas bifossatus* (= *Dryadophis bifossatus*; *Drymobius bifossatus*) [Pereira, 1929a]; *Philodryas* sp. [Rego & Vicente, 1988]; *Xenodon merremi* (= *Ophis merremii*; *Waglerophis merremi*) [Pereira, 1929a].

**URUGUAY:** *Philodryas patagoniensis* (= *Philodryas schotti*) [Mañé-Garzón & Gortari, 1965]. Sites: Mouth, oesophagus, stomach, lung, eye.

B: 6.05-7.50 mm x 1.55-1.85 mm; OS: 650-750 in diameter; VS: 650-900 in diameter; E: 27-34 x 13-15 (after Pereira, 1929a).

\*Remarks: Referred as *Distomum xenodontis* by Cordero & Vogelsang (1928), as *Opisthognimus philodryadum* by Pereira (1929a) and Travassos & Freitas (1941c) and as *Opisthognimus (Westella) philodryadum* by Artigas, Ruiz & Leão (1943).

\**Opisthognimus serpentis* Artigas, Ruiz & Leão, 1943 (Fig. 147)

**ARGENTINA:** *Helicops infrataeniatus* [Lunaschi & Sutton, 1985].

**BRAZIL:** *Helicops carinicaudus* (= *Helicops corinicauda*) [sic]; *Helicops infrataeniatus*; [Rodrigues & Santos, 1974]; *Liophis miliaris*; *Liophis poecilogyrus* (= *Leimadophis poecilogyrus*); *Thamnodynastes pallidus* (= *Dryophylax pallidus*); *Tomodon dorsatus*; *Xenodon merremi* [Artigas, Ruiz & Leão, 1943].

**URUGUAY:** *Philodryas patagoniensis* (= *Philodryas schotti*) [Mañé-Garzón & Gortari, 1965]. Sites: Mouth, oesophagus.

B: 4.60-6.70 mm x 0.93-1.20 mm; OS: 494-636 in diameter; VS: 395-452 in diameter; E: 25-30 x 14-16 (after Artigas, Ruiz & Leão, 1943).

\*Remarks: Referred as *Westella serpentis* by Artigas, Ruiz & Leão, (1943), Freitas (1956), Mañé-Garzón & Gortari (1965), Rodrigues & Santos (1974) and by Lunaschi & Drago (2007).

\**Opisthognimus sulina* (Artigas, Ruiz & Leão, 1942) (Fig. 148)

**BRAZIL:** *Philodryas patagoniensis* (= *Philodryas schotti*) [Artigas, Ruiz & Leão, 1942; 1943; Freitas, 1956].

**URUGUAY:** *Philodryas patagoniensis* (= *Philodryas schotti*) [Mañé-Garzón & Gortari, 1965].  
Sites: Mouth, oesophagus.

B: 6.93-7.53 mm x 1.33-1.91 mm; OS: 424-692 in diameter; VS: 537-636 in diameter; E: 18-28 x 11-17 (after Artigas, Ruiz & Leão, 1942).

\*Remarks: Referred as *Westella sulina* by Artigas, Ruiz & Leão (1942); Freitas (1956) and by Mañé-Garzón & Gortari (1965); as *Opisthogonimus (Westella) sulina* by Artigas, Ruiz & Leão (1943).

***Opisthogonimus uruguayensis*** Mañé-Garzón & Holcman-Spector, 1973 (Fig. 149)

**URUGUAY:** *Thamnodynastes hypoconia* (= *Thamnodynastes strigilis*) [Mañé-Garzón & Holcman-Spector, 1973b].

Site: Mouth.

B: 1.49-1.88 mm x 0.38-0.46 mm; OS: 210-220 x 200-210; VS: 170-180 in diameter; E: 17 x 7 (after Mañé-Garzón & Holcman-Spector, 1973b).

### **PARACOTYLETREMA Volonterio, Baletta & Meneghel, 2006**

***Paracotyletrema poncedeleoni*** Volonterio, Baletta & Meneghel, 2006 (Fig. 150)

**URUGUAY:** *Lygophis anomalus* (= *Liophis anomalus*) [Volonterio, Baletta & Meneghel, 2006].

Sites: Mouth, oesophagus.

B: 0.58-1.15 mm x 0.37-0.63 mm; OS: 257-383 x 210-341; VS: 157-273 x 131-267; E: 25-32 x 13-17 (after Volonterio, Baletta & Meneghel, 2006).

### **FAMILY PLAGIORCHIIDAE Lühe, 1901**

#### **ALLOPHARYNX Strom, 1928**

***Allopharynx daileyi*** Bursey, Goldberg & Vitt, 2005 (Fig. 151)

**BRAZIL:** *Uranoscodon superciliosus* [Bursey, Goldberg & Vitt, 2005; Ávila & Silva, 2013].

Site: Intestine.

B: 9.40-15.00 mm x 0.50-2.10 mm; OS: 179-230 in diameter; VS: 204-281 in diameter; E: 15-24 x 10-15 (after Bursey, Goldberg & Vitt, 2005).

#### **BIERIA Leão, 1946**

***Bieria artigasi*** Leão, 1946 (Fig. 152)

**BRAZIL:** *Liophis miliaris* (= *Liophis miliaris miliaris*) [Leão, 1946].

Site: Lungs.

B: 10.00-17.00 mm x 1.16-2.02 mm; OS: 405-552 x 478-662; VS: 644-959 x 607-1,067; E: 27-42 x 15-23 (after Leão, 1946).

### **GLOSSIDIELLA Travassos, 1927**

*Glossidiella ornata* Travassos, 1927 (Fig. 153)

**ARGENTINA:** *Hydrodynastes gigas* (= *Cyclagras gigas*) [Poumarau, 1968; Lunaschi & Sutton, 1985; Martínez, Troiano, Binda, Selles, Jara & Fescina, 1996].

**BRAZIL:** *Hydrodynastes gigas* (= *Cyclagras gigas*) [Travassos, 1928].

Site: Lungs, mouth.

B: 11.00-13.20 mm x 1.00-1.50 mm; OS: 470-550 in diameter; VS: 400-450 in diameter; E: 50-55 x 27-30 (after Travassos, 1928).

### **GLOSSIDIOIDES Yamaguti, 1958**

\**Glossidioides loossi* (Travassos, 1927) Yamaguti, 1958 (Fig. 154)

**ARGENTINA:** *Hydrodynastes gigas* (= *Cyclagras gigas*) [Poumarau, 1968; Lunaschi & Sutton, 1985; Martínez, Troiano, Binda, Selles, Jara & Fescina, 1996].

**BRAZIL:** *Hydrodynastes gigas* (= *Cyclagras gigas*) [Travassos, 1927; 1928a].

Sites: Lungs, trachea.

B: 3.60-4.50 mm x 1.00-1.40 mm; OS: 320-450 in diameter; VS: 300-370 in diameter; E: 47-50 x 30 (after Travassos, 1928).

\*Remarks: Referred as *Glossidium loossi* by Travassos (1927, 1928a) and by mistake the host as *Eunectes murinus*, which was rectified posteriorly.

### **HAPLOMETROIDES Odhner, 1910**

\**Haplometroides buccicola* Odhner, 1911 (Fig. 155)

**ARGENTINA:** *Micrurus corallinus* [Lunaschi & Drago, 2010]; *Micrurus frontalis* [Poumarau, 1968].

**BRAZIL:** *Amphisbaena alba*; *Epicrates crassus* (= *Epicrates cenchria crassus*) [Ruiz & Perez, 1959]; *Erythrolamprus aesculapii* [Artigas & Paulino, 1988]; *Micrurus corallinus* [Santos, Barrella, Zica & Silva, 2008]; *Phalotris lativittatus* [Silva, Andrade, Monteiro e Silva, Rossellini & Barrella, 2005; Santos, Barrella, Zica & Silva, 2008].

**PARAGUAY:** *Micrurus* sp. (= *Elaps* sp.) [Odhner, 1911; Masi-Pallarés & Benitez-Usher, 1973; Masi-Pallarés, Benitez-Usher & Maciel, 1976].

Sites: Oesophagus, mouth, lungs.

B: 6.07- 8.21 mm x 1.28-2.05 mm; OS: 521-638 x 521-660; VS: 319-489 x 351-489; E: 45-47 x 24-28 (after Ruiz & Perez, 1959).

B: 5.48-7.21 mm x 1.51-1.75 mm; OS: 477-569 x 463-569; E: 41-48 x 22-25 (after Artigas & Paulino, 1988).

\*Remarks: Referred as *Zeferinella vazi* by Artigas & Paulino (1988) and considered synonym of *H. buccicola* by Tkach (2008).

***Haplometroides intercaecalis*** Silva, Ferreira & Strüssmann, 2007 (Fig. 156)

**BRAZIL:** *Phalotris matogrossensis* [Silva, Beda & Ferreira, 2008; Silva, 2008]; *Phalotris nasutus* [Silva, Ferreira & Strüssmann, 2007].

Sites: Oesophagus, stomach, intestine.

B: 6.27-7.58 mm x 0.80-1.18 mm; OS: 302-334 x 321-357; VS: 266-307 x 272-328; E: 42-57 x 30-41 (after Silva, Ferreira & Strüssmann, 2007).

***Haplometroides odhneri*** Ruiz & Perez, 1959 (Fig. 157)

**BRAZIL:** *Micrurus frontalis* [Silva & Barrella, 2002]; *Micrurus lemniscatus* [Ruiz & Perez, 1959]; *Trilepida koppesi* (= *Leptotyphlops koppesi*) [Silva, Zica, Cruz, O' Reilly & Costa, 2005].

Site: Oesophagus.

B: 4.40-5.24 mm x 0.71-1.19 mm; OS: 330-415 x 298-393; VS: 298-393 x 191-351; E: 34-37 x 23-25 (after Ruiz & Perez, 1959).

### **PARAHAPLOMETROIDES Thatcher, 1963**

***Parahaplometroides basiliscae*** Thatcher, 1963 (Fig. 158)

**SOUTH AMERICA:** (unknown locality) *Basiliscus basiliscus* [Stunkard & Gandal, 1966].

Site: Mouth.

B: 4.00-6.00 mm x 1.30-2.50 mm; OS and VS: 600-650 in diameter; E: 38-45 x 18-20 (after Stunkard & Gandal, 1966).

### **PLAGIORCHIS Lühe, 1899**

***Plagiorchis freitasi*** Vicente, 1978 (Fig. 159)

**BRAZIL:** *Tropidurus torquatus* [Vicente, 1978].

Site: Intestine.

B: 10.20-12.03 mm x 1.53-1.63 mm; OS: 210-280 x 250-280; VS: 210-220 x 150-210; E: 21-25 x 10-14 (after Vicente, 1978).

\****Plagiorchis lühei*** (Travassos, 1927) Mehra, 1931 (Fig. 160)

**ARGENTINA:** *Hydrodynastes gigas* (= *Cyclagras gigas*) [Poumarau, 1968; Lunaschi & Sutton, 1985; Martínez, Troiano, Binda, Selles, Jara & Fescina, 1996; Lunaschi & Drago, 2010].

**BRAZIL:** *Hydrodynastes gigas* (= *Cyclagras gigas*) [Travassos, 1927; 1928a; Artigas & Campos, 1976/1977].

Sites: Oesophagus, mouth, intestine, lungs.

B: 4.70-5.00 mm x 1.30-1.70 mm; OS: 520-650 in diameter; VS: 500-650 in diameter; E: 45-50 x 25-35 (after Travassos, 1927).

\*Remarks: Referred as *Microderma lühei* (Travassos, 1927) Mehra, 1931 by Poumarau (1968), Travassos, Freitas & Kohn (1969) and by Martínez, Troiano, Binda, Selles, Jara & Fescina (1996).

***Plagiorchis vicentei*** Rodrigues, 1994 (Fig. 161)

**BRAZIL:** *Hemidactylus mabouia* [Rodrigues, 1994].

Site: Intestine.

B: 1.73-2.66 mm x 0.69-0.98 mm; OS: 190-240 x 210-250; VS: 170-180 x 160-180; E: 29-33 x 14-18 (after Rodrigues, 1994).

### **PNEUMOTREMA Bhalerao, 1937**

***Pneumotrema travassosi*** Bhalerao, 1937 (Fig. 162)

**BRAZIL:** *Amphisbaena alba* [Bhalerao, 1937].

Site: Lungs.

B: 9.00 mm x 1.40 mm; OS: 500 x 370; VS: 750 in diameter; E: 38-42 x 22-26 (after Bhalerao, 1937).

### **STICHOLECITHA Prudhoe, 1949**

***Sticholecitha serpentis*** Prudhoe, 1949 (Fig. 163)

**BRAZIL:** *Bothrops moojeni* [Barrella & Silva, 2003; Silva, Rossellini, Dal-Pai & Silva, 2005; Silva, Rossellini, Silva & Silva, 2005]; *Xenodon severus* [Freitas, 1956].

**SURINAM:** *Chironius carinatus* [Prudhoe, 1949].

Site: Oesophagus.

B: 5.20-8.40 mm x 1.20-2.00 mm; OS: 560-650 in diameter; VS: 450-460 in diameter; E: 25-27 x 12-15 (after Prudhoe, 1949).

### **STYPHLODORA Looss, 1899**

***Styphlodora condita*** Faria, 1911 (Fig. 164)

**ARGENTINA:** *Bothropoides diporus* (= *Bothrops neuwiedii diporus*) [Boero, Led & Brandetti, 1972]; *Bothropoides neuwiedi* (= *Bothrops neuwiedii*; *Bothrops neuwiedii meridionalis*) [Poumarau, 1968; Lunaschi & Sutton, 1985]; *Bothrops alternatus* [Poumarau, 1968; Lunaschi & Sutton, 1985]; *Eunectes notaeus* [Boero, Led & Brandetti, 1972]; *Hydrodynastes gigas* (= *Cyclagras gigas*) [Poumarau, 1968; Lunaschi & Sutton, 1985]; *Leptodeira annulata* (= *Leptodeira annulata annulata*; *Leptodeira annulata pulchriceps*); *Mastigodryas bifossatus* (= *Drymobius bifossatus*); *Mastigodryas bifossatus triseriatus* [Poumarau, 1968; Lunaschi & Sutton, 1985]; *Micrurus pyrrhocryptus* (= *Micrurus frontalis pyrrhocryptus*) [Poumarau, 1968; Lunaschi & Sutton, 1985]; *Oxyrhopus rhombifer*; *Phylodryas patagoniensis* (= *Phylodryas*

*schottii*) [Poumarau, 1968; Lunaschi & Sutton, 1985]; *Thamnodynastes* sp. (=cited as *Thamnodynastes pallidus*; *Thamnodynastes strigatus* of Lunaschi & Sutton, 1985) [Poumarau, 1968; Lunaschi & Sutton, 1985]; *Tupinambis rufescens* [Lunaschi & Sutton, 1985]; *Xenodon dorbignyi* (= *Lystrophis dorbignyi*) [Poumarau, 1968; Lunaschi & Sutton, 1985]; *Xenodon merremi* (= *Ophis merremii*; *Waglerophis merremii*) [Pereira, 1929a; Poumarau, 1968; Lunaschi & Sutton, 1985].

**BRAZIL:** *Bothrops moojeni* [Barrella & Silva, 2003]; *Spilotes pullatus* [Faria, 1911].

Sites: Intestine, ureter, kidneys.

B: 4.50 mm x 1.00 mm; OS: 360 in diameter; VS: 430 in diameter; E: 42 x 22 (after Faria, 1911).

*Styphlodora gili* Mañé-Garzón & Holcman-Spector, 1967 (Fig. 165)

**BRAZIL:** *Bothropoides jararaca* (= *Bothrops jararaca*) [Fábio, 1979]; *Chironius bicarinatus* [Pinto, Mati & Melo, 2012].

**URUGUAY:** *Bothrops alternatus* (= *Bothrops alternata*) [Mañé-Garzón & Holcman-Spector, 1967c; Poumarau, 1968; Lunaschi & Sutton, 1985].

Sites: Kidneys, ureter.

B: 3.37-4.04 mm x 0.60-0.74 mm; OS: 250 x 270-300; VS: 310 x 300; E: 32-35 x 17 (after Mañé-Garzón & Holcman-Spector, 1967c).

*Styphlodora horrida* (Leidy, 1850) Odhner, 1910

**COLOMBIA:** *Boa constrictor* (= *Constrictor constrictor*) [Thatcher, 1970, wmd].

Site: Ureters.

### TRAVTREMA Pereira, 1929

\**Travtrema stenocotyle* (Cohn, 1902) Goodman, 1951 (Fig. 166)

**ARGENTINA:** *Bothropoides diporus* (= *Bothrops neuwiedii diporus*) [Poumarau, 1968; Lunaschi & Sutton, 1985; Martínez, Troiano, Binda, Selles, Jara & Fescina, 1996]; *Bothropoides neuwiedi* (= *Bothrops neuwiedii meridionalis*) [Poumarau, 1968; Lunaschi & Sutton, 1985]; *Bothrops alternatus* [Poumarau, 1968; Lunaschi & Sutton, 1985]; *Clelia rustica* [Lunaschi & Sutton, 1985]; *Hypsiboas pulchellus* [Ostrowski de Núñez, 1979]; *Liophis jaegeri* [Lunaschi & Sutton, 1985]; *Liophis miliaris semiaureus* [Boero, Led & Brandetti, 1972]; *Liophis poecilogyrus* (= *Leimadophis porcilogyrus*) [Ostrowski de Núñez, 1979]; *Lygophis flavifrenatus* (= *Liophis flavifrenatus*) [Poumarau, 1968; Lunaschi & Sutton, 1985]; *Philodryas patagoniensis* [Lunaschi & Sutton, 1985]; *Philodryas* sp. [Lunaschi & Drago, 2010]; *Sibynomorphus ventrimaculatus*; *Thamnodynastes* sp. (=cited as *Thamnodynastes pallidus*, *Thamnodynastes strigilis* of Lunaschi & Sutton, 1985); *Xenodon dorbignyi* (= *Lystrophis dorbignyi*) [Poumarau, 1968; Lunaschi & Sutton, 1985]; *Xenodon merremi* (= *Waglerophis merremii*) [Poumarau, 1968; Lunaschi & Sutton, 1985].

**BRAZIL:** *Bothropoides neuwiedi* (= *Bothrops neuwiedii*) [Pinto, Mati & Melo, 2012]; *Bothrops moojeni* [Barrella & Silva, 2003; Pinto, Mati & Melo, 2012]; *Bothrops* sp. [Freitas



& Dobbin Jr., 1957]; *Chironius fuscus* [Cohn, 1902]; *Liophis miliaris* (= *Liophis miliaris miliaris*) [Ruiz & Leão, 1942b; Ruiz, 1951; Freitas & Dobbin Jr., 1957]; *Liophis poecilogyrus* (= *Leimadophis poecilogyrus*) [Ruiz & Leão, 1942b; Ruiz, 1951]; *Mastigodryas bifossatus trisseriatus* (= *Dryadophis bifossatus trisseriatus*) [Ruiz, 1951]; [Freitas & Dobbin Jr., 1957]; *Philodryas patagoniensis* (= *Philodryas schottii*) [Ruiz & Leão, 1942b; Freitas & Dobbin Jr., 1957]; *Philodryas* sp. [Freitas & Dobbin Jr., 1957]; *Thamnodynastes pallidus* (= *Dryophylax pallidus*) [Ruiz & Leão, 1942b; Ruiz, 1951]; *Tomodon dorsatus* [Ruiz & Leão, 1942b]; *Xenodon merremi* (= *Ophis merremii*; *Waglerophis merremii*) [Pereira, 1929b; Ruiz & Leão, 1942b]; *Xenodon* sp. (= *Lystrophis* sp.) [Ruiz, 1951].

**PARAGUAY:** *Mastigodryas bifossatus* (= *Drymobius bifossatus*); *Tomodon dorsatus* (= *Tomodon dorsatum*) [Masi-Pallarés & Benitez-Usher, 1973; Masi-Pallarés, Benitez-Usher & Vergara, 1973; Masi-Pallarés, Benitez-Usher & Maciel, 1976].

**SOUTH AMERICA:** *Chironius fuscus* (= *Herpetodryas fuscus*) [Cohn, 1902].

**URUGUAY:** *Lygophis anomalus* (= *Liophis anomalus*) [Mañé-Garzón & Gortari, 1965].

Sites: Intestine, oesophagus, rectum, cloaca.

B: 0.61-3.15 mm x 0.35-1.66 mm; OS: 149-358 x 183-465; VS: 183-531 x 216-714; E: 56-67 x 26-34 (after Freitas & Dobbin Jr., 1957).

\*Remarks: Referred as *Leptophyllum stenocotyle* by Cohn (1902) and by Mañé-Garzón & Gortari (1965), and as *Travtrema travtrema* by Pereira (1929b).

## FAMILY RENIFERIDAE Pratt, 1902

### DASYMETRA Nicoll, 1911

*Dasymetra tupinambis* Nasir & Diaz, 1971 (Fig. 167)

**VENEZUELA:** *Tupinambis teguixin* (= *Tupinambis nigropunctatus*) [Nasir & Diaz, 1971b].

Site: Intestine.

B: 2.28-3.14 mm x 0.58-0.65 mm; OS: 244-291 in diameter; VS: 206-253 in diameter; E: 24-27 x 9-12 (after Nasir & Diaz, 1971b).

### RENIFER Pratt, 1902

The genus *Ochetosoma* Braun, 1902 (preoccupied) was considered synonym of *Renifer* by Tkach (2008).

\**Renifer chironius* (Nasir & Diaz, 1971) (Fig. 168)

**VENEZUELA:** *Chironius carinatus* [Nasir & Diaz, 1971b].

Sites: Oesophagus, intestine.

B: 0.17-0.23 mm x 0.67-0.93 mm; OS: 225-319 in diameter; VS: 244-479 in diameter; E: 33-36 x 56-65 (after Nasir & Diaz, 1971b).

\*Remarks: Referred as *Ochetosoma chironius* by Nasir & Diaz (1971b).

\**Renifer heterocoelium* (Travassos, 1921) Caballero & Vogelsang, 1947 (Fig. 169)

**BRAZIL:** *Bothropoides jararaca* (= *Bothrops jararaca*) [Corrêa, Paulino, Buononato & Federsoni, 1990; Barrella & Silva, 2003]; *Bothropoides insularis* (= *Bothrops insularis*) [Corrêa, Paulino, Buononato & Federsoni, 1990]; *Bothropoides neuwiedi* (= *Bothrops neuwiedii*; *Lachesis neuwiedii*) [Travassos, 1921a]; *Bothrops cotiara* [Corrêa, Paulino, Buononato & Federsoni, 1990; Barrella & Silva, 2003]; *Bothrops moojeni* [Corrêa, Paulino, Buononato & Federsoni, 1990; Barrella & Silva, 2003]; *Bothrops neuwiedii mattogrossensis*; *Chironius bicarinatus* [Corrêa, Paulino, Buononato & Federsoni, 1990]; *Chironius exoletus* [Silva, Rodrigues, Stein, Sipoli, Pinhão & Lopes, 1999]; *Chironius fuscus* [Corrêa, Paulino, Buononato & Federsoni, 1990]; *Drymarchon corais* (= *Drymarchon corais corais*); *Erythrolampus aesculapii*; *Helicops modestus* [Corrêa, Paulino, Buononato & Federsoni, 1990]; *Leptodeira annulata* [Pinto, Mati & Melo, 2012]; *Liophis almadensis*; *Liophis miliaris*; *Liophis poecilogyrus*; *Liophis reginae*; *Lygophis typhlus*; *Mastigodryas bifossatus*; *Philodryas patagoniensis*; *Pseudoboa coronata* (= *Clelia occipitolutea*) [Corrêa, Paulino, Buononato & Federsoni, 1990, after Pinto, Mati & Melo, 2012]; *Xenodon merremi* (= *Waglerophis merremii*) [Corrêa, Paulino, Buononato & Federsoni, 1990; Pinto, Mati & Melo, 2012].

**COLOMBIA:** *Atractus lasallei*; *Bothriechis schlegelli*; *Bothrops asper*; *Chironius carinatus*; *Leptodeira septentrionalis*; *Leptophis ahaetulla*; *Porthidium nasutum* [Lenis, Arredondo & Calle, 2009].

Sites: Intestine, mouth, oral cavity, oesophagus.

B: 2.50-3.30 mm x 1.00-1.20 mm; OS: 360-430 in diameter; VS: 520-640 in diameter; E: 37-42 x 17-23 (after Travassos, 1921a).

B: 2.69-5.14 mm x 0.96-1.47 mm; OS: 390-600 x 370-570; VS: 450-780 x 450-900; E: 32-40 x 18-24 (after Lenis, Arredondo & Calle, 2009).

\*Remarks: Referred as *Heterocoelium heterocoelium* by Travassos (1921a) and as *Ochetosoma heterocoelium* by Caballero & Vogelsang (1947).

Thatcher (1993) in the catalogue of neotropical trematodes cited this species as also occurring in Venezuela but, we did not find any reference in the literature that it occurs in this country. Caballero & Vogelsang (1947) only referred *O. heterocoelium* among the species belonging to the genus *Ochetosoma*.

\**Renifer monstrosus* (Braun, 1901) (Fig. 170)

**VENEZUELA:** *Bothrops atrox* [Caballero & Vogelsang, 1947; Caballero & Diaz-Ungria, 1958; Diaz-Ungria, 1967; 1973].

Site: Oesophagus.

B: 5.58-7.54 mm x 0.36-1.74 mm; OS: 614 in diameter; VS: 730-830 x 664-730; E: 34 x 15-19 (after Caballero & Vogelsang, 1947).

\*Remarks: Referred as *Ochetosoma miladelarocai* by Caballero & Vogelsang (1947) and by Diaz-Ungria (1967; 1973) and considered synonym of *Ochetosoma monstrosus* by Dubois & Mahon (1959).

## FAMILY STYPHLOTREMATIDAE Baer, 1924

### STYPHLOTREMA Odhner, 1911

*Styphlotrema solitaria* Looss, 1899 (Fig. 171)

**BRAZIL:** *Eretmochelys imbricata* [Werneck & Silva, 2012].

Site: Intestine.

B: 4.70-4.99 mm x 1.60 x 1.85 mm; OS: 254-306 x 301-335; VS: 340-380 x 309-361; E: 40-54 x 17-25 (after Werneck & Silva, 2012).

## FAMILY TELORCHIIDAE (Looss, 1899) Dollfus, 1925

### LOEFGRENIA Travassos, 1919

*Loefgrenia loefgreni* Travassos, 1919 (Fig. 172)

**BRAZIL:** *Podocnemis unifilis* [Travassos, 1919b].

Site: Intestine.

B: 24-28 mm x 3.50 mm; OS: 1.10-1.30 mm in diameter; VS: 900-1,000 in diameter; E: 95 x 52 (after Travassos, 1919b).

### ORCHIDASMA Looss, 1900

*Orchidasma amphiorchis* (Braun, 1899) Looss, 1900 (Fig. 173)

**ARGENTINA:** *Caretta caretta* (= *Thalassochelys caretta*) [Boero & Led, 1974].

**BRAZIL:** *Chelonia mydas* [Freitas & Lent, 1938b].

**PERU:** *Chelonia mydas agassizzii* [Tantaléan, Sarmiento & Huiza 1992].

Sites: Stomach, intestine.

B: 4.47-13.15 mm x 1.24-1.74 mm; OS: 330-570 x 410-570; VS: 170-340 x 200-370; E: 40-62 x 27-40 (after Freitas & Lent, 1938b).

### PSEUDOTELORCHIS Yamaguti, 1971

*Pseudotelorchis caimanis* Catto & Amato, 1993 (Fig. 174)

**BRAZIL:** *Caiman crocodilus* [Ostrowski de Núñez, 2003]; *Caiman yacare* (= *Caiman crocodilus yacare*) [Catto & Amato, 1993b].

Site: Oviduct.

B: 6.00-8.82 mm x 1.28-2.15 mm; OS: 549-658 x 530-695; VS: 475-640 x 494-640; E: 25-35 x 11-13 (after Catto & Amato, 1993b).

\**Pseudotelorchis devincenzii* (Mañé-Garzón & Gil, 1961) Catto & Amato, 1993 (Fig. 175)

**URUGUAY:** *Hydromedusa tectifera* [Mañé-Garzón & Gil, 1961a].

Site: Intestine.

B: 3.96 mm x 0.55 mm; OS: 222 x 216; VS: 143 in diameter; E: 26 x 11 (after Mañé-Garzón & Gil, 1961a).

\*Remarks: Referred as *Telorchis devincenzii* by Mañé-Garzón & Gil, (1961a) and considered synonym of *Telorchis hagmanni* Lent & Freitas, 1937 by Nasir (1974).

*Pseudotelorchis yacarei* Catto & Amato, 1993 (Fig. 176)

**BRAZIL:** *Caiman crocodilus* [Ostrowski de Núñez, 2003]; *Caiman yacare* (= *Caiman crocodilus yacare*) [Catto & Amato, 1993b].

Site: Intestine.

B: 1.44-3.07 mm x 0.26-0.53 mm; OS: 204-394 x 190-423; VS: 94-182 x 87-182; E: 23-31 x 11-14 (after Catto & Amato, 1993b).

### **TELORCHIS Lühe, 1899**

*Telorchis achavali* Mañé-Garzón & Holcman-Spector, 1973 (Fig. 177)

**URUGUAY:** *Trachemys dorbigni* (= *Pseudemys dorbignyi*) [Mañé-Garzón & Holcman-Spector, 1973a].

Site: Intestine.

B: 8.26-10.52 mm x 0.50-0.77 mm; OS: 130-180 x 160-240; VS: 110-160 x 80-130; E: 30-32 x 15-17 (after Mañé-Garzón & Holcman-Spector, 1973a).

*Telorchis aculeatus* (Linstow, 1879) Braun, 1901 (Fig. 178)

**VENEZUELA:** *Podocnemis unifilis* (= *Podocnemis cayennensis*) [Nasir, 1974].

Site: Intestine.

B: 2.66-4.53 mm x 0.35-0.68 mm; OS and VS: 84-192 in diameter; E: mng (after Nasir, 1974)

*Telorchis bifurcus* (Braun, 1899) Braun, 1901 (Fig. 179)

**BRAZIL:** *Podocnemis expansa* [Braun, 1901].

Site: Intestine.

B: 10.00-13.00 mm x 1.40-1.60 mm; OS: 180-200 x 240-250; VS: 220-270 x 210-250; E: 23 x 14 (after Braun, 1901).

*Telorchis birabeni* Mañé-Garzón & Gil, 1961 (Fig. 180)

**ARGENTINA:** *Phrynops hilarii* [Lombardero & Moriena, 1977].

**URUGUAY:** *Phrynops hilarii* (= *Phrynops geoffroyana hilarii*) [Mañé-Garzón & Gil, 1961c].

Site: Intestine.

B: 3.97-5.02 mm x 0.53-0.57 mm; OS: 226-246 in diameter; VS: 80 x 120; E: 17 x 10 (after Mañé-Garzón & Gil, 1961c).

*Telorchis clava* (Diesing, 1850) Luehe, 1899 (Fig. 181)

**ARGENTINA:** *Eunectes notaeus* [Poumarau, 1968; Lunaschi & Drago, 2010]; *Hydrodynastes gigas* [Martínez, Troiano, Binda, Selles, Jara & Fescina, 1996].

**BRAZIL:** *Boiruna maculata* (= *Hydroscopis plumbeus*; *Oxyrhopus cloelia*; *Pseudoboa cloelia*) [Diesing, 1850; MacCallum, 1921]; *Constrictor constrictor* (= *Zamensis constrictor*) [after Travassos, Freitas & Kohn, 1969]; *Drymarchon corais* (= *Coluber flaviventris*; *Drymarchon corais corais*) [after Travassos, Freitas & Kohn, 1969]; *Eunectes murinus* (= *Boa scytale*, *Eunectes* sp.) [Gomes & Pinto, 1978].

Site: Intestine.

B: 6.00 mm x 1.40 mm; OS and VS: 560 in diameter; E: 80 x 40 (after MacCallum, 1921).

B: 4.96-5.54 mm x 1.50-1.66 mm; OS: 610-660 x 660-720; VS: 600-660 in diameter; E: 21-25 x 10-14 (after Gomes & Pinto, 1978).

\**Telorchis diaphanus* Freitas & Dobbin Jr., 1959 (Fig. 182)

**BRAZIL:** *Kinosternon scorpioides* (= *Kinosternon scorpioides scorpioides*) [Freitas & Dobbin Jr., 1959].

Site: Intestine.

B: 1.54-3.48 mm x 0.52-0.77 mm; OS: 92-139 x 139-168; VS: 109-155 x 122-160; E: 42-43 x 21-23 (after Freitas & Dobbin Jr., 1959).

\*Remarks: Considered synonym of *Telorchis aculeatus* (Von Linstow, 1879) Braun, 1901 by Nasir (1974).

*Telorchis dubius* Mañé-Garzón & Holcman-Spector, 1968 (Fig. 183)

**URUGUAY:** *Trachemys dorbigni* (= *Pseudemys dorbigni*) [Mañé-Garzón & Holcman-Spector, 1968].

Site: Intestine.

B: 3.44-4.00 mm x 0.67-0.93 mm; OS: 120-130 x 130-170; VS: 140-170 x 180-200; E: 32-35 x 18 (after Mañé-Garzón & Holcman-Spector, 1968).

*Telorchis hagmanni* Lent & Freitas, 1937 (Fig. 184)

**BRAZIL:** *Podocnemis expansa* [Lent & Freitas, 1937; Alho, 1965].

**COLOMBIA:** *Podocnemis lewyana* [Lenis & Vélez, 2011].

Sites: Stomach, intestine.

B: 18.50 mm x 2.26 mm; OS: 630 x 530; VS: 530 in diameter; E: 27-32 x 14-17 (after Lent & Freitas, 1937).

\**Telorchis parvus* Braun, 1901 (Fig. 185)

**BRAZIL:** *Emys orbicularis* (= *Cistudo lutaria*; *Emys lutaria*; *Testudo orbicularis*) [Braun, 1901].

Site: Intestine.

B: 2.00 mm x 0.34 mm; OS: 59 in diameter; VS: 59 x 54; E: 41 x 23 (after Braun, 1901).

\*Remarks: Considered synonym of *Telorchis diminutus* Stunkard, 1915 by Nasir (1974).

\**Telorchis platensis* Mañé-Garzón & Gil, 1961 (Fig. 186)

**URUGUAY:** *Hydromedusa tectifera* (= *Hidromedusa tectifera*) [Mañé-Garzón & Gil, 1961b].

Site: Intestine.

B: 2.18 mm x 0.44 mm; OS: 156 x 130; VS: 104 in diameter; E: 41 x 18 (after Mañé-Garzón & Gil, 1961b).

\*Remarks: Considered synonym of *Telorchis diminutus* Stunkard, 1915 by Nasir (1974).

*Telorchis pleroticus* (Braun, 1899) Braun, 1901 (Fig. 187)

**BRAZIL:** “freshwater turtle” [Braun, 1901].

Site: Intestine.

B: 6.00-8.00 mm x 0.26-0.39 mm; OS: 210-240 in diameter; VS: 100-110 in diameter; E: 19-23 x 9-13 (after Braun, 1901).

\**Telorchis productus* Mañé-Garzón & Gil, 1961 (Fig. 188)

**URUGUAY:** *Prynops hilarii* (= *Prynops geoffroyana hilarii*) [Mañé-Garzón & Gil, 1961b].

Site: Intestine.

B: 19.02 mm x 0.66 mm; OS: 308 in diameter; VS: 226 in diameter; E: 20 x 13 (after Mañé-Garzón & Gil, 1961b).

\*Remarks: Considered synonym of *Telorchis pleroticus* (Braun, 1899) Braun, 1901 by Nasir (1974).

\**Telorchis rapidulus* Dobbin Jr., 1957 (Fig. 189)

**BRAZIL:** *Kinosternon scorpioides* [Dobbin Jr., 1957c].

Site: Intestine.

B: 6.96 mm x 0.85 mm; OS: 150 in diameter; VS: 180 x 170; E: 36-48 x 19-29 (after Dobbin Jr., 1957c).

\*Remarks: Considered synonym of *Telorchis aculeatus* (Von Linstow, 1879) Braun, 1901 by Nasir (1974).

## FAMILY UROTREMATIDAE Poche, 1926

### UROTREMA Braun, 1900

*Urotrema shirleyae* Zamparo, Brooks & Tkach, 2005

**BRAZIL:** *Anolis fuscoauratus* (= *Norops fuscoauratus*) [Goldberg, Bursey & Vitt, 2006, wmd].

Site: Intestine.

**SUPERFAMILY PRONOCEPHALOIDEA Looss, 1899**

**FAMILY PRONOCEPHALIDAE Looss, 1902**

**ADENOGASTER Looss, 1901**

*Adenogaster serialis* Looss, 1901 (Fig. 190)

**PERU:** *Chelonia mydas agassizzii* [Tantaleán, Sarmiento & Huiza, 1992, wmd].  
Site: Intestine.

**CETIOSACCUS Gilbert, 1940**

*Cetiosaccus galapagensis* Gilbert, 1940 (Fig. 191)

**GALAPAGOS ISLANDS:** *Amblyrhynchus cristatus* [Gilbert, 1940; Vercammen-Grandjean & Lowenstein, 1967].

Site: Intestine.

B: 7.00-13.00 mm x 1.00-1.30 mm; OS: 300-420 in diameter; VS: absent; E: 19-23 x 9-11 (after Gilbert, 1940).

**CRICOCEPHALUS Looss, 1899**

*Cricocephalus albus* (Kuhl & Hasselt, 1822) Looss, 1899 (Fig. 192)

**BRAZIL:** *Chelonia mydas* [Ruiz, 1946].

Site: Digestive tract.

B: 5.31-6.81 mm x 1.34-2.05 mm; OS: 458-558 in diameter; VS: absent; E: 24-28 x 11-13 (after Ruiz, 1946).

**IGUANACOLA Gilbert, 1938**

*Iguanacola navicularius* Gilbert, 1940 (Fig. 193)

**GALAPAGOS ISLANDS:** *Amblyrhynchus cristatus* [Gilbert, 1940; Vercammen-Grandjean & Lowenstein, 1967].

Site: Intestine.

B: 6.63-8.80 mm x 1.30-2.00 mm; OS: 570 x 600; VS: absent; E: 18-22 x 8-13 (after Gilbert, 1940).

**METACETABULUM Freitas & Lent, 1938**

*Metacetabulum invaginatum* Freitas & Lent, 1938 (Fig. 194)

**BRAZIL:** *Chelonia mydas* (= *Chelone mydas*) [Freitas & Lent, 1938b; Ruiz, 1946].

Site: Intestine.

B: 6.05-7.50 mm x 1.22-1.34 mm; OS: 120-140 x 100-120; VS: 530 x 450; E: 35-40 x 11-13 (after Freitas & Lent, 1938b).

### **PLEUROGONIUS Looss, 1901**

*Pleurogonius linearis* Looss, 1901 (Fig. 195)

**BRAZIL:** *Chelonia mydas* (= *Chelone mydas*) [Ruiz, 1946].

Site: Intestine.

B: 3.65-4.66 mm x 9.93-2.20 mm; OS: 113-155 in diameter; VS: absent; E: 28-36 x 15-22 (after Ruiz, 1946).

*Pleurogonius lobatus* Looss, 1901 (Fig. 196)

**BRAZIL:** *Chelonia mydas* (= *Chelone mydas*) [Ruiz, 1946].

Sites: Stomach, intestine.

B: 4.70-4.90 mm x 0.80-0.99 mm; OS: 143-146 in diameter; VS: absent; E: 26-30 x 13-18 (after Ruiz, 1946).

*Pleurogonius longiusculus* (Beneden, 1859) Looss, 1901 (Fig. 197)

**BRAZIL:** *Chelonia mydas* (= *Chelone mydas*) [Ruiz, 1946].

Site: Intestine.

B: 8.00-8.30 mm x 0.89-1.05 mm; OS: 157 in diameter; VS: absent; E: 27 x 13 (after Ruiz, 1946).

*Pleurogonius trigonocephalus* (Rudolphi, 1809) Looss, 1901 (Fig. 198)

**BRAZIL:** *Chelonia mydas* (= *Chelone mydas*) [Ruiz, 1946].

Site: Digestive tract.

B: 3.00-3.50 mm x less than 1.00 mm; OS: mng; VS: absent; E: 28 x 14 (after Ruiz, 1946).

### **PRONOCEPHALUS Looss, 1899**

\**Pronocephalus obliquus* Looss, 1901 (Fig. 199)

**BRAZIL:** *Caretta caretta* (= *Halichelis atra*; *Thalassochelys caretta*) [after Viana, 1924]; *Chelonia mydas* [Ruiz, 1946].

Sites: Intestine, oesophagus, stomach.

B: 3.25-5.84 mm x 1.09-1.69 mm; OS: 157-240 in diameter; VS: absent; E: 19-24 x 11-14 (after Ruiz, 1946).

\*Remarks: Referred as *Pronocephalus trigonocephalus* Looss, 1899 by Ruiz (1946).



## **PYELOSOMUM Looss, 1899**

\**Pyelosomum amblyrhynchi* (Gilbert, 1940) (Fig. 200)

**GALAPAGOS ISLANDS:** *Amblyrhynchus cristatus* [Gilbert, 1940; Ruiz, 1946; Vercammen-Grandjean & Lowenstein, 1967].

Site: Intestine.

B: 3.27-4.66 mm x 1.45-1.78 mm; OS: 330 x 340; VS: absent; E: 59-63 x 25-36 (after Gilbert, 1940).

\*Remarks: Referred as *Myosaccus amblyrhynchi* by Gilbert (1940) and by Ávila & Silva (2010). The genus *Myosaccus* Gilbert, 1940 was considered synonym of *Pyelosomum* Looss, 1899 by Pérez Ponce de León & Brooks (1995) and accepted by Blair (2005a).

*Pyelosomum crassum* (Looss, 1901) Ruiz, 1946 (Fig. 201)

**BRAZIL:** *Chelonia mydas* [Ruiz, 1946].

Site: Intestine.

B: 8.37-8.93 mm x 2.13-2.26 mm; OS: 42-52 x 19-22; VS: absent; E: 42-52 x 19-22 (after Ruiz, 1946).

*Pyelosomum longiusculus* Looss, 1901

**BRAZIL:** *Chelonia mydas* [Thatcher, 1993].

Site: Intestine.

B: 8.00-10.00 mm x 0.80-0.90 mm; OS and VS: absent; E: 27-28 x 13-15 (after Thatcher, 1993).

## **RUICEPHALUS Skrjabin, 1955**

\**Ruicephalus minutus* (Ruiz, 1946) Skrjabin, 1955 (Fig. 202)

**BRAZIL:** probably *Chelonia mydas* [Ruiz, 1946].

Site: Intestine.

B: 2.56-2.69 mm x 0.99-1.03; OS: 155 in diameter; VS: absent; E: 14-19 x 11 (after Ruiz, 1946).

\*Remarks: Described as *Pronocephalus minutus* by Ruiz (1946).

## **SUPERFAMILY SCHISTOSOMATOIDEA Stiles & Hassall, 1898**

### **FAMILY SPIRORCHIIDAE Stunkard, 1921**

#### **AMPHIORCHIS Price, 1934**

*Amphiorchis caborojoensis* Fischthal & Acholonu, 1976 (Fig. 203)

**BRAZIL:** *Eretmochelys imbricata* [Werneck, Gallo & Silva, 2008b; Werneck, Lima, Gallo & Silva, 2011].

Sites: Intestine, liver.

B: 5.55-5.88 mm x 0.94-1.064 mm; OS: 212-221 x 196-210; VS: 277-294 x 212-300; E: 25-38 x 48-64 with filaments (after Werneck, Gallo & Silva, 2008b).

*Amphiorchis indicus* Mehrotra, 1973 (Fig. 204)

**BRAZIL:** *Chelonia mydas* [Werneck & Silva, 2013].

Sites: liver, oesophagus, stomach, intestine.

B: 3.59-5.04 mm x 0.29-0.39 mm; OS: 70-122 x 54-83; VS: 77-167 x 55-148; E: 138-197 x 20- (after Werneck & Silva, 2013).

*Amphiorchis solus* (Simha & Chattopadhyaya, 1970) Platt, 2002 (Fig. 205)

**BRAZIL:** *Chelonia mydas* [Werneck, Lima, Gallo & Silva, 2011].

Site: Heart.

B: 2.86 mm x 0.55 mm; OS: 160 x 140; VS: 230 x 970; E: mng.

### **CARETTACOLA** Manter & Larson, 1950

*Carettacola stunkardi* (Martin & Bamberger, 1952) Dailey, Fast & Balazs, 1991

**BRAZIL:** *Chelonia mydas* [Werneck, Baldassin, Torres, Trazi & Berger, 2013]; *Eretmochelys imbricata* [Werneck, Gallo & Silva, 2008b].

Site: Body wash.

B: 6.51 mm x 0.47 mm; OS: 100 x 91; VS: 248 x 257; E: mng (after Werneck, Gallo & Silva, 2008b).

### **LEAREDIUS** Price, 1934

*Learedius learedi* Price, 1934 (Fig. 206)

**BRAZIL:** *Chelonia mydas* [Werneck, Becker, Gallo & Silva, 2006].

Sites: Heart, liver, spleen, lungs, kidneys, mesenterium.

B: 3.25-5.43 mm x 0.681-1.248 mm; OS: 287-365 x 257-324; VS: 445-498 x 475-504; E: 279-356 x 44-60 (after Werneck, Becker, Gallo & Silva, 2006).

### **MONTICELLIUS** Mehra, 1939

*Monticellius indicum* Mehra, 1939 (Fig. 207)

**BRAZIL:** *Chelonia mydas* [Werneck, Gallo & Silva, 2008a].

Site: Heart.

B: 3.67-5.65 mm x 0.36-0.43 mm; OS: 86-185 x 144-194; VS: 164-318 x 131-194; E: mng  
(after Werneck, Gallo & Silva, 2008a).

### **Especies dubiae**

#### ***Distoma pyxidatum* Bremser in Rudolphi, 1819**

**BRAZIL:** *Caiman crocodilus* (= *Caiman sclerops*) [Bremser in Rudolphi, 1819].  
Site: Intestine.



# HOST-PARASITE LIST

## AMPHIBIA

### BUFONIDAE

*Anaxyrus terrestris* (Bonnaterre, 1789) [=*Bufo musicus* Bonnaterre, 1789]  
*Rauschiella linguatula*

*Atelopus bomolochus* Peters, 1973  
*Gorgoderina parvicava*

*Atelopus ignescens* (Cornalia, 1849) [=*Atelopus laevis* Cope, 1868]  
*Gorgoderina cryptorchis*  
*Rudolphitrema rudolphii*

*Incilius nebulifer* (Girard, 1854) [=*Bufo granulatus* Baird and Girard, 1852]  
*Choledocystus hepaticus*  
*Maicuru solitarium*  
*Mesocoelium monas*  
*Rauschiella linguatula*  
*Rauschiella palmipedis*

*Melanophryniscus stelzneri* (Weyenbergh, 1875)  
*Gorgoderina darwini*

*Rhinella arenarum* (Hensel, 1867) [=*Bufo arenarius* after Lutz, 1934; *Bufo arenarum* Hensel, 1867]  
*Gorgoderina rochalimai*  
*Glypthelmins festina*  
*Mesocoelium monas*

*Rhinella crucifer* (Wied-Neuwied, 1821) [=*Bufo crucifer* Wied-Neuwied, 1821]  
*Gorgoderina cryptorchis*  
*Gorgoderina parvicava*  
*Mesocoelium monas*  
*Rauschiella linguatula*  
*Rudolphitrema rudolphii*

*Rhinella dorbignyi* (Duméril & Bibron, 1841) [=*Bufo dorbigny* Cope 1885]  
*Gorgoderina cryptorchis*

**Rhinella fernandezae** (Gallardo, 1957) [=*Bufo fernandezae* Cei, 1964]

*Catadiscus inopinatus*  
*Catadiscus marinholutzi*  
*Gorgoderina festoni*  
*Gorgoderina parvicava*  
*Haematoloechus longiplexus*  
*Rauschiella palmipedis*

**Rhinella icterica** (Spix, 1824) [=*Bufo ictericus* Spix, 1824; *Bufo marinus ictericus* Müller, 1927; *Chaunus ictericus* (Spix, 1824)]

*Catadiscus cohni*  
*Gorgoderina parvicava*  
*Haematoloechus fuelleborni*  
*Mesocoelium meggitti*  
*Mesocoelium monas*  
*Rauschiella linguatula*  
*Rauschiella palmipedis*  
*Rudolphitrema rudolphii*

**Rhinella limensis** (Werner, 1901) [=*Bufo limensis* Werner, 1901, *Bufo spinulosus limensis* Vellard, 1959]

*Gorgoderina parvicava*

**Rhinella marina** (Linnaeus, 1758) [=*Bufo horribilis* Wiegmann, 1833; *Bufo marinus* Schneider, 1799; *Bufo marinus bimaculatus* (Schneider, 1799); *Bufo marinus marinus* Schmidt, 1932; *Bufo aqua* Latreille, 1801; *Chaunus marinus* (Linnaeus, 1758)]

*Catadiscus cohni*  
*Choledocystus hepaticus*  
*Creptotrema lynchi*  
*Gorgoderina cryptorchis*  
*Gorgoderina diaster*  
*Gorgoderina parvicava*  
*Haematoloechus fuelleborni*  
*Mesocoelium lanfrediae*  
*Mesocoelium meggitti*  
*Mesocoelium monas*  
*Mesocoelium waltoni*  
*Pseudosonsinotrema chabaudi*  
*Rauschiella linguatula*  
*Rauschiella palmipedis*  
*Rauschiella robusta*

**Rhinella schneideri** (Werner, 1894) [=*Bufo schneideri* Werner, 1894; *Bufo paracnemis* Lutz, 1925].

*Catadiscus freitaslenti*

*Gorgoderina parvicava*  
*Gorgoderina rochalimai*  
*Mesocoelium monas*  
*Rauschiella linguatula*  
*Rauschiella palmipedis*

*Rhinella* sp. [=*Bufo* sp.]  
*Mesocoelium meggitti*

## CAECILIIDAE

*Chthonerpeton indistinctum* Reinhardt & Lütken, 1862  
*Glyphelmins sanmartini*  
*Rauschiella linguatula*

*Siphonops annulatus* (Mikan, 1820)  
*Mesocoelium monas*

## CALYPTOCEPHALELIIDAE

*Calyptocephalella gayi* (Duméril & Bibron, 1841) [= *Caudiverbera caudiverbera* Myers, 1962]  
*Gorgoderina valdiviensis*

## CERATOPHRYIDAE

*Ceratophrys cornuta* Linnaeus, 1758  
*Rauschiella linguatula*

*Ceratophrys cranwelli* Barrio, 1980  
*Haematoloechus longipectus*

*Telmatobius brachydactylus* (Peters, 1873) [= *Batrachophrynus brachydactylus* Peters, 1873]  
*Gorgoderina chilensis*

*Telmatobius culeus* (Garman, 1876)  
*Gorgoderina parvicava*

*Telmatobius jelskii* (Peters, 1873)  
*Gorgoderina parvicava*

*Telmatobius macrostomus* (Peters, 1873) [=*Batrachophrynus macrostomus* Peters, 1873]

*Gorgoderina parvicava*

*Gorgoderina parvicava minuta*

*Telmatobius peruvianus* Wiegmann, 1834

*Gorgoderina parvicava*

*Haematoloechus arequipensis*

*Haematoloechus pukinensis*

*Telmatobius* sp.

*Gorgoderina parvicava*

*Gorgoderina* sp.

## CYCLORAMAPHIDAE

*Alsodes roseus* [=*Eupsophus roseus* (Duméril and Bibron, 1841)]

*Rudolphitrema chilensis*

*Odontophrynus americanus* (Duméril & Bibron, 1841)

*Rauschiella repandum*

*Rhinoderma darwini* Duméril and Bibron, 1841

*Gorgoderina chilensis*

## HEMIPHRACTIDAE

*Gastrotheca pseustes* Duellman & Hillis, 1987

*Pseudosonsinotrema megalorchis*

## HYLIDAE

*Dendropsophus leucophyllatus* (Beireis, 1783) [=*Hyla leucophyllata* (Beireis, 1783)]

*Glyphelmis parva*

*Hypsiboas crepitans* (Wied-Neuwied, 1824) [=*Hyla crepitans* Wied-Neuwied, 1824]

*Choledocystus hepaticus*

*Hypsiboas pulchellus* (Duméril & Bibron, 1841) [=*Hyla pulchella* Duméril & Bibron, 1841]

*Catadiscus hylae*

*Catadiscus uruguayensis*

*Traxtrema stenocotyle*



***Hypsiboas raniceps*** [=*Hyla raniceps* (Cope, 1862)]

*Choledocystus vitellinophilum*

***Lysapus limellum*** Cope, 1862 [=*Lysapus limellus* Cope, 1962]

*Catadiscus propinquus*

*Catadiscus uruguayensis*

*Choledocystus vitellinophilum*

***Phyllomedusa azurea*** Cope, 1862

*Catadiscus uruguayensis*

***Pseudis minuta*** Günther, 1858 [=*Lysapsus mantidactylus* Gallardo, 1961; *Pseudis mantidactylus* Boulenger, 1882; *Pseudis meridionalis* Miranda-Ribeiro, 1926]

*Catadiscus corderoi*

*Catadiscus uruguayensis*

*Choledocystus pseudium*

***Pseudis paradoxa*** (Linnaeus, 1758)

*Catadiscus pygmaeus*

*Choledocystus incurvatum*

*Gorgoderina diaster*

*Gorgoderina parvicava*

*Neohaematoloechus neivai*

*Rauschiella palmipedis*

***Pseudis platensis*** Gallardo, 1961

*Catadiscus propinquus*

*Rauschiella palmipedis*

***Scinax nasicus*** (Cope, 1862)

*Catadiscus inopinatus*

*Travtrema stenocotyle*

***Scinax pedromedinai*** (Henle, 1991)

*Glypthelmins parva*

***Trachycephalus coriaceus*** (Peters, 1867) [=*Phrynohyas coriacea* Duellman, 1968]

*Glypthelmins parva*

## HYLODIDAE

***Hylodes nasus*** (Lichtenstein, 1823) [=*Elosia nasus* Günther, 1866]

*Gorgoderina cedroi*

*Opisthioglyphe amplicavus*

## LEIUPERIDAE

*Physalaemus gracilis* (Boulenger, 1883)  
*Rudolphitrema physalaemi*

*Physalaemus santafecinus* Barrio, 1965  
*Catadiscus inopinatus*  
*Rauschiella repandum*

*Pseudopaludicola boliviana* Parker, 1927  
*Gorgoderina cryptorchis*  
*Haematoloechus longiplexus*

*Leptodactylus bolivianus* Boulenger, 1898  
*Rauschiella linguatula*  
*Rauschiella palmipedis*

*Leptodactylus bufonius* Boulenger, 1894  
*Catadiscus inopinatus*  
*Rauschiella repandum*

*Leptodactylus chaquensis* Cei, 1950  
*Catadiscus inopinatus*  
*Catadiscus propinquus*  
*Gorgoderina parvicava*  
*Gorgoderina rochalimai*  
*Haematoloechus longiplexus*  
*Rauschiella palmipedis*  
*Rauschiella repandum*

*Leptodactylus fuscus* (Schneider, 1799) [= *Leptodactylus sibilatrix* Fitzinger, 1826;  
*Leptodactylus typhonius* Boulenger, 1882]  
*Catadiscus freitaslenti*  
*Mesocoelium monas*  
*Rauschiella palmipedis*

*Leptodactylus labyrinthicus* (Spix, 1824) [= *Leptodactylus pentadactylus labyrinthicus*  
Müller, 1927]  
*Gorgoderina parvicava*  
*Neohaematoloechus neivai*  
*Rauschiella linguatula*  
*Rauschiella palmipedis*

*Leptodactylus laticeps* (Boulenger, 1918)  
*Rauschiella chaquensis*

***Leptodactylus latinasus*** Jiménez de la Espada, 1875

*Catadiscus inopinatus*

*Haematoloechus longipectus*

*Rauschiella repandum*

***Leptodactylus latrans*** (Steffen, 1815) [= *Leptodactylus caliginosus* Girard, 1853, *Leptodactylus ocellatus* (Linnaeus, 1758)]

*Catadiscus corderoi*

*Catadiscus eldoradiensis*

*Catadiscus freitaslenti*

*Catadiscus inopinatus*

*Catadiscus marinholutzi*

*Catadiscus uruguayensis*

*Choledocystus simulans*

*Gorgodera australiensis*

*Gorgoderina carioca*

*Gorgoderina cryptorchis*

*Gorgoderina megacysta*

*Gorgoderina parvicava*

*Gorgoderina pigulevskyi*

*Gorgoderina rochalimai*

*Glypthelmins biliaris*

*Glypthelmins parva*

*Haematoloechus freitasi*

*Haematoloechus fuelleborni*

*Haematoloechus legrandi*

*Haematoloechus longipectus*

*Haematoloechus ozorioi*

*Halipegus dubius*

*Mesocoelium monas*

*Neohaematoloechus neivai*

*Plagiorchis rangeli*

*Rauschiella lenti*

*Rauschiella linguatula*

*Rauschiella palmipedis*

*Rauschiella proxima*

*Rauschiella repandum*

***Leptodactylus martinezi*** (Bokermann, 1956)

*Brachycoelium salamandrae*

***Leptodactylus mystaceus*** (Spix, 1824)

*Mesocoelium monas*

***Leptodactylus mystacinus*** (Burmeister, 1861)  
*Mesocoelium monas*

***Leptodactylus pentadactylus*** (Laurenti, 1768)  
*Gorgoderina parvicava*  
*Halipegus dubius*  
*Mesocoelium monas*  
*Neohaematoloechus neivai*  
*Rauschiella linguatula*

***Leptodactylus podicipinus*** (Cope, 1862)  
*Catadiscus propinquus*  
*Infidum infidum*  
*Travtrema stenocotyle*

***Leptodactylus rhodomystax*** Boulenger, 1883  
*Brachycoelium salamandrae*

***Leptodactylus rhodonotus*** (Günther, 1869)  
*Gorgoderina parvicava*

## PIPIDAE

***Pipa carvalhoi*** (Miranda-Ribeiro, 1937) [= *Hemipipa carvalhoi* Miranda-Ribeiro, 1937]  
*Catadiscus mirandai*

## RANIDAE

***Rana palmipes*** (Spix, 1824)  
*Catadiscus propinquus*  
*Gorgoderina diaster*  
*Gorgoderina parvicava*  
*Haematoloechus lutzi*  
*Iquitos ceii*  
*Loxogenes macrocirra*  
*Neohaematoloechus iturbei*  
*Neohaematoloechus neivai*  
*Rauschiella palmipedis*

***Rana*** sp.  
*Gorgoderina diaster*  
*Haematoloechus lutzi*

# REPTILIA

## ALLIGATORIDAE

***Caiman crocodilus*** (Linnaeus, 1758) [=*Caiman sclerops* Schneider 1801, *Crocodilus* sp.]

*Acanthostomum scyphocephalum*

*Caimanicola marajoara*

*Cyathocotyle brasiliensis*

*Cystodiplostomum hollyi*

*Distoma pyxidatum*

*Herpetodiplostomum caimancola*

*Odhneriotrema microcephala*

*Pachypsolus sclerops*

*Paradiplostomum abbreviatum*

*Prolecithodiplostomum constrictum*

*Proterodiplostomum longum*

*Proterodiplostomum medusae*

*Proterodiplostomum tumidulum*

*Pseudotelorchis caimanis*

*Pseudotelorchis yacarei*

*Stephanoprora campomica*

*Stephanoprora jacaretinga*

*Stephanoprora* sp.

***Caiman crocodilus crocodilus*** (Linnaeus, 1758)

*Proterodiplostomum medusae*

*Timoniella incognita*

*Timoniella ostrowskiae*

***Caiman crocodilus fuscus*** (Cope, 1868)

*Massoprostatum longum*

*Prolecithodiplostomum constrictum*

***Caiman latirostris*** (Daudin, 1802)

*Cystodiplostomum hollyi*

*Herpetodiplostomum caimancola*

*Paradiplostomum abbreviatum*

***Caiman yacare*** (Daudin, 1802) [=*Caiman crocodilus yacare* Müller & Hellmich 1936]

*Caimanicola marajoara*

*Cyathocotyle brasiliensis*

*Cystodiplostomum hollyi*

*Herpetodiplostomum caimancola*

*Pachypsolus sclerops*  
*Paradiplostomum abbreviatum*  
*Proctocaecum dorsale*  
*Prolecithodiplostomum constrictum*  
*Proterodiplostomum breve*  
*Proterodiplostomum globulare*  
*Proterodiplostomum longum*  
*Proterodiplostomum medusae*  
*Proterodiplostomum tumidulum*  
*Pseudotelorchis caimanis*  
*Pseudotelorchis yacarei*  
*Stephanoprora jacaretinga*

**Caiman** sp. [=Crocodylus sp.]  
*Crocodylicola pseudostoma*  
*Mesodiplostomum gladiolum*  
*Prolecithodiplostomum constrictum*  
*Proterodiplostomum longum*  
*Proterodiplostomum medusae*

**Melanosuchus niger** Spix, 1825  
*Caballerotrema* sp.  
*Echinostoma* sp.  
*Herpetodiplostomum caimancola*  
*Mesodiplostomum gladiolum*  
*Proterodiplostomum longum*  
*Stephanoprora nattereri*

**Paleosuchus palpebrosus** (Cuvier, 1807)  
*Pachypsolus sclerops*

**Paleosuchus** sp. [=Crocodylus coroa]  
*Caimanicola marajoara*  
*Proterodiplostomum longum*

## AMPHISBAENIDAE

**Amphisbaena alba** Linnaeus, 1758  
*Haplometroides buccicola*  
*Pneumotrema trivasossi*

**Amphisbaena ridleyi** Boulenger, 1890  
*Mesocoelium monas*

*Amphisbaena* sp.  
*Mesocoelium monas*

*Leposternon microcephalum* Wagler, 1824  
*Mesocoelium monas*

## ANGUIDAE

*Diploglossus lessonae* (Peracca, 1890)  
*Mesocoelium monas*

## BOIDAE

*Boa constrictor* Linnaeus, 1758 [= *Constrictor constrictor* Stull, 1935]  
*Paradistomum boae*  
*Styphlodora horrida*

*Epicrates crassus* Cope, 1862 [= *Epicrates cenchria crassus* Amaral 1929]  
*Haplometroides buccicola*

*Eunectes deschauenseei* Dunn & Conant, 1936  
*Ophiodiplostomum spectabile*

*Eunectes murinus* (Linnaeus, 1758)  
*Infidum infidum*  
*Telorchis clava*

*Eunectes notaeus* Cope, 1862  
*Infidum infidum*  
*Styphlodora condita*  
*Telorchis clava*

## CHELIDAE

*Mesoclemmys nasuta* (Schweigger, 1812) [= *Batrachemys nasuta* Stejneger 1909; *Rhynemis nasuta* Boulenger 1889]  
*Nematophila grandis*

*Chelus fimbriatus* Schneider, 1783 [= *Chelus fimbriata* Schneider, 1783; *Chelys fimbriata* Günther, 1882; *Testudo matamata* Schneider, 1783]  
*Acanthostomum scyphocephalum*  
*Nematophila grandis*

***Hydraspis schopjii***

*Nematophila grandis*

***Hydromedusa tectifera*** Cope, 1869 [=*Hidromedusa tectifera* Cope, 1869]

*Nematophila grandis*

*Pseudotelorchis devincenzii*

*Telorchis platensis*

***Mesoclemmys gibba*** Gray, 1873 (= *Phrynops gibbus* (Gaffney, 1977; *Hydraspis gibba* Boulenger, 1889])

*Nematophila grandis*

***Phrynops geoffroanus*** (Schweigger, 1812) [= *Hydraspis geoffroyana* Boulenger, 1889; *Phrynops geoffroanus geoffroanus* (Schweigger, 1812); *Phrynops geoffroyana* (Mertens et al. 1934; *Phrynops geoffroyana geoffroyana* (Schweigger, 1812))]

*Nematophila grandis*

*Prionosomoides scalaris*

***Phrynops hilarii*** (Duméril & Bibron, 1835) [= *Phrynops geoffroyana hillarii* Lüling, 1984]

*Acanthostomum scyphocephalum*

*Caimanicola brauni*

*Cheloniodiplostomum testudinis*

*Nematophila grandis*

*Prionosoma phrynopsis*

*Prionosomoides scalaris*

*Telorchis birabeni*

*Telorchis productus*

*Timoniella ostrowskiae*

***Phrynops*** sp. (cited as probably *Hydraspis* sp.)

*Nematophila argentinum*

*Nematophila venezuelensis*

## CHELONIIDAE

***Caretta caretta*** (Linnaeus, 1758) [= *Halichelis atra*]; *Thalassochelys caretta* Boulenger 1886]

*Lophotaspis vallei*

*Orchidasma amphiorchis*

*Pronocephalus obliquus*

*Rhytidodes gelatinosus*

***Chelonia mydas*** Linnaeus, 1758 [= *Chelone mydas* Brongniart, 1805]

*Amphiorchis indicus*

*Amphiorchis solus*



*Carettacola stunkardi*  
*Cricocephalus albus*  
*Learedius learedi*  
*Metacetabulum invaginatum*  
*Monticellius indicum*  
*Orchidasma amphiorchis*  
*Pleurogonius linearis*  
*Pleurogonius lobatus*  
*Pleurogonius longiusculus*  
*Pleurogonius trigonocephalus*  
*Polyangium linguatula*  
*Pronocephalus obliquus*  
*Pyelosomum crassum*  
*Pyelosomum longiusculus*  
*Ruicephalus minutus*  
*Rhytidodes gelatinosus*

***Chelonia mydas agassizzii*** Bocourt, 1868  
*Adenogaster serialis*  
*Orchidasma amphiorchis*

***Eretmochelys imbricata*** (Linnaeus, 1758)  
*Amphiorchis caborojoensis*  
*Carettacola stunkardi*  
*Styphlotrema solitaria*

## COLUBRIDAE

***Atractus lasallei*** Amaral, 1931  
*Renifer heterocoelium*

***Boiruna maculata*** (Boulenger, 1896)  
*Opisthogonimus lecithonotus*  
*Telorchis clava*

***Chironius bicarinatus*** (Wied, 1820)  
*Opisthogonimus fonsecai*  
*Renifer heterocoelium*  
*Styphlodora gili*

***Chironius carinatus*** (Linnaeus, 1758)  
*Opisthogonimus afranioi*  
*Opisthogonimus pereirai*  
*Paradistomum parvissimum*

*Renifer chironius*  
*Renifer heterocoelium*  
*Sticholecitha serpentis*

***Chironius exoletus*** (Linnaeus, 1758)  
*Renifer heterocoelium*

***Chironius foveatus*** Bailey, 1955  
*Opisthognimus fonsecai*

***Chironius fuscus*** Linnaeus, 1758 [=*Herpetodryas fuscus* Duméril & Bibron, 1854]  
*Catadiscus dolichocotyle*  
*Leurosoma rudolfbarthi*  
*Renifer heterocoelium*  
*Travtrema stenocotyle*

***Clelia clelia*** (Daudin, 1803) [=*Cloelia cloelia* Daudin, 1803; =*Pseudoboa cloelia* Serio, 1921]  
*Opisthognimus lecithonotus*

***Clelia rustica*** (Cope, 1878)  
*Travtrema stenocotyle*

***Coluber*** sp.  
*Heterodiplostomum lanceolatum*  
*Ophiodiplostomum ancyloides*  
*Ophiodiplostomum spectabile*

***Drymarchon corais*** (Boie, 1827) [=*Drymarchon corais corais* Amaral, 1929]  
*Acanthostomum scyphocephalum*  
*Infidum similis*  
*Renifer heterocoelium*  
*Telorchis clava*  
*Timoniella incognita*  
*Timoniella ostrowskiae*

***Erythrolamprus aesculapii*** (Linnaeus, 1758)  
*Catadiscus freitaslenti*  
*Haplometroides buccicola*  
*Opisthognimus fonsecai*  
*Renifer heterocoelium*

***Helicops carinicaudus*** (Wied-Neuwied, 1825 [=*Helicops carinicauda* Boulenger, 1893])  
*Opisthognimus lecithonotus*  
*Opisthognimus serpentis*

***Helicops infrataeniatus*** Jan, 1865  
*Heterodiplostomum helicopsis*  
*Heterodiplostomum lanceolatum*  
*Opisthognomus lecithonotus*  
*Opisthognomus serpentis*

***Helicops leopardinus*** (Schlegel, 1837) [=*Helicops leopardina* Amaral, 1929]  
*Heterodiplostomum lanceolatum*  
*Opisthognomus lecithonotus*

***Helicops modestus*** Günther, 1861  
*Renifer heterocoelium*

***Hydrodynastes gigas*** (Duméril, Bibron & Duméril, 1854) [=*Cyclagras gigas* Cope, 1885]  
*Glossidiella ornata*  
*Glossidioides loossi*  
*Heterodiplostomum lanceolatum*  
*Infidum infidum*  
*Opisthognomus lecithonotus*  
*Plagiorchis lühei*  
*Styphlodora condita*  
*Telorchis clava*

***Leptodeira annulata*** (Linnaeus, 1758) [=*Leptodeira annulata pulchriceps* Duellman, 1958;  
*Leptodeira annulata annulata* (Linnaeus, 1758)]  
*Styphlodora condita*

***Leptodeira septentrionalis*** Kennicott, 1859  
*Renifer heterocoelium*

***Leptophis ahaetulla*** (Linnaeus, 1758)  
*Renifer heterocoelium*

***Liophis almadensis*** (Wagler, 1824) [=*Leimadophis almada* (Wagler, 1824)]  
*Catadiscus Freitaslenti*  
*Renifer heterocoelium*

***Liophis jaegeri*** (Günther, 1858)  
*Opisthognomus megabothrium*  
*Travtrema stenocotyle*

***Liophis miliaris*** (Linnaeus, 1758) [=*Liophis miliaris miliaris* (Linnaeus, 1758)]  
*Aliptrema riberoi*  
*Bieria artigasi*  
*Catadiscus dolichocotyle*

*Catadiscus freitaslenti*  
*Infidum similis*  
*Liophistrema pulmonale*  
*Ophiodiplostomum spectabile*  
*Opisthognomus fariai*  
*Opisthognomus fonsecai*  
*Opisthognomus megabothrium*  
*Opisthognomus serpentis*  
*Renifer heterocoelium*  
*Travtrema stenocotyle*

***Liophis miliaris semiaureus*** (Cope, 1862)

*Opisthognomus fonsecai*  
*Travtrema stenocotyle*

***Liophis poecilogyrus*** (Wied-Neuwied, 1825) [= *Leimadophis poecilogyrus* Amaral, 1944]

*Catadiscus uruguayensis*  
*Infidum similis*  
*Mesocoelium monas*  
*Ophiodiplostomum spectabile*  
*Opisthognomus fonsecai*  
*Opisthognomus serpentis*  
*Renifer heterocoelium*  
*Travtrema stenocotyle*

***Liophis poecilogyrus reticulatus*** (Parker, 1931)

*Heterodiplostomum lanceolatum*

***Liophis reginae*** (Linnaeus, 1758)

*Ophiodiplostomum spectabile*  
*Renifer heterocoelium*

***Liophis typhlus*** (Linnaeus, 1758) [= *Dromicus typhlus* (Linnaeus, 1758)]

*Catadiscus rochai*  
*Ophiodiplostomum spectabile*  
*Opisthognomus fonsecai*  
*Renifer heterocoelium*

***Lygophis anomalus*** (Günther, 1858) [= *Liphis anomalus* Amaral 1925]

*Opisthognomus lecithonotus*  
*Paracotyletrema poncedeleoni*  
*Travtrema stenocotyle*

***Lygophis flavifrenatus*** Cope, 1862 [= *Liophis flavifrenatus* Cope, 1862]

*Catadiscus freitaslenti*

*Opisthognomus lecithonotus*  
*Travtrema stenocotyle*

***Lygophis typhlus*** (Linnaeus, 1758) [= *Leimadophis typhlus* Peters & Orejas Miranda, 1970]  
*Opisthognomus artigasi*  
*Opisthognomus lecithonotus*

***Mastigodryas bifossatus*** (Raddi, 1820) [= *Dryadophis bifossatus* (Raddi, 1820), *Drymobius bifossatus* Boulenger, 1894; *Eudryas bifossatus*]  
*Heterodiplostomum lanceolatum*  
*Infidum similis*  
*Mesocoelium sibynomorphi*  
*Ophiodiplostomum spectabile*  
*Opisthognomus artigasi*  
*Opisthognomus fonsecai*  
*Opisthognomus lecithonotus*  
*Paradistomum parvissimum*  
*Rauschiella linguatula*  
*Styphlodora condita*  
*Travtrema stenocotyle*

***Mastigodryas bifossatus triseriatus*** (Amaral, 1931)  
*Opisthognomus artigasi*  
*Opisthognomus lecithonotus*  
*Styphlodora condita*  
*Travtrema stenocotyle*

***Oxyrhopus rhombifer*** Duméril, Bibron & Duméril, 1854  
*Styphlodora condita*

***Phalotris lativittatus*** Ferrarezzi, 1993  
*Haplometroides buccicola*

***Phalotris matogrossensis*** Lema, D'Agostini & Cappelari, 2005  
*Haplometroides intercaecalis*

***Phalotris nasutus*** (Gomes, 1915)  
*Haplometroides intercaecalis*

***Philodryas hoodensis*** (Van Denburgh, 1912) [= *Leimadophis chamissonis*, *Oreophis* (*Driomicus*) *hoodensis*]  
*Infidum luckeri*

***Philodryas olfersii*** (Lichtenstein, 1823)  
*Catadiscus longicoecalis*

*Infidum similis*

***Philodryas patagoniensis*** (Girard, 1857) [=*Philodryas schottii* Günther, 1858]

*Opisthognomus afranioi*  
*Opisthognomus fonsecai*  
*Opisthognomus interrogativus*  
*Opisthognomus lecithonotus*  
*Opisthognomus serpentis*  
*Opisthognomus sulina*  
*Paradistomum parvissimum*  
*Renifer heterocoelium*  
*Styphlodora condita*  
*Travtrema stenocotyle*

***Philodryas psammophidea*** [=*Philodryas psammophideus* Günther, 1872]

*Infidum infidum*  
*Opisthognomus artigasi*

***Philodryas* sp.**

*Catadiscus dolichocotyle*  
*Infidum similis*  
*Ophiodyplostomum spectabile*  
*Opisthognomus lecithonotus*  
*Travtrema stenocotyle*

***Pseudoboa coronata*** Schneider, 1801 [=*Clelia occipitolutea* (Duméril, Bibron & Duméril, 1854)]

*Opisthognomus fonsecai*  
*Renifer heterocoelium*

***Sibynomorphus mikanii*** Schlegel, 1837 [=*Sibynomorphus mikanii mikanii* Peters & Orejas-Miranda, 1970]

*Mesocoelium sibynomorphi*

***Sibynomorphus turgidus*** (Cope, 1868)

*Mesocoelium monas*

***Sibynomorphus ventrimaculatus*** (Boulenger, 1885)

*Mesocoelium monas*  
*Travtrema stenocotyle*

***Sibynomorphus* sp.**

*Mesocoelium monas*

***Spilotes pullatus*** (Linnaeus, 1758) [=*Coluber pullatus* Wagler, 1830]

*Cotylotretus rugosus*

*Styphlodora condita*

***Thamnodynastes hypoconia*** (Cope, 1860) [=*Thamnodynastes strigilis* Peters & Orejas-Miranda 1970]

*Opisthogonimus uruguayensis*

***Thamnodynastes pallidus*** (Linnaeus, 1758) [=*Dryophylax pallidus* (Linnaeus, 1758)]

*Opisthogonimus artigasi*

*Opisthogonimus lecithonotus*

*Opisthogonimus serpentis*

*Travtrema stenocotyle*

***Thamnodynastes strigatus*** (Günther, 1858)

*Liophistrema buccalis*

*Opisthogonimus lecithonotus*

*Styphlodora condita*

***Thamnodynastes*** sp. [cited as *Thamnodynastes pallidus* and *Thamnodynastes strigilis* by Lunaschi & Sutton, 1985]

*Opisthogonimus artigasi*

*Opisthogonimus lecithonotus*

*Styphlodora condita*

*Travtrema stenocotyle*

***Tomodon ocellatus*** Duméril, Bibron & Duméril, 1854

*Mesocoelium monas*

***Tomodon dorsatus*** Duméril, Bibron and Duméril, 1854 [=*Tomodon dorsatum* Duméril, Bibron and Duméril, 1854]

*Opisthogonimus serpentis*

*Travtrema stenocotyle*

***Xenodon dorbignyi*** (Duméril, Bibron & Duméril, 1854) [=*Lystrophis dorbignyi* Cope, 1855]

*Catadiscus Freitaslenti*

*Catadiscus longicoecalis*

*Opisthogonimus artigasi*

*Opisthogonimus lecithonotus*

*Styphlodora condita*

*Travtrema stenocotyle*

***Xenodon guentheri*** Boulenger, 1894

*Heterodiplostomum lanceolatum*

***Xenodon merremi*** (Wagler, 1824) [=*Ophis merremii* Wagler, 1824; *Waglerophis merremii* Romano & Hoge 1972]

*Catadiscus freitaslenti*  
*Ophiodiplostomum spectabile*  
*Opisthognomus fariai*  
*Opisthognomus fonsecai*  
*Opisthognomus interrogativus*  
*Opisthognomus lecithonotus*  
*Opisthognomus megabothrium*  
*Opisthognomus misionesensis*  
*Opisthognomus serpentis*  
*Renifer heterocoelium*  
*Styphlodora condita*  
*Travtrema stenocotyle*

*Xenodon severus* (Linnaeus, 1758)  
*Sticholecitha serpentis*

*Xenodon* sp.[=*Lystrophis* sp.]  
*Travtrema stenocotyle*

## CORYTOPHANIDAE

*Basiliscus basiliscus* (Linnaeus, 1758)  
*Parahaplometroides basiliscae*

## CROCODYLIDAE

*Crocodylus intermedius* (Graves, 1819)  
*Caimanicola marajoara*

## DACTYLOIDAE

*Anolis fuscoauratus* D'orbigny, 1837 [= *Norops fuscoauratus* Nicholson, 2002]  
*Mesocoelium monas*  
*Urotrema shirleyae*

*Anolis scypheus* Cope, 1864 [= *Anolis nitens* Wagler, 1830]  
*Brachycoelium salamandrae*

## ELAPIDAE

*Micrurus corallinus* (Merrem, 1820)  
*Haplometroides buccicola*



***Micrurus frontalis*** (Duméril, Bibron & Duméril, 1854)

*Haplometroides buccicola*

*Haplometroides odhneri*

***Micrurus lemniscatus*** (Linnaeus, 1758)

*Haplometroides odhneri*

***Micrurus pyrrhocryptus*** (Cope, 1862) [= *Micrurus frontalis pyrrhocryptus* Shreve 1953]

*Opisthogonimus lecithonotus*

*Styphlodora condita*

***Micrurus* sp.** [= *Elaps* sp.]

*Haplometroides buccicola*

## EMYDIDAE

***Emys orbicularis*** (Linnaeus, 1758) [= *Cistudo lutaria* Strauch, 1862; *Emys lutaria* Schweigger, 1812; *Testudo orbicularis* Linnaeus, 1758]

*Telorchis parvus*

***Trachemys callirostris callirostris*** (Gray 1855)

*Nematophila argentinum*

***Trachemys dorbigni*** (Duméril & Bibron, 1835) [= *Pseudemys dorbigni* Mertens et al. 1934]

*Telorchis achavali*

*Telorchis dubius*

## GEKKONIDAE

***Hemidactylus mabouia*** (Moreau De Jonnés, 1818)

*Paradistomum parvissimum*

*Plagiorchis vicentei*

***Lygodactylus klugei*** (Smith, Martin & Swain, 1977)

*Mesocoelium monas*

## GEOEMYDIDAE

***Rhinoclemmys areolata*** Duméril & Bibron, 1851

*Nematophila grandis*

***Rhinoclemmys punctularia*** (Daudin, 1801) [= *Geoemyda punctularia punctularia* Wettstein,

1934; *Rhinoclemmys punctularia punctularia* Daudin, 1801]

*Nematophila grandis*

***Rhinoclemmys nasuta*** (Boulenger, 1902)

*Halltrema heteroxenus*

*Nematophila grandis*

*Octangioides tlacotalpensis*

*Pseudocleptodiscus margaritae*

## GYMNOPHTHALMIDAE

***Alopoglossus angulatus*** Linnaeus, 1758

*Mesocoelium monas*

***Cercosaura eigenmanni*** (Griffin, 1917) [= *Prionodactylus eigenmanni* Griffin, 1917]

*Mesocoelium monas*

***Echinosaura horrida horrida*** Boulenger, 1890

*Sphaeridiotrema echinosauense*

***Leposoma osvaldoi*** Ávila-Pires, 1995

*Brachycoelium salamandrae*

## IGUANIDAE

***Amblyrhynchus cristatus*** Bell, 1825

*Cetiosaccus galapagensis*

*Iguanacola navicularius*

*Pyelosomum amblyrhynchi*

***Iguana iguana*** (Linnaeus, 1758) [= *Iguana tuberculata* Laurenti 1768; *Hypstophus tuberculatus*]

*Helicotrema asymmetricum*

*Helicotrema magniovatum*

*Helicotrema spirale*

*Paradistomum parvissimum*

*Pulchrosomoides elegans*

## KINOSTERNIDAE

***Kinosternon scorpioides*** (Linnaeus, 1766) [= *Kinosternon scorpioides scorpioides* (Linnaeus, 1766)]

*Nematophila grandis*  
*Telorchis diaphanous*  
*Telorchis rapidulus*

## LEPTOTYPHLOPIDAE

*Trilepida koppesi* (Amaral, 1955) [= *Leptotyphlops koppesi* Amaral, 1955]  
*Haplometroides odhneri*

## LIOLAEMIDAE

*Liolaemus lutzae* Mertens, 1938  
*Paradistomum parvissimum*

## PHYLLODACTYLIDAE

*Gymnodactylus geckoides* Spix, 1825  
*Paradistomum rabusculum*

*Thecadactylus solimoensis* Bergmann & Russell, 2007  
*Mesocoelium monas*

## PODOCNEMIDIDAE

*Peltocephalus dumerilianus* Schweigger, 1812 [= *Podocnemis dumeriliana* (Duméril & Bibron, 1835); *Podocnemis tracaxa* Boulenger 1889]  
*Helicotrema spirale*  
*Nematophila grandis*

*Podocnemis erythrocephala* (Spix, 1824)  
*Nematophila grandis*

*Podocnemis expansa* (Schweigger, 1812)  
*Braunotrema pulvinatum*  
*Halltrema avitellina*  
*Nematophila grandis*  
*Oriximinatrema noronhae*  
*Podocnemitrema papillosus*  
*Rhytidodes gelatinosus*  
*Telorchis bifurcus*  
*Telorchis hagmanni*

**Podocnemis lewyana** Duméril, 1852

*Nematophila argentinum*

*Nematophila venezuelensis*

*Neodeuterobaris pritchardae*

*Pseudonematophila ovalis*

*Telorchis hagmanni*

**Podocnemis unifilis** Troschel, 1848 [= *Podocnemis cayennensis* Schweigger, 1812]

*Loefgrenia loefgreni*

*Nematophila grandis*

*Telorchis aculeatus*

**Podocnemis vogli** Müller, 1935

*Nematophila grandis*

**Podocnemis** sp.

*Halltrema avitellina*

*Halltrema heteroxenus*

*Nematophila grandis*

*Pseudonematophila ovalis*

## SCINCIDAE

**Mabuya agilis** (Raddi, 1823)

*Paradistomum parvissimum*

**Mabuya macrorhyncha** Hoge, 1946

*Paradistomum parvissimum*

*Pulchrosomoides elegans*

**Trachylepis atlantica** (Schmidt, 1945) [= *Mabuya maculata* Anderson, 1900]

*Mesocoelium monas*

## TEIIDAE

**Ameiva ameiva** (Linnaeus, 1758)

*Paradistomum parvissimum*

**Kentropyx calcarata** Spix, 1825

*Paradistomum parvissimum*

**Tupinambis rufescens** (Günther, 1871)

*Styphlodora condita*

*Tupinambis teguixin* (Linnaeus, 1758) [=*Tupinambis nigropunctatus* Spix, 1825]  
*Dasymetra tupinambis*  
*Paradistomum parvissimum*  
*Pulchrosomoides elegans*

## TESTUDINIDAE

*Chelonoidis denticulata* (Linnaeus, 1766) [=*Geochelone denticulata* (Linnaeus, 1766);  
*Testudo denticulata* Linnaeus, 1766; *Testudo tabulata* Walbaum, 1782]  
*Halltrema avitellina*  
*Helicotrema spirale*

*Kinixys erosa* (Schweigger, 1812)  
*Nematophila grandis*

*Testudo* sp.  
*Cheloniodiplostomum testudinis*

## TROPIDURIDAE

*Plica plica* Linnaeus, 1758  
*Mesocoelium monas*  
*Paradistomum parvissimum*

*Tropidurus torquatus* (Wied, 1820) [=*Tropidurus torquatus torquatus* (Wied, 1820)]  
*Mesocoelium monas*  
*Paradistomum parvissimum*  
*Plagiorchis freitasi*

*Uranoscodon superciliosus* (Linnaeus, 1758)  
*Allopharynx daileyi*  
*Mesocoelium monas*  
*Mesocoelium sibynomorphi*  
*Paradistomum parvissimum*

## VIPERIDAE

*Bothriechis schlegelii* (Berthold, 1846)  
*Renifer heterocoelium*

*Bothropoides diporus* (Cope, 1862) [=*Bothrops neuwiedi diporus* Cochran, 1961]  
*Catadiscus freitaslenti*

*Catadiscus longicoecalis*  
*Infidum infidum*  
*Opisthognomus lecithonotus*  
*Styphlodora condita*  
*Travtrema stenocotyle*

***Bothropoides insularis*** (Amaral, 1921) [= *Bothrops insularis* Amaral, 1929]  
*Renifer heterocoelium*

***Bothropoides jararaca*** (Wied, 1824) [= *Bothrops jararaca* Duméril & Bibron, 1854]  
Infidum similis

*Opisthognomus artigasi*  
*Opisthognomus fonsecai*  
*Opisthognomus interrogativus*  
*Opisthognomus lecithonotus*  
*Paradistomum parvissimum*  
*Renifer heterocoelium*  
*Styphlodora gili*

***Bothropoides neuwiedi*** (Wagler, 1824) [= *Bothrops neuwiedii* Wagler, 1824; *Bothrops neuwiedi meridionalis* Amaral, 1930; *Lachesis neuwiedii* Berg, 1898].

*Catadiscus freitaslenti*  
*Catadiscus longicoecalis*  
*Opisthognomus afranioi*  
*Opisthognomus fonsecai*  
*Opisthognomus lecithonotus*  
*Renifer heterocoelium*  
*Styphlodora condita*  
*Travtrema stenocotyle*

***Bothrops alternatus*** Duméril, Bibron & Duméril, 1854 [= *Bothrops alternata* Amaral, 1925]

*Catadiscus freitaslenti*  
*Heterodiplostomum lanceolatum*  
*Opisthognomus fonsecai*  
*Opisthognomus lecithonotus*  
*Styphlodora condita*  
*Styphlodora gili*  
*Travtrema stenocotyle*

***Bothrops asper*** (Garman, 1884)  
*Renifer heterocoelium*

***Bothrops atrox*** (Linnaeus, 1758)  
*Opisthognomus lecithonotus*  
*Renifer monstruosum*

***Bothrops cotiara*** Gomes, 1913

*Renifer heterocoelium*

***Bothrops jararacussu*** (Lacerda, 1884)

*Opisthogonimus lecithonotus*

***Bothrops moojeni*** Hoge, 1966

*Infidum infidum*

*Opisthogonimus artigasi*

*Opisthogonimus fonsecai*

*Opisthogonimus lecithonotus*

*Renifer heterocoelium*

*Sticholecitha serpentis*

*Styphlodora condita*

*Travtrema stenocotyle*

***Bothrops neuwiedii mattogrossensis*** (Amaral, 1925)

*Renifer heterocoelium*

***Bothrops*** sp.

*Opisthogonimus lecithonotus*

*Travtrema stenocotyle*

***Crotalus durissus terrificus*** (Laurenti, 1768)

*Ophiodiplostomum spectabile*

***Porthidium nasutum*** (Bocourt, 1868)

*Renifer heterocoelium*

“crocodiles”

*Travtrema stenocotyle*

“freshwater turtle”

*Telorchis pleroticus*

“marine turtle”

*Neoctangium travassosi*





## REFERENCES

- Alho, C. J. R., 1964. Dois novos hospedadores de *Nematophila grande* Diesing, 1839 (Trematoda, Paramphistomidae). *Boletim do Museu Paraense Emílio Goeldi (Zoologia)*, 52: 1-4.
- Alho, C. J. R., 1965. Contribuição ao conhecimento da fauna helmintológica de quelônios do estado do Pará, Brasil. *Boletim do Museu Paraense Emílio Goeldi (Zoologia)*, 58: 1-8.
- Alho, C. J. R. & Vicente, J. J., 1964. *Podocnemitrema papillosus* g. n. sp. n. e nova organização da sistemática da família Microscaphiidae Travassos, 1922 (Trematoda: Paramphistomoidae). *Revista Brasileira de Biologia*, 24: 17-22.
- Álvarez, A., Lenis, C. & Vélez, I. 2005. First report of two species of trematodes (Digenea: Proterodiplostomidae) for Colombia in *Caiman crocodilus fuscus* (Reptilia: Crocodylia). *Caldasia*, 27: 287-291.
- AmphibiaWeb: Information on amphibian biology and conservation. [web application]. 2013. Berkeley, California: AmphibiaWeb. Available: <http://amphibiaweb.org/>. (Accessed: Apr 25, 2013).
- Anjos, L. A., Bezerra, C. H., Passos, D. C., Zanchi, D. & Galdino, C. A. B., 2011. Helminth fauna of two gecko lizards, *Hemidactylus agrius* and *Lygodactylus klugei* (Gekkonidae), from Caatinga Biome, Northeastern Brazil. *Neotropical Helminthology*, 5 (2): 285-290.
- Araujo, T. L., 1941. Nota sobre um trematóide Aspidogastridae de tartaruga marinha. *Boletim da Industria Animal, São Paulo*, 4 (3-4): 184-186.
- Artigas, P. T. & Campos, M. S., 1976/1977. Considerações sobre *Plagiorchis luhei* Travassos, 1927. (*Microderma luehi* Mehra, 1931) (Trematoda, Plagiorchiidae) parasito de *Hydrodynastes gigas* Dum et Brib. (Reptilia, Colubridae). *Memórias do Instituto Butantan*, 40-41: 265-279.
- Artigas, P. T. & Paulino, R. C., 1988. *Zeferinella vazi* gen. nov., sp. nov. (Plagiorchidae), a parasite from the mouth and oesophagus of *Erythrolamprus aesculapsi* (Serpentes, Colubridae). *Memórias do Instituto Butantan*, 50 (3): 63-69.
- Artigas, P. T. & Pérez, M. D., 1964. *Catadiscus eldoradiensis* n. sp., Trematoda, Paramphistomata de *Leptodactylus ocellatus*. *Memórias do Instituto Butantan*, 31: 5-8.
- Artigas, P. T., Ruiz, J. M. & Leão, A. T., 1942. Trematódeos de ofídeos. *Liophistrema pulmonalis*, n. g., n. sp. (Plagiorchiidae). *Memórias do Instituto Butantan*, 16: 157-165.
- Artigas, P. T., Ruiz, J. M. & Leão, A. T., 1943. Algumas notas sobre o genero *Opisthgonimus* Luehe, 1900. Descrição de *Opisthgonimus serpentis* sp. n., trematóide de ofídeo. *Memórias do Instituto Butantan*, 17:47-59.

- Artigas, P. T. & Zerpa, M. G., 1961. *Plagiorchis rangeli* n. sp. parasito de *Leptodactylus ocellatus* (Trematoda, Plagiorchiidae). *Anais da Faculdade de Farmácia e Odontologia, Universidade de São Paulo*, 18 (1): 25-28.
- Ávila, R. W. & Silva, R. J., 2010. Checklist of helminths from lizards and amphisbaenians (Reptilia, Squamata) of South America. *Journal of Venomous Animals and Toxins including Tropical Diseases*, 16 (4): 543-572.
- Ávila, R. W. & Silva, R. J., 2013. Helminths of lizards from the municipality of Aripuanã in the southern Amazon region of Brazil. *Journal of Helminthology*, 87 (1): 12-16.
- Barrella, T. H. & Silva, R. J., 2003. Digenetic trematodes infection in a *Bothrops moojeni* (Viperidae) population from a fauna rescue in Porto Primavera, São Paulo, State. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, 55 (2): 243-245.
- Bechara, M. & Vélez, I., 2010. Some digeneans of *Rhinella marina* (Anura: Bufonidae) in Colombia. *Revista Mexicana de Biodiversidad*, 81: 39-42.
- Bechara-Escudero, M. & Asprilla-Murillo, S., 2007. Tremátodos digéneos de *Chaunus marinus* (Anura: Bufonidae) en el Municipio de Quibdó, Chocó. *Revista Institucional Universidad Tecnológica del Chocó: Investigación, Biodiversidad y Desarrollo*, 2: 13-17.
- Bhalerao, G. D., 1937. On *Pneumotrema travassosi* gen. et sp. nov. and two other trematode parasites from animals dying in the Zoological Society's garden during 1936-1937. *Zoological Society of London*, C. 107 (9): 34, B., 107 (3): 365-369.
- Blair, D., 2005. Family Pronocephalidae Looss, 1899. In: Jones, A., Bray, R. A. & Gibson, D. I. (Eds) *Keys to the Trematoda*. Vol. 2. CABI Publishing and The Natural History Museum, Wallingford: 361-380.
- Boero, J. J. & Led, J. E., 1971. Parasites of autochthonous fauna. V. Parasite of Argentine birds. VI. Parasite of Argentine ophidians. VII. Parasite of Argentine bats. *Analeceta Veterinaria*, 3 (1/3): 91-103.
- Boero, J. J. & Led, J. E., 1974. Parasites of the autochthonous fauna. *Revista de Agronomía y de Veterinaria, Buenos Aires*, 3 (1): 16-17.
- Boero, J. J., Led, J. E. & Brandetti, E., 1972. El parasitismo de la fauna autóctona. *Revista de Agronomía y de Veterinaria, Buenos Aires*, 1 (8): 17-29.
- Brandes, G., 1888. Die Familie der *Holostomeae*. Ein Prodrömus zu einer Monographie derselben. Diss. 72 pp. Reudnitz-Leipzig.
- Brandes, G., 1890. Die Familie der Holostomiden. Bronn's Klassen und Ordnungen des Thier-Reichs, Leipzig, 4 (1): 567-925.

- Braun, M., 1899. I. Wissenschaftliche Mittheilungen. 1. Über *Clinostomum* Leidy. *Zoologischer Anzeiger*, 22: 489-493.
- Braun, M., 1901. Trematoden der Chelonier. *Mitteilungen aus dem Zoologischen Museum in Berlin*, 2:5-58
- Bray, R. A., Gibson, D. I. & Jones, A., 2008. *Keys to the Trematoda*. Vol. 3. Eds. R. A. Bray, D. I. Gibson, & A. Jones. CABI Publishing, Wallingford, UK, and The Natural History Museum, London: 824 pp.
- Brooks, D. R., 1976a. *Neodeuterobaris pritchardae* gen. et sp. n. (Digenea: Microscaphidiidae) a sideneck turtle, *Podocnemis lewyana* Dumeril 1852, from Colombia. *Journal of Parasitology*, 62 (3): 426-428.
- Brooks, D. R., 1976b. Five species of Platyhelminthes from *Bufo marinus* L. (Anura: Bufonidae) in Colombia with descriptions of *Creptotrema lynchi* sp. n. (Digenea: Allocreadiidae) and *Glypthelmins robustus* (Digenea: Macroderoididae). *Journal of Parasitology*, 62 (3): 429-433.
- Brooks, D. R., 1980. Revision of the Acanthostominae Poche, 1926 (Digenea: Cryptogonimidae). *Zoological Journal of the Linnean Society*, 70: 313-382.
- Bursey, C. R. & Goldberg, S. R., 2004. *Cosmocerca vrcibradici* n. sp. (Ascaridida: Cosmocercidae), *Oswaldocruzia vitti* n. sp. (Strongylida: Molineoidae), and other helminths from *Prionodactylus eigenmanni* and *Prionodactylus oshaughnessyi* (Sauria: Gymnophthalmidae) from Brazil and Ecuador. *Journal of Parasitology*, 90 (1): 140-145.
- Bursey, C. R., Goldberg, S. R. & Parmelee, J. R., 2001. Gastrointestinal helminths of 51 species of anurans from Reserva Cuzco Amazonico, Peru. *Comparative Parasitology*, 68 (1): 21-35.
- Bursey, C. R., Goldberg, S. R. & Vitt, L. J., 2005. New species of *Allopharynx* (Digenea: Plagiorchiidae) and other helminths in *Uranoscodon superciliosus* (Squamata: Tropiduridae) from Amazonian Brazil. *Journal of Parasitology*, 91 (6): 1395-1398.
- Caballero, C. E. & Diaz-Ungria, C., 1958. Intento de un Catálogo de los Tremátodos Digéneos registrados en territorio Venezolano. *Memoria de la Sociedad de Ciencias Naturales La Salle*, 18: 19-36.
- Caballero, C. E. & Vogelsang, E. G., 1947. Fauna helmintologica venezolana. I. *Ochetosoma miladelarocai* n. sp. de *Bothrops atrox* L. y hallazgo de *Physaloptera retusa* (Rud., 1819) em *Cnemidophorus lemniscatus*. *Revista de Medicina Veterinaria y Parasitología, Caracas*, 6 (1-4): 53-62.
- Caballero, C. E., Vogelsang, E. G. & Zerecero, D. M. C., 1953. Fauna helmintológica venezolana. (IV). Algunos tremátodos de batracios y mamíferos. *Revista de Medicina Veterinaria y Parasitología, Caracas*, 12 (1-4): 195-208.

- Caira, J. N. & Bogéa, T., 2005. Family Allocreadiidae Looss, 1902. In: Eds. Jones, A., Bray, R. A. & Gibson, D. I. (Eds) *Keys to the Trematoda. Vol. 2*. CABI Publishing and The Natural History Museum, Wallingford: 417-436.
- Campião, K. M., Delatorre, M., Rodrigues, R. B., Silva, R. J. & Ferreira, V. L., 2012. The effect of local environmental variables on the helminth parasite communities of the point-belly frog *Leptodactylus podicipinus* from ponds in the Pantanal wetlands. *Journal of Parasitology*, 98 (2): 229-235.
- Campião, K. M., Silva, R. J. & Ferreira, V. L., 2009. Helminth parasites of *Leptodactylus podicipinus* (Anura: Leptodactylidae) from south-eastern Pantanal, State of Mato Grosso do Sul, Brazil. *Journal of Helminthology*, 83 (4): 345-349.
- Campião, K. M., Silva, R. J. & Ferreira, V. L., 2010. Helminth component community of the paradoxal frog *Pseudis platensis* Gallardo, 1961 (Anura: Hylidae) from south-eastern Pantanal, Brazil. *Parasitology Research*, 106 (3): 747-751.
- Carter, A. S. & Etges, F. J., 1972. A redescription of *Acanthostomum marajoarum* (Teixeira de Freitas & Lent, 1938) with notes on the subfamily Acanthostominae (Nicoll, 1914) Huges, Higginbotham and Clary, 1942 (Trematoda: Acanthostomatidae). *Proceedings of the Helminthological Society of Washington*, 39 (2): 234-239.
- Catto, J. B. & Amato, J. F. R., 1993a. Digenetic trematodes (Cryptogonimidae, Acanthostominae) parasites of the caiman, *Caiman crocodilus yacare* (Reptilia, Crocodylia) from the Pantanal Mato-Grossense, Brazil, with the description of a new species. *Memórias do Instituto Oswaldo Cruz*, 88: 435-440.
- Catto, J. B. & Amato, J. F. R., 1993b. Two new species of *Pseudotolorchis* (Digenea, Telorchidae), parasites of the caiman, *Caiman crocodilus yacare* (Reptilia, Crocodylia) from the Pantanal Mato-Grossense, Brazil. *Memórias do Instituto Oswaldo Cruz*, 88 (4): 561-566.
- Catto, J. B. & Amato, J. F. R., 1994a. Helminth community structure of the caiman, *Caiman crocodilus yacare* (Crocodylia, Alligatoridae) in the Brazilian "Pantanal". *Revista Brasileira de Parasitologia Veterinária*, 3 (2): 109-118.
- Catto, J. B. & Amato, J. F. R., 1994b. Proterodiplostome parasites (Digenea, Proterodiplostomidae) of the caiman, *Caiman crocodilus yacare* (Reptilia, Crocodylia) in the Pantanal Mato-Grossense, Brazil, with the description of two new species. *Memórias do Instituto Oswaldo Cruz*, 89 (4): 539-551.
- Cohn, L., 1902. Zwei neue Distomen. *Centralblatt für Bakteriologie*, 32 (12): 877-885.
- Cohn, L., 1903. Helminthologische Mitteilungen. *Archiv für Naturgeschichte*, 69: 47-68.

- Cordero, E. H., 1942. La validez de *Halipegus dubius* (Trematoda, fam. Hemiuridae). *Anais da Academia Brasileira de Ciências*, 14 (2): 127-134.
- Cordero, E. H., 1944. Dos nuevas especies de tremátodos del género *Glyphelmins* de los batracios del Uruguay. *Anais da Acadademia Brasileira de Ciências*, 16 (1): 1-8.
- Cordero, E. M. & Vogelsang, E. G., 1928. *Distomum xenodontis* n. sp. Nuevo trematode del intestino de *Xenodon merremii* (Wagler) de Jujuy. In *Cuarta Reunión de la Sociedad Argentina de Patología Regional del Norte*.
- Cordero, E. M. & Vogelsang, E. G., 1939. Nuevos tremátodos. I. Dos especies del genero *Pneumonoeces* Looss, del pumon de *Rana palmipes* Spix de Venezuela. *Revista de Medicina Veterinária y Parasitología, Caracas*, 1 (2-4): 173-178.
- Cordero, E. H. & Vogelsang, E. G., 1940. Nuevos tremátodos, II. Cuatro Paramphistomidae de los quelonios sudamericanos. *Revista de Medicina Veterinaria y Parasitología, Caracas*, 1 (2-4): 3-15.
- Correa, A. A. S. & Artigas, P. T., 1978/1979. *Catadiscus rochai* n. sp. (Trematoda, Paramphistomidae) parasito de *Dromicustyphlus* (L.) (Ophidia: Colubridae). *Memórias do Instituto Butantan*, 42-43: 145-150.
- Corrêa, F. M. A., Paulino, R. C., Buononato, M. A. & Federsoni, P. A., 1990. *Ochetosoma heterocoelium* (Travassos, 1921) (Trematoda: Digenea: Ochetosomatidae) em novo hospedeiro. *Memórias do Instituto Butantan*, 52 (1): 11-16.
- Dias, M. T. & Nasir, P., 1969. Intraespecific variations in *Proterodiplostomum longum* (Trematoda: Proterodiplostomidae). *Acta Científica Venezolana*, 20 (3/4): 126.
- Diaz-Ungria, C., 1967. Helminths da Venezuela. *Barquisimito: Centro experimental de Estudios superiores, Escuela de Ciencias Veterinarias, Parasitología y Zoología Medica*: 88 pp.
- Diaz-Ungria, C., 1973. Endoparasitic helminths of Venezuela. *Ciencia e Veterinaria*, 3 (1/2): 37-242.
- Diaz-Ungria, C., 1978. Helminths parásitos de vertebrados em el Estado Zulia. Algunas especies nuevas para Venezuela. *Kasmera*, 6 (1/4): 207-233.
- Diaz-Ungria, C., 1979. Algunas especies de helminths nuevas para Venezuela. *Revista Iberica de Parasitología*, 39: 313-336.
- Diesing, K. M., 1850. *Systema Helminthum*, 1, XIII + 679 pp. Vindobonae.
- Dione, W., 1947. *Gorgoderina chilensis* n. sp., trematodo de la vejiga urinaria del sapito vaquero, *Rhinoderma darwini* D. B. *Comunicaciones Zoológicas del Museo de Historia Natural de Montevideo*, 2 (39): 1-11.

- Dobbin Jr., J. E., 1957a. Fauna helmintológica de batráquios de Pernambuco. I. Trematoda. *Anais da Sociedade de Biologia de Pernambuco*, 15 (1): 29-61.
- Dobbin Jr., J. E., 1957b. Nota sobre as espécies de *Haematoloecus* Looss, 1899 que ocorrem na América do Sul. *Memórias do Instituto Oswaldo Cruz*, 55 (2): 167-189.
- Dobbin Jr., J. E., 1957c. Sobre uma nova especie do gênero *Telorchis* Lühe, 1900 (Trematoda: Telorchidae). *Revista Brasileira de Biologia*, 17 (4): 509-512.
- Dobbin Jr., J. E., 1958. *Glypthelmins vitellinophilum* sp. n., parasito de *Hyla raniceps* (Cape). *Memórias do Instituto Oswaldo Cruz*, 56 (1): 153-157.
- Dollfus, R. P., 1935. Sur *Crocodilicola* et autres Hémistomes de Crocodiliens. *Extrait d'Archive du Museum d' Histoire Naturelle, Paris, vol. du Tricentenaire*, 6 série, 12: 637-646.
- Dronen, N. O, Calhoun, D. M & Simcik, S. R., 2012. *Mesocoelium* Odhner, 1901 (Digenea: Mesocoelidae) revisited; a revision of the family and re-evaluation of species composition in the genus. *Zootaxa*, 3387:1-96.
- Dubois, G., 1936. Les diplostomes de reptiles (Trematoda: Proterodiplostomidae nov. fam.) du musée de Vienne. *Bulletin de la Société de Science Naturelle de Neuchâtel*, 61: 5-80.
- Dubois, G., 1938. Liste systématique des Strigéidés du Brésil et du Venezuela. *Livro Jubilar do Professor Lauro Travassos*: 145-156.
- Dubois, G., 1948. Sur trois diplostomes de crocodiliens (Trematoda: Strigeida). *Annales de Parasitologie Humaine et Comparée*, 23: 5-13.
- Dubois, G., 1953. Systématique des Strigeida. Complément de la monographie. *Memórias de la Societé Neuchâteloise des Sciences Naturelles*, 81(2): 1-141.
- Dubois, G., 1979. Révision et nouvelle clé de détermination des diplostomes de reptiles (Trematoda: Proterodiplostomidae Dubois, 1936). *Bulletin de la Société Neuchâteloise des Sciences Naturelles*, 102: 39-48.
- Dubois, G., 1988. Quelques Strigeoidea (Trematoda) récoltés au Paraguay par les expéditions du Muséum d'Histoire Naturelle de Genève, au cours des années 1979, 1982 et 1985. *Revue Suisse de Zoologie*, 95 (2): 521-532.
- Dubois, G. & Mahon, J., 1959. Étude de quelques trématodes Nord-Américains suivie d'une revision des genres *Galactosomum* Looss, 1899 et *Ochetosoma* Braun, 1901. *Bulletin de la Société de Science Naturelle de Neuchâtel*, 82: 191-229.
- Dyer, W. G., 1986. Trematodes of some Ecuadorian amphibians. *Transactions of the Illinois State Academy of Science*, 79 (1-2): 137-140.

- Dyer, W. G. & Altig, R., 1977. Helminths of some Ecuadorian anurans. *Herpetologica*, 33: 293-296.
- Dyer, W. G. & Carr, J. L., 1990. Some digeneans of the Neotropical turtle genus *Rhinoclemmys* in Mexico and South America. *Journal of the Helminthological Society of Washington*, 57 (1): 12-14.
- Fábio, S. P., 1979. Considerações sistemáticas sobre algumas espécies do gênero *Styphlodora* Looss, 1899 (Trematoda). *Revista Brasileira de Biologia*, 39 (1): 229-232.
- Fábio, S. P., 1982. Helminths of sympatric populations of some species of anurans of the family Leptodactylidae. *Arquivos da Universidade Federal Rural do Rio de Janeiro*, 5 (1): 69-83.
- Fábio, S. P. & Pinheiro, N. L., 2001. Histological aspects of *Neohaematoloechus neivai* (Travassos and Artigas, 1927) (Trematoda: Haematoloechidae) in the lungs of *Leptodactylus ocellatus* (Linnaeus, 1758) (Anura: Leptodactylidae). *Boletim do Museu Nacional do Rio de Janeiro, Zoologia*, 458: 1-14.
- Fábio, S. P. & Rolas, F. J. T., 1974. Sobre alguns helmintos parasitos de *Dryadophis bifossatus* (Raddi). *Memórias do Instituto Oswaldo Cruz*, 72 (1/2): 49-61.
- Fahel, J., 1952. Fauna helmintológica de “Gias” de Salvador. (*Leptodactylus pentadactylus*). *Anais da Academia Brasileira de Ciências*, 24 (4): 389-436.
- Faria, G., 1910. Contribuição para a sistemática helmintológica brasileira. II: *Dicrocoelium infidum* n. sp. Parasito da vesícula biliar de *Eunectes murina* L. *Memórias do Instituto Oswaldo Cruz*, 2 (1): 22-28.
- Faria, G., 1911. Contribuições para a helmintologia brasileira. *Styphlodora condita* n. sp. *Memórias do Instituto Oswaldo Cruz*, 3 (1): 40-45.
- Faria, M. J., 1978. Prevalência de trematódeos parasitas de anfíbios anuros no Estado do Rio de Janeiro. *Atas da Sociedade de Biologia do Rio de Janeiro*, 19: 55-57.
- Fernandes, J. C., 1958. Notas sobre algumas espécies do gênero *Gorgoderina* Looss, 1902 (Trematoda, Gorgoderidae). *Memórias do Instituto Oswaldo Cruz*, 56 (1): 1-15.
- Flowers, J. R., Law, M. & Carvajal-Endara, S., 2011. *Pseudosonsinotrema megalorchis* n. sp. (Digenea: Pleurogenidae) from the Paramo marsupial frog, *Gastrotheca pseustes* (Anura: Hemiphractidae), Ecuador. *Comparative Parasitology*, 78 (1): 15-20
- Font, W. F. & Lotz, J. M., 2008. Family Telorchhiidae Looss, 1899. In: Bray, R. A., Gibson, D. I. & Jones, A. (Eds) *Keys to the Trematoda*. Vol. 3. CABI Publishing and The Natural History Museum, Wallingford: 425-436.

- Fortes, E. & Hoffmann, R. P., 1987/1988. Record of platyhelminths in snakes in Rio Grande do Sul. *Arquivos da Faculdade Veterinária da Universidade Federal do Rio Grande do Sul*, 15/16: 23-25.
- Freitas, J. F. T., 1941a. Sobre alguns trematódeos parasitos de rãs. *Revista Brasileira de Biologia*, 1 (1): 31-40.
- Freitas, J. F. T., 1941b. Novo trematódeo paranfistomídeo parasito de rã - *Catadiscus inopinatus* n. sp. *Revista Brasileira de Biologia*, 1 (2): 121-123.
- Freitas, J. F. T., 1943. *Catadiscus mirandai* n. sp., parasito de *Hemipipa carvalhoi* Mir.-Rib. *Revista Brasileira de Biologia*, 3 (4): 411-412.
- Freitas, J. F. T., 1956. Breve nota sobre Opisthogoniminae Travassos, 1928 e grupos afins (Trematoda, Plagiorchioidea). *Revista Brasileira de Biologia*, 16 (2): 141-144.
- Freitas, J. F. T., 1958. Breve nota sobre o *Distoma monas* Rudolphi, 1819 (Trematoda). *Revista Brasileira de Biologia*, 18 (2): 171-174.
- Freitas, J. F. T., 1960a. Sobre um novo parasita de anfíbio: *Maicuru solitarium* g. n., sp. n. (Trematoda, Plagiorchiidae). *Boletim do Museu Paraense Emilio Goeldi, n. s., Zoologia*, 30: 1-4.
- Freitas, J. F. T., 1960b. Rápidas informações sobre hospedadores e distribuição geográfica de alguns trematódeos parasitos de batráquios. *Atas da Sociedade de Biologia do Rio de Janeiro*, 4 (3): 29-32.
- Freitas, J. F. T., 1961. Primeira espécie brasileira do gênero *Brachycoelium* Dujardin, 1845. *Boletim do Museu Paraense Emilio Goeldi, n. s., Zoologia*, 32: 1-5.
- Freitas, J. F. T., 1963. Revisão da família Mesocoeliidae Dollfus, 1933 (Trematoda). *Memórias do Instituto Oswaldo Cruz*, 61 (2): 177-311.
- Freitas, J. F. T. & Dobbin Jr., J. E., 1956. Novo parasita de rã: *Catadiscus propinquus* sp. n. (Trematoda, Paramphistomoidea). *Revista Brasileira de Biologia*, 4: 439-441.
- Freitas, J. F. T. & Dobbin Jr., J. E., 1957. Sobre *Travtrema stenocotyle* (Cohn, 1902). *Boletim do Museu Nacional*, 170: 1-25.
- Freitas, J. F. T. & Dobbin Jr., J. E., 1959. *Telorchis diaphanus* sp. n. trematódeo parasito de quelônio. *Anais da Sociedade de Biologia de Pernambuco*, 16 (1): 191-199.
- Freitas, J. F. T. & Dobbin Jr., J. E., 1967. Sobre um novo trematódeo Echinostomatidae parasito de quelônio. *Memórias do Instituto Oswaldo Cruz*, 65 (1): 37-39.



- Freitas, J. F. T. & Lent, H., 1937. Sobre um novo trematódeo parasito de *Iguana tuberculata* (Laur.). *Memórias do Instituto Oswaldo Cruz*, 32 (1): 55-58.
- Freitas, J. F. T. & Lent, H., 1938a. Pesquisas helmintológicas realizadas no Estado do Pará. II. Dois novos trematódeos de *Caiman sclerops* Gray. *Memórias do Instituto Oswaldo Cruz*, 33 (1): 53-56.
- Freitas, J. F. T. & Lent, H., 1938b. Sobre alguns trematódeos parasitos de *Chelone mydas* (L.), principalmente Paramphistomoidea. *Memórias do Instituto Oswaldo Cruz*, 33 (1): 79-87.
- Freitas, J. F. T. & Lent, H., 1939a. Considerações sobre algumas espécies americanas do gênero *Haematoloechus* Looss, 1899. *Livro em Homenagem ao Professor Álvaro e Miguel Osório de Almeida*: 246-256.
- Freitas, J. F. T. & Lent, H., 1939b. Revisão do gênero *Catadiscus* Cohn, 1904 (Trematoda, Paramphistomoidae). *Boletim Biológico*, 4 (2): 305-315.
- Freitas, J. F. T. & Lent, H., 1942. A propósito de *Halltrema avitellina* Lent & Freitas, 1939. *Revista Brasileira de Biologia*, 2 (91): 115-116.
- Fróes, A. M. & Lima, D. F., 1974. Ocorrência de *Haematoloechus freitasi* Mané-Garzón & Solares, 1959 (Trematoda, Plagiorchiidae) no Brasil. *Arquivos da Faculdade de Veterinária da Universidade Federal do Rio Grande do Sul*, 2 (1): 21-23.
- Frost, D. R., 2011. Amphibian Species of the World: an Online Reference. Version 5.5 (31 January, 2011). Electronic Database accessible at <http://research.amnh.org/vz/herpetology/amphibia/> American Museum of Natural History, New York, USA.
- García, L. & Tantaleán, V. M., 1987. *Gorgoderina* sp. y *Batrachotaenia* sp. parásitos de *Batrachophrynus macrostomus* Peters, 1873 (Amphibios) de la laguna de Huicra, Pasco – Perú. *Biota*, 13: 34-39.
- Gibson, D. I., Jones, A. & Bray, R. A., 2002. *Keys to the Trematoda*. Vol. 1. Eds. D. I. Gibson, A. Jones & R. A. Bray. CABI Publishing, Wallingford, UK, and The Natural History Museum, London, xvi + 521 pp.
- Gilbert, P. T., 1940. Three new trematodes from the Galapagos Islands marine iguana *Amblyrhynchus cristatus*. *Allan Hancock Pacific Expedition*, 2 (6): 91-108.
- Goldberg, S. R., Bursey, C. R., Caldwell, J. P. & Shepard, D. B., 2009. Gastrointestinal helminths of six sympatric species of *Leptodactylus* from Tocantins State, Brazil. *Comparative Parasitology*, 76 (2): 258-266.

- Goldberg, S. R., Bursey, C. R., Caldwell, J. P., Vitt, L. J. & Costa, G. C., 2007. Gastrointestinal helminths from six species of frogs and three species of lizards, sympatric in Para State, Brazil. *Comparative Parasitology*, 74 (2): 327-342.
- Goldberg, S. R., Bursey, C. R. & Vitt, L. J., 2006. Helminths of the brown-eared anole, *Norops fuscoauratus* (Squamata, Polychrotidae), from Brazil and Ecuador, South America. *Phyllomedusa Journal of Neotropical Herpetology*, 5 (1): 83-86.
- Goldberg, S. R., Bursey, C. & Vitt, L. J., 2007. Parasite communities of two lizard species, *Alopoglossus angulatus* and *Alopoglossus atriventris*, from Brazil and Ecuador. *Herpetological Journal*, 17 (4): 269-272.
- Goldberg, S. R., Bursey, C. R. & Vitt, L. J., 2009. Diet and parasite communities of two lizard species, *Plica plica* and *Plica umbra* from Brazil and Ecuador. *Herpetological Journal*, 19 (1): 49-52.
- Gomes, D. C. & Pinto, R. M., 1978. Contribuição ao conhecimento da fauna helmintológica da região amazônica – Trematódeos. *Atas da Sociedade de Biologia do Rio de Janeiro*, 19: 43-46.
- Gomes, T., F., F., Melo, F. T. V., Giese, E. G., Furtado, A. P., Gonçalves, E. C. & Santos, J. N., 2013. A new species of *Mesocoelium* (Digenea: Mesocoeliidae) found in *Rhinella marina* (Amphibia: Bufonidae) from Brazilian Amazonia. *Memórias do Instituto Oswaldo Cruz*, 108 (2): 186-191.
- González, C. E. & Hamann, M. I., 2006. Helminth parasites of *Leptodactylus bufonis* Boulenger, 1894 (Anura: Leptodactylidae) from Corrientes, Argentina. *Revista Española de Herpetología*, 20: 39-46.
- Hamann, M. I., 2004. Seasonal maturation of *Catadiscus propinquus* (Digenea: Diplodiscidae) in *Lysapsus limellus* (Anura: Pseudidae) from an argentinean subtropical permanent pond. *Physis*, 59: 29-36.
- Hamann, M. I., 2006. Seasonal maturation of *Glythelmins vitellinophilum* (Trematoda: Digenea) in *Lysapsus limellus* (Anura: Pseudidae) from em Argentinian subtropical permanent pond. *Brazilian Journal of Biology*, 66: 85-93.
- Hamann, M. I., González, C. E. & Kehr, A. I., 2006. Helminth community structure of the oven frog *Leptodactylus latinasus* (Anura, Leptodactylidae) from Corrientes, Argentina. *Acta Parasitologica*, 51 (4): 294-299.
- Hamann, M. I. & Kehr, A. I., 1997. *Lysapsus limellus*. Parasitism. *Herpetological Review*, 28: 85.

- Hamann, M. I. & Kehr, A. I., 1999. Population dynamics and ecological relationships between *Glyphelmims vitellinophilum* Bobbin, 1958 (Trematoda, Macroderoididae) and the host *Lysapsus limellus* Cope, 1862 (Anura, Pseudidae) in a semipermanent pond of Corrientes, Argentina. *Physis sec. B*, 57 (132/133): 17-24.
- Hamann, M. I., Kehr, A. I. & González, C. E., 2006. Species affinity and infracommunity ordination of helminths of *Leptodactylus chaquensis* (Anura: Leptodactylidae) in two contrasting environments from Northeastern Argentina. *Journal of Parasitology*, 92 (6): 1171-1179.
- Hamann, M. I., Kehr, A. I. & González, C. E. 2009. Niche specificity of two *Glyphelmims* (Trematoda) congeners infecting *Leptodactylus chaquensis* (Anura: Leptodactylidae) from Argentina. *Journal of Parasitology*, 95 (4): 817-822.
- Hamann, M. I., Kehr, A. I. & González, C. E., 2010. Helminth community structure of *Scinax nasicus* (Anura: Hylidae) from a South American subtropical area. *Diseases of Aquatic Organisms*, 93 (1): 71-82.
- Hamann, M. I., Kehr, A. I. & González, C. E. 2012. Community structure of helminth parasites of *Leptodactylus bufonius* (Anura: Leptodactylidae) from northeastern Argentina. *Zoological Studies*, 51 (8): 1454-1465.
- Hamann, M. I., Kehr, A. I., González, C. E., 2013a. Biodiversity of trematodes associated with amphibians from a variety of habitats in Corrientes Province, Argentina. *Journal of Helminthology*, 87 (3): 286-300
- Hamann, M. I., Kehr, A. I., González, C. E., 2013b. Helminth communities in the burrowing toad, *Rhinella fernandezae*, from northeastern Argentina. *Biologia*, 68: 1155-1162.
- Hamann, M. I., Kehr, A. I., González, C. E., Duré, M. I. & Schaefer, E. F., 2009. Parasite and reproductive features of *Scinax nasicus* (Anura: Hylidae) from a South American subtropical area. *Interciencia*, 34 (3): 214-218.
- Hamann, M. I. & Pérez, D. V., 1999. Presencia de *Haematoloechus longipectus* Stafford, 1902 (Trematoda, Haematoloechidae) em anfíbios argentinos. *Facena*, 15: 157-162.
- Heyneman, D., Brenes, M. R. R. & Diaz-Ungria, C., 1960. Trematodos de Venezuela – II. Tremátodos de peces, reptiles y aves con description de em nueva especies del género *Lubens*. *Memoria de la Sociedad de Ciencias Naturales La Salle, Caracas, Venezuela*, 20 (56): 138-149.
- Holcman-Spector, B. & Mañé-Garzón, F., 1973. A new species of the genus *Liophistrema* Ruiz and Leão, 1942 (Digenea, Ophisthognimidae). *Neotropica*, 19 (58): 11-14.

- Hoppe, E. G. L., Pedrassani, D., Hoffmann-Inocente, A. C., Tebaldi, J. H., Storti, L. F., Zannuzzo, E. S., Avancini, N. & Nascimento, A. A., 2008. Ecological studies on helminthic taxocenosis of sympatric *Chaunus ictericus* (Spix, 1824) and *Chaunus schneideri* (Werner, 1894) (Anura: Bufonidae) captured on São Cristóvão district, Três Barras county, Santa Catarina State, Brazil. *Brazilian Journal of Veterinary Parasitology*, 17 (supl. 1): 166-169.
- Iannacone, J., 2003a. Helminths parasites of *Atelopus bomolochus* Peters, 1973 (Anura: Bufonidae) from Piura, Peru. *Gayana*, 67 (1): 9-15.
- Iannacone, J., 2003b. Helminth parasites of *Telmatobius jelskii* (Peters) (Anura, Leptodactylidae) from Lima, Peru. *Revista Brasileira de Zoologia*, 20 (1): 131-134.
- Ibáñez, N. H., 1980. Fauna helmintologica peruana. *Rudolphitrema rudolphii* (Travassos, 1924) parasito de *Atelopus levis* Guenther de Cajamarca, Peru. *Boletín Peruano de Parasitología*, 2 (1/2): 38-45.
- Ibáñez, N. H., 1998. Mención de algunos trematodos em la fauna helmintológica peruana. *Revista Peruana de Parasitología*, 13: 90-97.
- Ibáñez, N. H. & Córdova, E. B., 1979. Algunos trematodos de *Telmatobius* del sur del Perú. *Boletín Peruano de Parasitología*, 1 (2): 54-66
- Incorvaia, I. S., 1983. *Catadiscus hylae* sp. n. (Trematoda: Paramphistomidae) parasite intestinal de *Hyla pulchella* Dumeril y Bibron, 1841 (Anura – Hylidae) de la provincia de Buenos Aires – Argentina. *Neotropica*, 29: 91-95.
- Jones, A., 2005. Superfamily Paramphistomoidea Fishoeder, 1901. In: Jones, A., Bray, R. A. & Gibson, D. I. (Eds) *Keys to the Trematoda*. Vol. 2. CABI Publishing and The Natural History Museum, Wallingford: 221-356.
- Jones, A., Bray, R. A. & Gibson, D. I., 2005. *Keys to the Trematoda*. Vol. 2. Eds. A. Jones, R. A. Bray & D. I. Gibson. CABI Publishing, UK, and The Natural History Museum, London, xvi + 745 pp.
- Kehr, A. I., Manly, B. F. J. & Hamann, M. I., 2000. Coexistence of helminth species in *Lysapsus limellus* (Anura: Pseudidae) from a Argentinean subtropical area: influence of biotic and abiotic factors. *Oecologia*, 125 (4): 549-558.
- Knoff, M., Brooks, D. R., Mullins, M. C. & Gomes, D. C., 2012. A new genus and a new species of Cladorchiidae (Digenea: Dadayiinae) from *Podocnemis expansa* (Chelonia) of the neotropical region, State of Para, Brazil. *Journal of Parasitology*, 98 (2): 378-381.
- Kohn A, Fernandes B. M. M., 1976. Sobre uma nova espécie do gênero *Leurosoma* Ozaki, 1932 (Trematoda, Allocreadiidae) parasito de ofídeo. *Atas da Sociedade de Biologia do Rio de Janeiro*, 18: 87-89.

- Kohn, A. & Fernandes, B. M. M., 1988. Revision of the Brazilian species of the genus *Halipegus* Looss, 1899 (Trematoda: Derogenidae). *Systematic Parasitology*, 11: 129-137.
- Kohn, A., Fernandes, B. M. M. & Cohen, S. C., 2007. South American Trematodes Parasites of Fishes. Rio de Janeiro: Editora Imprinta, 318 pp.
- Kostadinova, A., 2005a. Family Echinostomatidae Looss, 1899 In: Jones, A., Bray, R. A. & Gibson, D. I. (Eds) *Keys to the Trematoda*. Vol. 2. CABI Publishing and The Natural History Museum, Wallingford: 9-64.
- Krasnolobova, T. A., 1977. Principy sistematiki trematodroda *Plagiorchis* Luehe, 1899. *Trudy Gel'mintologicheskoi Laboratorii Akademii Nauk SSR*, 27: 65-110.
- Leão, A. T., 1946. Sobre um novo gênero de Liosphistrematinae Artigas, Ruiz & Leão, 1942 (Trematoda, Plagiorchiidae). *Memórias do Instituto Butantan*, 19: 33-40.
- Leão, A. T. & Ruiz, J. M. 1943. Notas helmintológicas 7. *Opisthogonimus fariai* n. sp. (Trematoda, Opisthogoniminae). *Anais da Faculdade de Farmácia e Odontologia da Universidade de São Paulo*, 3: 96-104.
- Led, J. E. & Boero, J. J., 1973. Parasitoses of the autochthonous fauna. *Revista de Agronomia y de Veterinaria, Buenos Aires*, 2 (4): 17-18.
- Lenis, C., Arredondo, J. C. & Calle, J. I., 2009. *Ochetosoma heterocoelium* (Digenea: Plagiorchiidae) in snakes from Colombia. *Revista Mexicana de Biodiversidad*, 80 (3): 603-609.
- Lenis, C. & Vélez, I., 2011. Digeneans in *Trachemys callirrostris callirrostris* and *Podocnemis lewyana* (Testudinata) from the Magdalena River, Colombia: description of *Pseudonematophila* n. gen. and amendment of *Nematophila* Travassos, 1934 (Cladorchiidae: Schizamphistominae). *Zootaxa*, 3095: 49-62.
- Lent, H. & Freitas, J. F. T., 1937. Pesquisas helmintológicas realizadas no Estado do Pará. I. Trematoda: Fascioloidea. *Memórias do Instituto Oswaldo Cruz*, 32: 449-460.
- Lent, H. & Freitas, J. F. T., 1938. Pesquisas helmintológicas realizadas no Estado do Pará. III. Um raro parasito de tartarugas fluviais do Amazonas. *Memórias do Instituto Oswaldo Cruz*, 33 (1): 57-61.
- Lent, H. & Freitas, J. F. T., 1939. Pesquisas helmintológicas realizadas no Estado do Pará. VII. Trematoda, Paramphistomoidea. *Boletim de Biologia*, 4 (1): 82-86.
- Lent, H., Freitas, J. F. T. & Proença, M. C., 1946. Alguns helmintos de batráquios colecionados no Paraguai. *Memórias do Instituto Oswaldo Cruz*, 44 (1): 195-214.

- Lombardero, O. J. & Moriena, R. A., 1977. Nuevos trematodos para la Argentina en *Phrynosops hilarii* (Duméril y Briçon). *Revista de Medicina Veterinaria*, 58: 64-68.
- Lunaschi, L. I. & Drago, F. B., 2001. Opisthognimid digeneans parasites of *Waglerophis merremii* (Reptilia) from Argentina, with the description of *Opisthognimus misionesensis* sp. nov. *Physis, Sec. A, B y C.*, 58 (134/135) 31-37.
- Lunaschi, L. I. & Drago, F. B., 2002. Primer registro de *Catadiscus uruguayensis* Freitas y Lent, 1939 (Digenea: Diplodiscidae) como parásito de reptiles. *Neotropica*, 48: 65-67.
- Lunaschi, L. I. & Drago, F. B., 2007. Checklist of digenean parasites of amphibians and reptiles from Argentina. *Zootaxa*, 1476: 51-68.
- Lunaschi, L. I. & Drago, F. B., 2010. Platyhelminthes, Trematoda, Digenea Carus, 1863: Distribution extension in Argentina and new Anura and Ophidia hosts. *Check List*, 6 (3): 447-450.
- Lunaschi, L. I. & Sutton, C. A., 1985. Trematodes de reptiles incorporados a la colección helmintológica del Museo de La Plata. *Neotropica*, 31 (85): 69-81.
- Lunaschi, L. I. & Sutton, C. A., 1990. Presencia de *Paradiplostomum abbreviatum* (Brandes, 1888) (Digenea, Proterodiplostomidae) em *Caiman latirostris* (Daud.) em Argentina. *Neotropica*, 36: 123-127.
- Luque, J. L., Martins, A. N. & Tavares, L. E. R., 2005. Community structure of metazoan parasites of the yellow Cururu toad, *Bufo ictericus* (Anura, Bufonidae) from Rio de Janeiro, Brazil. *Acta Parasitologica*, 50 (3): 215-220.
- Lutz, A., 1926. Societé de Biologie de Rio de Janeiro, Session de 15 de september de 1926. *Memórias do Instituto Oswaldo Cruz*, 19: 237-238.
- Lutz, A., 1928. Estudios sobre trematodes observados em Venezuela. *Estudios de Zoologia y Parasitologia Venezolanas*: 133 pp.
- MacCallum, W. G., 1921. Studies in Helminthology. *Zoopathologica*, 1 (6): 140-284.
- Mañé-Garzón, F., 1958. Em nouveau trematode des batraciens de l'Uruguay. *Catadiscus corderoi* n. sp. *Comunicaciones Zoologicas del Museo de Historia Natural de Montevideo*, 4: 1-3.
- Mañé-Garzón, F. & Alonso, A., 1976. Em nueva especie de Digenea Strigeoidea. *Heterodiplostomum helicopsis* n. sp. del Instituto de la culebra de agua *Helicops carinicaudus* (Wied, 1825). *Revista de Biología del Uruguay*, 4: 85-91.
- Mañé-Garzón, F. & Gil, A., 1959. Dos nuevas especies del género *Haematoloechus* Looss, 1899, de ranas del Uruguay. *Actas y Trabajos del Primer Congreso Sudamericano de Zoología, La Plata*: 217-222.

- Mañé-Garzón, F. & Gil, A., 1961a. Trematodos de las tortugas del Uruguay II. *Comunicaciones Zoológicas del Museo de Historia Natural de Montevideo*, 5: 1-6.
- Mañé-Garzón, F. & Gil, A., 1961b. Trematodos de las tortugas del Uruguay IV. Tres nuevas especies del género *Telorchis* Lühe, 1900. *Comunicaciones Zoológicas del Museo de Historia Natural de Montevideo*, 5: 1-7.
- Mañé-Garzón, F. & Gil, A., 1961c. Trematodos de las tortugas del Uruguay III. Una nueva especie del género *Telorchis* Lühe, 1900 (Trematoda, Telorchidae). *Neotropica*, 7: 38-42.
- Mañé-Garzón, F. & Gil, A., 1963. A new trematode of *Rana palmipes* of Peru. *Iquitos ceii* n. gen. n. sp. *Neotropica*, 9: 124-128.
- Mañé-Garzón, F. & Gonzáles, L. E., 1978a. *Gorgoderina darwini* n. sp. digenean parasite from the urinary bladder of *Melanophryniscus stelzneri*. *Revista de Biología del Uruguay*, 6: 39-43.
- Mañé-Garzón, F. & Gonzáles, L. E., 1978b. Two species of *Gorgoderina* (*Gorgoderimma*) from the urinary bladder of *Leptodactylus ocellatus* from Uruguay. *Revista de Biología del Uruguay*, 6: 45-50.
- Mañé-Garzón, F. & Gortari, A. M. 1965. Sobre algunos trematodos de ofideos del Uruguay. *Comunicaciones Zoológicas del Museo de Historia Natural de Montevideo*, 8: 1-21.
- Mañé-Garzón, F. & Holcman-Spector, B., 1967a. A nueva especie del género *Margeana* (Digenea), parasita de *Pseudis mantidactylus* (Cope). *Comunicaciones Zoológicas del Museo de Historia Natural de Montevideo*, 9 (113): 1-4.
- Mañé-Garzón, F. & Holcman-Spector, B., 1967b. *Margeana chaquensis* n. sp. (Digenea) parasita de *Leptodactylus laticeps* del chaco argentino. *Comunicaciones Zoológicas del Museo de Historia Natural de Montevideo*, 115 (9): 1-4.
- Mañé-Garzón, F. & Holcman-Spector, B., 1967c. *Styphlodora gili* nov. sp. (Digenea) parasito de las vías urinarias de *Bothrops alternata*. *Comunicaciones Zoológicas del Museo de Historia Natural Montevideo*, 9 (114): 1-6.
- Mañé-Garzón, F. & Holcman-Spector, B., 1968. Trematodos de las tortugas del Uruguay VIII. Em nueva especie del género *Telorchis* (Lühe, 1900) del intestino de *Pseudemys dorbignyi* (Dum & Bib). *Comunicaciones Zoológicas del Museo de Historia Natural de Montevideo*, 9: 1-4.
- Mañé-Garzón, F. & Holcman-Spector, B., 1973a. Trematodes of turtles of Uruguay, X. *Telorchis achavali* n. sp. from the small intestine of *Pseudemys dorbignyi*. *Revista de Biología del Uruguay*, 1 (1): 5-9.

- Mañé-Garzón, F. & Holcman-Spector, B., 1973b. A new species of the genus *Opisthogonimus* (Digenea, Opisthogonimidae). *Neotropica*, 19 (60): 118-121.
- Mañé-Garzón, F. & Holcman-Spector, B., 1974. Trematodes of amphibians of Uruguay I. Species of the genera *Margeana* Cort, 1919, *Glypthelmins* Stafford, 1905 and *Plagiorchis* Lühe, 1899. *Revista de Biología del Uruguay*, 2 (2): 101-117.
- Mañé-Garzón, F. & Ponce de León, R., 1976. *Rudolphitrema physalaemi* n. sp. nueva especie de digenea Plagiorchiidae del intestino de la ranita gato *Physalaemus gracilis* (Boulenger) del Uruguay. *Revista de Biología del Uruguay*, 4 (2): 93-97.
- Martínez, F. A., Troiano, J. C., Binda, J. L., Selles, D. E., Jara, D. & Fescina, N., 1996. Trematodes of some ophidians of the north east of Argentina. *Cuadernos de Herpetología*, 9 (2): 85-94.
- Masi-Pallarés, R. & Benitez-Usher, C. A., 1973. List of helminth parasites of fish and reptiles in Paraguay. *Revista Paraguaya de Microbiología*, 8 (1): 113-118.
- Masi-Pallarés, R., Benitez-Usher, C. A. & Maciel, S., 1976. Lista dos helmintos del Paraguay. *Revista Paraguaya de Microbiología*, 11 (1): 43-59.
- Masi-Pallarés, R., Benitez-Usher, C. A. & Vergara, G., 1973. Helminths of fish and reptiles in Paraguay (Part I). *Revista Paraguaya de Microbiología*, 8 (1): 67-112.
- Masi-Pallarés, R. & Maciel, S., 1974. Helminthes en batracios del Paraguay (Ira. Parte), con descripción de una nueva especie *Aplectana pudenda* (Oxyuridae: Cosmocercinae). *Revista Paraguaya de Microbiología*, 9: 55-60.
- McIntosh, A., 1939. A new dicrocoeliid trematode collected on the Presidential Cruise of 1938. *Smithsonia Miscellaneous Collection*, 98 (16): 2pp.
- Mehra, R. K., 1939. New monostomes of the family Pronocephalidae Looss, 1902. *Proceedings of the National Academy of Sciences, India*, 9: 99-130.
- Miyazaki, I., Kifune, T., Habe, S. & Uyema, N., 1978. Reports of Fukuoka University Scientific Expeditions to Peru, 1976. Part 1. General account of the expedition and records of helminth parasites of wild mammals, molusks and insects. *Ocassional Publication, Department of Parasitology of the School of Medicine of the Fukuoka University*, 1: 1-28.
- Nasir, P., 1966. Two new species of digenetic trematodes from Venezuelan amphibians. *Proceedings of the Helminthological Society of Washington*, 33 (2): 166-170
- Nasir, P., 1974. Revision of the genera *Acanthostomum* Looss, 1899 and *Telorchis* Lühe, 1899 (Trematoda: Digenea) with redescription of *Acanthostomum* (*Acanthostomum*) *scy-*



- phocephalum* (Braun, 1899) and *Telorchis aculeatus* (Von Linstow, 1879) Braun, 1901. *Rivista di Parassitologia*, 35 (1): 1-22.
- Nasir, P. & Diaz, M. T., 1970. A redescription of *Glyphelminis linguatula* (Rudolphi, 1819) Travassos, 1924 and *G. vesicalis* (Ruiz and Leão, 1942) Yamaguti, 1958 with a key to the valid species. *Rivista di Parassitologia*, 31 (4): 261-274.
- Nasir, P. & Diaz, M. T., 1971a. A redescription of *Mesocoelium monas* (Rudolphi, 1819) Freitas, 1958, and specific determination in genus *Mesocoelium* Odhner, 1910 (Trematoda, Digenea). *Rivista di Parasitologia*, 32 (3): 149-158.
- Nasir, P. & Diaz, M. T., 1971b. Flukes from Venezuelan reptiles with observations on intra-specific variations. *Rivista di Parassitologia*, 32 (4): 233-248.
- Nasir, P. & Rodriguez, M. L., 1967. *Proterodiplostomum intermedium* n. sp. (Trematoda: Digenea) from the crocodile *Caiman crocodilus crocodilus* (L.) in Venezuela. *Proceedings of the Helminthological Society of Washington*, 34 (2): 144-146.
- Niewiadomska, K., 2002. Family Proterodiplostomidae Dubois, 1936. In: Gibson, D. I., Jones, A. & Bray, R. A., 2002. *Keys to the Trematoda*. Vol. 1. Eds. D. I. Gibson, A. Jones & R. A. Bray. CABI Publishing, Wallingford, UK, and The Natural History Museum, London: 215-229.
- Noronha, D., M., R., Sá, M., Knoff, M., Muniz-Pereira, L. C. & Pinto, R. M., 2009. *Adolfo Lutz e a Coleção Helminológica do Instituto Oswaldo Cruz*. Museu Nacional, Rio de Janeiro: 154 pp.
- O'Brien, R. T., Sidner, R. A. & Etges, F. J., 1979. *Sphaeridiotrema echinosaurense* sp. n. (Trematoda, Psilostomidae) from *Echinosaura horrida* in Ecuador. *Proceedings of the Helminthological Society of Washington*, 46 (2): 185-187.
- Odhner, T., 1902. Mitteilung zur Kenntnis der Distomen II. *Zentralblatt für Bakteriologie, Parasitenkunde und Infektionskrankheiten*, 31: 152-162.
- Odhner, T., 1911. Zum natürlichen System der digenen Trematoden. *Zoologischer Anzeiger*, 37 (8-9) 181-191.
- Odhner, T., 1912. Zum natürlichen System der digenen Trematoden. V. *Zoologischer Anzeiger*, 41 (2): 54-71.
- Olmos, V. & Muñoz, G., 2006. Current state of knowledge of eumetazoan parasites of Chilean freshwater ecosystems. *Gayana*, 70 (1): 122-139.
- Ostrowski de Núñez, M., 1978/1979. Studies on the freshwater fauna of Argentina. IX. On members of the family Paramphistomatidae (Trematoda). *Physis, B*, 38 (95): 55-62

- Ostrowski de Núñez, M., 1979. Ungewöhnlich Xiphidiocercariae aus *Ampullaria canaliculata* nebst Bemerkungen über *Travtrema stenocotyle*. *Angewandte Parasitologie*, 33: 733-755.
- Ostrowski de Núñez, M., 1984a. Redescription of *Acanthostomum marajoarum* from the original specimens. *Physis*, 42 (102): 25-27.
- Ostrowski de Núñez, M., 1984b. Beiträge zur Gattung *Acanthostomum* (Trematoda, Acanthostomidae) und zu den Entwicklungszyklen von *A. marajoarum* (Freitas & Lent, 1938) und *A. loossi* (Perez Viguera, 1957) in Venezuela. *Mitteilungen aus dem Zoologischen Museum in Berlin*, 60 (2): 179-201.
- Ostrowski de Núñez, M., 1986. *Acanthostomum scyphocephalum* (Braun, 1899) Hughes, Higginbotham und Clary, 1941: Neubeschreibung des typischen material aus dem Naturhistorischen Museum in Wien. *Annalen des Naturhistorischen Museums in Wien*, 87: 331-337.
- Ostrowski de Núñez, M., 1987. Der Entwicklungszyklus von *Acanthostomum brauni* Mañé-Garzón und Gil, 1961 (Trematoda, Acanthostomatidae). *Zoologischer Anzeiger*, 218 (5/6): 273-286.
- Ostrowski de Núñez, M., 2003. Digenean trematodes of crocodiles collected by Johann Natterer in Brazil, deposited in the Natural History Museum, Vienna. *Annalen des Naturhistorischen Museums in Wien Serie B, Botanik und Zoologie*, 104B: 399-413.
- Paraense, W. L., 1992. *Halipegus dubius* Klein, 1905 (Trematoda, Hemiuridae): a redescription, with notes on the working of the ovarian complex. *Memórias do Instituto Oswaldo Cruz*, 87, Supp. I: 179-190.
- Pereira, C., 1928. Fauna helmintológica dos ophideos brasileiros III. *Boletim Biológico*, 12: 50-54.
- Pereira, C., 1929a. Revisão do gênero *Opisthogonimus* (Trematoda). *Revista do Museu Paulista*, 16: 995-1014.
- Pereira, C., 1929b. *Travtrema travtrema* n. gen. e n. sp., trematóide parasito do intestino de cobra. *Boletim Biológico*, 16: 92-96.
- Pereira, C. & Cuocolo, R., 1940a. Trematóides brasileiros do gênero *Mesocoelium* Odhner. *Arquivos do Instituto de Biologia*, São Paulo, 11 (43): 399-412.
- Pereira, C. & Cuocolo, R., 1940b. Trematóides vesicais de anfíbios do nordeste brasileiro. *Arquivos do Instituto de Biologia*, São Paulo, 11 (44): 413-420.
- Pereira, C. & Cuocolo, R., 1941. Processo papilomatoso das vias biliares de *Leptodactylus ocellatus* (L.), determinado por *Choledocystus eucharis* n. g., n. sp. (Trematoda: Plagiorchiidae). *Arquivos do Instituto de Biologia*, São Paulo, 12 (23): 311-324.

- Perez, M. D., 1964. Trematódeos digenéticos parasitos de anura (Amphibia) da América do Sul. *Faculdade de Farmácia e Bioquímica da Universidade de São Paulo*: 152.
- Pérez Ponce de León, G. & Brooks, D. R., 1995. Phylogenetic Relationships among the species of *Pyelosomum* Looss, 1899 (Digenea: Pronocephalidae). *Journal of Parasitology*, 81(2): 278-280.
- Pinto, H. A., Mati, V. L. T. & Melo, A. L., 2012. New hosts and localities for trematodes of snakes (Reptilia: Squamata) from Minas Gerais State, Southeastern Brazil. *Comparative Parasitology*, 79 (2): 238-246.
- Pinto, R. M. & Noronha, D., 1972. Contribuição ao conhecimento da fauna helmintológica do Município de Alfenas, Estado de Minas Gerais. *Memórias do Instituto Oswaldo Cruz*, 70 (3): 391-407.
- Poumarau, E. M. C., 1965. *Catadiscus longicoecalis* nueva especie parasita de ofidios (Trematoda; Paramphistomidae) con una lista de especies del género *Catadiscus* Cohn, 1904. *Physis, Buenos Aires*, 25: 277-282.
- Poumarau, E. M. C., 1968. Trematodes de ofidios de la Argentina. *Revista Museo Argentino de Ciencias Naturales Bernardino Rivadavia*, 1: 1-129.
- Prudhoe, S., 1949. Some roundworms and flatworms from west Indies and Surinam III. Trematodes. *Journal of the Linnaean Society Zoology*, 41 (281): 415-419.
- Puga, S. R., 1979. *Gorgoderina valdiviensis* n. sp., a new digenetic trematode (Gorgoderidae) parasite of Chilean frog *Caudiverbera caudiverbera*. *Studies on Neotropical Fauna and Environment*, 14 (4): 227-232.
- Puga, S. R., 1982. Two platyhelminths from *Caudiverba caudiverba* (Anura: Leptodactylidae) in Southern Chile. *Herpetological Review*, 13 (1): 13-14.
- Puga, S. R., 1986. *Rudolphitrema chilensis* sp. nov., a new digenetic trematode parasite from the Chilean anuran *Eusophus roseus* (Leptodactylidae). *Boletín Chileno de Parasitología*, 41 (1-2): 13-16.
- Puga, S. R., 1994. Helminthological fauna of Chilean anura. *Boletín Chileno de Parasitología*, 49: 81-84.
- Puga, S. R. & Torres, P., 1999. Helminth parasites of *Eupsophus roseus* (Anura: Leptodactylidae) from Southern Chile. *Memórias do Instituto Oswaldo Cruz*, 94 (6): 725-726.
- Ramalho, A. C. O., Silva, R. J., Schwartz, H. O. & Peres Jr., A. K., 2009. Helminths from an introduced species (*Tupinambis merianae*), and two endemic species (*Trachylepis atlantica* and *Amphisbaena ridleyi*) from Fernando de Noronha Archipelago, Brazil. *Journal of Parasitology*, 95 (4): 1026-1028.

- Razo-Mendivil, U. J., León-Règagnon, V. & Pérez-Ponce de León, G. P., 2006. Monophyly and systematic position of *Glyphelminis* (Digenea), based on partial 18S rDNA sequences and morphological evidence. *Organisms Diversity and Evolution*, 6 (4): 308-320.
- Rego, A. A. & Vicente, J. J., 1988. Scientific expedition to collect helminths in Pantanal, Mato Grosso. *Ciência e Cultura*, 40 (1): 65-68.
- Rodrigues, H. O., 1968. Novos hospedeiros de *Paradistomum parvissimum* (Travassos, 1918) Travassos, 1919 (Trematoda, Dicrocoeliidae). *Atas da Sociedade de Biologia do Rio de Janeiro*, 11 (5): 167-168.
- Rodrigues, H. O., 1970. Estudo da fauna helmintológica de *Hemidactylus mabouia* (M. de J.) no Estado da Guanabara. *Atas da Sociedade de Biologia do Rio de Janeiro*, 12 suppl.: 15-23.
- Rodrigues, H. O., 1986. Contribuição ao estudo da fauna helmintológica de vertebrados de Nova Iguaçu, RJ. *Atas da Sociedade de Biologia do Rio de Janeiro*, 26: 27-28.
- Rodrigues, H. O., 1992. *Pseudocapillaria* (*Ichthyocapillaria*) *maricaensis* n. sp. (Nematoda, Capillariidae) and remarks on the helminthological fauna of *Liolaemus lutzae* Mertens, 1938 (Lacertilia, Iguanidae). *Memórias do Instituto Oswaldo Cruz*, 87 (2): 297-300.
- Rodrigues, H. O., 1994. *Plagiorchis vicentei* sp. n. (Trematoda, Plagiorchiidae), a new trematode from *Hemidactylus mabouia* (Moreau de Jonnes) (Lacertilia, Gekkonidae). *Revista Brasileira de Zoologia*, 11 (4): 669-672.
- Rodrigues, H. O., Rodrigues, S. S. & Cristóforo, R., 1978. Subsídios ao estudo dos trematódeos parasitos de anfíbios de Barra do Piraí, Estado do Rio de Janeiro. *Atas da Sociedade de Biologia do Rio de Janeiro*, 19: 25-29.
- Rodrigues, H. O., Rodrigues, S. S., Cristóforo, R., 1982. Contribuição ao conhecimento da fauna helmintológica de anfíbios de Barra do Piraí, Estado do Rio de Janeiro. *Atas da Sociedade de Biologia do Rio de Janeiro*, 23: 5-8.
- Rodrigues, H. O., Rodrigues, S. S. & Faria, Z., 1990. Contribuição ao conhecimento da fauna helmintológica dos vertebrados de Maricá, Estado do Rio de Janeiro, Brasil. *Memórias do Instituto Oswaldo Cruz*, 85 (1): 115-116.
- Rodrigues, H. O. & Santos, E., 1974. Notas sobre dois helmintos parasitos de répteis. *Atas da Sociedade de Biologia do Rio de Janeiro*, 17 (2): 55-58.
- Rudolphi, C. A., 1819. *Entozoorum synopsis cui accedunt mantissa duplex et indices locuple tissimi*, Berolini: 811 pp.

- Ruiz, J. M., 1943a. *Catadiscus Freitaslenti* sp. n. (Trematoda: Paramphistomoidea), parasito de ofídeo neotrópico: observações sobre a presença de dois canais eferentes no gênero *Catadiscus* Cohn, 1904. *Memórias do Instituto Butantan*, 17: 29-33.
- Ruiz, J. M., 1943b. *Neoctangium travassosi*, gen. n., sp. n., (Trematoda: Paramphistomoi-dea), parasito de quelônio marinho. Chave dos gêneros da família Microscaphidiidae Travassos, 1922. *Memórias do Instituto Butantan*, 17: 35-45.
- Ruiz, J. M., 1946. Pronocephalida (Trematoda). Estudo das espécies brasileiras e revisão da família. *Memórias do Instituto Butantan*, 19 : 249-372.
- Ruiz, J. M., 1949. Considerações sobre o gênero *Choledocystus* Pereira & Cuocolo, 1941 (Trematoda: Plagiorchiidae). *Revista Brasileira de Biologia*, 9 (2): 167-174.
- Ruiz, J. M., 1951. Estudo do sistema excretor de *Leptophyllum stenocotyle* Cohn, 1902 (Trematoda: Plagiorchiidae). *Memórias do Instituto Butantan*, 23: 45-49.
- Ruiz, J. M. & Leão, A. T., 1942a. Notas helmintológicas: Três novas espécies de *Opisthogoni-mus* parasitas de ofídeos brasileiros (Trematoda: Plagiorchiidae). *Memórias do Instituto Butantan*, 16: 171-185.
- Ruiz, J. M. & Leão, A. T., 1942b. Notas helmintológicas. 2. Algumas considerações em torno do gênero *Leptophyllum* Cohn, 1902 (Trematoda: Plagiorchiidae). *Memórias do Instituto Butantan*, 16: 187-195.
- Ruiz, J. M. & Leão, A. T., 1942c. Notas helmintológicas. 4. *Choledocystus vesicalis* n. sp., parasita de vesícula biliar de *Bufo marinus* (L.) (Trematoda: Plagiorchiidae). *Memórias do Instituto Butantan*, 16 : 209-212.
- Ruiz, J. M. & Leão, A. T., 1943a. Notas helmintológicas. 6. *Cyathocotyle brasiliensis* n. sp. (Trematoda, Cyathocotylidae), parasito de *Caiman sclerops* (Gray) do Brasil. *Revista Brasileira de Biologia*, 3 (2): 191-198.
- Ruiz, J. M. & Leão, A. T., 1943b. Notas helmintológicas III. Nova espécie de trematóide do gênero *Infidum* Travassos, 1916 (Dicrocoellidae) parasito de ofídeo brasileiro. *Memórias do Instituto Oswaldo Cruz*, 16: 203-207.
- Ruiz, J. M. & Leão, A. T., 1955. Notas helmintológicas. *Aliptrema riberoi* n. gen., n. sp. (Trematoda: Plagiorchiidae), parasita de ofídeo brasileiro. *Arquivos do Museu Nacional*, 42 (2): 485-487.
- Ruiz, J. M. & Perez, M. D., 1959. Gênero *Haplometroides*, redescrição da espécie tipo e de-scrição de *H. odhneri* sp. n. (Trematoda, Plagiorchiidae). *Anais da Faculdade de Farmácia e Odontologia da Universidade de São Paulo*, 16: 87-91.

- Ruiz, J. M. & Rangel, J. M., 1954. Estrigêidas de répteis brasileiros (Trematoda: Strigeata). *Memórias do Instituto Butantan*, 26: 257-278.
- Salízar, P. & Sánchez, L., 2004. First record for Peru of *Nematophila grandis* (Diesing, 1839) Travassos, 1934 (Trematoda, Diplodiscidae) in *Podocnemis unifilis* (Troschel, 1848) (Testudines, Pelomedusidae). *Revista Peruana de Biología*, 11 (1): 37-40.
- Sánchez, N., Tantaleán, M., Vela, D. & Méndez, A., 2006. Gastrointestinal parasites of taricaya, *Podocnemis unifilis* (Troschel, 1848) (Testudines: Podocnemididae) from Iquitos, Peru. *Revista Peruana de Biología*, 13 (1): 119-120.
- Santos, K. R., Barrella, T. H., Zica, E. O. P. & Silva, R. J., 2008. New reports on parasitism by *Haplometroides buccicola* (Digenea, Plagiorchiidae) in Brazilian snakes. *Journal of Venomous Animals and Toxins Including Tropical Diseases*, 14 (3): 527-532.
- Santos, V. G. T. & Amato, S. B., 2010. Helminth fauna of *Rhinella fernandezae* (Anura, Bufonidae) from the Rio Grande do Sul Coastland, Brazil - analysis of the parasitic community. *Journal of Parasitology*, 96 (4): 823-826.
- Santos, V. G. T., Amato, S. B. & Borges-Martins, M., 2013. Community structure of helminth parasites of the “Cururu” toad, *Rhinella icterica* (Anura: Bufonidae) from southern Brazil. *Parasitology Research*, 112 (3):1097-1103.
- Savazzini, L. A., 1930. Contribución el estudio de parásitos de los aparatos circulatorio y digestivo de nuestro *Leptodactylus ocellatus*. Nuevas especies de nematodes, cestodes y trematodes. *Tesis de la Escola de Farmacia de la Universidad del Litoral, Buenos Aires*: 43 pp.
- Schaefer, E. F., Hamann, M. I., Kehr, A. I., Gonzalez, C. E. & Duré, M. I., 2006. Trophic, reproductive and parasitological aspects of the ecology of *Leptodactylus chaquensis* (Anura: Leptodactylidae) in Argentina. *Herpetological Journal*, 16 (4): 387-394.
- Silva, R. J., 2004. Note on the feeding habits of *Opisthogonimus lecithonotus* (Trematoda, Digenea, Plagiorchiidae). *Parasitology Research*, 94 (6): 471-472.
- Silva, R. J., 2005. Copulation of *Opisthogonimus fonsecai* Ruiz & Leão, 1942 (Trematoda, Digenea, Plagiorchiidae), parasite of *Bothrops moojeni* Hoge, 1966 (Serpentes, Viperidae). *The Journal of Venomous Animals and Toxins including Tropical Diseases*, 11 (1): 68-75.
- Silva, R. J., 2008. New record of *Haplometroides intercaecalis* (Digenea, Plagiorchiidae) infecting a Brazilian snake. *Journal of Venomous Animals and Toxins including Tropical Diseases*, 14 (1): 161-165.

- Silva, R. J., Andrade, P. A., Monteiro e Silva, H. A., Rossellini, M. & Barrella, T. H., 2005. Report on the occurrence of *Haplometroides buccicola* (Trematoda, Digenea, Plagiorchiidae) infecting *Phalotris lativittatus* (Serpentes, Colubridae) in Brazil. *The Journal of Venomous Animals and Toxins including Tropical Diseases*, 11 (3): 373-379.
- Silva, R. J. & Barrella, T. H., 2002. *Micrurus frontalis* as a new host recorded for *Haplometroides odhneri* (Trematoda, Digenea, Plagiorchiidae). *Revista Brasileira de Parasitologia Veterinária*, 11 (1): 47-48.
- Silva, R. J., Beda, A. F. & Ferreira, V. L. 2008. New record of *Haplometroides intercaecalis* (Digenea, Plagiorchiidae) infecting a Brazilian snake. *The Journal of Venomous Animals and Toxins including Tropical Diseases*, 14 (1): 161-165
- Silva, R. J., Ferreira, V. L. & Strüssmann, C., 2007. New species of *Haplometroides* (Digenea: Plagiorchiidae) from *Phalotris nasutus* (Gomes, 1915) (Serpentes, Colubridae). *Journal of Parasitology*, 93 (4): 917-921.
- Silva, R. J., Rodrigues, R. R., Stein, M. F. B., Sipoli, G. P.M., Pinhão, R. & Lopes, C. A. M., 1999. The detection of *Ochetosoma heterocoelium* (Travassos, 1921) (Trematoda: Digenea: Ochetosomatidae) in *Chironius exoletus* (Linnaeus, 1758) (Ophidia: Colubridae). *The Journal of Venomous Animals and Toxins including Tropical Diseases*, 5 (1): 85-90.
- Silva, R. J., Zica, E., O., P., Cruz, M., O' Reilly, J. C. & Costa, M. C., 2005. Occurrence of *Haplometroides odhneri* (Trematoda, Digenea, Plagiorchiidae) infecting *Leptotyphlops koppesi* (Serpentes, Leptotyphlopidae). *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, 57 (supl. 2): 267-269.
- Silva, T. B., Rossellini, M., Dal-Pai, S. M. & Silva, R. J., 2005. Histological characterization of *Sticholecitha serpentis* Prudhoe, 1949 (Digenea, Bieriidae, Sticholecithinae), parasite of *Bothrops moojeni* Hoge, 1966 (Serpentes, Viperidae). *Journal of Venomous Animals and Toxins including Tropical Diseases*, 11 (4): 510-531.
- Silva, T. B., Rossellini, M., Silva, M. D. P. & Silva, R. J., 2005. Histological characterization of *Sticholecitha serpentis* Prudhoe, 1949 (Digenea, Bieriidae, Sticholecithinae), parasite of *Bothrops moojeni* Hoge, 1966 (Serpentes, Viperidae). *Journal of Venomous Animals and Toxins including Tropical Diseases*, 11 (4): 510-531.
- Stumpf, I. V. K., 1981/1982. Helminths in *Leptodactylus ocellatus* (L. 1758) in Curitiba, Brazil. *Acta Biológica Paranaense*, 10/11: 215-218.
- Stunkard, H. W. & Gandal, C. P., 1966. A digenetic trematode *Parahaplometroides basiliscae* Thatcher, 1963, from the mouth of the crested lizard, *Basiliscus basiliscus*. *Zoologica*, 51 (3): 91-95.

- Sullivan, J. J., 1976. Redescription of *Choledocystis incurvatum* (Nasir, 1966) n. comb. (Digenea: Plagiorchiidae), a parasite of *Pseudis paradoxa* (L.) in Venezuela. *Rivista di Parasitologia*, 37 (2-3): 241-245.
- Sullivan, J. J., 1977a. Revision of the genus *Rauschiella* Babero, 1951 (Digenea: Plagiorchiidae) with a redescription of *R. palmipedis* (Lutz, 1928) n. comb. from Venezuelan frogs. *Proceedings of the Helminthological Society of Washington*, 44 (1): 82-86.
- Sullivan J. J., 1977b. Redescription of *Choledocystis hepaticus* (Lutz, 1928) n. comb., and the status of *C. linguatula* (Rudolphi, 1819) (Trematoda: Plagiorchioidea). *Proceedings of the Helminthological Society of Washington*, 44 (2): 162-171.
- Suriano, D. M., 1965a. Redescrición de *Gorgoderina parvicava* Travassos trematode de la vejiga urinaria de *Leptodactylus ocellatus* (L.) de la República Argentina. *Neotropica*, 11: 19-22.
- Suriano, D. M., 1965b. Sobre *Gorgoderina australiensis* Johnston (Trematoda) parásita de *Leptodactylus ocellatus* (L.). *Neotropica*, 11: 89-94.
- Suriano, D. M., 1968. *Glypthelmins biliaris* sp. nov. (Trematoda: Brachycoelidae) parasita de *Leptodactylus ocellatus* (Amphibia, Leptodactylidae) de la Republica Argentina. *Neotropica*, 14 (43): 27-31.
- Suriano, D. M., 1970. Estudio sobre la fauna parasitaria de *Leptodactylus ocellatus* (L.) (Anfibia-Leptodactylidae) de la República Argentina. *Revista del Museo Argentino de Ciencias Naturales Bernardino Rivadavia, Zoologia*, 10 (15): 215-239.
- Suriano, D. M., 1978. Estudio sobre la fauna parasitaria de *Leptodactylus ocellatus* (L.) (Amphibia-Leptodactylidae) de la República Argentina. *Revista Museo Argentino de Ciencias Naturales Bernardino Rivadavia, Zoologia*, 10: 215-239.
- Tantaleán, M. V. & García, L., 1993. Trematodes de la familia Gorgoderidae en anfibios Leptodactylidae de la region altoandina del Peru. *Boletín de Lima*, 15 (85): 25-27.
- Tantaleán, M. V., Martínez, R. & Juárez, D., 1974/1975. Estudio de algunos trematodos del Peru. *Revista Peruana de Medicina Tropical*, 3-4: 46-56.
- Tantaleán, M. V., Sarmiento, L. B. & Huiza, A. F., 1992. Digeneos (Trematoda) del Peru. *Boletín de Lima*, 80: 47-84.
- Thatcher, V. E., 1970. Some plagiorchiid trematodes from Paraná and Colombia including *Phillandrophyllus magnacirrus* n. g., n. sp. from a marsupial and a review of *Parallopharynx*. *Transactions of the American Microscopical Society*, 89 (3): 349-354.



- Thatcher, V. E., 1993. *Trematódeos Neotropicais*. Instituto Nacional de Pesquisas da Amazônia, Manaus, Amazonas: 553 pp.
- Tkach, V. V., 2008. Family Opisthogonimidae Travassos, 1928 In: Bray, R. A., Gibson, D. I. & Jones, A., 2008. *Keys to the Trematoda*. Vol. 3. Eds. R. A. Bray, D. I. Gibson, & A. Jones. CABI Publishing, Wallingford, UK, and The Natural History Museum, London: 401-405.
- Travassos, L., 1916. Trematodeos novos. *Brazil Médico*, 30 (1): 257-258.
- Travassos, L., 1918. Helmitos parasitos de animais domésticos. I. Dicrocoelidae. *Revista de Veterinária e Zootecnia*, 8 (1): 3-15.
- Travassos, L., 1919a. Contribuição para a sistemática dos Dicrocoelinae Looss, 1899. *Aquivos da Escola Superior de Agricultura e Medicina Veterinária*, 3: 7-24.
- Travassos, L., 1919b. Novo tipo de Telorchinae. *Revista da Sociedade Brasileira de Ciências*, 3: 183-187.
- Travassos, L., 1921a. Trematódeos novos. II. *Brazil Medico*, 35 (1): 179-180.
- Travassos, L., 1921b. Trematódeos novos. III. *Brazil Medico*, 35 (1): 221-222.
- Travassos, L., 1922a. Informações sobre a fauna helmintológica de Mato Grosso. *Folha Médica*, 3 (24): 187-190.
- Travassos, L., 1922b. Contribuição para o conhecimento da fauna helmintológica brasileira. XVII. Gorgoderidae brasileiras. *Memórias do Instituto Oswaldo Cruz*, 15 (1): 220-234.
- Travassos, L., 1924a. Contribuição para o conhecimento dos helmintos dos batráquios do Brasil I. Trematódeos intestinais. *Sciencia Médica*, 2 (11): 618-628.
- Travassos, L., 1924b. Contribuição para o conhecimento dos helmintos dos batráquios do Brasil II. Trematódeos vesicais. *Sciencia Médica*, 2 (12): 746-748.
- Travassos, L., 1926a. Trematódeos novos (V). *Boletim Biologico*, 1: 16-20.
- Travassos, L., 1926b. *Catadiscus cohni* nova espécie, novo trematódeo de batrachio. *Sciencia Medica*, 4 (6): 278-279.
- Travassos, L. 1927. Trematódeos novos V. *Boletim Biológico*, 7: 95-101.
- Travassos, L. 1928. Fauna helmintológica de Mato Grosso (Trematódeos – 1ª parte). *Memórias do Instituto Oswaldo Cruz*, 21 (2): 309-341.

- Travassos, L., 1930. Pesquisas helmintológicas realizadas em Hamburgo. IV. Notas sobre o gênero *Opisthioglyphe* Looss, 1899 e gêneros próximos. *Memórias do Instituto Oswaldo Cruz*, 24 (1): 1-17.
- Travassos, L., 1934. Sinopse dos Paramphistomoidea. *Memórias do Instituto Oswaldo Cruz*, 29 (1): 19-178.
- Travassos, L., 1944a. Relatório da excursão do Instituto Oswaldo Cruz ao Município de Santa Teresa, no Estado do Espírito Santo, em agosto e setembro de 1943. *Memórias do Instituto Oswaldo Cruz*, 40 (2): 121-128.
- Travassos, L., 1944b. Revisão da família Dicrocoeliidae Odhner, 1910. *Monografia Instituto Oswaldo Cruz*, 2: 357 pp.
- Travassos, L., 1945. Relatório da excursão realizada no vale do rio Itaúnas, norte do Estado do Espírito Santo, nos meses de setembro e outubro de 1944. *Memórias do Instituto Oswaldo Cruz*, 42 (3): 487-502.
- Travassos, L., 1951. O gênero *Pulchrosoma* Travassos, 1916 e sua situação no sistema de trematódeos. *Arquivos de Zoologia do Estado de São Paulo*, 7 (9): 465-492.
- Travassos, L. & Artigas, P., 1927. *Pneumonescus neivai* n. sp., trematódeo do pulmão de rã. *Boletim Biológico*, 10: 212-214.
- Travassos, L. & Darriba, A. R., 1930. Pesquisas helmintológicas realizadas em Hamburgo. III. Trematódeos dos gêneros *Pneumonoeces* e *Ostiolum*. *Memórias do Instituto Oswaldo Cruz*, 23 (5): 237-253.
- Travassos, L. & Freitas, J. F. T., 1941a. Relatório da terceira excursão do Instituto Oswaldo Cruz, realizada à zona da Estrada de Ferro Noroeste do Brasil, em janeiro de 1941. II. Pesquisas parasitológicas. *Memórias do Instituto Oswaldo Cruz*, 36 (3): 272-295.
- Travassos, L. & Freitas, J. F. T., 1941b. Relatório da terceira excursão do Instituto Oswaldo Cruz, realizada à zona da Estrada de Ferro Noroeste do Brasil, em fevereiro e março de 1940. II. Pesquisas parasitológicas. *Memórias do Instituto Oswaldo Cruz*, 35 (3): 610-634.
- Travassos, L. & Freitas, J. F. T., 1941c. Relatório da quarta excursão do Instituto Oswaldo Cruz, realizada à zona da Estrada de Ferro Noroeste do Brasil, em agosto e setembro de 1940. Pesquisas parasitológicas. *Memórias do Instituto Oswaldo Cruz*, 35 (4): 705-721.
- Travassos, L. & Freitas, J. F. T., 1942. Relatório da sexta excursão do Instituto Oswaldo Cruz, realizada à zona da Estrada de Ferro Noroeste do Brasil, em novembro de 1941. *Memórias do Instituto Oswaldo Cruz*, 37 (3): 259-286.

- Travassos, L. & Freitas, J. F. T., 1964. Pesquisas helmintológicas realizadas em Maicuru, Estado do Pará. *Boletim do Museo Paraense Emílio Goeldi, Publicações avulsas*, 2: 1-16.
- Travassos, L., Freitas, J. F. T. & Kohn, A., 1969. Trematódeos do Brasil. *Memórias do Instituto Oswaldo Cruz*, 67 (fasc. único): 886 pp.
- Travassos, L., Freitas, J. F. T. & Mendonça, J. M., 1964. Relatório da excursão do Instituto Oswaldo Cruz ao Parque de Reserva e Refúgio Soóretama, no Estado do Espírito Santo, em outubro de 1963. *Boletim do Museu de Biologia Professor Mello Leitão, Zoologia*, 23: 1-26.
- Travassos, L., Freitas, J. F. T., Mendonça, J. M. & Rodrigues, H. O., 1962. Segunda excursão a Cabo Frio, Estado do Rio de Janeiro. *Atas da Sociedade Brasileira de Biologia do Rio de Janeiro*, 6 (4): 37-38.
- Travassos, L., Pinto, C. & Muniz, J., 1928. Excursão científica ao Estado de Mato Grosso na zona do Pantanal (margens dos rios S. Lourenço e Cuyabá) realizada em 1922. *Memórias do Instituto Oswaldo Cruz*, 20: 249-269.
- Ucrós, H., 1959. Contribución al estudio de la fauna helmintológica colombiana. *Anales de la Sociedad de Biología de Bogotá*, 8 (1): 1-12.
- Uetz, P. (ed.), The Reptile Database, <http://www.reptile-database.org>, accessed Aug 3, 2012
- Uribe-Piedrahita, C., 1948. Contribuciones al estudio de la parasitología en Colombia, II. *Caldasia*, 5: 211-219.
- Vercammen-Grandjean, P. H. & Lowenstein, J., 1967. The intestinal Pronocephalidae (Vermes, Trematoda) of the marine iguana from the Galapagos Islands; a possible case of pure mutualism. *Annales de Parasitologie Humane et Comparee*, 42 (4): 435-41.
- Viana, L., 1924. Tentativa de catalogação das espécies brasileiras de trematódeos. *Memórias do Instituto Oswaldo Cruz*, 17 (1): 95-227.
- Vicente, J. J., 1978. Helminhos de *Tropidurus* (Lacertilia, Iguanidae) da Coleção Helmintológica do Instituto Oswaldo Cruz. 1. Trematoda, Cestoda, Acanthocephala, Linguatulida. *Atas da Sociedade de Biologia do Rio de Janeiro*, 19: 71-77.
- Vicente, J. J. & Santos, E., 1976. Fauna helmintológica de *Leptodactylus ocellatus* (L., 1758) de Volta Redonda, Estado do Rio de Janeiro. *Atas da Sociedade de Biologia do Rio de Janeiro*, 18 (1): 27-42.
- Volonterio, O., Baletta, S. & Meneghel, M., 2006. A new genus and species of Opisthogonimid (Digenea) of *Liophis anomalus* (Serpentes: Colubridae) from Uruguay. *Journal of Parasitology*, 92 (5): 1058-1063.

- Vrcibradic, D., Rocha, C. F. D., Bursey, C. R. & Vicente, J. J., 2002. Helminth communities of two sympatric skinks (*Mabuya agilis* and *Mabuya macrorhyncha*) from two “restinga” habitats in southeastern Brazil. *Journal of Helminthology*, 76 (4): 355-361.
- Walton, A. C., 1951. Parasites of the Amphibia. Trematoda I. *Journal of Parasitology*, 37 (5) Sect 2, Suppl.: 23.
- Werneck, M. R., Baldassin, P., Torres, F., Trazi, A. & Berger, B., 2013. Report of *Carettacola stunkardi* (Martin & Bamberger, 1952) Dailey, Fast & Balazs, 1991 (Digenea: Spirorchiidae) infecting green turtle *Chelonia mydas* Linnaeus, 1758 (Testudines, Cheloniidae) in Brazil. *Brazilian Journal of Biology*, 73 (3): 675-676.
- Werneck, M. R., Becker, J. H., Gallo, B. G. & Silva, R. J., 2006. *Leare dius learedi* Price 1934 (Digenea, Spirorchiidae) in *Chelonia mydas* Linnaeus 1758 (Testudines, Cheloniidae) in Brazil: case report. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, 58 (4): 550-555.
- Werneck, M. R., Gallo, B. M. G. & Silva, R. J., 2008a. First report of *Monticellius indicum* Mehra, 1939 (Digenea: Spirorchiidae) infecting *Chelonia mydas* Linnaeus, 1758 (Testudines: Cheloniidae) from Brazil. *Brazilian Journal of Biology*, 68 (2): 455-456.
- Werneck, M. R., Gallo, B. M. G. & Silva, R. J., 2008b. Spirorchiids (Digenea: Spirorchiidae) infecting a Hawksbill sea turtle *Eretmochelys imbricata* (Linnaeus 1758) from Brazil. *Arquivo Brasileiro de Medicina Veterinária e Zootecnia*, 60 (3): 663-666.
- Werneck, M. R., Lima, E. H. S. M., Gallo, B. M. G. & Silva, R. J., 2011. Occurrence of *Amphiorchis solus* (Simha & Chattopadhyaya, 1970) (Digenea: Spirorchiidae) infecting the green turtle *Chelonia mydas* Linnaeus, 1758 (Testudines: Cheloniidae) in Brazil. *Comparative Parasitology*, 78 (1): 200-203.
- Werneck, M. R. & Silva, R. J., 2012. *Styphlotrema solitaria* Looss, 1899 (Digenea, Styphlotrematidae) infecting *Eretmochelys imbricata* Linnaeus 1758 (Testudines, Cheloniidae) in Brazil. *Neotropical Helminthology*, 6 (1): 121-126.
- Werneck, M. R. & Silva, R. J., 2013. Occurrence of *Amphiorchis indicus* Mehrotra, 1973 (Digenea, Spirorchiidae) infecting green turtle *Chelonia mydas* Linnaeus, 1758 (Testudines, Cheloniidae) in Brazil. *Brazilian Journal of Biology*, 73 (1): 225-227
- Yamaguti, S., 1971. *Synopsis of Digenetic Trematodes of Vertebrates*. Keigaku Publishing, Tokyo, 590 pp.



Fig. 1: *Creptotrema lynchi* after Brooks, 1976. Bar = 250µm

Fig. 2: *Maicuru solitarium* after Freitas, 1960. Bar = 0,1mm

Fig. 3: *Iquitos ceii* after Mañé-Garzón & Gil, 1963. Bar = 0,015mm

Fig. 4: *Gorgoderia australiensis* after Suriano, 1978. Bar = 1mm

Fig. 5: *Gorgoderina carioca* after Fernandes, 1958. Bar = 1mm



Fig. 6. *Gorgoderina cedroi* after Travassos, 1924. Bar = 0,1mm

Fig. 7: *Gorgoderina chilensis* after Dioni, 1947. Bar = 1mm

Fig. 8: *Gorgoderina cryptorchis* after Travassos, 1924. Bar = 0,1mm

Fig. 9: *Gorgoderina darwini* after Mañé-Garzón & Gonzáles, 1978. Bar = 1mm

Fig. 10: *Gorgoderina diaster* after Fernandes, 1958. Bar = 0,5mm

Fig. 11: *Gorgoderina megacysta* after Mañé-Garzón & Gonzáles, 1978. Bar = 1mm

Fig. 12: *Gorgoderina parvicava* after Fernandes, 1958. Bar = 1mm

Fig. 13: *Gorgoderina parvicava minuta* after Tantaleán & García, 1993. Bar = 1mm



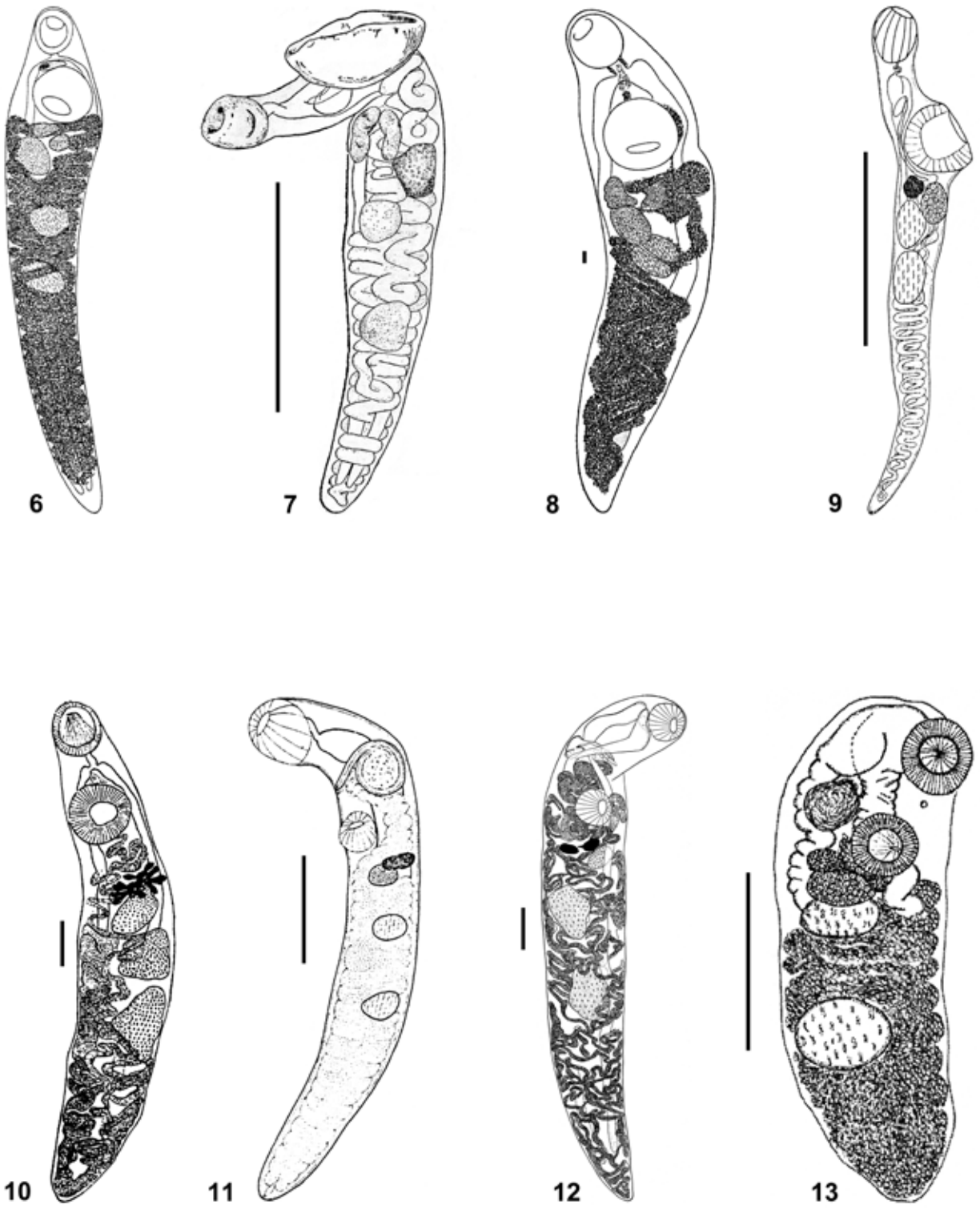


Fig. 14: *Gorgoderina pigulevskyi* after Fernandes, 1958.

Fig. 15: *Gorgoderina rochalimai* after Pereira & Cuocolo, 1940. Bar = 3mm

Fig. 16: *Gorgoderina valdiviensis* after Puga, 1979. Bar = 2mm

Fig. 17: *Gorgoderina* sp. after Ibáñez & Córdova, 1979. Bar = 1mm

Fig. 18: *Mesocoelium lanfrediae* after Gomes, Melo, Giese, Furtado, Gonçalves & Santos, 2013. Bar = 200µm

Fig. 19: *Mesocoelium meggitti* after Pereira & Cuocolo, 1940. Bar = 0,2mm

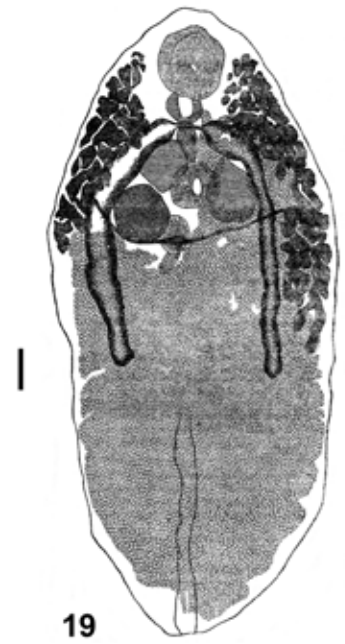
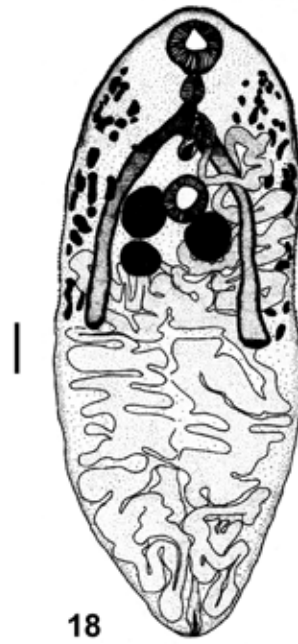
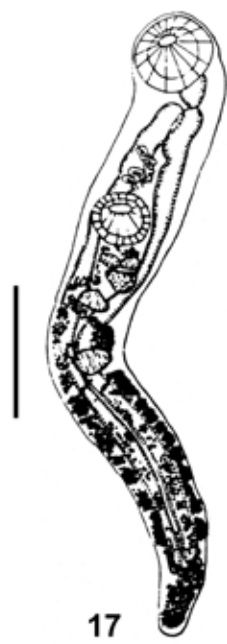
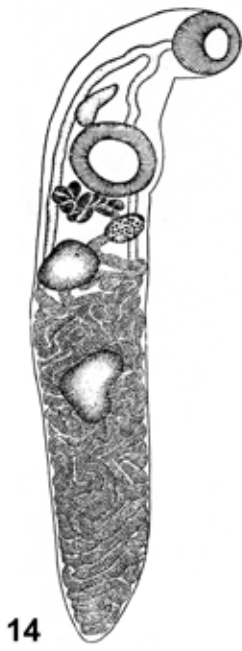


Fig. 20: *Mesocoelium monas* after Freitas, 1963. Bar = 0,2mm

Fig. 21: *Mesocoelium waltoni* after Pereira & Cuocolo, 1940. Bar = 0,2mm

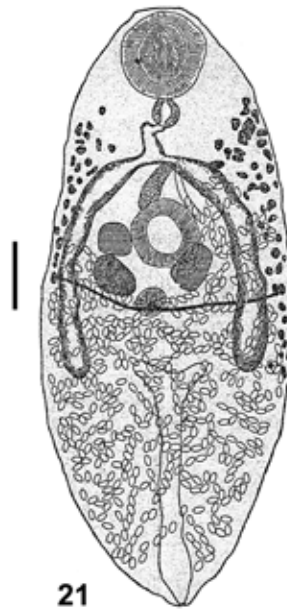
Fig. 22: *Halipegus dubius* after Kohn & Fernandes, 1988. Bar = 1mm

Fig. 23: *Pseudosonsinotrema chabaudi* after Bechara & Vélez, 2010. Bar = 0,2mm

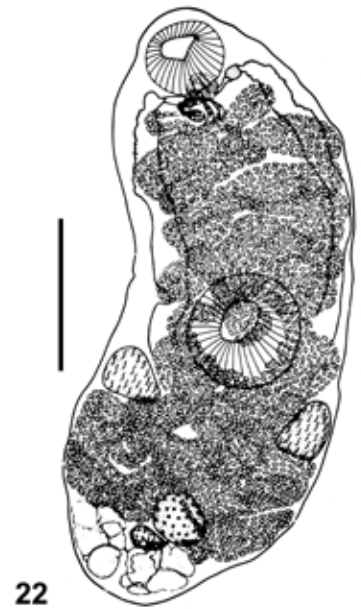
Fig. 24: *Pseudosonsinotrema megalorchis* after Flowers, Law & Carvajal-Endara, 2011. Bar = 100µm



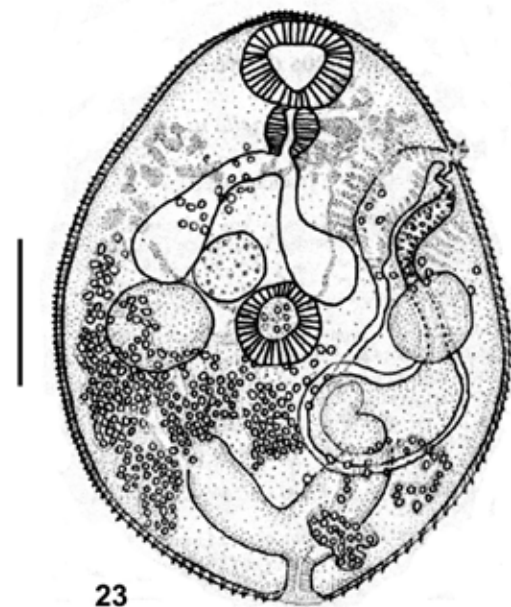
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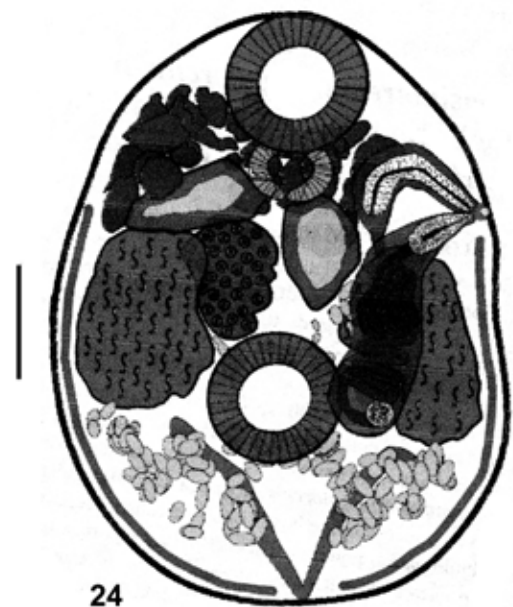
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Fig. 25: *Catadiscus cohni* after Freitas & Lent, 1939. Bar = 1mm

Fig. 26: *Catadiscus corderoi* after Mañé-Garzón, 1958. Bar = 1mm

Fig. 27: *Catadiscus eldoradiensis* after Artigas & Pérez, 1964.

Fig. 28: *Catadiscus freitaslenti* after Lent, Freitas & Proença, 1946. Bar = 1mm

Fig. 29: *Catadiscus hylae* after Incorvaia, 1983. Bar = 0,5mm

Fig. 30: *Catadiscus inopinatus* after Freitas, 1941. Bar = 1mm

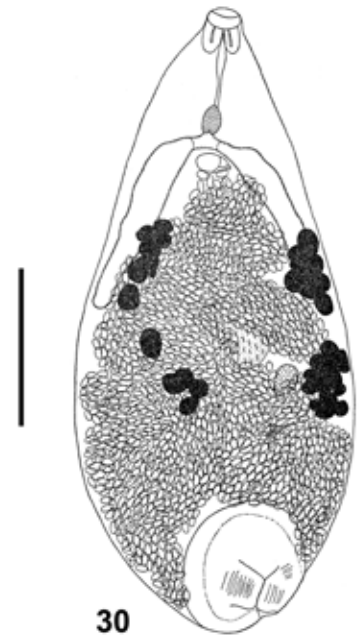
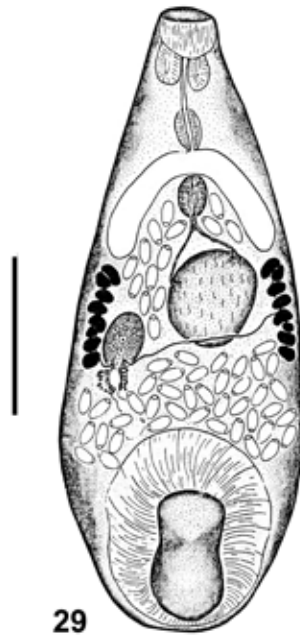
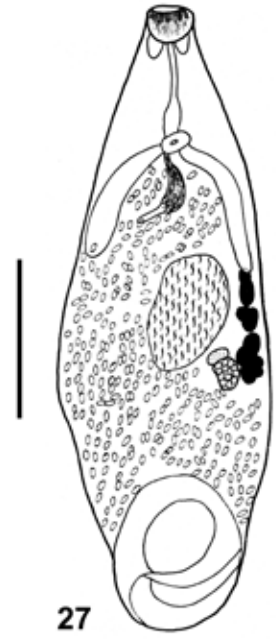
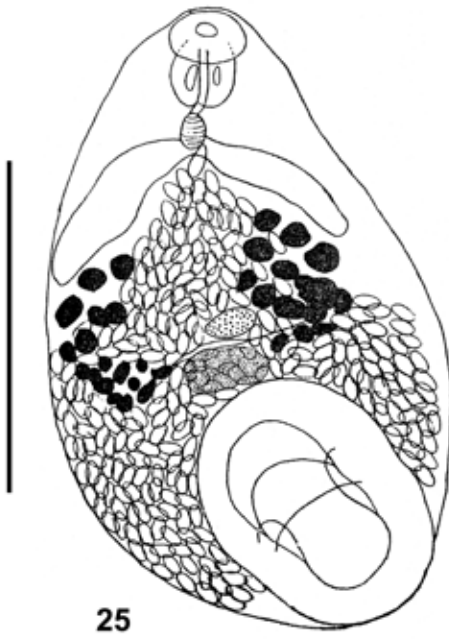


Fig. 31: *Catadiscus marinholutzi* after Freitas & Lent, 1939. Bar = 1mm

Fig. 32: *Catadiscus mirandai* after Freitas, 1943. Bar = 1mm

Fig. 33. *Catadiscus propinquus* after Freitas & Dobbin Jr., 1956.

Fig. 34. *Catadiscus pygmmaeus* after Freitas & Lent, 1939. Bar = 1mm

Fig. 35: *Catadiscus uruguayensis* after Freitas & Lent, 1939. Bar = 1mm

Fig. 36: *Glypthelmins biliaris* after Suriano, 1968. Bar = 1mm



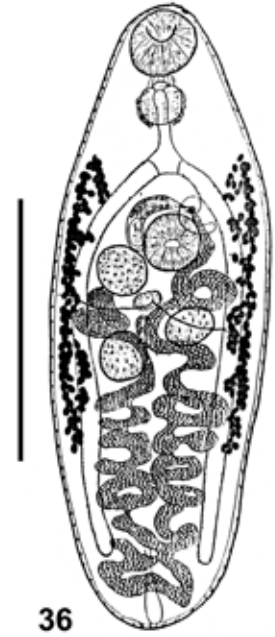
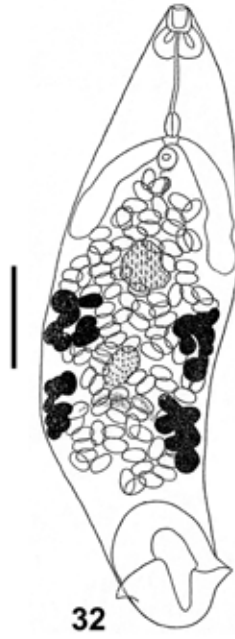


Fig. 37. *Glyphelmins festina* after Cordero, 1944. Bar = 1mm

Fig. 38. *Glyphelmins parva* after Travassos, 1924. Bar = 0,2mm

Fig. 39: *Glyphelmins sanmartini* after Mañé-Garzón & Holcman-Spector, 1974. Bar = 1mm

Fig. 40: *Haematoloechus arequipensis* after Ibañez & Córdova, 1979. Bar = 1mm

Fig. 41: *Haematoloechus freitasi* after Mañé-Garzón & Gil, 1959. Bar = 1mm

Fig. 42: *Haematoloechus fuelleborni* after Travassos & Darriba, 1930. Bar = 1mm

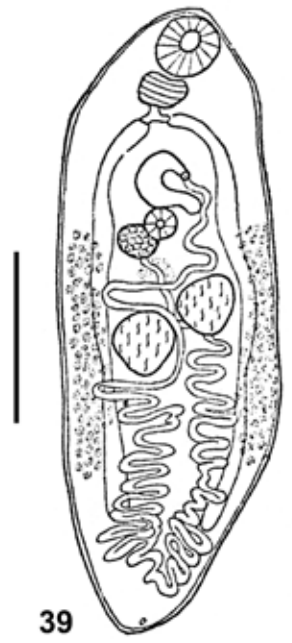
Fig. 43: *Haematoloechus longiplexus* after Hamann & Pérez, 1999. Bar = 500 µm



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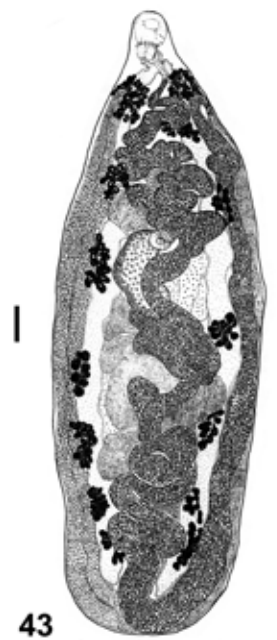
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Fig. 44: *Haematoloechus lutzi* after Freitas & Lent, 1939. Bar = 1mm

Fig. 45: *Haematoloechus ozorioi* after Freitas & Lent, 1939. Bar = 1mm

Fig. 46: *Haematoloechus pukinensis* after Ibañez & Córdova, 1979. Bar = 1mm

Fig. 47: *Neohaematoloechus iturbei* after Dobbin Jr., 1957. Bar = 1mm

Fig. 48: *Neohaematoloechus neivai* after Travassos & Darriba, 1930. Bar = 1mm

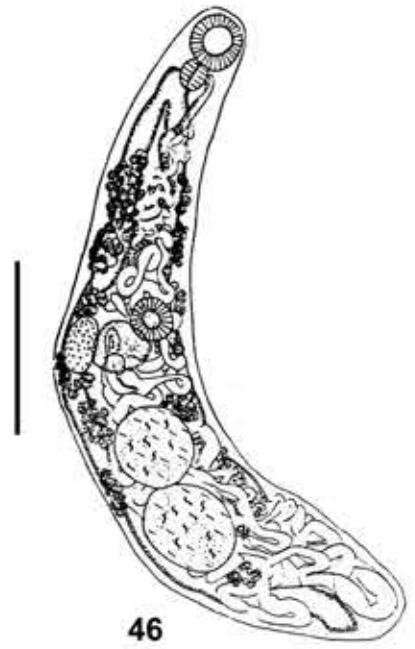
Fig. 49: *Rauschiella chaquensis* after Mañé-Garzón & Holcman-Spector, 1967. Bar = 0,5mm



44



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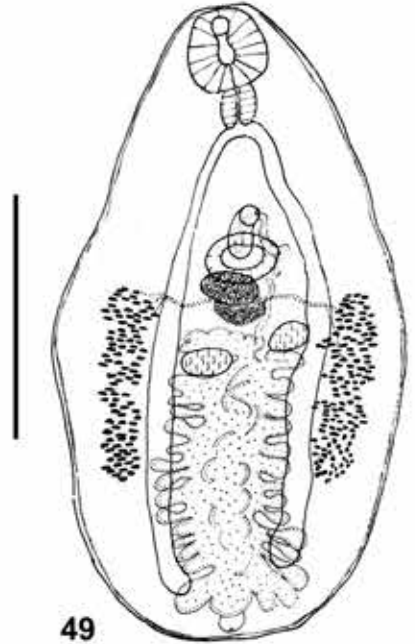
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49

Fig. 50: *Rauschiella lenti* after Freitas, 1941. Bar = 1mm

Fig. 51: *Rauschiella linguatula* after Travassos, 1924. Bar = 0,2mm

Fig. 52: *Rauschiella palmipedis* after Freitas, 1941. Bar = 1mm

Fig. 53: *Rauschiella proxima* after Freitas, 1941. Bar = 1mm

Fig. 54: *Rauschiella repandum* after Travassos, 1924.

Fig. 55: *Rauschiella robusta* after Brooks, 1976. Bar = 500µm



Fig. 56: *Choledocystus hepaticus* after Sullivan, 1977. Bar = 0,5mm

Fig. 57: *Choledocystus incurvatum* after Nasir, 1966. Bar = 0,3mm

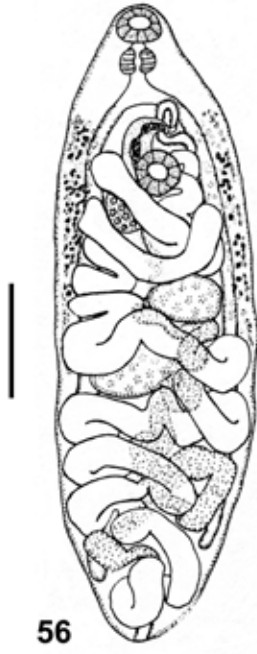
Fig. 58: *Choledocystus pseudium* after Mañé-Garzón & Holcman-Spector, 1967. Bar = 1mm

Fig. 59: *Choledocystus simulans* after Freitas, 1941. Bar = 1mm

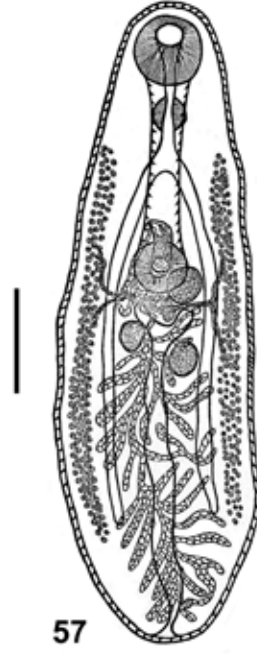
Fig. 60: *Choledocystus vitellinophilum* after Dobbin, 1958. Bar = 1mm

Fig. 61: *Plagiorchis rangeli* after Artigas & Zerpa, 1961. Bar = 1mm





56



57



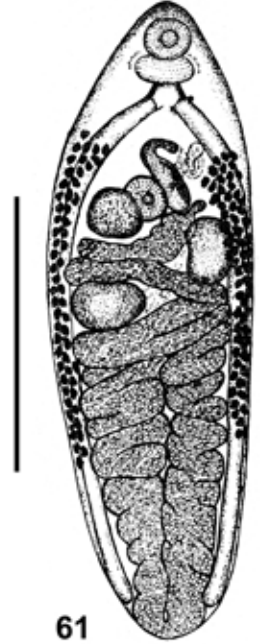
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59



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61

Fig. 62: *Rudolphitrema chilensis* after Puga, 1986. Bar = 0,5mm

Fig. 63: *Rudolphitrema physalaemi* after Mañé-Garzón & Ponce de León, 1976. Bar = 0,2mm

Fig. 64: *Rudolphitrema rudolphii* after Travassos, 1924. Bar = 0,2mm

Fig. 65: *Opisthioglyphe amplicavus* after Travassos, 1930.

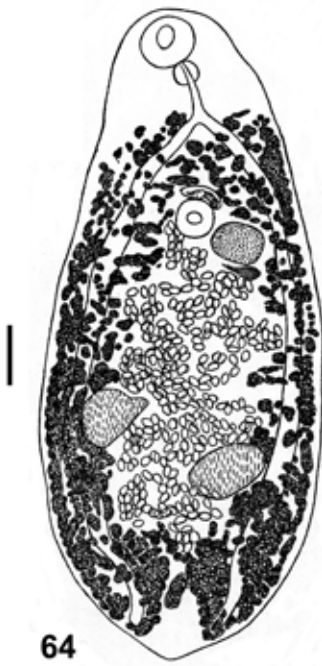
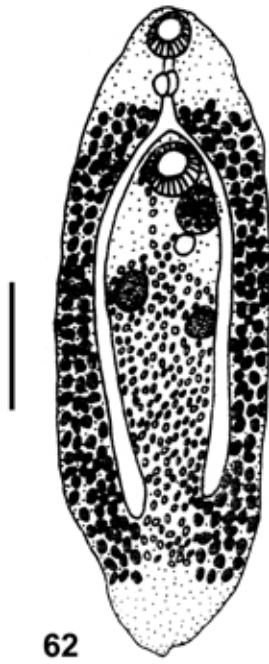


Fig. 66: *Lophotaspis vallei* after Araujo, 1941. Bar = 1mm

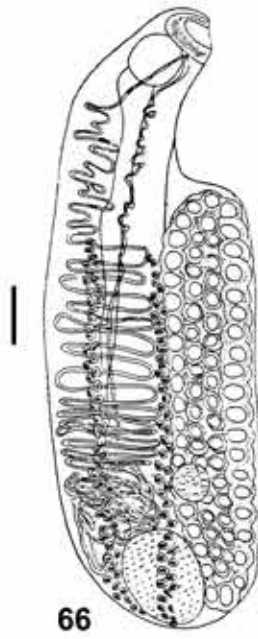
Fig. 67: *Leurosoma rudolfbarthi* after Kohn & Fernandes, 1976. Bar = 1mm

Fig. 68: *Odhneriotrema microcephala* after Travassos, 1929. Bar = 2mm

Fig. 69: *Helicotrema asymmetricum* after Travassos, 1928. Bar = 2mm

Fig. 70: *Helicotrema magniovatum* after Odhner, 1912.

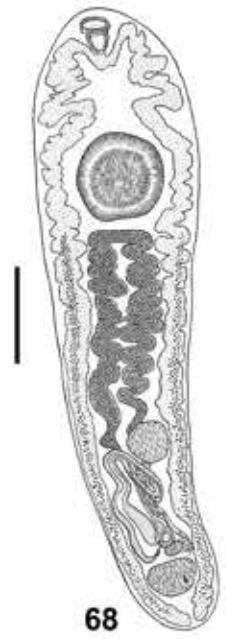
Fig. 71: *Helicotrema spirale* after Diesing, 1855.



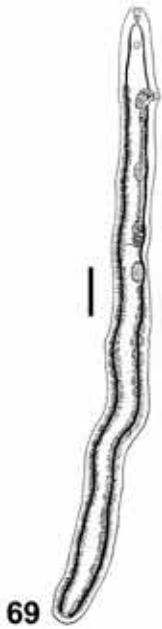
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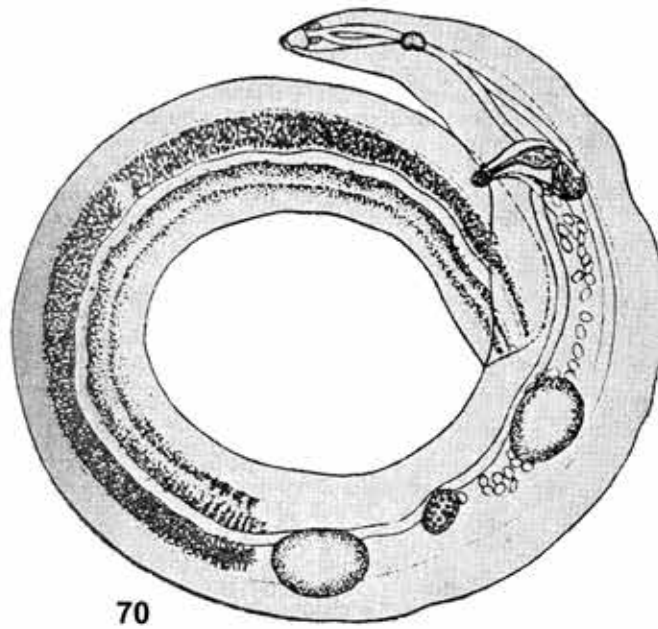
67



68



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70



71

Fig. 72: *Cyathocotyle brasiliensis* after Ruiz & Leão, 1943. Bar = 1mm

Fig. 73: *Cheloniodiplostomum testudinis* after Dubois, 1938.

Fig. 74: *Crocodicicola pseudostoma* after Dubois, 1938. s/ escala

Fig. 75: *Cystodiplostomum hollyi* after Ruiz & Rangel, 1954. Bar = 0.5mm

Fig. 76: *Herpetodiplostomum caimancola* after Dubois, 1938.

Fig. 77: *Heterodiplostomum helicopsis* after Mañé-Garzón & Alonso, 1976. Bar = 1mm

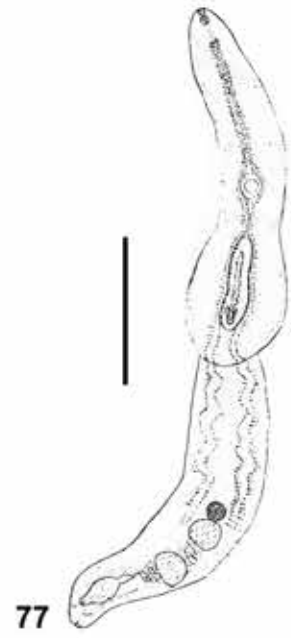
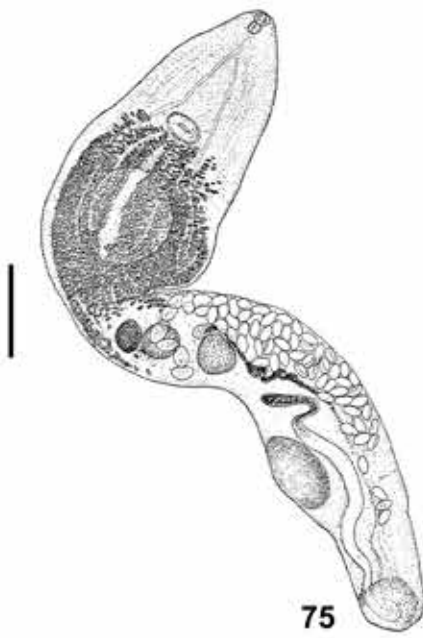


Fig. 78: *Heterodiplostomum lanceolatum* after Ruiz & Rangel, 1954. Bar = 0.2mm

Fig. 79: *Massoprostatum longum* after Álvarez, Lenis & Vélez, 2005. Bar = 0.25mm

Fig. 80: *Mesodiplostomum gladiolum* after Dubois, 1938.

Fig. 81: *Ophiodiplostomum ancyloides* after Dubois, 1938.

Fig. 82: *Ophiodiplostomum spectabile* after Ruiz & Rangel, 1954. Bar = 0.5mm

Fig. 83: *Paradiplostomum abbreviatum* after Dubois, 1938.





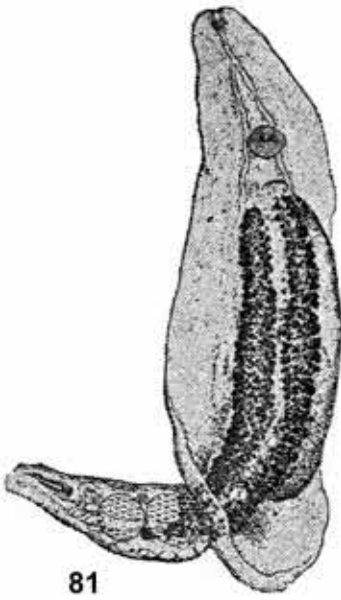
78



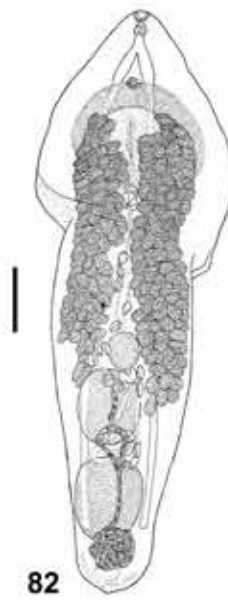
79



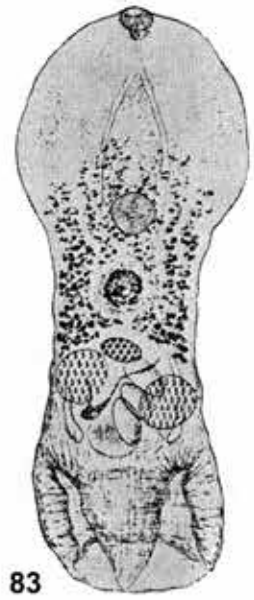
80



81



82



83

Fig. 84: *Prolecithodiplostomum constrictum* after Dubois, 1938.

Fig. 85: *Proterodiplostomum breve* after Catto & Amato, 1994. Bar = 500µm

Fig. 86: *Proterodiplostomum globulare* after Catto & Amato, 1994. Bar = 1mm

Fig. 87: *Proterodiplostomum longum* after Dubois, 1938.

Fig. 88: *Proterodiplostomum medusae* after Ruiz & Rangel, 1954. Bar = 0.5mm

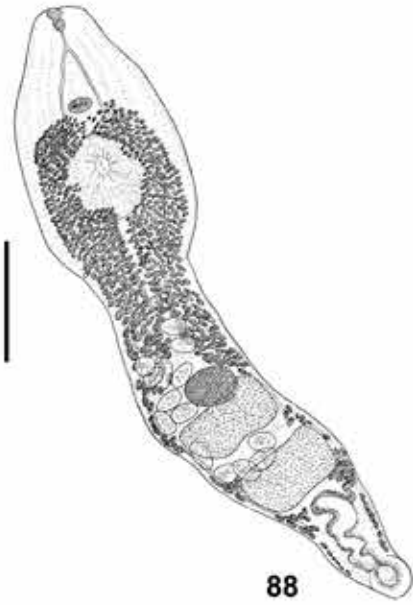
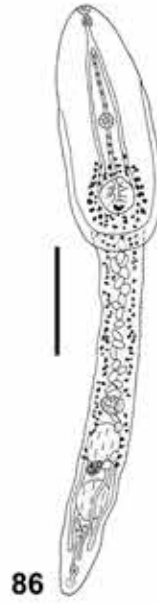


Fig. 89: *Proterodiplostomum tumidulum* after Dubois, 1938.

Fig. 90: *Caballerotrema* sp. after Ostrowski de Núñez, 2003. Bar = 1mm

Fig. 91: *Echinostoma* sp. after Ostrowski de Núñez, 2003. Bar = 1mm

Fig. 92: *Prionosoma phrynopsis* after Mañé-Garzón & Gil, 1961. Bar = 1mm

Fig. 93: *Prionosomoides scalaris* after Dobbin Jr., 1967. Bar = 1mm

Fig. 94: *Pulchrosomoides elegans* after Freitas & Lent, 1937. Bar = 3mm

Fig. 95: *Stephanoprora campomica* after Nasir & Diaz, 1971. Bar = 1mm

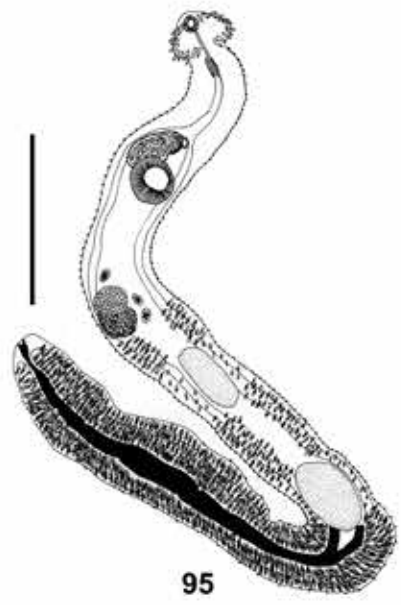
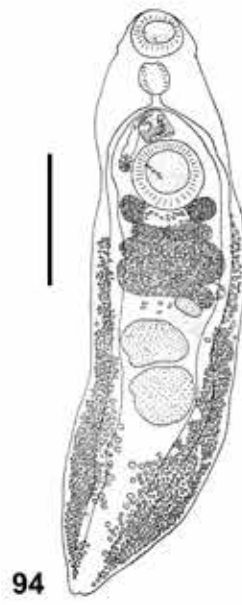
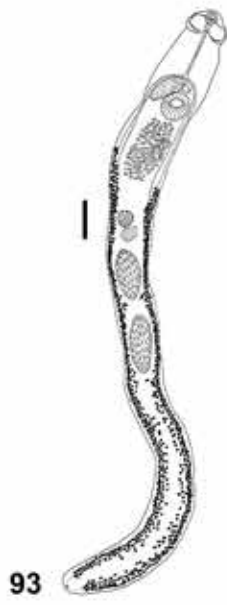
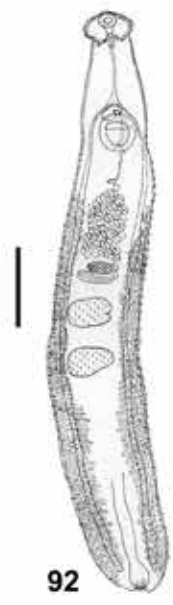
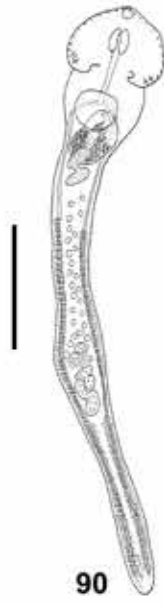


Fig. 96: *Stephanoprora jacaretinga* after Freitas & Lent, 1938. Bar = 0.6mm

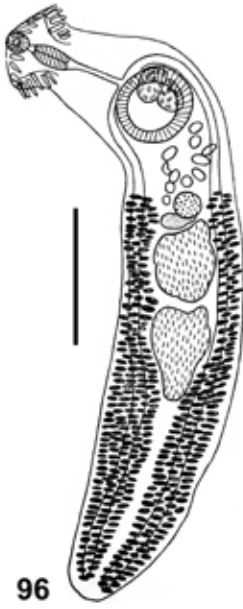
Fig. 97: *Stephanoprora nattereri* after Ostrowski de Núñez, 2003. Bar = 2mm

Fig. 98: *Stephanoprora* sp. after Ostrowski de Núñez, 2003. Bar = 0.5mm

Fig. 99: *Cotylotretus rugosus* after Odhner, 1902.

Fig. 100: *Sphaeridiotrema echinosauense* after O'Brien, Sidner & Etges, 1979. Bar = 0.2mm

Fig. 101: *Rhytidodes gelatinosus* after Braun, 1901.



96



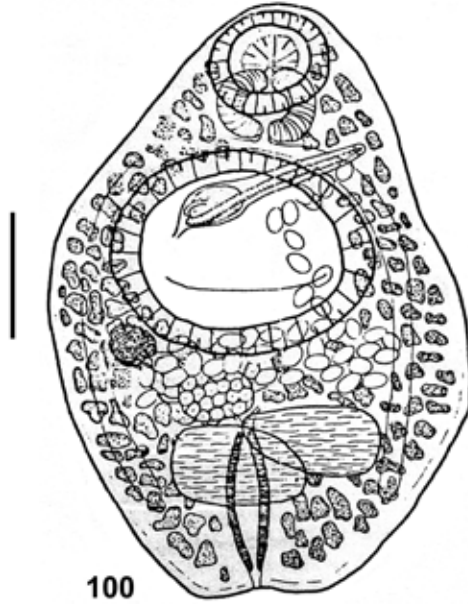
97



98



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100



101

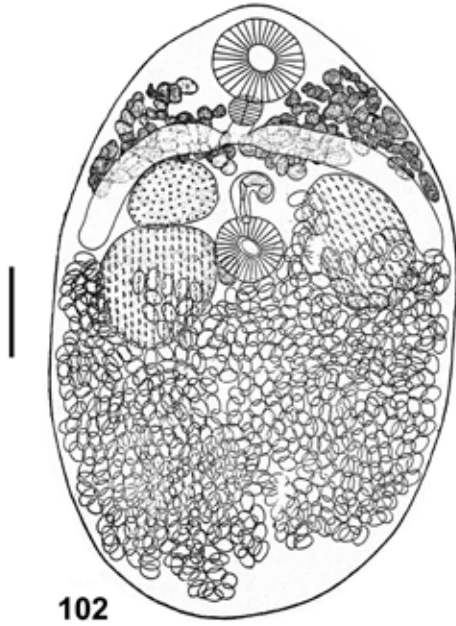
Fig. 102: *Brachycoelium salamandrae* after Freitas, 1961. Bar = 0.2mm

Fig. 103: *Braunotrema pulvinatum* after Lent & Freitas, 1938. Bar = 1mm

Fig. 104: *Infidum infidum* after Travassos, 1944. Bar = 1mm

Fig. 105: *Infidum luckeri* after McIntosh, 1939. Bar = 1mm





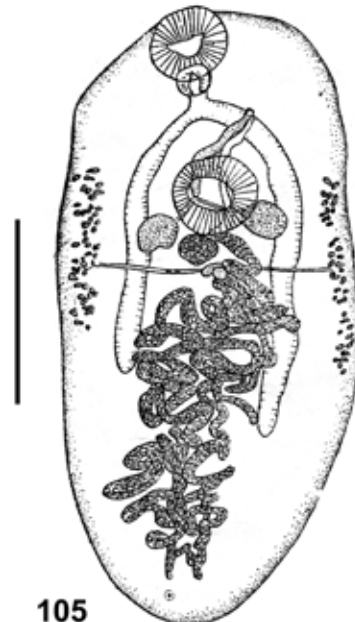
102



103



104



105

Fig. 106: *Infidum similis* after Travassos, 1944. Bar = 1mm

Fig. 107: *Paradistomum boae* ter McCallum, 1921.

Fig. 108: *Paradistomum parvissimum* after Travassos, 1944. Bar = 0.5mm

Fig. 109: *Paradistomum rabusculum* after Kossack, 1910.

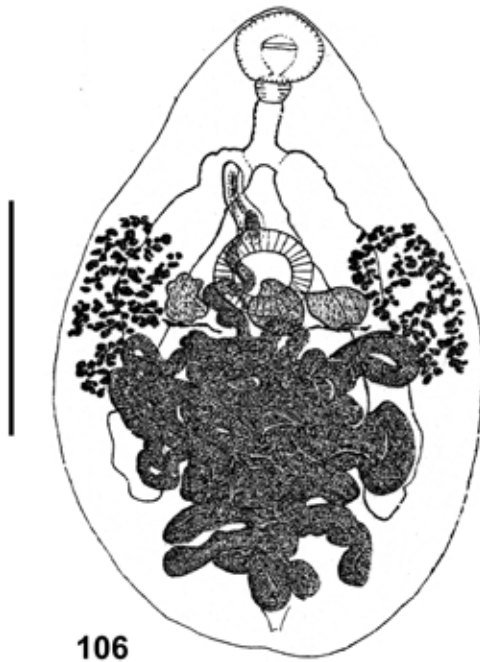


Fig. 110: *Mesocoelium monas* after Pereira & Cuocolo, 1940. Bar = 0.2mm

Fig. 111: *Mesocoelium sibynomorphi* after Ruiz & Leão, 1943. Bar = 1mm

Fig. 112: *Pachysolus sclerops* after Travassos, 1928. Bar = 0.2 mm

Fig. 113: *Aliptrema ribeiroi* after Ruiz & Leão, 1955. Bar = 1mm

Fig. 114: *Neoctagium travassosi* after Ruiz, 1943. Bar = 2mm

Fig. 115: *Neodeuterobaris pritchardae* after Brooks, 1976. Bar = 1mm

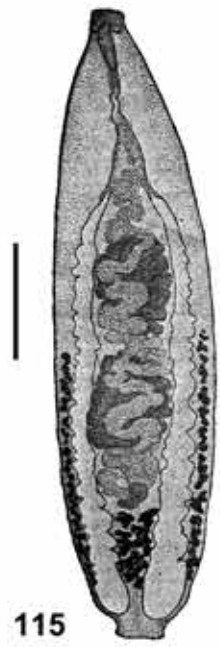
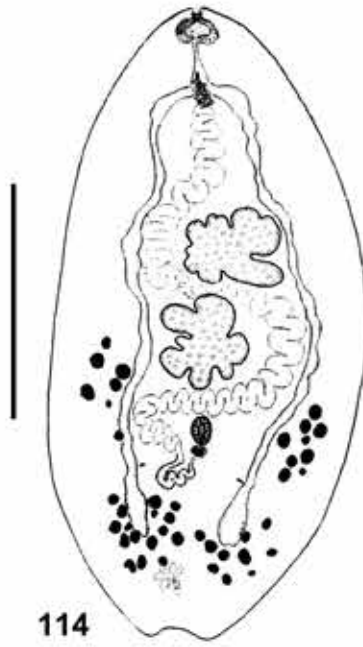
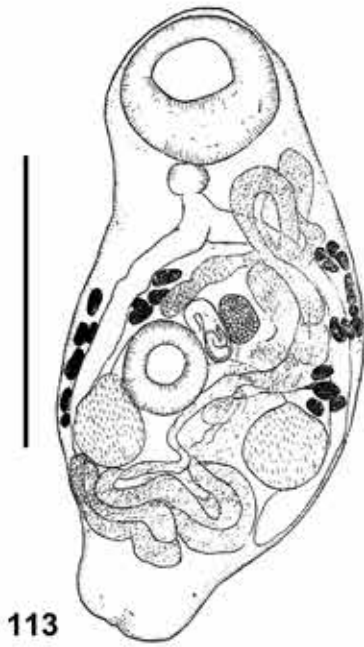
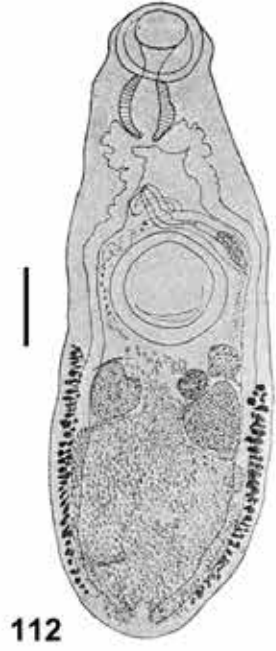
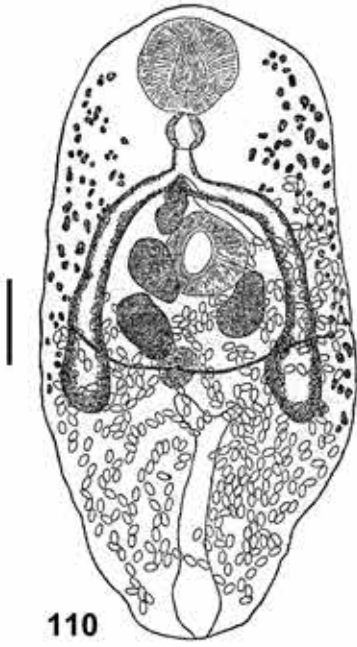


Fig. 116: *Podocnemitrema papillosus* after Alho & Vicente, 1964. Bar = 1mm

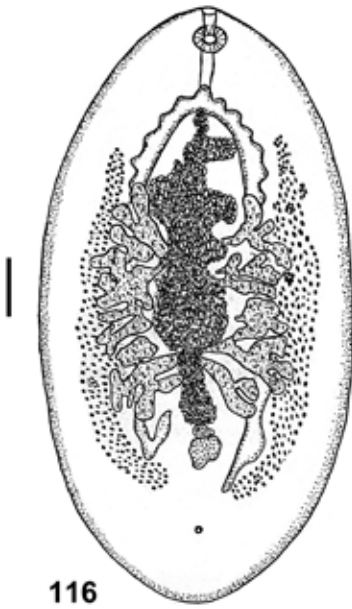
Fig. 117: *Polyagium linguatula* after Freitas & Lent, 1938. Bar = 1mm

Fig. 118: *Acanthostomum scyphocephalum* after Nasir, 1974. Bar = 0.4mm

Fig. 119: *Caimanicola brauni* after Mañé-Garzón & Gil, 1961. Bar = 0.5mm

Fig. 120: *Caimanicola marajoara* after Freitas & Lent, 1938. Bar = 0.2mm

Fig. 121: *Proctocaecum dorsale* after Catto & Amato, 1993. Bar = 500µm



116



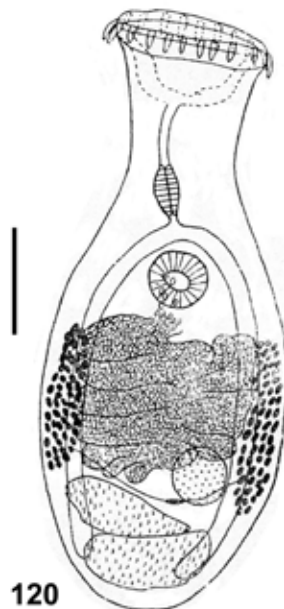
117



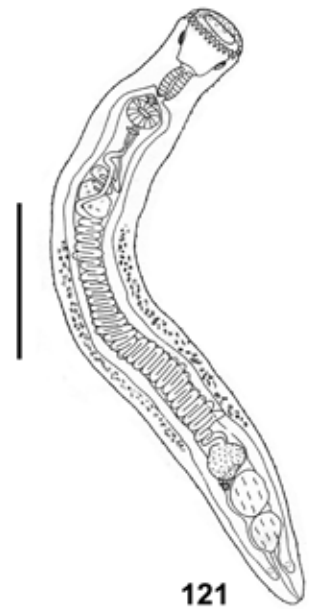
118



119



120



121

Fig. 122: *Timoniella incognita* after Nasir, 1974. Bar = 0.4mm

Fig. 123: *Timoniella ostrowskiae* after Mañé-Garzón & Gil, 1961. Bar = 1mm

Fig. 124: *Halltrema avitellina* after Alho, 1965. Bar = 1mm

Fig. 125: *Halltrema heteroxenus* after Cordero & Vogelsang, 1940.

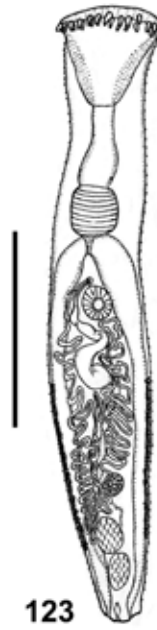
Fig. 126: *Nematophila argentinum* after Lenis & Vélez, 2011. Bar = 2mm

Fig. 127: *Nematophila grandis* after Lenis & Vélez, 2011. Bar = 2mm





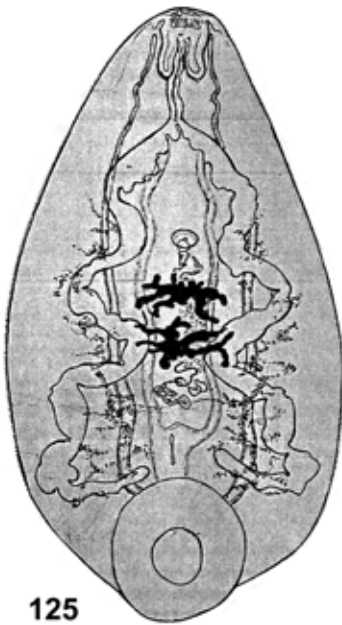
122



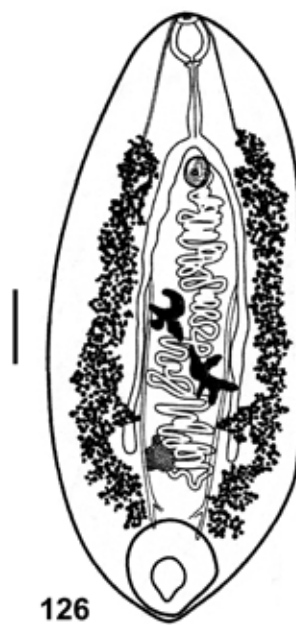
123



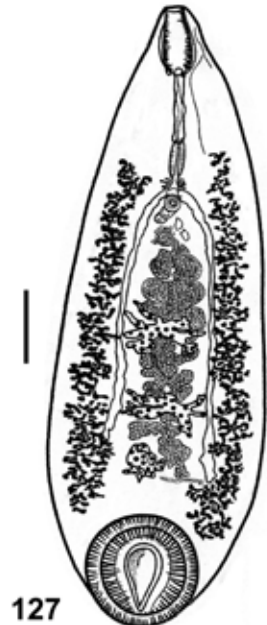
124



125



126



127

Fig. 128: *Nematophila venezuelensis* after Lenis & Vélez, 2011. Bar = 2mm

Fig. 129: *Oriximinatrema noronhae* after Knoff, Brooks, Mullins & Gomes, 2012. Bar = 0.25 mm

Fig. 130: *Pseudonematophila ovalis* after Lenis & Vélez, 2011. Bar = 2mm

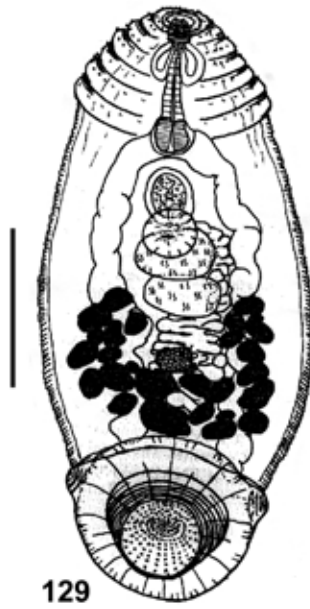
Fig. 131: *Catadiscus dolichocotyle* after Mañé-Garzón & Gortari, 1965. Bar = 0.5mm

Fig. 132: *Catadiscus freitaslenti* after Ruiz, 1943. Bar = 1mm

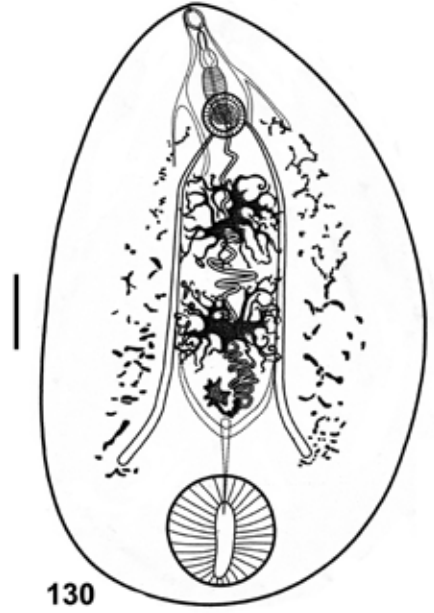
Fig. 133: *Catadiscus longicoecalis* after Poumarau, 1965. Bar = 0.5mm



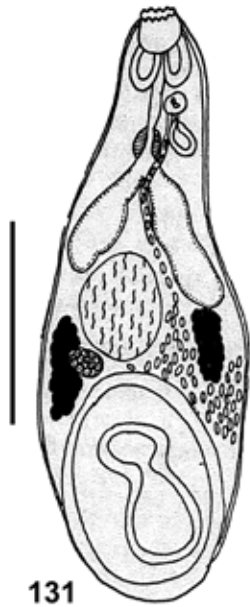
128



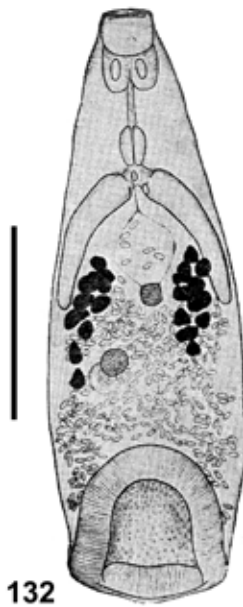
129



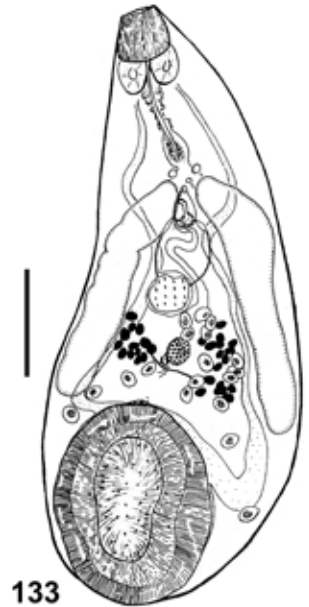
130



131



132



133

Fig. 134: *Catadiscus rochai* after Correa & Artigas, 1978/1979. Bar = 1mm

Fig. 135: *Catadiscus uruguayensis* after Lunaschi & Drago, 2002. Bar = 300µm

Fig. 136: *Liophistrema buccalis* after Mañé-Gárzon, 1973.

Fig. 137: *Liophistrema pulmonale* after Artigas, Ruiz & Leão, 1942. Bar = 1mm

Fig. 138: *Opisthognimus afranioi* after Pereira, 1929. Bar = 0.5mm

Fig. 139: *Opisthognimus artigasi* after Ruiz & Leão, 1942. Bar = 1mm

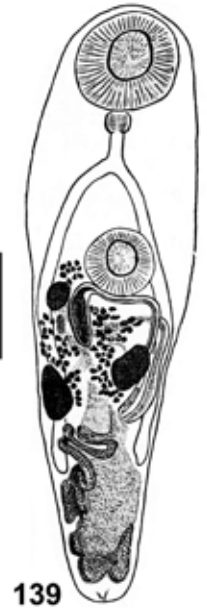
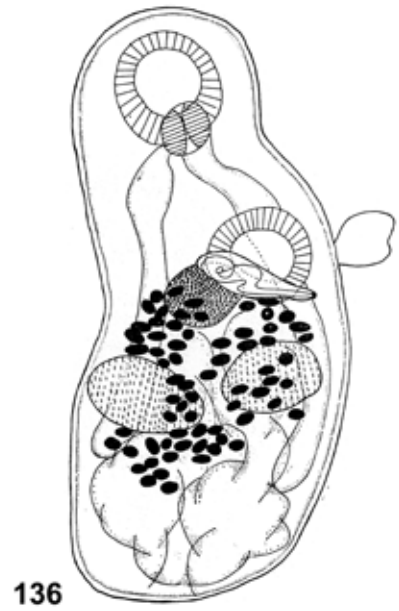
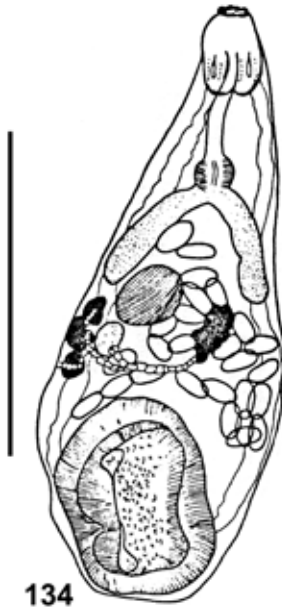


Fig. 140: *Opisthgonimus fariai* after Leão & Ruiz, 1943. Bar = 0.5mm

Fig. 141: *Opisthgonimus fonsecai* after Ruiz & Leão, 1942. Bar = 0.5mm

Fig. 142: *Opisthgonimus interrogativus* after Pereira, 1929. Bar = 0.5mm

Fig. 143: *Opisthgonimus megabothrium* after Pereira, 1929. Bar = 0.3mm

Fig. 144: *Opisthgonimus misionesensis* after Lunaschi & Drago, 2001. Bar = 1mm

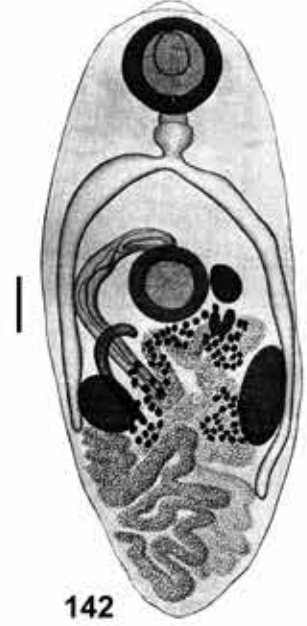
Fig. 145: *Opisthgonimus pereirai* after Ruiz & Leão, 1942. Bar = 1mm



140



141



142



143



144



145

Fig. 146: *Opisthgonimus lecithonotus* after Pereira, 1929. Bar = 0.5mm

Fig. 147: *Opisthgonimus serpentis* after Artigas, Ruiz & Leão, 1943. Bar = 1mm

Fig. 148: *Opisthgonimus sulina* after Artigas, Ruiz & Leão, 1942. Bar = 1mm

Fig. 149: *Opisthgonimus uruguayensis* after Mañé-Garzón & Holcman-Spector, 1973. Bar = 50mm

Fig. 150: *Paracotylotrema pocedeleoni* after Volonterio, Baletta & Meneghel, 2006. Bar = 300  $\mu$ m

Fig. 151: *Allopharynx daileyi* after Bursey, Goldberg & Vitt, 2005. Bar = 1mm



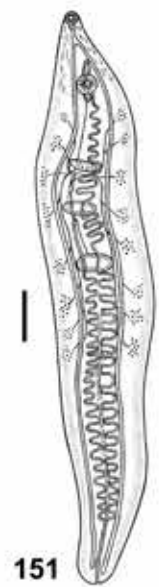
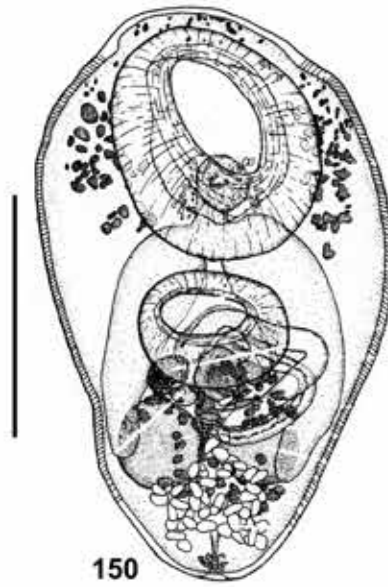
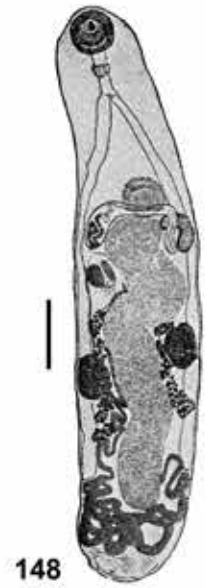


Fig. 152: *Bieria artigasi* after Leão, 1946.

Fig. 153: *Glossidiella ornata* after Travassos, 1928 Bar = 0.3mm

Fig. 154: *Glossidioides loossi* after Travassos, 1928. Bar = 0.3mm

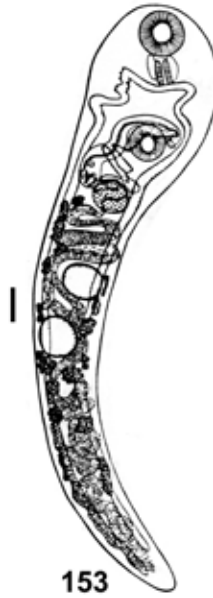
Fig. 155: *Haplometroides buccicola* after Artigas & Paulino, 1988. Bar = 1mm

Fig. 156: *Haplometroides intercaecalis* after Silva, Ferreira & Strüssmann, 2007. Bar = 1mm

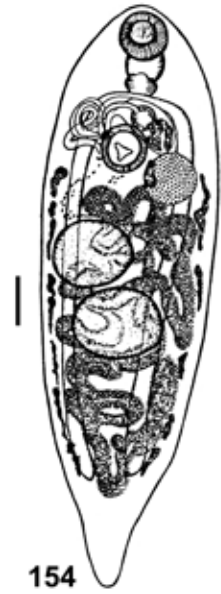
Fig. 157: *Haplometroides odhneri* after Ruiz & Perez, 1959. Bar = 1mm



152



153



154



155



156



157

Fig. 158: *Parahaplometroides basiliscae* after Stunkard & Gandal, 1966.

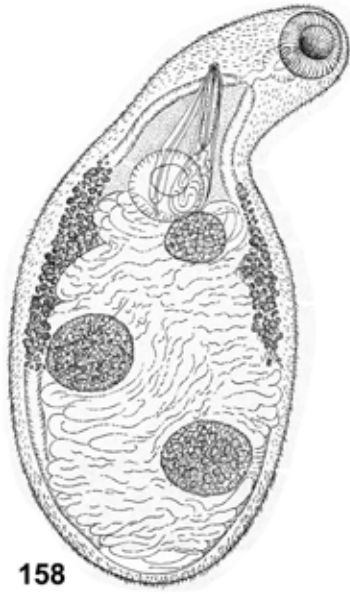
Fig. 159: *Plagiorchis freitasi* after Vicente, 1978. Bar = 0.2mm

Fig. 160: *Plagiorchis luhei* after Travassos, 1928. Bar = 0.3mm

Fig. 161: *Plagiorchis vicentei* after Rodrigues, 1994. Bar = 0.4mm

Fig. 162: *Pneumotrema travassosi* after Bhalerao, 1937.

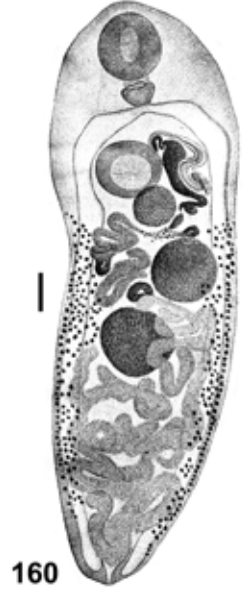
Fig. 163: *Sticholecitha serpentis* after Prudhoe, 1949. Bar = 1mm



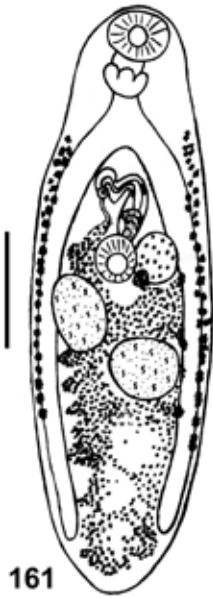
158



159



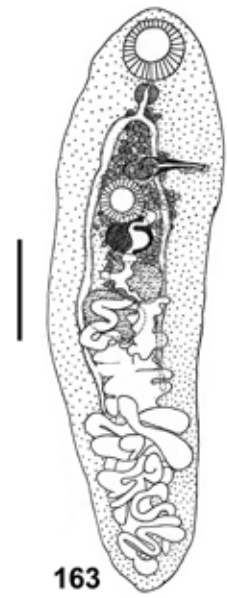
160



161



162



163

Fig. 164: *Styphlodora condita* after Faria, 1911.

Fig. 165: *Styphlodora gili* after Mañe-Garzón & Holcman-Spector, 1967. Bar = 1mm

Fig. 166: *Travtrema stenocotyle* after Freitas & Dobbin Jr., 1957. Bar = 0.2mm

Fig. 167: *Dasymetra tupinambis* after Nasir & Diaz, 1971. Bar = 1mm

Fig. 168: *Renifer chironius* after Nasir & Diaz, 1971. Bar = 1mm

Fig. 169: *Renifer heterocoelium* after Travassos, 1921.

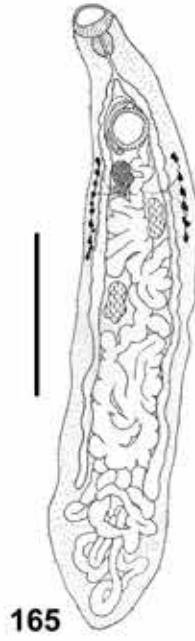


Fig. 170: *Renifer monstrosus* after Caballero & Vogelsang, 1947. Bar = 7.540mm

Fig. 171: *Styphlotrema solitaria* after Werneck & Silva, 2012. Bar = 1mm

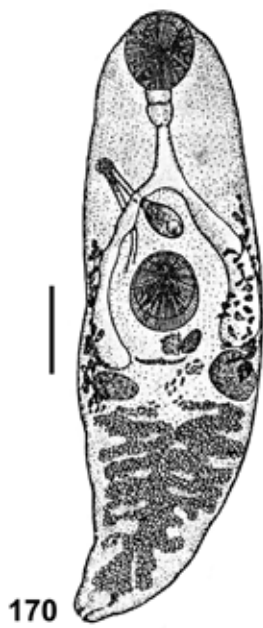
Fig. 172: *Loefgrenia loefgrenia* after Travassos, 1919.

Fig. 173: *Orchidasma amphiorchis* after Freitas & Lent, 1938. Bar = 1mm

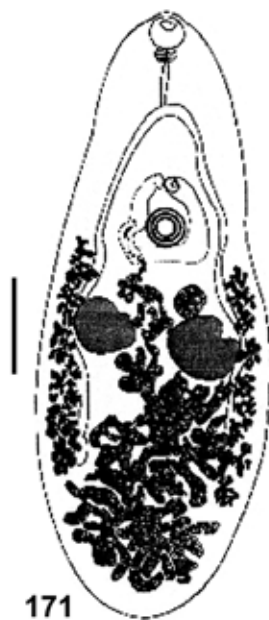
Fig. 174: *Pseudotelorchis caimanis* after Catto & Amato, 1993. Bar = 2mm

Fig. 175: *Pseudotelorchis devicenzii* after Mañé-Garzón & Gil, 1961. Bar = 1mm





170



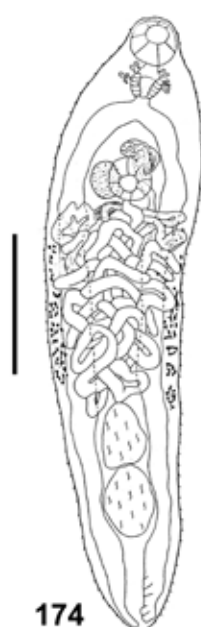
171



172



173



174



175

Fig. 176: *Pseudotelorchis yacarei* after Catto & Amato, 1993. Bar = 250µm

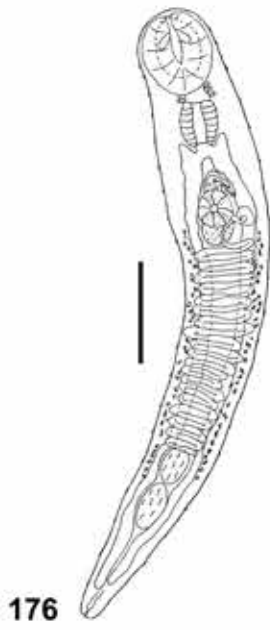
Fig. 177: *Telorchis achavali* after Mañé-Garzón & Holcman-Spector, 1973. Bar = 10µm

Fig. 178: *Telorchis aculeatus* after Nasir, 1974. Bar = 0.8mm

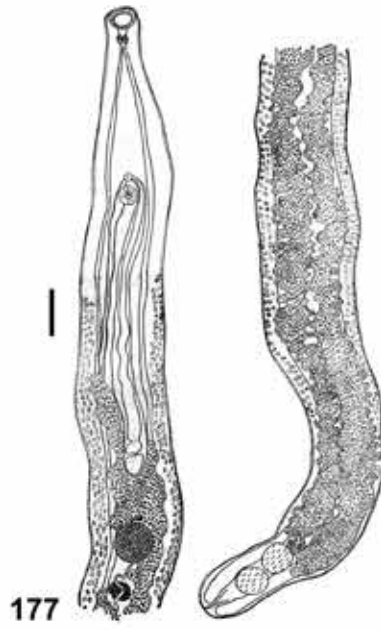
Fig. 179: *Telorchis bifurcus* after Braun, 1901.

Fig. 180: *Telorchis birabeni* after Mañé-Garzón & Gil, 1961. Bar = 1mm

Fig. 181: *Telorchis clava* after Poumarau, 1968. Bar = 1mm



176



177



178



179



180



181

Fig. 182: *Telorchis diaphanus* after Freitas & Dobbin Jr., 1959. Bar = 0.4mm

Fig. 183: *Telorchis dubius* after Mañé-Garzón & Holcman-Spector, 1968. Bar = 1mm

Fig. 184: *Telorchis hagmanni* after Lent & Freitas, 1937. Bar = 2mm

Fig. 185: *Telorchis parvus* after Braun, 1901.

Fig. 186: *Telorchis platensis* after Mañé-Garzón & Gil, 1961. Bar = 0.25mm

Fig. 187: *Telorchis pleroticus* after Braun, 1901.

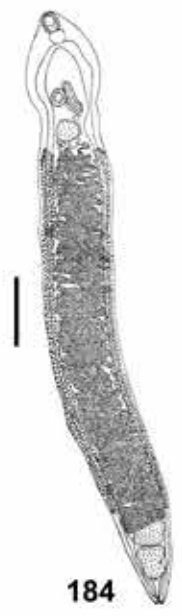
Fig. 188: *Telorchis productus* after Mañé-Garzón & Gil, 1961. Bar = 1mm



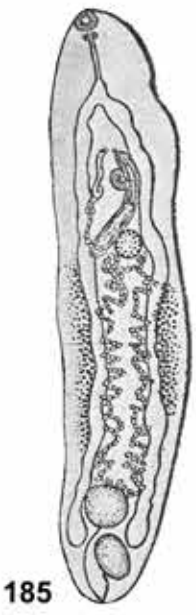
182



183



184



185



186



187



188

Fig. 189: *Telorchis rapidulus* after Dobbin Jr., 1957. Bar = 1mm

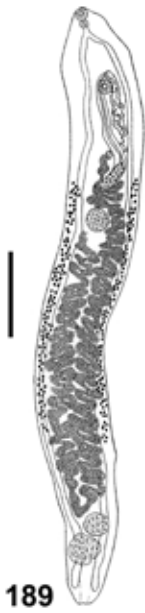
Fig. 190: *Adenogaster serialis* after Salas & Bolaños, 1977. Bar = 3mm

Fig. 191: *Cetiosaccus galapagensis* after Gilbert, 1940. Bar = 2 mm

Fig. 192: *Cricocephalus albus* after Ruiz, 1946. Bar = 1mm

Fig. 193: *Iguanacola navicularius* after Gilbert, 1940. Bar = 2mm

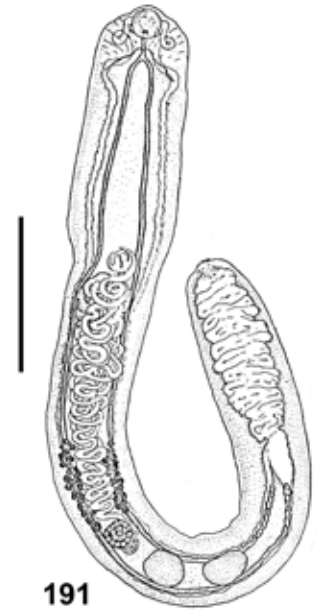
Fig. 194: *Metacetabulum invaginatium* after Freitas & Lent, 1938. Bar = 1mm



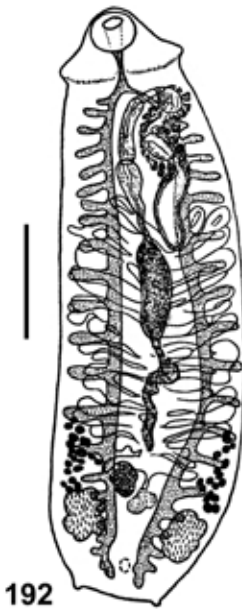
189



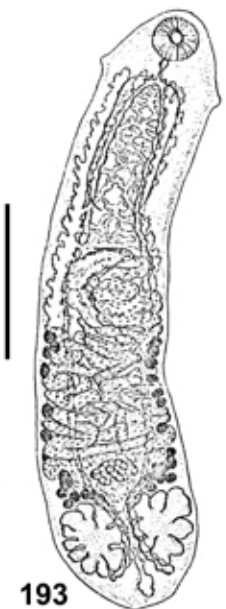
190



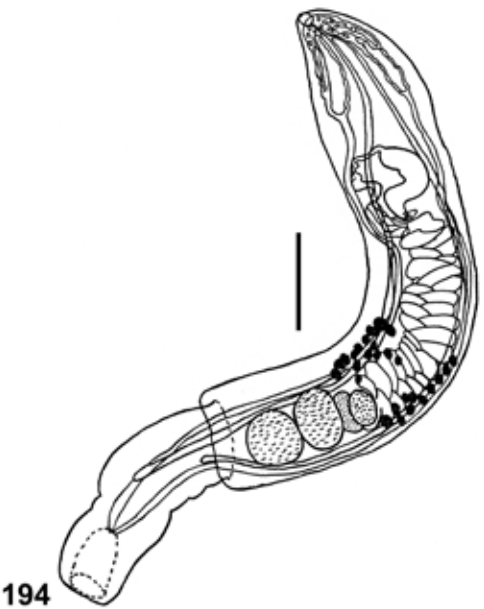
191



192



193



194

Fig. 195: *Pleurogonius linearis* after Ruiz, 1946. Bar = 1mm

Fig. 196: *Pleurogonius lobatus* after Ruiz, 1946.

Fig. 197: *Pleurogonius longiusculus* after Ruiz, 1946.

Fig. 198: *Pleurogonius trigonocephalus* after Ruiz, 1946. Bar = 0.5mm

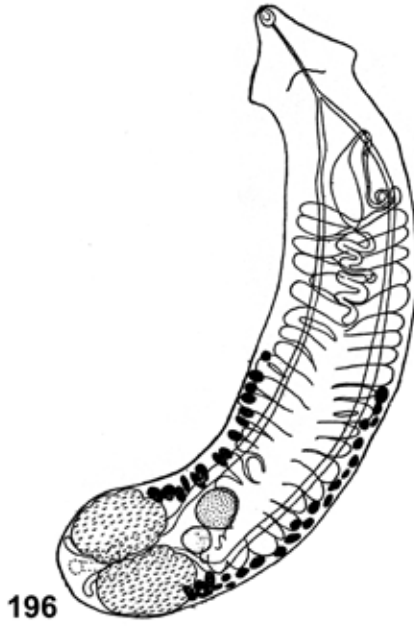
Fig. 199: *Pronocephalus obliquus* after Ruiz, 1946. Bar = 1mm

Fig. 200: *Pyelosomum amblyrhynchi* after Gilbert, 1940. Bar = 1mm

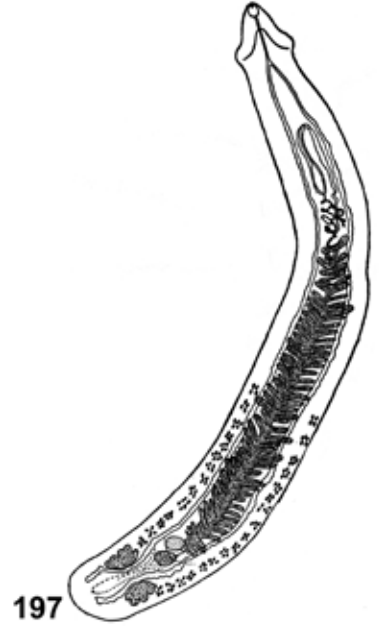




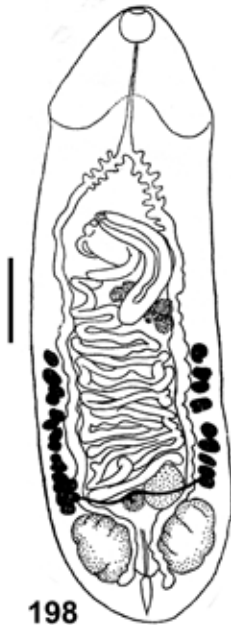
195



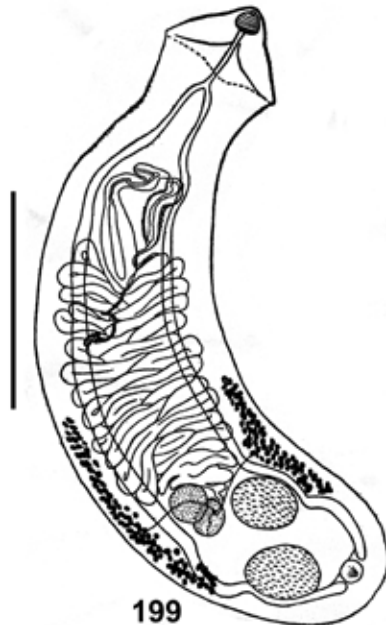
196



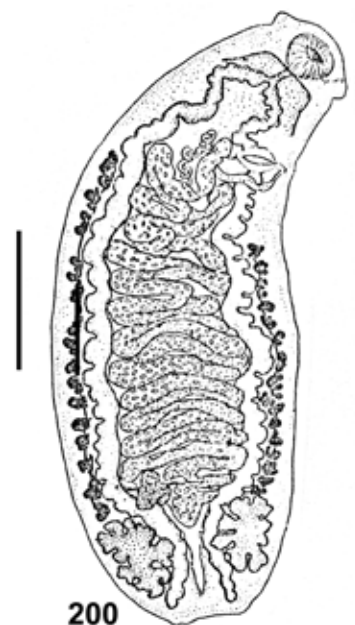
197



198



199



200

Fig. 201: *Pyelosomum crassum* after Ruiz, 1946. Bar = 1mm

Fig. 202: *Ruicephalus minutus* after Ruiz, 1946. Bar = 1mm

Fig. 203: *Amphiorchis caborojoensis* after Werneck, Lima, Gallo & Silva, 2011. Bar = 1mm

Fig. 204: *Amphiorchis indicus* after Werneck & Silva, 2013. Bar = 1mm

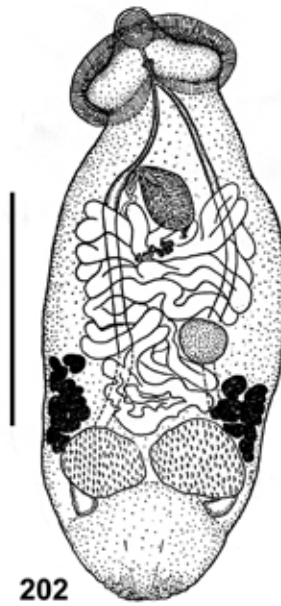
Fig. 205: *Amphiorchis solus* after Werneck, Lima, Gallo & Silva, 2011. Bar = 1mm

Fig. 206: *Learedius learedii* after Werneck, Becker, Gallo & Silva, 2006. Bar = 1mm

Fig. 207: *Monticellius indicum* after Werneck, Gallo & Silva, 2009. Bar = 0.5mm



201



202



203



204



205



206



207



# INDEX TO TREMATODA

## A

<i>Acanthostomum</i> .....	51, 52, 85, 87, 88, 90, 108, 120, 122, 174
<i>Acanthostomum (Acanthostomum) scyphocephalum</i> .....	51
<i>Acanthostomum brauni</i> .....	51, 122
<i>Acanthostomum marajoarum</i> .....	51, 108, 122
<i>Acanthostomum scyphocephalum</i> .....	51, 52, 85, 87, 88, 90, 122, 174
<i>Adenogaster</i> .....	71, 89, 198
<i>Adenogaster serialis</i> .....	71, 89, 198
<i>Aliptrema</i> .....	49, 91, 125, 172
<i>Aliptrema riberoi</i> .....	49, 91, 125
<i>Allassostoma venezuelensis</i> .....	54
<i>Allocreadiidae</i> .....	17, 107, 108, 116
<i>Allopharynx</i> .....	60, 101, 107, 184
<i>Allopharynx daileyi</i> .....	60, 101, 184
<i>Amphiorchis</i> .....	73, 74, 88, 89, 132, 202
<i>Amphiorchis caborojoensis</i> .....	73, 89, 202
<i>Amphiorchis indicus</i> .....	74, 88, 132, 202
<i>Amphiorchis solus</i> .....	74, 88, 132, 202
<i>Amphistoma grande</i> .....	54
<i>Amphistomum dolichocotyle</i> .....	55
<i>Aspidogastrea</i> .....	13, 14
<i>Aspidogastridae</i> .....	105

## B

<i>Bieria</i> .....	60, 91, 186
<i>Bieria artigasi</i> .....	60, 91, 186
<i>Brachycoeliidae</i> .....	17, 46
<i>Brachycoelium</i> .....	17, 46, 83, 84, 96, 98, 112, 168
<i>Brachycoelium mesocoeliiformis</i> .....	46
<i>Brachycoelium salamandrae</i> .....	17, 46, 83, 84, 96, 98, 168
<i>Braunotrema</i> .....	46, 99, 168
<i>Braunotrema pulvinatum</i> .....	46, 99, 168

## C

<i>Caballerotrema sp.</i> .....	43, 86, 164
<i>Caimanicola</i> .....	51, 85, 86, 88, 96, 174
<i>Caimanicola brauni</i> .....	51, 88, 174
<i>Caimanicola marajoara</i> .....	51, 85, 86, 96, 174
<i>Carettacola</i> .....	74, 89, 132
<i>Carettacola stunkardi</i> .....	74, 89, 132
<i>Catadiscus</i> .....	24, 25, 26, 55, 78, 80, 81, 82, 83, 84, 90, 91, 92, 93, 94, 95, 96, 101, 102, 105, 109, 112, 113, 114, 116, 118, 123, 125, 129, 142, 144, 178, 180
<i>Catadiscus cohni</i> .....	24, 78, 129, 142

<i>Catadiscus corderoi</i> .....	24, 81, 83, 118, 142
<i>Catadiscus dolichocotyle</i> .....	55, 90, 91, 94, 178
<i>Catadiscus eldoradiensis</i> .....	25, 83, 105, 142
<i>Catadiscus freitaslenti</i> .....	25, 55, 78, 82, 83, 90, 91, 92, 95, 96, 101, 102, 125, 142, 178
<i>Catadiscus hylae</i> .....	25, 80, 116, 142
<i>Catadiscus inopinatus</i> .....	25, 78, 81, 82, 83, 112, 142
<i>Catadiscus longicoecalis</i> .....	55, 93, 95, 102, 123, 178
<i>Catadiscus marinholutzi</i> .....	26, 78, 83, 144
<i>Catadiscus mirandai</i> .....	26, 84, 112, 144
<i>Catadiscus propinquus</i> .....	26, 81, 82, 84, 112, 114, 144
<i>Catadiscus pygmaeus</i> .....	26
<i>Catadiscus rochai</i> .....	55, 92, 109, 180
<i>Catadiscus uruguayensis</i> .....	26, 55, 80, 81, 83, 92, 118, 144, 180
<i>Cephalogonimus americanus</i> .....	36
<i>Cetiosaccus</i> .....	71, 98, 198
<i>Cetiosaccus galapagensis</i> .....	71, 98, 198
<i>Cheloniodiplostomum</i> .....	39, 88, 101, 158
<i>Cheloniodiplostomum testudinis</i> .....	39, 88, 101, 158
<i>Choledocystus</i> .....	31, 33, 34, 77, 78, 80, 81, 83, 122, 125, 128, 152
<i>Choledocystus elegans</i> .....	31
<i>Choledocystus eucharis</i> .....	31, 122
<i>Choledocystus hepaticus</i> .....	33, 77, 78, 80, 128, 152
<i>Choledocystus incurvatum</i> .....	34, 81, 152
<i>Choledocystus intermedius</i> .....	34
<i>Choledocystus pseudium</i> .....	34, 81, 152
<i>Choledocystus simulans</i> .....	34, 83, 152
<i>Choledocystus vesicalis</i> .....	31, 125
<i>Choledocystus vitellinophilum</i> .....	34, 81, 152
<i>Cladorchiidae</i> .....	116, 117
<i>Cladorchis heteroxenus</i> .....	53
<i>Cotylotretus</i> .....	45, 94, 166
<i>Cotylotretus rugosus</i> .....	45, 94, 166
<i>Creptotrema</i> .....	17, 78, 107, 134
<i>Creptotrema lynchi</i> .....	17, 78, 107, 134
<i>Cricocephalus</i> .....	71, 89, 198
<i>Cricocephalus albus</i> .....	71, 89, 198
<i>Crocodicola</i> .....	39, 40, 86, 110, 158
<i>Crocodicola caimancola</i> .....	40
<i>Crocodicola pseudostoma</i> .....	39, 86, 158
<i>Cryptogonimidae</i> .....	107, 108
<i>Cyathocotyle</i> .....	38, 85, 125, 158
<i>Cyathocotyle brasiliensis</i> .....	38, 85, 125, 158
<i>Cyathocotylidae</i> .....	38, 125
<i>Cystodiplostomum</i> .....	39, 85, 158
<i>Cystodiplostomum hollyi</i> .....	39, 85, 158

## D

<i>Dasymetra</i> .....	65, 101, 190
<i>Dasymetra tupinambis</i> .....	65, 101, 190
<i>Derogenidae</i> .....	117
<i>Dicrocoeliidae</i> .....	124, 130
<i>Dicrocoelium infidum</i> .....	47, 111
<i>Diplodiscidae</i> .....	114, 118, 126
<i>Diplodiscus pygmaeus</i> .....	26
<i>Diplostomum abbreviatum</i> .....	42
<i>Diplostomum longum</i> .....	42
<i>Distoma gelatinosum</i> .....	46
<i>Distoma pyxidatum</i> .....	75, 85
<i>Distomum pulvinatum</i> .....	46
<i>Distomum xenodontis</i> .....	59, 109
<i>Dolichosaccus amplicava</i> .....	36

## E

<i>Echinostoma</i> .....	43, 45, 86, 164
<i>Echinostoma jacaretinga</i> .....	45
<i>Echinostoma sp.</i> .....	43, 86, 164
<i>Echinostomatidae</i> .....	112, 117
<i>Eurytrema parvum</i> .....	48

## G

<i>Gastris sclerops</i> .....	49
<i>Glossidiella</i> .....	61, 91, 186
<i>Glossidiella ornata</i> .....	61, 91, 186
<i>Glossidioides</i> .....	61, 91, 186
<i>Glossidioides loossi</i> .....	61, 91, 186
<i>Glossidium loossi</i> .....	61
<i>Glypthelmins</i> .....	27, 31, 32, 33, 34, 35, 77, 79, 80, 81, 83, 107, 109, 110, 114, 115, 120, 121, 124, 128, 144, 146
<i>Glypthelmins biliaris</i> .....	27, 83, 128, 144
<i>Glypthelmins elegans</i> .....	31
<i>Glypthelmins festina</i> .....	27, 77, 146
<i>Glypthelmins hepaticus</i> .....	34
<i>Glypthelmins incurvatum</i> .....	34
<i>Glypthelmins linguatula</i> .....	31, 34, 121
<i>Glypthelmins palmipedis</i> .....	32
<i>Glypthelmins parva</i> .....	27, 80, 81, 83, 146
<i>Glypthelmins proximus</i> .....	33
<i>Glypthelmins pseudium</i> .....	34
<i>Glypthelmins ramitesticularis</i> .....	34
<i>Glypthelmins repandum</i> .....	33
<i>Glypthelmins robustus</i> .....	33, 107

<i>Glyphelmins sanmartini</i> .....	27, 79, 146
<i>Glyphelmins simulans</i> .....	34
<i>Glyphelmins vesicalis</i> .....	31
<i>Glyphelmins vitellinophilum</i> .....	35, 110, 114, 115
<i>Glyphelminthidae</i> .....	27
<i>Gorgodera</i> .....	18, 83, 128, 134
<i>Gorgodera australiensis</i> .....	18, 83, 128, 134
<i>Gorgoderidae</i> .....	18, 111, 123, 128, 129
<i>Gorgoderina</i> .....	18, 19, 20, 21, 77, 78, 79, 80, 81, 82, 83, 84, 109, 111, 113, 119, 123, 128, 134, 136, 138
<i>Gorgoderina carioca</i> .....	18, 83, 134
<i>Gorgoderina cedroi</i> .....	18, 81, 136
<i>Gorgoderina chilensis</i> .....	19, 79, 80, 109, 136
<i>Gorgoderina cryptorchis</i> .....	19, 77, 78, 82, 83, 136
<i>Gorgoderina darwini</i> .....	19, 77, 119, 136
<i>Gorgoderina diaster</i> .....	19, 78, 81, 84, 136
<i>Gorgoderina festoni</i> .....	20, 78
<i>Gorgoderina (Gorgoderina) cryptorchis</i> .....	19
<i>Gorgoderina (Gorgoderina) diaster</i> .....	19
<i>Gorgoderina (Gorgoderina) rochalimai</i> .....	21
<i>Gorgoderina (Gorgoderina) parvicava</i> .....	20
<i>Gorgoderina megacysta</i> .....	20, 83, 136
<i>Gorgoderina (Metagorgoderina) carioca</i> .....	18
<i>Gorgoderina (Metagorgoderina) diaster</i> .....	19
<i>Gorgoderina (Metagorgoderina) pigulevskyi</i> .....	21
<i>Gorgoderina (Metagorgoderina) rochalimai</i> .....	21
<i>Gorgoderina (Neogorgoderina) chilensis</i> .....	19
<i>Gorgoderina parvicava</i> .....	20, 21, 77, 78, 79, 80, 81, 82, 83, 84, 128, 136
<i>Gorgoderina parvicava minuta</i> .....	21, 80, 136
<i>Gorgoderina permagna</i> .....	20
<i>Gorgoderina pigulevskyi</i> .....	21, 83, 138
<i>Gorgoderina rochalimai</i> .....	21, 77, 79, 82, 83, 138
<i>Gorgoderina sp.</i> .....	21, 80, 113, 138
<i>Gorgoderina valdiviensis</i> .....	21, 79, 123, 138

## H

<i>Haematolechus travdarribus</i> .....	36
<i>Haematoloechidae</i> .....	28, 111, 115
<i>Haematoloechus</i> .....	28, 29, 30, 78, 79, 80, 82, 83, 84, 113, 115, 118, 146, 148
<i>Haematoloechus arequipensis</i> .....	28, 80, 146
<i>Haematoloechus freitasi</i> .....	28, 83, 113, 146
<i>Haematoloechus fuelleborni</i> .....	28, 78, 83, 146
<i>Haematoloechus (Haematoloechus) lutzi</i> .....	29
<i>Haematoloechus iturbei</i> .....	30
<i>Haematoloechus legrandi</i> .....	28, 83
<i>Haematoloechus longiplexus</i> .....	28, 78, 79, 82, 83, 115, 146



<i>Haematoloechus lutzi</i> .....	29, 84, 148
<i>Haematoloechus medioplexus</i> .....	30
<i>Haematoloechus neivai</i> .....	30
<i>Haematoloechus ozorioi</i> .....	29, 83, 148
<i>Haematoloechus pukinensis</i> .....	29, 80, 148
<i>Halipegus</i> .....	23, 83, 84, 109, 117, 122, 140
<i>Halipegus dubius</i> .....	23, 83, 84, 109, 122, 140
<i>Halipegus similis</i> .....	23
<i>Halltrema</i> .....	52, 53, 98, 99, 100, 101, 113, 176
<i>Halltrema avitellina</i> .....	52, 99, 100, 101, 113, 176
<i>Halltrema heteroxenus</i> .....	53, 98, 100, 176
<i>Haplometra palmipedis</i> .....	32
<i>Haplometroides</i> .....	61, 62, 86, 87, 90, 93, 96, 97, 99, 125, 126, 127, 186
<i>Haplometroides buccicola</i> .....	61, 86, 87, 90, 93, 96, 97, 126, 127, 186
<i>Haplometroides intercaecalis</i> .....	62, 93, 126, 127, 186
<i>Haplometroides odhneri</i> .....	62, 97, 99, 127, 186
<i>Helicometra asymmetrica</i> .....	38
<i>Helicotrema</i> .....	38, 98, 99, 101, 156
<i>Helicotrema asymmetricum</i> .....	38, 98, 156
<i>Helicotrema magniovatum</i> .....	38, 98, 156
<i>Helicotrema spirale</i> .....	38, 98, 99, 101, 156
<i>Herpetodiplostomum</i> .....	39, 40, 85, 86, 158
<i>Herpetodiplostomum caimancola</i> .....	39, 40, 85, 86, 158
<i>Herpetodiplostomum testudinis</i> .....	39
<i>Heterocoelium heterocoelium</i> .....	66
<i>Heterodiplostomum</i> .....	40, 90, 91, 92, 93, 95, 102, 118, 158, 160
<i>Heterodiplostomum helicopsis</i> .....	40, 91, 118, 158
<i>Heterodiplostomum lanceolatum</i> .....	40, 90, 91, 92, 93, 95, 102, 160

## I

<i>Iguanacola</i> .....	71, 98, 198
<i>Iguanacola navicularius</i> .....	71, 98, 198
<i>Infidum</i> .....	18, 46, 47, 84, 87, 90, 91, 92, 93, 94, 102, 103, 125, 168, 170
<i>Infidum infidum</i> .....	18, 47, 84, 87, 91, 94, 102, 103, 168
<i>Infidum intermedium</i> .....	47
<i>Infidum luckeri</i> .....	47, 93, 168
<i>Infidum similis</i> .....	47, 90, 92, 93, 94, 102, 170
<i>Iquitos</i> .....	18, 84, 119, 126, 134
<i>Iquitos ceii</i> .....	18, 84, 119, 134

## L

<i>Learedius</i> .....	74, 89, 132, 202
<i>Learedius learedi</i> .....	74, 89, 132
<i>Leptophyllum stenocotyle</i> .....	65, 125
<i>Leurosoma</i> .....	37, 90, 116, 156
<i>Leurosoma rudolfbarthi</i> .....	37, 90, 156

<i>Liophistrema</i> .....	56, 92, 95, 105, 115, 180
<i>Liophistrema pulmonale</i> .....	56, 92, 180
<i>Liophistrema pulmonalis</i> .....	56, 105
<i>Loefgrenia</i> .....	67, 100, 192
<i>Loefgrenia loefgreni</i> .....	67, 100
<i>Lophotaspis</i> .....	14, 37, 88, 156
<i>Lophotaspis vallei</i> .....	14, 37, 88, 156
<i>Loxogenes</i> .....	24, 84
<i>Loxogenes macrocirra</i> .....	24, 84

## M

<i>Macroderoididae</i> .....	107, 115
<i>Maicuru</i> .....	17, 77, 112, 131, 134
<i>Maicuru solitarium</i> .....	17, 77, 112, 134
<i>Margeana chaquensis</i> .....	30, 119
<i>Margeana linguatula</i> .....	31
<i>Margeana proximus</i> .....	33
<i>Margeana pseudium</i> .....	34
<i>Margeana sanmartini</i> .....	28
<i>Margeana sera</i> .....	31
<i>Massoprostatum</i> .....	40, 85, 160
<i>Massoprostatum longum</i> .....	40, 85, 160
<i>Mesocoeliidae</i> .....	112, 114
<i>Mesocoelium</i> .....	14, 22, 23, 48, 49, 77, 78, 79, 82, 83, 84, 86, 87, 92, 93, 94, 95, 96, 97, 98, 99, 100, 101, 110, 114, 121, 122, 138, 140, 172
<i>Mesocoelium incognitum</i> .....	23
<i>Mesocoelium lanfrediae</i> .....	22, 78, 138
<i>Mesocoelium meggitti</i> .....	22, 78, 79, 138
<i>Mesocoelium monas</i> .....	14, 22, 48, 77, 78, 79, 82, 83, 84, 86, 87, 92, 94, 95, 96, 97, 98, 99, 100, 101, 121, 140, 172
<i>Mesocoelium sibynomorphi</i> .....	49, 93, 94, 101, 172
<i>Mesocoelium sociale</i> .....	23
<i>Mesocoelium sp.</i> .....	23
<i>Mesocoelium travassosi</i> .....	22
<i>Mesocoelium waltoni</i> .....	23, 78, 140
<i>Mesodiplostomum</i> .....	41, 86, 160
<i>Mesodiplostomum gladiolum</i> .....	41, 86, 160
<i>Metacetabulum</i> .....	71, 89, 198
<i>Metacetabulum invaginatum</i> .....	71, 89, 198
<i>Metorchis leptodactylus</i> .....	32
<i>Microderma lühei</i> .....	63
<i>Microscaphidiidae</i> .....	107, 125
<i>Monostomum spirale</i> .....	38
<i>Monticellius</i> .....	74, 89, 132, 202
<i>Monticellius indicum</i> .....	74, 89, 132, 202
<i>Myosaccus amblyrhynchi</i> .....	73

## N

<i>Nematophila</i> .....	53, 54, 87, 88, 97, 98, 99, 100, 101, 105, 117, 126, 176, 178
<i>Nematophila argentinum</i> .....	53, 88, 97, 100, 176
<i>Nematophila grande</i> .....	54, 105
<i>Nematophila grandis</i> .....	53, 87, 88, 97, 98, 99, 100, 101, 126, 176
<i>Nematophila ovalis</i> .....	54
<i>Nematophila venezuelensis</i> .....	54, 88, 100, 178
<i>Neotangium</i> .....	50, 103, 125
<i>Neotangium travassosi</i> .....	50, 103, 125
<i>Neodeuterobaris</i> .....	50, 100, 107, 172
<i>Neodeuterobaris pritchardae</i> .....	50, 100, 107, 172
<i>Neohaematoloechus</i> .....	29, 30, 81, 82, 83, 84, 111, 148
<i>Neohaematoloechus iturbei</i> .....	29, 84, 148
<i>Neohaematoloechus neivai</i> .....	30, 81, 82, 83, 84, 111, 148
<i>Nephrocephalus microcephalus</i> .....	38

## O

<i>Ochetosoma chironius</i> .....	65
<i>Ochetosoma heterocoelium</i> .....	66, 109, 117, 127
<i>Ochetosoma miladelarocai</i> .....	66, 107
<i>Ochetosoma monstrosus</i> .....	66
<i>Octangiodes</i> .....	50
<i>Octangioides tlacotalpensis</i> .....	50, 98
<i>Odhneriotrema</i> .....	37, 85, 156
<i>Odhneriotrema microcephala</i> .....	37, 156
<i>Ophiodiplostomum</i> .....	41, 87, 90, 92, 93, 94, 96, 103, 160
<i>Ophiodiplostomum ancyloides</i> .....	41, 90, 160
<i>Ophiodiplostomum spectabile</i> .....	41, 87, 90, 92, 93, 94, 96, 103, 160
<i>Opisthioglyphe</i> .....	36, 81, 130, 154
<i>Opisthioglyphe amplicavus</i> .....	36, 81, 154
<i>Opisthognimidae</i> .....	120, 129
<i>Opisthognimus afranioi</i> .....	56, 89, 94, 102, 180
<i>Opisthognimus artigasi</i> .....	57, 93, 94, 95, 102, 103, 180
<i>Opisthognimus fariai</i> .....	57, 92, 96, 117, 182
<i>Opisthognimus fonsecai</i> .....	57, 89, 90, 92, 93, 94, 96, 102, 103, 126, 182
<i>Opisthognimus interrogativus</i> .....	57, 94, 96, 102, 182
<i>Opisthognimus lecithonotus</i> .....	58, 89, 90, 91, 92, 93, 94, 95, 96, 97, 102, 103, 126, 184
<i>Opisthognimus megabothrium</i> .....	58, 91, 92, 96, 182
<i>Opisthognimus misionesensis</i> .....	58, 96, 118, 182
<i>Opisthognimus (Opisthognimus) megabothrium</i> .....	58
<i>Opisthognimus pereirai</i> .....	58, 89, 182
<i>Opisthognimus philodryadum</i> .....	59
<i>Opisthognimus serpentis</i> .....	59, 90, 91, 92, 94, 95, 96, 184
<i>Opisthognimus sulina</i> .....	59, 94, 184
<i>Opisthognimus uruguayensis</i> .....	60, 95, 184

<i>Opisthogonimus (Westella) philodryadum</i> .....	59
<i>Opisthogonimus (Westella) sulina</i> .....	60
<i>Opistogonimus</i> .....	105
<i>Orchidasma</i> .....	67, 88, 89, 192
<i>Orchidasma amphiorchis</i> .....	67, 88, 89, 192
<i>Oriximinatrema</i> .....	54, 99, 178
<i>Oriximinatrema noronhae</i> .....	54, 99, 178

## P

<i>Pachypsolus</i> .....	49, 85, 86, 172
<i>Pachypsolus sclerops</i> .....	49, 85, 86, 172
<i>Paracotyletrema</i> .....	60, 92
<i>Paracotyletrema poncedeleoni</i> .....	60, 92
<i>Paradiplostomum</i> .....	41, 85, 86, 118, 160
<i>Paradiplostomum abbreviatum</i> .....	41, 85, 86, 118, 160
<i>Paradistomum</i> .....	47, 48, 87, 89, 93, 94, 97, 98, 99, 100, 101, 102, 124, 170
<i>Paradistomum boae</i> .....	47, 87, 170
<i>Paradistomum lutzi</i> .....	48
<i>Paradistomum magnum</i> .....	48
<i>Paradistomum parvissimum</i> .....	48, 89, 93, 94, 97, 98, 99, 100, 101, 102, 170
<i>Paradistomum rabusculum</i> .....	48, 99, 170
<i>Parahaplometroides</i> .....	62, 96, 127, 188
<i>Parahaplometroides basiliscae</i> .....	62, 96, 127, 188
<i>Paramphistomum argentinum</i> .....	53
<i>Petalodiplostomum ancyloides</i> .....	41
<i>Petalodiplostomum aristoterisi</i> .....	41
<i>Plagiorchiidae</i> .....	33, 105, 106, 107, 112, 113, 117, 120, 122, 124, 125, 126, 127, 128
<i>Plagiorchis</i> .....	31, 34, 35, 62, 63, 83, 91, 97, 101, 105, 106, 117, 120, 124, 152, 188
<i>Plagiorchis freitasi</i> .....	62, 101, 188
<i>Plagiorchis hepaticus</i> .....	34
<i>Plagiorchis lenti</i> .....	31
<i>Plagiorchis lühei</i> .....	62, 91
<i>Plagiorchis rangeli</i> .....	35, 83, 106, 152
<i>Plagiorchis vicentei</i> .....	63, 97, 124, 188
<i>Pleurogenidae</i> .....	111
<i>Pleurogonius</i> .....	72, 89, 200
<i>Pleurogonius linearis</i> .....	72, 89, 200
<i>Pleurogonius lobatus</i> .....	72, 89, 200
<i>Pleurogonius longiusculus</i> .....	72, 89, 200
<i>Pleurogonius trigonocephalus</i> .....	72, 89, 200
<i>Pneumonesces neivai</i> .....	30, 130
<i>Pneumonoeces fuelleborni</i> .....	28
<i>Pneumonoeces iturbei</i> .....	30
<i>Pneumonoeces ozorioi</i> .....	29
<i>Pneumonoeces planorbis</i> .....	30
<i>Pneumonoeces pseudis</i> .....	30

<i>Pneumonoeces schulzei</i> .....	36
<i>Pneumonoeces tejeræ</i> .....	29
<i>Pneumotrema</i> .....	63, 86, 106, 188
<i>Pneumotrema travasossi</i> .....	86
<i>Podocnemitrema</i> .....	50, 99, 105, 174
<i>Podocnemitrema papillosus</i> .....	50, 99, 105, 174
<i>Polyangium</i> .....	50, 89
<i>Polyangium linguatula</i> .....	50, 89
<i>Prionosoma</i> .....	44, 88, 164
<i>Prionosoma phrynopsis</i> .....	44, 88, 164
<i>Prionosomoides</i> .....	44, 88, 164
<i>Prionosomoides scalaris</i> .....	44, 88, 164
<i>Proctocaecum</i> .....	52, 86, 174
<i>Proctocaecum dorsale</i> .....	52, 86, 174
<i>Prohemistomum babai</i> .....	40
<i>Prolecithodiplostomum</i> .....	42, 85, 86, 162
<i>Prolecithodiplostomum cavum</i> .....	42
<i>Prolecithodiplostomum constrictum</i> .....	42, 85, 86, 162
<i>Pronocephalidae</i> .....	71, 106, 120, 123, 131
<i>Pronocephalus</i> .....	72, 73, 88, 89, 200
<i>Pronocephalus minutus</i> .....	73
<i>Pronocephalus obliquus</i> .....	72, 88, 89, 200
<i>Pronocephalus trigonocephalus</i> .....	72
<i>Proterodiplostomidae</i> .....	105, 108, 109, 110, 118, 121
<i>Proterodiplostomum</i> .....	42, 43, 85, 86, 109, 121, 162, 164
<i>Proterodiplostomum brasiliensis</i> .....	43
<i>Proterodiplostomum breve</i> .....	42, 86, 162
<i>Proterodiplostomum globulare</i> .....	42, 86, 162
<i>Proterodiplostomum intermedium</i> .....	43, 121
<i>Proterodiplostomum longum</i> .....	42, 85, 86, 109, 162
<i>Proterodiplostomum medusæ</i> .....	43, 85, 86, 162
<i>Proterodiplostomum tumidulum</i> .....	43, 85, 86, 164
<i>Pseudallassostoma</i> .....	53
<i>Pseudallassostoma heteroxenus</i> .....	53
<i>Pseudocleptodiscus</i> .....	54, 98
<i>Pseudocleptodiscus margaritæ</i> .....	54, 98
<i>Pseudonematophila</i> .....	54, 100, 117, 178
<i>Pseudonematophila ovalis</i> .....	54, 100, 178
<i>Pseudoneodiplostomum brasiliensis</i> .....	43
<i>Pseudosonsinotrema</i> .....	24, 78, 80, 111, 140
<i>Pseudosonsinotrema chabaudi</i> .....	24, 78, 140
<i>Pseudosonsinotrema megalorchis</i> .....	24, 80, 111, 140
<i>Pseudotelorchis</i> .....	67, 68, 85, 86, 88, 108, 192, 194
<i>Pseudotelorchis caimanis</i> .....	67, 85, 86, 192
<i>Pseudotelorchis devincenzii</i> .....	67, 88

<i>Pseudotelorchis yacarei</i> .....	68, 85, 86, 194
<i>Psilostomidae</i> .....	45, 121
<i>Pulchrosomoides</i> .....	44, 98, 100, 101, 164
<i>Pulchrosomoides elegans</i> .....	44, 98, 100, 101, 164
<i>Pyelosomum</i> .....	73, 89, 98, 123, 200, 202
<i>Pyelosomum amblyrhynchi</i> .....	73, 98, 200
<i>Pyelosomum crassum</i> .....	73, 89, 202
<i>Pyelosomum longiusculus</i> .....	73, 89

## R

<i>Rauschiella</i> .....	28, 30, 31, 32, 33, 56, 77, 78, 79, 80, 81, 82, 83, 84, 93, 128, 148, 150
<i>Rauschiella chaquensis</i> .....	30, 82, 148
<i>Rauschiella lenti</i> .....	30, 83, 150
<i>Rauschiella linguatula</i> .....	31, 56, 77, 78, 79, 82, 83, 84, 93, 150
<i>Rauschiella palmipedis</i> .....	31, 77, 78, 79, 81, 82, 83, 84, 150
<i>Rauschiella proxima</i> .....	32, 83, 150
<i>Rauschiella repandum</i> .....	33, 80, 82, 83, 150
<i>Rauschiella robusta</i> .....	33, 78, 150
<i>Renifer</i> .....	65, 66, 89, 90, 91, 92, 94, 96, 101, 102, 103, 190, 192
<i>Renifer chironius</i> .....	65, 90, 190
<i>Renifer monstrosus</i> .....	66, 102, 192
<i>Rhytidodes</i> .....	14, 46, 88, 89, 99, 166
<i>Rhytidodes gelatinosus</i> .....	14, 46, 88, 89, 99, 166
<i>Rhytidodidae</i> .....	46
<i>Rudolphiella rudolphi</i> .....	35
<i>Rudolphitrema</i> .....	35, 77, 78, 80, 82, 116, 120, 123, 154
<i>Rudolphitrema chilensis</i> .....	35, 80, 123, 154
<i>Rudolphitrema physalaemi</i> .....	35, 82, 120, 154
<i>Rudolphitrema rudolphii</i> .....	35, 77, 78, 116, 154
<i>Ruicephalus</i> .....	73, 89, 202
<i>Ruicephalus minutus</i> .....	73, 89, 202

## S

<i>Sphaeridiotrema</i> .....	45, 98, 121, 166
<i>Spirorchiidae</i> .....	132
<i>Stephanoprora</i> .....	44, 45, 85, 86, 164, 166
<i>Stephanoprora campomica</i> .....	44, 85, 164
<i>Stephanoprora jacaretinga</i> .....	44, 85, 86, 166
<i>Stephanoprora nattereri</i> .....	45, 86, 166
<i>Stephanoprora sp.</i> .....	45, 85, 166
<i>Sticholecitha</i> .....	63, 90, 96, 103, 127, 188
<i>Sticholecitha serpentis</i> .....	63, 90, 96, 103, 127, 188
<i>Styphlodora</i> .....	63, 64, 87, 89, 91, 93, 94, 95, 96, 97, 100, 102, 103, 111, 119, 190
<i>Styphlodora condita</i> .....	63, 87, 91, 93, 94, 95, 96, 97, 100, 102, 103, 111, 190
<i>Styphlodora gili</i> .....	64, 89, 102, 119, 190
<i>Styphlodora horrida</i> .....	64, 87

<i>Styphlotrema</i> .....	67, 89, 132, 192
<i>Styphlotrema solitaria</i> .....	67, 89, 132, 192
<i>Styphlotrematidae</i> .....	132

## T

<i>Telorchiidae</i> .....	108, 110, 111, 119
<i>Telorchis</i> .....	68, 69, 70, 87, 88, 89, 90, 91, 97, 99, 100, 103, 110, 112, 119, 120, 121, 194, 196, 198
<i>Telorchis achavali</i> .....	68, 97, 119, 194
<i>Telorchis aculeatus</i> .....	68, 69, 70, 100, 121, 194
<i>Telorchis bifurcus</i> .....	68, 99, 194
<i>Telorchis birabeni</i> .....	68, 88, 194
<i>Telorchis clava</i> .....	69, 87, 89, 90, 91, 194
<i>Telorchis devincenzii</i> .....	68
<i>Telorchis diaphanus</i> .....	69, 112, 196
<i>Telorchis diminutus</i> .....	70
<i>Telorchis dubius</i> .....	69, 97, 196
<i>Telorchis hagmanni</i> .....	68, 69, 99, 100, 196
<i>Telorchis parvus</i> .....	69, 97, 196
<i>Telorchis platensis</i> .....	70, 88, 196
<i>Telorchis pleroticus</i> .....	70, 103, 196
<i>Telorchis productus</i> .....	70, 88, 196
<i>Telorchis rapidulus</i> .....	70, 99, 198
<i>Thaumatocotyle pulvinatum</i> .....	46
<i>Timoniella</i> .....	52, 85, 88, 90, 176
<i>Timoniella incognita</i> .....	52, 85, 90, 176
<i>Timoniella ostrowskiae</i> .....	52, 85, 88, 90, 176
<i>Travtrema</i> .....	35, 36, 64, 65, 80, 81, 84, 90, 91, 92, 93, 94, 95, 96, 102, 103, 112, 122, 190
<i>Travtrema aff. stenocotyle</i> .....	36
<i>Travtrema stenocotyle</i> .....	35, 64, 80, 81, 84, 90, 91, 92, 93, 94, 95, 96, 102, 103, 112, 122, 190
<i>Travtrema travtrema</i> .....	65, 122

## U

<i>Urotrema</i> .....	70, 96
-----------------------	--------

## W

<i>Westella afranioi</i> .....	57
<i>Westella serpentis</i> .....	59
<i>Westella sulina</i> .....	60

## Z

<i>Zeferinella vazi</i> .....	62, 105
<i>Zoogonoides boae</i> .....	48





# INDEX TO HOSTS

## A

<i>Alopoglossus angulatus</i> .....	49, 98, 114
<i>Alsodes roseus</i> .....	35, 80
<i>Amblyrhynchus cristatus</i> .....	71, 73, 98, 113
<i>Ameiva ameiva</i> .....	48, 100
<i>Amphisbaena alba</i> .....	61, 63, 86
<i>Amphisbaena ridleyi</i> .....	48, 86, 123
<i>Amphisbaena sp.</i> .....	48, 87
<i>Anaxyrus terrestris</i> .....	31, 77
<i>Anolis fuscoauratus</i> .....	48, 70, 96
<i>Anolis nitens</i> .....	46, 96
<i>Anolis scypheus</i> .....	46
<i>Atelopus bomolochus</i> .....	20, 77, 116
<i>Atelopus ignescens</i> .....	19, 35, 77
<i>Atractus lasallei</i> .....	66, 89

## B

<i>Basiliscus basiliscus</i> .....	62, 96, 127
<i>Batrachemys nasuta</i> .....	53, 87
<i>Batrachophrynus brachydactylus</i> .....	19, 79
<i>Batrachophrynus macrostomus</i> .....	20, 21, 80, 113
<i>Boa constrictor</i> .....	47, 64, 87
<i>Boa scytale</i> .....	69
<i>Boiruna maculata</i> .....	59, 69, 89
<i>Bothriechis schlegelli</i> .....	66
<i>Bothropoides diporus</i> .....	47, 55, 58, 63, 64, 101
<i>Bothropoides insularis</i> .....	66, 102
<i>Bothropoides jararaca</i> .....	47, 48, 57, 58, 64, 66, 102
<i>Bothropoides neuwiedi</i> .....	55, 56, 57, 58, 63, 64, 66, 102
<i>Bothrops alternata</i> .....	40, 58, 64, 102, 119
<i>Bothrops alternatus</i> .....	40, 55, 57, 63, 64, 102
<i>Bothrops asper</i> .....	66, 102
<i>Bothrops atrox</i> .....	59, 66, 102, 107
<i>Bothrops cotiara</i> .....	66, 103
<i>Bothrops insularis</i> .....	66, 102
<i>Bothrops jararaca</i> .....	47, 48, 57, 58, 64, 66, 102
<i>Bothrops jararacussu</i> .....	59, 103
<i>Bothrops moojeni</i> .....	47, 57, 58, 59, 63, 64, 66, 103, 106, 126, 127
<i>Bothrops neuwiedi diporus</i> .....	47, 101
<i>Bothrops neuwiedii</i> .....	55, 56, 57, 58, 63, 64, 66, 102, 103
<i>Bothrops neuwiedii mattogrossensis</i> .....	66, 103
<i>Bothrops neuwiedii meridionalis</i> .....	55, 58, 63, 64
<i>Bothrops sp.</i> .....	59, 64, 103

<i>Bufo agua</i> .....	20, 31, 32, 78
<i>Bufo arenarius</i> .....	21, 77
<i>Bufo crucifer</i> .....	19, 20, 22, 31, 35, 77
<i>Bufo dorbigny</i> .....	19, 77
<i>Bufo fernandezae</i> .....	29, 78
<i>Bufo granulatus</i> .....	17, 22, 31, 32, 33, 77
<i>Bufo horribilis</i> .....	22, 78
<i>Bufo ictericus</i> .....	20, 22, 78, 118
<i>Bufo limensis</i> .....	78
<i>Bufo marinus</i> .....	17, 19, 20, 22, 23, 24, 28, 31, 32, 33, 78, 107, 125
<i>Bufo marinus bimaculatus</i> .....	22, 31, 78
<i>Bufo marinus ictericus</i> .....	22, 31, 78
<i>Bufo marinus marinus</i> .....	22, 78
<i>Bufo musicus</i> .....	31, 77
<i>Bufo paracnemis</i> .....	20, 21, 22, 25, 31, 32, 78
<i>Bufo schneideri</i> .....	20, 21, 22, 25, 31, 32, 78
<i>Bufo sp.</i> .....	22, 79
<i>Bufo spinulosus limensis</i> .....	20, 78

## C

<i>Caiman crocodilus</i> .....	14, 37, 38, 39, 40, 41, 42, 43, 44, 45, 49, 51, 52, 67, 68, 75, 85, 105, 108, 121
<i>Caiman crocodilus crocodilus</i> .....	43, 52, 85, 121
<i>Caiman crocodilus fuscus</i> .....	40, 42, 85, 105
<i>Caiman crocodilus yacare</i> .....	38, 39, 41, 42, 43, 44, 49, 51, 52, 67, 68, 85, 108
<i>Caiman latirostris</i> .....	39, 41, 85, 118
<i>Caiman sclerops</i> .....	37, 38, 39, 41, 42, 43, 44, 49, 51, 75, 85, 113, 125
<i>Caiman sp.</i> .....	39, 41, 42, 43, 86
<i>Caiman yacare</i> .....	38, 39, 41, 42, 43, 44, 49, 51, 52, 67, 68, 85
<i>Calyptocephalella gayi</i> .....	21, 79
<i>Caretta caretta</i> .....	14, 37, 46, 67, 72, 88
<i>Caudiverbera caudiverbera</i> .....	21, 79, 123
<i>Ceratophrys cornuta</i> .....	31, 79
<i>Ceratophrys cranwelli</i> .....	28, 79
<i>Cercosaura eigenmanni</i> .....	48, 98
<i>Chaunus ictericus</i> .....	28, 32, 78, 116
<i>Chaunus marinus</i> .....	22, 78, 106
<i>Chelone mydas</i> .....	50, 71, 72, 88, 113
<i>Chelonia mydas agassizii</i> .....	67, 71, 89
<i>Chelonoidis denticulata</i> .....	38, 52, 101
<i>Chelus fimbriata</i> .....	87
<i>Chelus fimbriatus</i> .....	51, 53, 87
<i>Chelys fimbriata</i> .....	51, 53, 87
<i>Chironius bicarinatus</i> .....	57, 64, 66, 89
<i>Chironius carinatus</i> .....	48, 56, 58, 63, 65, 66, 89
<i>Chironius exoletus</i> .....	66, 90, 127

<i>Chironius foveatus</i> .....	57, 90
<i>Chironius fuscus</i> .....	37, 55, 65, 66, 90
<i>Chthonerpeton indistinctum</i> .....	27, 31, 79
<i>Cistudo lutaria</i> .....	69, 97
<i>Clelia occipitolutea</i> .....	57, 66, 94
<i>Clelia rustica</i> .....	64, 90
<i>Cloelia cloelia</i> .....	59, 90
<i>Coluber flaviventris</i> .....	69
<i>Coluber pullatus</i> .....	45, 94
<i>Coluber sp.</i> .....	40, 41, 90
<i>Constrictor constrictor</i> .....	47, 64, 69, 87
<i>crocodiles</i> .....	103, 122
<i>Crocodylus coroa</i> .....	42, 86
<i>Crocodylus sp.</i> .....	39, 41, 42, 85, 86
<i>Crotalus durissus terrificus</i> .....	41, 103
<i>Cyclagras gigas</i> .....	40, 47, 59, 61, 62, 63, 91
<i>Cystignatus ocellatus</i> .....	33

## D

<i>Dendropsophus leucophyllatus</i> .....	27, 80
<i>Diploglossus lessonae</i> .....	48, 87
<i>Dromicus typhlus</i> .....	55, 92
<i>Dryadophis bifossatus</i> .....	41, 47, 48, 49, 56, 59, 65, 93, 111
<i>Drymarchon corais</i> .....	47, 52, 66, 69, 90
<i>Drymarchon corais corais</i> .....	47, 66, 69, 90
<i>Drymobius bifossatus</i> .....	41, 47, 57, 59, 63, 65, 93
<i>Dryophylax pallidus</i> .....	57, 59, 65, 95

## E

<i>Echinosaura horrida horrida</i> .....	45, 98
<i>Elaps sp.</i> .....	61, 97
<i>Elosia nasus</i> .....	18, 36, 81
<i>Emys lutaria</i> .....	69, 97
<i>Emys orbicularis</i> .....	69, 97
<i>Epicrates cenchria crassus</i> .....	61, 87
<i>Epicrates crassus</i> .....	61, 87
<i>Eretmochelys imbricata</i> .....	67, 74, 89, 132
<i>Erythrolamprus aesculapii</i> .....	57, 61, 90
<i>Erythrolamprus aesculapii</i> .....	55, 66
<i>Eudryas bifossatus</i> .....	47, 93
<i>Eunectes deschauenseei</i> .....	41, 87
<i>Eunectes murinus</i> .....	47, 61, 69, 87
<i>Eunectes notaeus</i> .....	47, 63, 69, 87
<i>Eunectes sp.</i> .....	69
<i>Eusophus roseus</i> .....	35, 123

**F**

“freshwater turtle” ..... 70, 103

**G**

*Gastrotheca pseustes* ..... 24, 80, 111  
*Geochelone denticulata* ..... 38, 101  
*Geoemyda punctularia punctularia* ..... 53, 97  
*Gymnodactylus geckoides* ..... 48, 99

**H**

*Halichelis atra* ..... 72, 88  
*Helicops carinicauda* ..... 59, 90  
*Helicops carinicaudus* ..... 59, 90, 118  
*Helicops corinicauda* ..... 59  
*Helicops infrataeniatus* ..... 40, 59, 91  
*Helicops leopardina* ..... 40, 91  
*Helicops leopardinus* ..... 40  
*Helicops modestus* ..... 66, 91  
*Hemidactylus mabouia* ..... 48, 63, 97, 124  
*Hemipipa carvalhoi* ..... 26, 84, 112  
*Herpetodryas fuscus* ..... 55, 65, 90  
*Hidromedusa tectifera* ..... 70, 88  
*Hydraspis geoffroyana* ..... 53, 88  
*Hydraspis gibba* ..... 53, 88  
*Hydraspis schopjii* ..... 53, 88  
*Hydraspis sp.* ..... 53, 54, 88  
*Hydrodynastes gigas* ..... 40, 47, 59, 61, 62, 63, 69, 91, 105  
*Hydromedusa tectifera* ..... 53, 68, 70, 88  
*Hydroscopis plumbeus* ..... 69  
*Hyla crepitans* ..... 33, 80  
*Hyla leucophyllata* ..... 27, 80  
*Hyla pulchella* ..... 25, 26, 80, 116  
*Hyla raniceps* ..... 34, 81, 110  
*Hylodes nasus* ..... 18, 36, 81  
*Hypsiboas crepitans* ..... 33, 80  
*Hypsiboas pulchellus* ..... 25, 26, 64, 80  
*Hypsiboas raniceps* ..... 34, 81  
*Hypstophus tuberculatus* ..... 38, 98

**I**

*Iguana iguana* ..... 38, 44, 48, 98  
*Iguana tuberculata* ..... 38, 44, 98, 113  
*Incilius nebulifer* ..... 17, 22, 31, 32, 33, 77

**K**

<i>Kentropyx calcarata</i> .....	48, 100
<i>Kinixys erosa</i> .....	53, 101
<i>Kinosternon scorpioides</i> .....	53, 69, 70, 98
<i>Kinosternon scorpioides scorpioides</i> .....	53, 69, 98

**L**

<i>Lachesis newwiedii</i> .....	66, 102
<i>Leimadophis almada</i> .....	55, 91
<i>Leimadophis chamissonis</i> .....	47, 93
<i>Leimadophis poecilogyrus</i> .....	41, 47, 48, 59, 65, 92
<i>Leimadophis typhlus</i> .....	57, 59, 93
<i>Leposoma osvaldoi</i> .....	46, 98
<i>Leposternon microcephalum</i> .....	48, 87
<i>Leptodactylus bolivianus</i> .....	31, 32, 82
<i>Leptodactylus bufonius</i> .....	25, 33, 82, 115
<i>Leptodactylus caliginosus</i> .....	26, 32, 83
<i>Leptodactylus chaquensis</i> .....	20, 21, 25, 26, 28, 31, 33, 82, 115, 126
<i>Leptodactylus fuscus</i> .....	22, 25, 32, 82
<i>Leptodactylus labyrinthicus</i> .....	20, 30, 31, 82
<i>Leptodactylus laticeps</i> .....	30, 82, 119
<i>Leptodactylus latinasus</i> .....	25, 29, 33, 83, 114
<i>Leptodactylus latrans</i> .....	14, 18, 19, 20, 21, 22, 23, 24, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 83
<i>Leptodactylus martinezi</i> .....	17, 83
<i>Leptodactylus mystaceus</i> .....	22, 83
<i>Leptodactylus mystacinus</i> .....	22, 84
<i>Leptodactylus ocellatus</i> .....	18, 19, 20, 21, 22, 23, 25, 26, 27, 28, 29, 30, 31, 32, 33, 34, 35, 83, 105, 106, 111, 119, 122, 127, 128, 131
<i>Leptodactylus pentadactylus</i> .....	20, 22, 23, 30, 31, 32, 82, 84, 111
<i>Leptodactylus pentadactylus labyrinthicus</i> .....	20, 30, 31, 32, 82
<i>Leptodactylus podicipinus</i> .....	18, 26, 36, 84, 108
<i>Leptodactylus rhodomystax</i> .....	17, 84
<i>Leptodactylus rhodonotus</i> .....	20, 84
<i>Leptodactylus sibilatrix</i> .....	22, 82
<i>Leptodactylus typhonius</i> .....	25, 32, 82
<i>Leptodeira annulata</i> .....	63, 66, 91
<i>Leptodeira annulata annulata</i> .....	63, 91
<i>Leptodeira annulata pulchriceps</i> .....	63, 91
<i>Leptodeira septentrionalis</i> .....	66, 91
<i>Leptophis ahaetulla</i> .....	66, 91
<i>Leptotyphlops koppesi</i> .....	62, 99, 127
<i>Liolaemus lutzae</i> .....	48, 99, 124
<i>Liophis almadensis</i> .....	55, 66, 91
<i>Liophis anomalus</i> .....	60, 65, 131

<i>Liophis flavifrenatus</i> .....	55, 64, 92
<i>Liophis jaegeri</i> .....	58, 64, 91
<i>Liophis merremii</i> .....	58
<i>Liophis miliaris</i> .....	41, 47, 49, 55, 56, 57, 58, 59, 60, 64, 65, 66, 91, 92
<i>Liophis miliaris miliaris</i> .....	41, 47, 49, 55, 56, 57, 58, 60, 65, 91
<i>Liophis miliaris semiaureus</i> .....	57, 64, 92
<i>Liophis poecilogyrus</i> .....	40, 41, 47, 48, 55, 57, 59, 64, 65, 66, 92
<i>Liophis poecilogyrus reticulatus</i> .....	40, 92
<i>Liophis reginae</i> .....	41, 66, 92
<i>Liphis anomalus</i> .....	59, 92
<i>Lisapsus limellum</i> .....	26, 34
<i>Lygodactylus klugei</i> .....	48, 97, 105
<i>Lygophis anomalus</i> .....	59, 60, 65, 92
<i>Lygophis flavifrenatus</i> .....	55, 59, 64, 92
<i>Lygophis typhlus</i> .....	55, 57, 59, 66, 93
<i>Lysapsus limellus</i> .....	26, 34, 114, 115, 116
<i>Lysapsus mantidactylus</i> .....	26, 81
<i>Lystrophis dorbignyi</i> .....	55, 57, 59, 64, 95

## M

<i>Mabuya agilis</i> .....	48, 100, 132
<i>Mabuya macrorhyncha</i> .....	44, 48, 100, 132
<i>Mabuya maculata</i> .....	48, 100
<i>marine turtle</i> .....	103
<i>Mastigodryas bifossatus</i> .....	40, 41, 47, 48, 49, 56, 57, 59, 63, 65, 66, 93
<i>Mastigodryas bifossatus triseriatus</i> .....	57, 59, 63, 93
<i>Melanophryniscus stelzneri</i> .....	19, 77
<i>Melanosuchus niger</i> .....	39, 41, 42, 43, 45, 86
<i>Mesoclemmys gibba</i> .....	53, 88
<i>Mesoclemmys nasuta</i> .....	53, 87
<i>Micrurus corallinus</i> .....	61, 96
<i>Micrurus frontalis</i> .....	61, 62, 63, 97, 127
<i>Micrurus frontalis pyrrhocryptus</i> .....	63, 97
<i>Micrurus lemniscatus</i> .....	62, 97
<i>Micrurus pyrrhocryptus</i> .....	59, 63, 97
<i>Micrurus sp.</i> .....	61, 97

## N

<i>Norops fuscoauratus</i> .....	70, 96, 114
----------------------------------	-------------

## O

<i>Odontophrynus americanus</i> .....	33, 80
<i>Ophis merremii</i> .....	58, 59, 64, 65, 95
<i>Oreophis (Driomicus) hoodensis</i> .....	47, 93
<i>Oxyrhophus cloelia</i> .....	69
<i>Oxyrhopus rhombifer</i> .....	63, 93

**P**

<i>Paleosuchus palpebrosus</i> .....	49, 86
<i>Paleosuchus</i> sp. ....	42, 51, 86
<i>Peltocephalus dumerilianus</i> .....	38, 53, 99
<i>Phalotris lativittatus</i> .....	61, 93, 127
<i>Phalotris matogrossensis</i> .....	62, 93
<i>Phalotris nasutus</i> .....	62, 93, 127
<i>Philodryas hoodensis</i> .....	47, 93
<i>Philodryas olfersii</i> .....	47, 55, 93
<i>Philodryas patagoniensis</i> .....	48, 56, 57, 58, 59, 60, 64, 65, 66, 94
<i>Philodryas psammophideus</i> .....	47, 57, 94
<i>Philodryas schotti</i> .....	56, 59, 60
<i>Philodryas schottii</i> .....	48, 58, 59, 65, 94
<i>Philodryas</i> sp.....	41, 47, 55, 59, 64, 65, 94
<i>Phrynohyas coriacea</i> .....	27, 81
<i>Phrynops geoffroanus</i> .....	44, 53, 88
<i>Phrynops geoffroanus geoffroanus</i> .....	44, 88
<i>Phrynops gibbus</i> .....	53, 88
<i>Phrynops hilarii</i> .....	39, 44, 51, 53, 68, 88, 118
<i>Phrynops</i> sp.....	53, 54, 88
<i>Phyllomedusa azurea</i> .....	26, 81
<i>Phylodryas patagoniensis</i> .....	63
<i>Phylodryas schottii</i> .....	63
<i>Physalaemus gracilis</i> .....	35, 82, 120
<i>Physalaemus santafecinus</i> .....	25, 33, 82
<i>Pipa carvalhoi</i> .....	26, 84
<i>Plica plica</i> .....	48, 114
<i>Podocnemis cayennensis</i> .....	68, 100
<i>Podocnemis dumeriliana</i> .....	38, 53, 99
<i>Podocnemis erythrocephala</i> .....	53, 99
<i>Podocnemis expansa</i> .....	14, 46, 50, 52, 53, 54, 68, 69, 99, 116
<i>Podocnemis lewyana</i> .....	50, 53, 54, 69, 100, 107, 117
<i>Podocnemis</i> sp.....	52, 53, 54, 100
<i>Podocnemis tracaxa</i> .....	38, 53, 99
<i>Podocnemis unifilis</i> .....	53, 67, 68, 100, 126
<i>Podocnemis vogli</i> .....	53, 100
<i>Porthidium nasutum</i> .....	66, 103
<i>Prionodactylus eigenmanni</i> .....	48, 98, 107
<i>Pseudemys dorbigni</i> .....	69, 97, 119
<i>Pseudemys dorbignyi</i> .....	68
<i>Pseudis mantidactylus</i> .....	34, 81, 119
<i>Pseudis meridionalis</i> .....	25, 81
<i>Pseudis minuta</i> .....	25, 26, 34, 81
<i>Pseudis paradoxa</i> .....	19, 20, 26, 30, 34, 81, 128
<i>Pseudis platensis</i> .....	26, 32, 81, 108

<i>Pseudoboa cloelia</i> .....	59, 69, 90
<i>Pseudoboa coronata</i> .....	66, 94
<i>Pseudopaludicola boliviana</i> .....	19, 29, 82

## R

<i>Rana palmipes</i> .....	18, 19, 20, 24, 26, 29, 30, 32, 84, 109, 119
<i>Rana sp.</i> .....	19, 29, 36, 84
<i>Rhinella arenarum</i> .....	21, 22, 27, 77
<i>Rhinella crucifer</i> .....	19, 20, 22, 31, 35, 77
<i>Rhinella dorbignyi</i> .....	19, 77
<i>Rhinella fernandezae</i> .....	20, 25, 26, 29, 32, 78, 115, 126
<i>Rhinella icterica</i> .....	20, 22, 24, 28, 31, 32, 35, 78, 126
<i>Rhinella limensis</i> .....	20, 78
<i>Rhinella marina</i> .....	19, 22, 23, 24, 28, 31, 32, 33, 78, 106, 114
<i>Rhinella schneideri</i> .....	20, 21, 22, 25, 31, 32, 78
<i>Rhinella sp.</i> .....	22, 79
<i>Rhinoclemmys areolata</i> .....	53, 97
<i>Rhinoclemmys nasuta</i> .....	50, 53, 54, 98
<i>Rhinoclemmys punctularia</i> .....	53, 97, 98
<i>Rhinoclemmys punctularia punctularia</i> .....	53, 98
<i>Rhinoderma darwinii</i> .....	19, 80
<i>Rhynemis nasuta</i> .....	53, 87

## S

<i>Scinax nasicus</i> .....	25, 35, 81, 115
<i>Scinax pedromedinai</i> .....	27, 81
<i>Sibynomorphus mikanii</i> .....	49, 94
<i>Sibynomorphus mikanii mikanii</i> .....	49, 94
<i>Sibynomorphus sp.</i> .....	48, 94
<i>Sibynomorphus turgidus</i> .....	48, 94
<i>Sibynomorphus ventrimaculatus</i> .....	48, 64, 94
<i>Siphonops annulatus</i> .....	14, 22, 79
<i>Spilotes pullatus</i> .....	45, 64, 94

## T

<i>Telmatobius brachydactylus</i> .....	19, 79
<i>Telmatobius culeus</i> .....	20, 79
<i>Telmatobius jelskii</i> .....	20, 79, 116
<i>Telmatobius macrostomus</i> .....	20, 21, 80
<i>Telmatobius peruvianus</i> .....	20, 28, 29, 80
<i>Telmatobius sp.</i> .....	20, 21, 80
<i>Testudo denticulata</i> .....	52, 101
<i>Testudo matamata</i> .....	51, 87
<i>Testudo orbicularis</i> .....	69, 97
<i>Testudo sp.</i> .....	39, 101
<i>Testudo tabulata</i> .....	38, 101



<i>Thalassochelis caretta</i> .....	46
<i>Thamnodynastes hypoconia</i> .....	60, 95
<i>Thamnodynastes pallidus</i> .....	57, 59, 64, 65, 95
<i>Thamnodynastes sp.</i> .....	57, 59, 64, 95
<i>Thamnodynastes strigatus</i> .....	56, 59, 64, 95
<i>Thamnodynastes strigilis</i> .....	60, 64, 95
<i>Thecadactylus solimoensis</i> .....	48, 99
<i>Tomodon dorsatum</i> .....	65, 95
<i>Tomodon dorsatus</i> .....	59, 65, 95
<i>Tomodon ocellatus</i> .....	48, 95
<i>Trachemys callirostris callirostris</i> .....	53, 97
<i>Trachemys dorbignii</i> .....	68, 69, 97
<i>Trachycephalus coriaceus</i> .....	27, 81
<i>Trachylepis atlantica</i> .....	48, 100, 123
<i>Trilepida koppei</i> .....	62, 99
<i>Tropidurus torquatus</i> .....	48, 62, 101
<i>Tropidurus torquatus torquatus</i> .....	48, 101
<i>Tupinambis nigropunctatus</i> .....	65, 101
<i>Tupinambis rufescens</i> .....	64, 100
<i>Tupinambis teguixin</i> .....	44, 48, 65, 101

## U

<i>Uranoscodon superciliosus</i> .....	48, 49, 60, 101, 107
--	----------------------

## W

<i>Waglerophis merremi</i> .....	55, 59
<i>Waglerophis merremii</i> .....	57, 58, 64, 65, 66, 95, 118

## X

<i>Xenodon dorbignyi</i> .....	55, 57, 59, 64, 95
<i>Xenodon guentheri</i> .....	40, 95
<i>Xenodon merremi</i> .....	41, 55, 57, 58, 59, 64, 65, 66, 95
<i>Xenodon severus</i> .....	63, 96

## Z

<i>Zamensis constrictor</i> .....	69
-----------------------------------	----

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