Review Article
Experimental Chemotherapy for Chagas Disease: A Morphological, Biochemical, and Proteomic Overview of Potential Trypanosoma cruzi Targets of Amidines Derivatives and Naphthoquinones

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Chagas disease (CD), caused by Trypanosoma cruzi, affects approximately eight million individuals in Latin America and is emerging in nonendemic areas due to the globalisation of immigration and nonvectorial transmission routes. Although CD represents an important public health problem, resulting in high morbidity and considerable mortality rates, few investments have been allocated towards developing novel anti-T. cruzi agents. The available therapy for CD is based on two nitro derivatives (benznidazole (Bz) and nifurtimox (Nf)) developed more than four decades ago. Both are far from ideal due to substantial secondary side effects, limited efficacy against different parasite isolates, long-term therapy, and their well-known poor activity in the late chronic phase. These drawbacks justify the urgent need to identify better drugs to treat chagasic patients. Although several classes of natural and synthetic compounds have been reported to act in vitro and in vivo on T. cruzi, since the introduction of Bz and Nf, only a few drugs, such as allopurinol and a few sterol inhibitors, have moved to clinical trials. This reflects, at least in part, the absence of well-established universal protocols to screen and compare drug activity. In addition, a large number of in vitro studies have been conducted using only epimastigotes and trypomastigotes instead of evaluating compounds’ activities against intracellular amastigotes, which are the reproductive forms in the vertebrate host and are thus an important determinant in the selection and identification of effective compounds for further in vivo analysis. In addition, due to pharmacokinetics and absorption, distribution, metabolism, and excretion characteristics, several compounds that were promising in vitro have not been as effective as Nf or Bz in animal models of T. cruzi infection. In the last two decades, our team has collaborated with different medicinal chemistry groups to develop preclinical studies for CD and investigate the in vitro and in vivo efficacy, toxicity, selectivity, and parasite targets of different classes of natural and synthetic compounds. Some of these results will be briefly presented, focusing primarily on diamidines and related compounds and naphthoquinone derivatives that showed the most promising efficacy against T. cruzi.

1. Chagas Disease and Its Treatment
Chagas disease (CD) is caused by the intracellular obligatory parasite Trypanosoma cruzi. The life cycle of this parasite involves haematophagous triatomine insect vectors, diverse vertebrate mammalian hosts, and different developmental forms. Briefly, after bloodstream trypomastigotes are ingested by an insect, they are converted to epimastigotes, which proliferate and differentiate into metacyclic forms within the posterior intestine of the triatomine. These infective forms, released within the faeces and urine, can invade vertebrate cells and undergo another round of differentiation into the intracellular amastigote forms, which proliferate and then transform back to trypomastigotes, the form that disseminates the infection. CD is the major cause of infectious cardiopathy and represents an important public
health problem; it is broadly dispersed in 18 developing
countries in South and Central America [1]. It affects
approximately eight million people in Latin America, of
whom 30–40% either have or will develop cardiomyopathy,
digestive megasystems, or both [2]. CD transmission
occurs primarily via the vector (90%) but may also occur
through blood transfusion and congenital transmission as
well as laboratory accidents [3], organ transplantation [4],
and ingestion of contaminated food and beverages [5].
Recently, CD has become a major concern due to globali-
sation, which results in immigration of infected individuals
to nonendemic regions, thus spreading the disease [6].
Although both vectorial and transfusional transmission have
sharply declined in the past 20 years due to Southern
Cone countries’ policies, several challenges still need to
be overcome, including those related to sustainable public
health initiatives, vector control strategies, and educational
approaches [7, 8]. Thus, because epidemiological and trans-
mission control characteristics of CD may vary according
to each country’s ecological conditions and adopted health
policies, continuous epidemiological survey in conjunction
with efficient and universal therapy of the infected individ-
uals must be performed to maintain and even reduce the
number of new acute cases [9].

CD affects mainly poor, rural and forgotten populations
and has two consecutive clinical phases: the acute phase that
appears shortly after infection, which ranges from flu-like
symptoms to intense myocarditis (in approximately 10% of
infected people), and the chronic symptomatic phase, which
develops in approximately one-third of infected individuals
after an asymptomatic period (indeterminate form) lasting
years or decades [10, 11]. The main clinical manifesta-
tions of the chronic stage include cardiac and/or gastrointestinal
involvement, and the variability in CD outcome has been
related to host response and parasite heterogeneity [12].
Although its pathology is poorly understood, growing
evidence has shown that parasitic persistence within the
target organs associated with an unregulated host immune
response is involved in pathogenesis, disease progression,
and outcome [1, 13, 14].

Since its discovery by Chagas more than a hundred
years ago [15], CD still poses many challenges, including
its peculiar epidemiology, characterised by a variety of risk
factors (diverse vectors and reservoirs and different forms
of transmission and parasite isolates present in domiciliar,
peridomiciliar, and sylvatic environments), and the lack of
prophylactic therapies and effective chemotherapeutic
schemes [10, 16, 17].

Nifurtimox ([3-methyl-4-(5′-nitrofurfurylideneamine)
tetrahydro-4H-1,4-tiazine-1,1-dioxide], Nf) and benzida-
azole (N-benzyl-2-nitroimidazole acetamide, Bz) were empir-
ically introduced into the clinical therapy regime for CD over
four decades ago. Neither drug is ideal because they present
variable results depending on the phase of the disease (they
are only effective in the acute and recent chronic phases of
the infection), the dose and duration of the treatment, patient
age, and endemic region in addition to showing undesirable
secondary side effects [18, 19]. Additionally, differences in
the susceptibility and natural resistance of different T. cruzi
isolates to both nitroderivatives have also been reported
[20]. It has been suggested that CD must be treated in
all its stages, including acute (acquired or congenital),
chronic reactivated (under immunosuppressive conditions),
determinate, and early chronic phases, as determined by
the presence of parasitic DNA using PCR analysis [21].
Additionally, although there is still no criterion of cure
for symptomatic late chronic cases because most treated
individuals show positive serology, recent data suggest the
benefits of Bz therapy for chronic patients through the arrest
of cardiac damage and a decrease in serology titres [22,
23]. These results reinforce a goal for identifying parasitic
targets that is strengthened by the concept that chagasic
cardiomyopathy is related to parasitic persistence within
the target organs along with an unbalanced host immune
response, which could be useful for a new CD therapy [24].

Since the introduction of Bz and Nf, only allopurinol
and a few azoles, such as itraconazole, fluconazole, and
ketoconazole, have moved to clinical trials [25–27]. In fact,
drug development efforts for CD are almost exclusively
in preclinical research, although phase II studies for the
antifungal drug posaconazole and a produrg of ravuconazole
are being planned [28]. In addition, clinical data have
proven a positive effect of posaconazole in the therapy
of a chronic chagasic patient with systemic lupus erythe-
matosus [29]. However, the high costs of posaconazole may
impair its use in CD.

The gap between preclinical studies and clinical trials
may be associated with the small amount of investments by
pharmaceutical industries due to the low monetary return
and to the previous mistaken concept that during the later
stages of CD, parasitism is absent [30]. In addition, the lack
of standardised protocols and the use of epimastigotes for
drug screening may represent significant impairments for the
discovery of novel anti-T. cruzi candidates [31]. Based on
current knowledge regarding parasite and host physiology,
a promising trypanocidal drug would include the following
characteristics: (i) high activity against the parasitic forms
present in mammalian hosts (intracellular amastigotes and
bloodstream trypomastigotes), (ii) high activity against
diverse T. cruzi strains for use in different endemic regions,
(iii) efficacy against both acute and chronic infections,
(iv) oral bioavailability in few doses, (v) low toxicity and
improved safety with low potential for genotoxicity and
teratogenicity given the potential use in children and women
of reproductive age, (vi) low cost and high stability for
a long shelf-life in tropical temperatures, (vii) high levels
of tissue accumulation and long terminal half-lives, (viii)
low risk for cardiotoxicity because the heart is the primary
organ affected in chagasic patients, and (ix) low risk for
interactions with hepatic cytochrome P450s to avoid drug-
drug interactions because many patients use antiarrhythmic
drugs and anticoagulants [28, 32].

2. Parasite Targets and Lead Compounds
for New Drugs

Advances in proteomics, biochemistry, and in understanding
the biological aspects of T. cruzi infection have allowed for
the development of new approaches to identify parasite-specific targets and, thus, the design of novel potential drugs [33, 34]. It has been proposed that a rational therapy for *T. cruzi* should be directed against different parasitic metabolic targets, including ergosterol biosynthesis, trypanothione metabolism, cysteine protease, pyrophosphate metabolism, protein or purine synthesis, and DNA [21, 28, 35–37]. The combination of different drugs with the aim of achieving higher efficiency and lower toxicity is also an interesting therapeutic strategy [38]. Recent studies have demonstrated the successful synergism of posaconazole with amiodarone, an antiarrhythmic drug, against *T. cruzi in vitro* and *in vivo* [39]. Another relevant approach is the aetiological therapy of CD using carrier molecules, such as ruthenium complexes, that bind 14a-demethylase inhibitors [40] or benznidazole [41], improving both solubility and parasite specificity [28]. A recent study has shown the successful use of ruthenium complexes to deliver nitric oxide to *T. cruzi*-infected cells [42].

Proteomic approaches have been extensively applied for the evaluation of the expression, structure, and function of proteins on a large-scale, including their physiological role, expression regulation, and validity of genome annotations [43]. In trypanosomatids, the control of gene expression is particularly important because all protein-encoding genes are organised in large polycistronic transcription units, producing the RNA that will be processed by trans-splicing. Furthermore, the modulation of protein expression and, consequently, its function is directly related to posttranslational modifications in these protozoa [44, 45]. Proteomic studies have been performed to evaluate the mechanisms of *T. cruzi* resistance/susceptibility to drugs [44–46].

In the present review, we summarise *in vitro* and *in vivo* results on the efficacy, toxicity, selectivity, and cellular targets of aromatic diamidines and naphthoquinone derivatives, two groups of compounds with promising efficacy against *T. cruzi*. In this framework, morphological techniques, such as light (confocal and fluorescence) and electron (transmission and scanning) microscopy, have been employed. Other cellular (flow cytometry), biochemical (respirometry and fluorimetry), and proteomic (bidimensional electrophoresis and mass spectrometry) approaches have also been employed to identify specific targets in the parasite.

2.1. Aromatic Diamidines and DNA Damages. Diamidines, such as pentamidine, propamidine, and diminazene aceturate, have been successfully used in human and veterinary medicine, and they are the first class of drug extensively employed for early-stage human African trypanosomiasis and for cutaneous leishmaniasis caused by *Leishmania guyanensis* [47–49]. Their major drawbacks are poor oral bioavailability and severe side effects. To overcome these issues, new dicaticionic analogues and prodrugs have been synthesised by different medicinal chemistry groups and widely assayed *in vitro* and *in vivo* [50]. One of the most promising compounds is an orally effective prodrug of furamidine (DB75) named DB289, which has been in phase III clinical trials for African trypanosomiasis [51]. Unfortunately, recent results at an extended dosage led to the withdrawal of DB289 from human trials due to toxicity issues.

Despite the strong activity of these dicaticionic compounds against African trypanosomes, few have been assayed as anti-*T. cruzi* candidates [50]. Our team has recently been working on the potential effect of diamidines and congeners against this parasite using both *in vitro* and in vivo models to compare analogues with different structures, cationic centres, and effective motifs [19]. Our data have clearly shown the promising activity of some of these compounds, which displayed high therapeutic windows [52–56].

Although DB75 and its N-phenyl-substituted analogue (DB569) display equivalent DNA-binding properties, DB569 exhibited higher *in vitro* activity against different strains and stages of *T. cruzi*, with IC<sub>50</sub> values in the low micromolar range [57]. Due to the characteristic fluorescence of these diamidines, their localisation in DNA-enriched organelles was determined due to strong labelling of the kDNA [58]. Flow cytometry and transmission electron microscopy (TEM) analysis also demonstrated that DB75 and DB569 disturb parasite mitochondria and nuclei, leading to morphological characteristics of programmed cell death, such as higher levels of apoptotic-like parasites observed after the treatment with DB569 [57, 59]. These findings stimulated further *in vivo* analysis with this analogue, which showed a reduction in the number of parasites and CD8<sup>+</sup> T cells in heart tissues and reversion of electrocardiogram (ECG) alterations in acutely *T. cruzi*-infected mice, leading to an increase in the survival rates [60]. The ECG protection provided by DB569 was also found during the chronic infection of experimental animals, suggesting that the reversion observed in treated animals may be associated with the reduction in cardiac CD8<sup>+</sup> lymphocyte infiltration and parasitism, ultimately contributing to their survival [59, 60].

A diarylihydrophenamine diamidine (DB1362) was evaluated against amastigotes and bloodstream trypanomastigotes of *T. cruzi* and showed good efficacy *in vitro* at submicromolar concentrations, inducing low host cytotoxicity. This diamidine presented a dose-dependent trypanocidal effect after incubation in the presence of plasma constituents (mouse blood), exhibiting IC<sub>50</sub> values similar to those found in the absence of blood, pointing to its potential prophylactic application in blood banks. TEM and flow cytometry have shown that in bloodstream parasites the most important alterations were in kinetoplast organisation and mitochondrial membrane potential [54]. In an acute *T. cruzi* experimental mouse model, treatment with two doses of 25 mg/kg DB1362 (at the onset and at the parasitaemia peak) led to a 40% decrease in the circulating trypanomastigotes and cardiac parasitism (similar levels to Bz) and protected against ECG alterations, leading to a 100% survival rate [54].

Studies on the biological and ultrastructural effect and subcellular localisation of six novel diamidines in *T. cruzi* confirmed their low toxicity towards mammalian cells (LC<sub>50</sub> > 96 μM) and demonstrated that small linear molecules (DB1627, DB1646, and DB1670) were not effective. However, the other three diamidines (DB1645, DB1582, and DB1651) were active, with IC<sub>50</sub> values between 0.15 and 13.3 μM against bloodstream and intracellular amastigotes [61].
Several potential transporters of diamidines have been described in other parasites, such as African trypanosomes, *Leishmania* species and *Plasmodium falciparum* [62–64]. However, the mechanism of diamidine uptake in *T. cruzi* is unknown and requires further investigation. The intrinsic fluorescence of some of these compounds allows for monitoring their localisation, as previously reported in studies with African trypanosomes [50, 65, 66]. Some of these diamidines, like DB1582 and DB1651, were localised in parasitic nuclei and kDNA (with higher intensity in kDNA) and within punctate non-DNA-containing cytoplasmic organelles usually localised in the anterior portion of trypomastigotes and near the nuclei and kinetoplasts in amastigotes, which are possibly acidocalcisomes, as previously described for *T. brucei* [65]. As previously suggested for African trypanosomes, the localisation of these compounds within these acidic organelles could play a role in their mechanism of action and/or act as storage sites [65, 66], but additional studies are needed to clarify this matter. Batista et al. [61] demonstrated that these diamidines caused striking alterations in the mitochondria and kinetoplasts of *T. cruzi*, and some of them also induced disorganisation of microtubules, with the appearance of multiple axoneme structures in trypomastigotes [52, 61]. No major alterations have been reported in either subpellicular or flagellar microtubules of *T. cruzi* treated with drugs that target microtubules, such as taxol, colchicines, and vinblastine, possibly due to the high content of acetylated tubulin and/or poly-glutamylation of tubulin in these parasites [67]. Because these structures are more resistant to microtubule disrupters in trypanosomatids compared to mammalian cells, they may represent interesting targets for drug development and justify further investigations.

Although the exact mechanism of action of diamidines on *T. cruzi* and other trypanosomatids has not been clearly demonstrated, it is likely that multiple modes of action may be responsible and that compound uptake represents a fundamental step in their action and selectivity [50, 68]. One of the long-hypothesised mechanisms of diamidines is related to their selective binding to sequences rich in adenosine and thymine (AT) of kDNA minicircles, leading to kinetoplast destruction and parasite death [58, 69]. Because the kDNA of trypanosomatids contains high numbers of AT-binding sites in thousands of repeated minicircles, it is possible that these structures represent potential specific targets for diamidines [50]. However, although these compounds are excellent minor groove DNA-binders, this interaction alone cannot fully explain their biological activity. Recent reports have suggested that their association with DNA could represent an initial step followed by topological changes leading to molecular instability and destruction and/or modification of DNA–protein interactions, leading to replication errors, DNA degradation, and parasite death [70]. TEM studies have shown that the organisation of mitochondria and kinetoplasts in *T. cruzi* is highly altered by several diamidines and related compounds, such as arylimidamides (AIs), at concentrations that do not affect mammalian host cells [19, 51, 58]. AIs, previously known as reversed amidines, have extraordinary activity against both *Leishmania* [71–73] and *T. cruzi* [52, 53, 55, 56, 74]. They differ from other furan analogues because the amide is bound to the central aromatic linker via a nitrogen atom rather than a carbon atom [72].

Flow cytometry data have confirmed that diamidines and AIs target the mitochondria-kinetoplast complex of *T. cruzi* through interference with the mitochondrial membrane potential [53, 54]. *In vitro* screening of novel diamidines against *T. cruzi* has shown that these compounds localise to a higher extent within the kinetoplast than in the nucleus, and no correlation was found between trypanocidal activity and higher kDNA accumulation [75, 76], as previously reported in *T. brucei* [65]. Other targets for diamidines that also have been proposed include the inhibition of tyrosyl-DNA phosphodiesterase, topoisomerases, protein kinase A, proteases, and polymerases [51, 77, 78].

To better understand the mechanism of action of these aromatic compounds, a study of the possible correlation between kDNA-binding properties of 13 amidines with their trypanocidal efficacy against *T. cruzi* was performed. Four diamidines (DB75, DB569, DB1345, and DB829), eight arylimidamides (DB766, DB749, DB889, DB709, DB613A, DB1831, DB1852, and DB2002), and one guanylhydrazone (DB1080) were assayed using thermal denaturation (*Tm*) and circular dichroism (CD) studies using both whole *T. cruzi* purified kDNA and a conserved synthetic parasitic sequence corresponding to the biological activity of each compound [79]. The findings suggest that the strong interaction of amidines with kDNA may not be sufficient to generate and trigger their trypanocidal activity, and other mechanisms of action may be involved and/or associated.

AIs have potent *in vitro* dose-dependent activity against *T. cruzi*, showing superior trypanocidal activity compared to diguanidinocationic groups and other classical diamidines [52, 56]. Recently, a monoamidine, an arylimidamide, and a guanylhydrazone were evaluated, and the data showed that all compounds exerted, at low micromolar doses, a trypanocidal effect upon both intracellular and bloodstream parasites [74]. However, the potency and selectivity of DB613A, an AI, towards intracellular parasites (with a selective index >126) corroborated previous results that demonstrated the high promising trypanocidal activity of these compounds.

*In vitro* and *in vivo* studies conducted with a novel AIA, DB766, showed its strong trypanocidal activity and excellent selectivity for intracellular amastigotes and trypomastigotes (Y strain), the two relevant parasite forms present in mammalian hosts, exhibiting IC₅₀ values of 25 and 60 nM, respectively [61]. DB766 also exerted striking effects on a wide panel of different parasite strains, including those naturally resistant to Nf and Bz, displaying higher activity *in vitro* than Bz and gentian violet, which are important requirements for identifying a potential anti-*T. cruzi* agent. It is also important to point that DB766 was active against parasite isolates that circulate in peridomiciliar and sylvatic ecotopes from two different regions in Brazil: (i) the northeast (Jaguaribe Valley, Ceará state) that represents an important area for CD surveillance, where high rates of natural triatomine infection are observed (mostly *T. cruzi*...
Six novel aromatic amidinic compounds were tested in vitro to determine activity against the infective and intracellular stages of *T. cruzi* and evaluate their selectivity and toxicity towards primary cultures of cardiomyocytes [56]. The data demonstrated that all of the aromatic amidines were active against *T. cruzi* in vitro and that the arylimidamide DB1470 was the most effective compound, presenting IC$_{50}$ values at submicromolar levels and a good selectivity index and maintaining significant trypanocidal activity at 4°C in the presence of blood constituents [56]. Interestingly, AIAs, such as DB1470 and DB766, exhibited potent trypanocidal activity against *T. cruzi* in the presence of blood constituents [52, 55]. This characteristic is highly desirable for new potential trypanocidal agents for use in blood banks in endemic areas. Unfortunately, although transfusional control has led to a decline in the number of new blood bank-related infections, it is not universally performed. The only trypanocidal agent available for chemical prophylaxis of blood in areas of high endemcity is gentian violet, which is a toxic cationic dye that gives a purple colour to the blood and stains the skin and mucosa of the recipients [10, 81]. These limitations encourage the search for new compounds that could be used in blood bank prophylaxis; thus, AIAs such as DB766 represent promising agents for further evaluation for this purpose [55]. In summary, the efficacy of diamidines and congeners, like AIAs, against *T. cruzi* requires further studies to help establish a valuable scheme of therapy for CD.

2.2. Naphthoquinone Derivatives and Mitochondrial Dysfunction. Naphthoquinones are compounds present in different families of plants that serve as vital links in the electron transport chains in the metabolic pathway and participate in multiple biological oxidative processes [82, 83]. They are considered privileged structures in medicinal chemistry due to their biological activities and structural properties [84]. The redox cycling of quinones may be initiated by either a one- or two-electron reduction. The one electron reduction leads to the formation of semiquinones, unstable intermediates that react rapidly with molecular oxygen, generating free radicals. All of these highly reactive oxygen species (ROS) may react directly with DNA or other cellular macromolecules, such as lipids and proteins, leading to cell damage [85]. This reaction results in shunting electrons toward oxygen, an ineffective pathway for reduction equivalents otherwise used for cytochrome P450 reductase-dependent reactions [86–89]. Another alternative is reduction by two electrons, leading to the formation of hydroquinones, mediated by DT-diphorase [90, 91]. This enzyme reduces toxic, reactive, and unstable quinones, bypassing the creation of toxic intermediates (e.g., a semiquinone radical), and sparing the cell from ROS formation. Whether the two-electron reduction of a quinone leads to detoxification or to activation of oxidative stress depends upon the rate of autoxidation of the formed hydroquinone [86]. If this rate is low under physiological conditions, conjugation may occur before oxidation. As a consequence, the two-electron reduction will lead to detoxification, and an increase in the DT-diphorase activity in tissues would be expected to decrease the toxicity of the quinone. If, however, the hydroquinone
is rapidly oxidised, only a minor fraction may be conjugated before oxidation occurs, and hydroquinone formation would constitute an activation reaction. As a result, enhanced tissue levels of NQO1 would be expected to increase the toxicity of the quinone [92]. Quinones are oxidants and electrophiles, and the relative contribution of these properties to both their toxic and therapeutic activities is influenced by their chemical structure [93]. Two major mechanisms of quinone cytotoxicity have been proposed: stimulation of oxidative stress and alkylation of cellular nucleophiles, which are the mechanisms of action of encompass a large range of biomolecules [94]. Cellular damage can also occur through the alkylation of crucial proteins and nucleic acids.

In addition to their widespread presence in nature, the great interest in the study and mechanisms of action of compounds with a quinoidal structure is due to their multiple roles in organisms. Several quinonoids isolated from traditional medicinal plants are being investigated for their anticancer properties [95]. The antiprotozoal activities of naphthoquinones have been reported, and several of them have been identified as possible leads for drug development [96–98].

Lapachol is easily isolated from the heartwood of trees of the Bignoniaceae family abundant in tropical rainforests, while both α-lapachone and β-lapachone are present only in small amounts. In Brazil, more than 46 types of such woods, popularly known by the name “ipes” (Tabebuia sp.), have been described. In folk medicine, especially among Indian populations, plants containing naphthoquinones have been employed for the treatment of different diseases, such as cancer [99, 100]. The inner bark of Tabebuia avellanedae, commonly known as “pau d’arco” (lapacho, taheebo), is used as an analgesic, an anti-inflammatory, an antineoplastic, and a diuretic by the local people in the northeastern regions of Brazil [101].

Previous reports have shown that against T. cruzi epimastigotes, β-lapachone increases the generation of reactive oxygen species through formation of the semiquinone radical, leading to lipid peroxidation and inhibition of nucleic acid and protein synthesis [102–106]. T. cruzi is known to be deficient in reactive oxygen and nitrogen species detoxification and for being especially sensitive to oxidative stress conditions [107]. Its single mitochondrion, containing a branched electron transport chain and a specialised kDNA region [77], is an extraordinary drug target [33]. The ultrastructural injuries observed in β-lapachone-treated epimastigotes [108] together with the increase in the generation of hydrogen peroxide clearly demonstrates the mitochondrial susceptibility of T. cruzi to naphthoquinones. Unfortunately, no trypanocidal effect was observed in suspensions containing foetal calf serum or rabbit haemoglobin solution, suggesting that β-lapachone could be inactivated by either reduction in the presence of oxhaemoglobin or interaction with serum proteins [109].

Due to the easy access to natural sources of quinones from Brazilian flora and the synthetic mechanisms developed by the group of Dr. Pinto (NPPN/UFRJ) exploring the electrophilicity of 1,2-quinoidal carboxyls [110–113], naphthoquinones have been used as starting points for medicinal chemistry studies. Since the 90s, our group has been studying the anti-T. cruzi activity of this class of chemicals [114]. An initial screening was performed on 60 derivatives obtained through the reaction of several naphthoquinones with common reagents from heterocyclic chemistry, leading to 14 oxazolic, 30 imidazolic, and 10 other related heterocyclic compounds [115–119]. Comparing the activity of the original naphthoquinones and their derivatives, we concluded that minor structural features involved with an increase in lipophilicity, such as the furane moiety, the presence of a methoxyl group, and an aliphatic side chain, led to an increase in the effect on T. cruzi. It is possible that a lipophilic character allows better penetration of the compound through the plasma membrane of the parasite. The activity of the synthesised compounds on T. cruzi showed no uniform behaviour and was in some cases higher, lower, or similar to the activity of the original naphthoquinones from which they were obtained. For the naphthoazoxoles assayed, there was no correlation between biological activity and the type of the mono-oxygenated ring (pyrane versus furane). As shown for naphthoquinones, a lipophilic characteristic, introduced by this appendage, and the presence of a methoxyl or a phenyl group increased the trypanocidal activity. A characteristic of the synthesised naphthoimidazoles was that most of them had aromatic groups containing electron-releasing or electron-withdrawing groups attached to the imidazole ring, and the most active compounds against T. cruzi were obtained from β-lapachone (see Section 2.3) [115, 119].

Another group of naphthoquinone derivatives was also synthesised and assayed on trypomastigote forms, including β-lapachone- and nor-β-lapachone-based 1,2,3-triazoles and 3-arylmino-nor-β-lapachones [120, 121]. 1,2,3-Triazoles are an important class of heterocyclic compounds due to their wide range of biological activities. The strategy of molecular hybridisation linking them to naphthoquinones resulted in compounds endowed with redox properties and a trypanocidal profile. The 1,2,3-triazole derivatives of nor-β-lapachone were more active than the original quinone, and the apolar phenyl-substituted derivative (2,2-dimethyl-3-(4-phenyl-1,2,3-triazol-1-yl)-2,3-dihydro-naphtho[1,2-b]furan-4,5-dione) was the most active compound (IC50/24 h = 17.3 ± 2.0 μM) [120]. Such activity could be due to its higher lipophilic character, which allows better penetration through the parasite’s plasma membrane. In addition, the key intermediate azides used for the synthesis of both the β-lapachone and nor-β-lapachone series of 1,2,3-triazoles displayed higher activity than Bz (IC50/24 h = 103.6 ± 0.6 μM). In the case of nor-β-lapachones-3-arylaminosubstituted compounds, the insertion of chlorine, bromine, nitro, and methoxy groups into the alylamo ring intensified the trypanocidal activity.

Using nor-lapachol as a starting point, substituted ortho-naphthofuranquinones, a nonsubstituted para-naphthofuranquinone, an oxyrane and an azide were prepared. Using α-lapachone as a base, a new nonsubstituted para-naphthofuranquinone was prepared. The most active compounds were three ortho-naphthofuranquinones with trypanocidal activity higher than that of Bz [122]. In another
set of experiments, three new naphthofuranquinones were synthesised and assayed on *T. cruzi*. Two of them were obtained by the addition of iodine to C-allyl-lawsone (2-hydroxy-3-allyl-naphthoquinone) followed by cyclisation, generating a furan ring; the third was obtained through an acid-catalysed reaction by dissolution of the original quinone in sulphuric acid. These compounds were active on bloodstream trypomastigote and epimastigote forms with IC$_{50}$ values between 165–640 and 2.5–25 μM, respectively [123]. The treatment of infected murine macrophages caused a dose-dependent decrease in the percentage of infection with low toxicity to host mammalian cells (over 100 μM), with IC$_{50}$/24h values of 30.7 ± 2.5 μM for intracellular amastigotes between 1.2 and 3.5 μM [124]. An ultrastructural analysis of treated epimastigotes and trypomastigotes indicated a potent effect of the three naphthofuranquinones on parasitic mitochondria, which appeared drastically swollen and with a washed-out matrix profile. Fluorescence-activated cell sorting analysis of rhodamine-123-stained *T. cruzi* showed that these naphthofuranquinones caused a potent dose-dependent collapse of the mitochondrial membrane potential (ΔΨ$_{m}$), especially in epimastigote forms. Such a collapse represented a 30–60% decrease in the parasitic ΔΨ$_{m}$. These compounds also specifically decreased mitochondrial complex I-III activities parallel to a reduction in succinate-induced oxygen consumption between 64–75% in epimastigotes and 72–92% in trypomastigotes. Mitochondrial hydrogen peroxide formation was also increased 1.3–4.5-fold in epimastigotes after treatment with naphthofuranquinones. Our results indicated that the trypanocidal action of these quinones was associated with mitochondrial dysfunction, leading to increased reactive oxygen species generation and parasitic death [124]. The easy synthetic route of these compounds in the laboratory opens the possibility of large-scale production with high yields for assays in experimental mouse models.

2.3. Naphthoimidazoles and Putative Targets. Among the different classes of naphthoquinone derivatives screened against *T. cruzi* (see Section 2.2), the most active derivatives against bloodstream trypomastigotes were three naphthoimidazoles derived from β-lapachone with the aromatic moieties phenyl (N1), 3-indolyl (N2), and methyl-p-phenyl (N3), which were selected for further studies [84, 85]. They were also active against intracellular amastigotes and epimastigotes (Table 1) and showed toxicity to the host cell in concentrations greater than 100 μM. The most susceptible form of the parasite was the intracellular amastigote, with an IC$_{50}$/24h between 6.5 and 9.0 μM [125, 126]. The highest activity against bloodstream forms was observed for N2, with IC$_{50}$/24h values of 12.3 ± 1.2 and 61.6 ± 3.6 μM for 0% and 100% blood, respectively. In epimastigotes, N3 was the most effective, with IC$_{50}$/24h values of 30.7 ± 3.6 μM. All three compounds also blocked the cell cycle (up to 96% inhibition of DNA duplication), inhibited succinate cytochrome c reductase (16–42%) and metacyclogenesis (IC$_{50}$/96h values between 0.35–0.66 μM) and induced extensive morphological damage to the mitochondria, Golgi complex and reservosomes. In treated trypomastigotes, an altered kinetoplast network, mitochondrial swelling, plasma membrane blebbing and DNA fragmentation were found [125, 126]. DNA fragmentation was also evaluated using total DNA electrophoresis and flow cytometry techniques and showed a maximum of 75% TUNEL+ trypomastigotes after treatment with the naphthoimidazoles.

An investigation into their mode of action led to the characterisation of mitochondria, reservosomes, and DNA as their main targets and stimulated further studies about death pathways. Ultrastructural analysis revealed both autophagic (autophagosomes) and apoptotic-like (membrane blebbing) phenotypes. In epimastigotes and trypomastigotes, the naphthoimidazoles induced the formation of concentric membranes, autophagosomes with a loss of matrix electron density, and endoplasmic reticulum profiles surrounding different structures [127]. Apoptosis-like features, such as the release of mitochondrial cytochrome c to the cytosol, has also been detected in N3-treated parasites. Flow cytometry analysis showed a small increase in phosphatidylserine exposure in N2-treated trypomastigotes and a large increase in the percentage of necrosis caused by N1 and N2. These death phenotypes were not detected in treated epimastigotes. A strong increase in the labelling of monodansyl cadaverine (a well-known autophagic marker) was observed up to 45 and 71% in treated trypomastigotes and epimastigotes, respectively. The inhibition of the trypanocidal action of the three naphthoimidazoles by wortmannin or 3-methyladenine together with the overexpression of ATG3, ATG4, ATG7, and ATG8 genes in treated epimastigotes and the ultrastructural evidence pointed to autophagy as the predominant phenotype induced by these compounds [127, 128].

To assess the mechanism of action of the naphthoimidazoles N1, N2, and N3, treated epimastigotes were submitted to two-dimensional gel electrophoresis (2-DE) and mass spectrometry to monitor changes in the protein patterns of different important pathways in *T. cruzi*. Results showed that 9 of the 30 proteins altered were mitochondrial, reinforcing previous morphological and biochemical studies that showed this organelle as the main target of these drugs [125, 126]. Treatment with the compounds led to an upregulation of three proteins: heat-shock protein 85, enolase 1 and trypanothione synthetase (Table 2). The overmodulation of the trypanothione pathway strongly suggests that increased scavenging is necessary due to the increased thiol content in treated parasites, as previously described for trypanosomatids [129].

The downregulation of 26 proteins was observed: 7 in N1, 14 in N2, and 15 in N3. Table 2 summarises all modulated proteins after treatment. A strong decrease (20–80%) in protein levels was detected after treatment with the three naphthoimidazoles. Energy metabolism is an important target of these compounds. The expression of enzymes, such as enolase, pyruvate dehydrogenase, and cytochrome c oxidase, were altered in treated epimastigotes. Interference with ATP generation occurred in different stages of the pathway, including glycolysis, the citric acid cycle, and the mitochondrial electron transport chain. The main structural target of naphthoimidazoles was microtubules. A remarkable
Table 1: IC_{50} values/1 d (μM) for the effect of N1, N2, and N3 against T. cruzi.a.

<table>
<thead>
<tr>
<th></th>
<th>Trypomastigotes</th>
<th>Epimastigotes</th>
<th>Amastigotes</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>0% blood</td>
<td>100% blood</td>
<td>Extracellular</td>
</tr>
<tr>
<td>N1</td>
<td>35.8 ± 1.2d</td>
<td>62.1 ± 3.0</td>
<td>13.4 ± 1.1</td>
</tr>
<tr>
<td>N2</td>
<td>12.3 ± 1.2</td>
<td>61.6 ± 3.6</td>
<td>12.4 ± 1.9</td>
</tr>
<tr>
<td>N3</td>
<td>28.2 ± 0.9</td>
<td>68.3 ± 7.3</td>
<td>9.7 ± 0.2</td>
</tr>
</tbody>
</table>

a References [123, 124].
b Tissue-cultured derived amastigotes.
c Number of amastigotes/100 peritoneal macrophages.
d Mean ± SD of at least 3 independent experiments.

Table 2: Modulated proteins in naphthoimidazoles-treated epimastigotes.a.

<table>
<thead>
<tr>
<th>Protein description</th>
<th>Expression status</th>
<th>Treatment</th>
</tr>
</thead>
<tbody>
<tr>
<td>Trypanothione synthetase</td>
<td>upregulated</td>
<td>N1, N2, N3</td>
</tr>
<tr>
<td>Mitochondrial heat shock</td>
<td>downregulated</td>
<td>N1</td>
</tr>
<tr>
<td>Heat shock protein 60</td>
<td>downregulated</td>
<td>N2</td>
</tr>
<tr>
<td>Elongation factor 1-α</td>
<td>downregulated</td>
<td>N1</td>
</tr>
<tr>
<td>Enolase 1</td>
<td>upregulated</td>
<td>N3</td>
</tr>
<tr>
<td>Glutamamyl carboxypeptidase</td>
<td>downregulated</td>
<td>N1</td>
</tr>
<tr>
<td>Heat shock protein 85</td>
<td>upregulated/downregulated</td>
<td>N1, N2, N3</td>
</tr>
<tr>
<td>Tyrosine aminotransferase</td>
<td>downregulated</td>
<td>N3</td>
</tr>
<tr>
<td>Cytochrome C oxidase subunit IV</td>
<td>downregulated</td>
<td>N2</td>
</tr>
<tr>
<td>Activated PKC receptor</td>
<td>downregulated</td>
<td>N3</td>
</tr>
<tr>
<td>Hypothetical protein</td>
<td>downregulated</td>
<td>N2</td>
</tr>
<tr>
<td>β-tubulin</td>
<td>downregulated</td>
<td>N1, N2</td>
</tr>
<tr>
<td>Pyruvate dehydrogenase E1 β subunit</td>
<td>downregulated</td>
<td>N3</td>
</tr>
<tr>
<td>Sterol 24-C-methyltransferases</td>
<td>downregulated</td>
<td>N3</td>
</tr>
<tr>
<td>Elongation factor 1-β</td>
<td>downregulated</td>
<td>N2, N3</td>
</tr>
<tr>
<td>Proteosome α 7 subunit</td>
<td>downregulated</td>
<td>N1</td>
</tr>
<tr>
<td>Peroxiredoxin</td>
<td>downregulated</td>
<td>N2</td>
</tr>
<tr>
<td>Hypothetical protein</td>
<td>downregulated</td>
<td>N1</td>
</tr>
<tr>
<td>IgE-dependent histamine-releasing factor</td>
<td>downregulated</td>
<td>N3</td>
</tr>
<tr>
<td>Elongation factor 2</td>
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<td>N2</td>
</tr>
<tr>
<td>Dehydrogenase</td>
<td>downregulated</td>
<td>N3</td>
</tr>
<tr>
<td>Cystathionine β-synthase 6</td>
<td>downregulated</td>
<td>N2</td>
</tr>
<tr>
<td>α-tubulin</td>
<td>downregulated</td>
<td>N3</td>
</tr>
</tbody>
</table>

a Reference [128].

Decrease in the α- and β-tubulin content was detected in epimastigotes treated with the three compounds. Interestingly, previous ultrastructural data showed no damage to subpellicular and flagellar conformations after treatment [125, 126]. ELISA showed a decrease in tyrosinated tubulin content, suggesting interference with intracellular vesicle traffic and/or mitotic spindle formation, data reinforced by the blockage of mitosis in naphthoimidazole-treated epimastigotes [126, 130]. Amino acid metabolism was one of the most important targets of the naphthoimidazoles. The levels of tyrosine aminotransferase, elongation factors and different heat-shock proteins were affected by N1, N2, and N3, leading to a decrease in the amino acid synthesis and, consequently, an imbalance in proteins important for cell survival. N2 and N3 also arrested the latter steps of sterol biosynthesis through the downregulation of sterol 24-C-methyltransferase (Table 2). The partial impairment of this pathway could lead to alterations in the lipid composition, with a loss of membrane fluidity. Proteomic and ultrastructural data showed no evidence of necrosis or apoptosis-like cell death in treated parasites [127, 130]. A further detailed study on metabolic interactions is crucial to further elucidate the mechanisms through which naphthoimidazoles act.

3. Concluding Remarks

Despite the drawbacks found during the therapy of chagasic patients with Bz and Nf, in the last decades, only a few compounds have moved to clinical trials, possibly due to the low investments allocated to this area and the lack of standardised protocols for drug screening [31]. As previously noted, “the more leads/approaches that progress to investigational drug candidates, the better chance that new treatments for this often fatal infection will be available to patients in the
near future” [131]. Thus, the recent implementation of high-throughput compound screening against *T. cruzi* will allow for the rapid evaluation of several thousands of compounds per month against intracellular amastigotes, which represent an important tool that may yield the identification of novel promising compounds that can move to clinical trials [28].

However, these promising trypanocidal candidates will need more complete pharmacological and safety test analyses to be considered for clinical trials. However, consistent care must be provided through the adoption of a biopsychosocial model, considering patient therapy in the context of biological, psychological, and social factors, and economic difficulties, which can compromise quality of life [132, 133]. Finally, another important limitation related to moving new compounds towards clinical trials is the absence of feasible markers to monitor the progression of the chronic disease and affordable, efficient, and accessible diagnostic tests. Thus, the development of new drugs for most parasitic diseases requires a multidisciplinary approach involving diverse research areas, such as molecular and cellular biology, chemistry and biochemistry, pharmacology, and toxicology, to provide new insights related to the development and discovery of more selective compounds that could be used for Chagas disease therapy.

As concluded by Abad-Franch et al. [8], improved specific chemotherapy, including more practical formulations (e.g., paediatric) or combinations of existing drugs, and a better understanding of pathogenesis as well as the relative contribution of the parasite and host genetic makeup are clearly needed. New strategies for drug design have been improved by the recent results in *T. cruzi* biochemistry, allowing for better elucidation of the effects of trypanocidal agents.

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