Serological evidence of canine exposure to arthropod-borne pathogens in different landscapes in Rio de Janeiro, Brazil

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1. Introduction

Dirofilaria immitis, Ehrlichia canis, Anaplasma phagocytophilum, Borrelia burgdorferi and Leishmania spp., are important arthropod-transmitted pathogens of medical and veterinary concern (Labarte et al., 2003; Dantas-Torres, 2008; Bowman et al., 2009; Villeneuve et al., 2011; Cardoso et al., 2012). It is known that D. immitis is one of the most important nematodes in veterinary medicine due to the high numbers of infected domestic and wild animals (Labarte et al., 2003; AHS, 2014). This parasite is vectored by several culicidae species that may present hemi-synanthropic (e.g., Aedes taeniorhynchus) or synanthropic (e.g. Culex quinquefasciatus) behavior (Labarte et al., 1998). Tick-borne pathogens such as E. canis, A. phagocytophilum, and B. burgdorferi are the cause of important diseases for humans and domestic or wild animals (Little et al., 2014; Sykes, 2014). Rhipicephalus sanguineus, known as the brown dog tick, is their principal vector species in Brazil (Labruna and Pereira, 2001).

Similarly, Leishmania spp. have been included in the list of the top 5 parasites that affect an incalculable number of domestic, synanthropic, or wild animals, and is considered one of the most prevalent neglected human infections. It is believed that approximately 200,000 to 400,000 new cases of human visceral leishmaniasis occur annually, with >90% of these cases occurring in 6 developing countries, including Brazil (WHO, 2015). From 0.7 to 1.3 million new cases of cutaneous leishmaniasis are reported annually, with approximately 95% of cases occurring in the Americas, the Mediterranean basin, the Middle East, and Central Asia. In Brazil, both cutaneous (caused by Leishmania braziliensis) and visceral forms are endemic (Aguilar et al., 1987), and outbreaks are usually related to disorderly land occupation (Kawa and Sabroza, 2002; Aguilar et al., 2014).

The occurrence of infected dogs with D. immitis or tick-borne parasites is high in tropical and subtropical areas and vectors are prevalent throughout the year in those areas (Labarte et al., 1997; Labarte et al., 2002; Genchi et al., 2009; Willi et al., 2012; Little et al., 2014).
Brazil *D. immitis* infections are known to be frequent in coastal areas where nature is better conserved, providing better conditions for development of culicidiae (Labarthe et al., 2014). Similarly, as with the other vector-borne pathogens, the distribution of *Leishmania* spp. depends on vector populations, and therefore, the expansion of phlebotomies sand flies habitats is directly related to the distribution of leishmaniasis (Killick-Kendrick, 1999; Maia-Elkhoury et al., 2008). Undoubtedly, the environment plays an important role in the occurrence of vector-borne pathogens, since it is strictly related to the development of their vectors. Therefore, the present study assessed the prevalence of *D. immitis*, *E. canis*, *A. phagocytophilum*, *B. burgdorferi* and *Leishmania* spp. in dogs living in different landscape settings in a Brazilian tropical area.

2. Materials and methods

2.1. Ethical aspect and study area

This study was approved by the Comissão de Ética no Uso de Animais – CEUA of the Fundação Oswaldo Cruz (protocol number: LW-33/11). This study was conducted from August 2011 to January 2012 in 3 different landscape areas of the state of Rio de Janeiro (22° 54′ S 43° 10′ W), Brazil. The landscapes were located along a 60-km line from a sandbank section to the mountain region in the eastern area of the state of Rio de Janeiro.

In the sandbank area (sea level) (Site 1), one locale was between the lagoon and the seashore and the other at the opposite margin of the lagoon. In the plains (50 m above sea level) (Site 2), the area studied was between the sandbank and the mountain regions. Finally, in the mountain landscape, 2 locales were included, one at 140 m and the other at 840 m above sea level (Site 3) (Fig. 1). The main features considered for all 3 landscapes were distance from the coast, land use, altitude, human population density, and environmental conservation status. The landscape margins and surface features were determined by visual analysis using Google Earth. Locations where dogs were sampled were acquired by accessing the American global navigation satellite system (GNSS) using a global position system receiver (GPiMAP 62 receiver, GARMIN). Acquired locations were subsequently processed in ArcGIS 10 software.

In the sandbank area, human population density was intermediate and conserved areas were scarce. The locale between the lagoon and the seashore consists basically of homes that are devoid of public water supply, sewage treatment, and paved streets, although public power had recently been installed. The houses had an unfinished aspect, with filthy yards and no trees. At the opposite margin of the sandbank region, houses were brick and well-finished. Roads were partially paved, and public power, water supplies, and public sewage treatment were the norm.

In the plains, the human occupation was dense, leading to intense destruction of natural resources in the region. The population in the area was expanding as people were abandoning urban settings and moving to these more rural areas. Streets were generally unpaved and houses were unfinished; however, public power and water supplies were available, but there was no public sewage treatment.

In the mountain landscape, the balance of nature was conserved, and human density was the lowest of the 3 regions. The 140-m point was set in a small village surrounded by Atlantic forest and some rural properties, where only the principal streets were paved, public power and water supplies were present, but sewage treatment was not. The 840-m point was a rural village with unpaved streets, water supply from local sources, and poor public power supply, situated in Atlantic forest.

2.2. Animals and blood sampling

Dogs (*n* = 333) estimated to be at least one year of age were used in this study. The study aimed to evaluate as many dogs as possible in a given area. To be included in the research, dogs had to have lived within the study area for at least 6 months and owners had to complete an Informed Consent Form giving permission for the dog’s participation in the study. Whole blood samples were collected from each dog through the puncture of the cephalic vein and stored in sterile plastic tubes containing anticoagulant (EDTA). Afterwards, each sample was divided in an aliquot of whole blood and from the other was obtained the plasma.

2.3. Laboratorial procedures

Whole blood samples were used to detect the presence of microfilariae by using a modified Knott test (Newton and Wright, 1956). In addition, plasma samples were tested for the presence of *D. immitis*, *E. canis*, *A. phagocytophilum*, and *B. burgdorferi* antigen using a commercial ELISA test (SNAP 4Dx®, IDEXX Laboratories, Maine, USA) following the manufacturer’s instructions. Dogs were considered infected with *D. immitis* when microfilariae or antigens were detected and with *E. canis*, *A. phagocytophilum*, or *B. Burgdorferi* when antibodies were detected.

The detection of *Leishmania* spp. infection was performed by the ELISA (ELIE Leishmaniose Canina – BioManguinhos/Fiocruz, Rio de Janeiro, Brazil) and immunofluorescence antibody test (IFAT) (Leishmaniose Canina - BioManguinhos/Fiocruz, Rio de Janeiro, Brazil). In addition, an immunocromatographic assay (TR DPP® canine visceral leishmaniasis - BioManguinhos/Fiocruz, Rio de Janeiro, Brazil) was performed. Dogs were considered positive for cutaneous leishmaniasis by ELISA and IFAT and for visceral leishmaniasis when tested positive by ELISA, IFAT and TR DPP.

2.4. Data analysis

Data were entered into EPI INFO 2000 data forms generated for this study, and data entry was verified for accuracy by 2 researchers. Non-parametric analysis was performed by chi square or Fisher's exact tests.

3. Results

After receiving owners’ consent, dogs were tested for *D. immitis* infection and tick-borne parasite seroprevalence; however, only 56.7% (189/333) could be tested for seroprevalence to *Leishmania* spp. because the ethical permission limited the volume of blood that could be collected. Therefore, only surpluses could be used.

The overall prevalence of the pathogens studied was 37% (123/333) for *D. immitis* infection; 46.8% (156/333) for tick-borne parasite sero-prevalence and 27% (51/189) for *Leishmania* spp. seroprevalence. It is important to note that *B. burgdorferi* was not detected in the dogs tested (Table 1).

In the sandbank area, the most prevalent parasite was *D. immitis* (68.9%; 115/167) when compared with tick-borne parasites (43.7%; 73/167; *x^2* = 21.465; df = 1; *P* < 0.0001) or *Leishmania* spp. (34.5%; 39/113; *x^2* = 32.128; df = 1; *P* < 0.0001). There was no difference in the seroprevalence between tick-borne pathogens and *Leishmania* spp. (*x^2* = 2.376; df = 1; *P* = 0.1564) (Table 1).

In the plains, tick-borne pathogens had a significantly higher prevalence (61.7% 37/60) compared with *D. immitis* infection (0/60; *x^2* = 53.494; df = 1; *P* < 0.0001) or *Leishmania* spp. (22.2%; 10/45; *x^2* = 16.181; df = 1; *P* = 0.0001). In this landscape a significant difference between *D. immitis* infection and *Leishmania* spp. seroprevalence was observed (*x^2* = 14.737; df = 1; *P* = 0.0005) (Table 1).

In the mountain area, the seroprevalence of tick-borne pathogens was highest (43.4%; 46/106) compared with the prevalence of *D. immitis* infection (7.5%; 8/106; *x^2* = 35.880; df = 1; *P* < 0.0001) or *Leishmania* spp. (6.4%; 2/33; *x^2* = 14.383; df = 1; *P* = 0.0003). No difference in *D. immitis* infection and *Leishmania* spp. was detected (*x^2* = 14.383; df = 1; *P* = 0.8523) (Table 1).
Of 123 dogs positive for *D. immitis* infection, 93.5% (115/123) were diagnosed at the sandbank. Likewise, of all 51 dogs diagnosed with *Leishmania* spp., 76.5% (39/51) were from the sandbank region (Table 1). Of the 115 dogs positive for *D. immitis* in the sandbank area, 39 also were co-infected with *E. canis* and 15 with *E. canis* and *A. phagocytophilum*. Finally, in the mountain region, 4 of the 8 dogs infected with *D. immitis* were co-infected with *E. canis* and *A. phagocytophilum*.

4. Discussion

Landscape characteristics are of paramount importance for arthropod populations and different arthropod communities can be highly affected by slight environmental changes. Considering those concepts, it is interesting to note that according to the detection of mosquito-borne infections and antibodies of tick-borne pathogens, ticks and

### Table 1

<table>
<thead>
<tr>
<th>Parasite</th>
<th>No. positive/total (%)</th>
<th>Sandbank</th>
<th>Plains</th>
<th>Mountain</th>
<th>Overall</th>
</tr>
</thead>
<tbody>
<tr>
<td>Mosquito-borne *</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Dirofilaria immitis</em></td>
<td>115/167 (68.9)*</td>
<td></td>
<td>0/60 (0)</td>
<td>8/106 (7.5)</td>
<td>123/333 (37.0)</td>
</tr>
<tr>
<td>Tick-borne **</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td><em>Ehrlichia canis</em></td>
<td>51/167 (30.5)</td>
<td>25/60 (41.7)</td>
<td>30/106 (28.3)</td>
<td>106/333 (31.8)</td>
<td></td>
</tr>
<tr>
<td><em>E. canis + Anaplasma phagocytophilum</em></td>
<td>22/167 (41.7)</td>
<td>12/60 (20.0)</td>
<td>14/106 (13.2)</td>
<td>48/333 (14.4)</td>
<td></td>
</tr>
<tr>
<td><em>A. phagocytophilum</em></td>
<td>0/167</td>
<td>0/60</td>
<td>2/106 (1.9)</td>
<td>2/333 (0.6)</td>
<td></td>
</tr>
<tr>
<td>Combined</td>
<td>73/167 (43.7)*</td>
<td>37/60 (61.7)*</td>
<td>40/106 (39.3)</td>
<td>156/333 (46.8)</td>
<td></td>
</tr>
<tr>
<td>Leishmania braziliensis</td>
<td>38/113 (33.6)</td>
<td>9/45 (20.0)</td>
<td>2/31 (6.4)</td>
<td>40/189 (21.5)</td>
<td></td>
</tr>
<tr>
<td>Leishmania infantum</td>
<td>1/113 (0.9)</td>
<td>1/45 (2.2)</td>
<td>0/31</td>
<td>2/189 (1.1)</td>
<td></td>
</tr>
<tr>
<td>Combined</td>
<td>39/113 (34.5)*</td>
<td>10/45 (22.2)*</td>
<td>2/31 (6.4)</td>
<td>51/189 (27.0)</td>
<td></td>
</tr>
</tbody>
</table>

Different letters within rows indicate significant difference (*P* < 0.001). Different symbols within columns indicate significant difference (*P* < 0.001).

* Tested by modified Knott test and ELISA commercial test (SNAP 4Dx®, IDEXX Laboratories, Maine, USA).

** Tested by ELISA commercial test (SNAP 4Dx®).

*** Tested by ELISA (EIE Leishmaniose Canina – BioManguinhos/Fiocruz, Rio de Janeiro, Brazil), indirect immunofluorescence – IFA (Leishmaniose Canina-BioManguinhos/Fiocruz) and immunocromatographic assay (TR DPP® canine visceral leishmaniasis - BioManguinhos/Fiocruz).
mosquitoes seemed to share the same habitats. These results suggest that within a landscape there are different habitats, which provide environmental conditions for development of the vector. Mosquitoes are water-borne and need warm temperatures and humidity, while *Lutzomyia* spp. need organic matter substratum in places with shade and cool temperatures. Results of the present study indicate that tick-borne diseases, and hence tick vectors, are most frequently found in the dry, warm and heavily populated suburban areas with substandard housing and squallor (Labarthe et al., 1998; Killick-Kendrick, 1999; Labruna and Pereira, 2001).

*Dirofilaria immitis* infection was higher in the coastal environment, as it has been previously observed in other Brazilian regions (Labarthe et al., 2014). The coastal environment provides richness of salinity that favors populations of the competent vector *Aedes taeniorhynchus* (Labarthe et al., 1998). However, results from the plains may be somewhat unexpected since the prevalence of *D. immitis* infections declined in dogs further from the coast. The squalor and the degradation of natural resources of the plains could be the main reason for decline in the establishment of semi-synanthropic mosquito populations, resulting in the absence of infection in the landscape. However, the well-conserved Atlantic forest resources of the mountains provided sufficient mosquito vectors to transmit *D. immitis* to the dogs, although at low rates, as has been observed previously (Labarthe et al., 2014). The source of infection to the mosquitoes and from the mosquitoes to the canine population of the mountains is unknown. However, microfilaricidal dogs travelling with their owners or sylvatic microfilariae Mustelidae or Canidae cannot be disregarded as a source of infective microfilariae (Bowman et al., 2009; Brown et al., 2012).

*Anaplasma phagocytophilum* is rarely reported in dogs in Brazil. This infection is vectored by *Ixodes* spp. ticks, which are rarely found on dogs in Brazil. Anti-*A. phagocytophilum* antibodies were frequently detected in association with anti-*E. canis* antibodies in this study, suggesting that both parasites are transmitted by the same tick species. However, it is also possible that cross-reaction among different *Anaplasmataceae* species may have occurred (Santos et al., 2011). The only landscape where dogs harbored anti-*A. phagocytophilum* antibodies and no anti-*E. canis* antibodies was in the mountains, where *Amblyomma cajennense*, the wood tick, occurs frequently and has been suggested as a vector for the bacteria. Therefore, it is possible that dogs that were allowed to roam free by the owners invade the natural environment and become infected (Labruna and Pereira, 2001; Onofrio et al., 2009; Santos et al., 2011).

The highest seroprevalence of *E. canis* was observed in the plains where substandard houses probably provide niches for its principal vector, *R. sanguineus*. Although the rubbish found in the plains areas certainly contributed to enhance Anaplasmataceae transmission, the well-conserved environment of the mountains or the sandbank also provided sufficient resources for *R. sanguineus* populations (Aguiar et al., 2007), and the seroprevalence of *E. canis* was relatively high in both of those regions.

Detection of anti-*Leishmania* antibodies in canine blood samples was not surprising, and the integumentary form is endemic in Rio de Janeiro, especially where substandard land occupation has occurred (Kawa and Sabroza, 2002; Aguiar et al., 2014). The detection of low antibody levels to *Leishmania* in the canine blood samples (ELISA and IFA tests) were mainly from the 2 less-conserved study areas (sandbank and plains), suggesting that *L. braziliensis* is a frequent parasite at those sites, and further confirming previous reports of how land occupation influences *Leishmania* spp. prevalence and distribution. Furthermore, the presence of 2 dogs infected with *L. infantum* (positive by ELISA, IFA and TR DPP® tests) in the crude occupied lowland areas provided evidence of the presence of the vector *Lutzomyia longipalpis* (Brazil et al., 1989, 2012). The confirmed autochthonous infection nearby (Paula et al., 2009) provides additional evidence of the ongoing risk for infection with this life-threatening zoonotic parasite.

The present study provides evidence that the lack of basic sanitation of homes and the neglected conditions in the nearby natural environment play an important role in the provision of resources for arthropod-borne canine infections. Furthermore, dogs in the eastern state of Rio de Janeiro were shown to be at risk of becoming infected by preventable life-threatening parasites with zoonotic potential. These data also provide evidence that enable veterinarians to advise their clients to use appropriate and effective preventive parasite control treatments for their pets, especially when they travel to high-risk areas and to routinely screen their patients for parasites. Further studies must be conducted to better understand the relationship among the environment, vectors, parasites and hosts.

**Ethical standards**

All of the studies reported herein were performed in compliance with current applicable local laws and regulations.

**Conflict of interest**

Liliane Willi Monteiro, Celeste Freitas de Souza, Tainá Laeta, Marcia Nobre de Miranda and Fabiana Batalha Knackfuss have no competing interests. Flavya-Mendes-de-Almeida is a consultant for Idexx Laboratorios. Jonimar Paiva is a consultant for Zoetis in Brazil. Norma Labarthe is a consultant for Bayer Animal Health, Idexx Laboratories, and Zoetis in Brazil.

**Acknowledgements**

The authors acknowledge Marcia Chame for the contribution to study design and Carolina Haje, Daniel Marques, Daniel Paiva, Daniel Ribeiro, Marcela Machado, Monique Paiva and Thiago Gomes for assistance in the acquisition of data. The authors thank Kathleen Newcomb, Nathalie, VA, USA for editorial assistance in the preparation of this manuscript. The authors are grateful to the owners and the dogs of the study. The authors also acknowledge Bayer S.A. and Linavet for providing part of the kits.

**References**


