VISCERAL LEISHMANIASIS CAUSED BY Leishmania (Viannia) braziliensis IN A PATIENT INFECTED WITH HUMAN IMMUNODEFICIENCY VIRUS

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SUMMARY

The current article reports the case of a 19-month-old-girl, from the state of Minas Gerais, Brazil, with visceral leishmaniasis, by Leishmania (Viannia) braziliensis, and Human Immunodeficiency Virus (HIV) co-infection. The child’s mother and father, aged 22 and 27 years old, respectively, were both HIV positive. The child was admitted to the General Pediatric Center, in Belo Horizonte, presenting high fever, fatigue, weight loss and enlargement of liver and spleen. Indirect immunofluorescent test revealed a titer of 1:320 for Leishmania. Such result was confirmed by the presence of amastigotes in bone marrow aspirate samples and culture of promastigote forms. Parasites were identified as being Leishmania (Viannia) braziliensis through PCR, using a L. braziliensis complex primer and a generic primer, followed by hybridization. Specific leishmaniasis therapy (Glucantime® antimonial) was intravenously administered.

KEYWORDS: Visceral leishmaniasis; Leishmania (Viannia) braziliensis; Human immunodeficiency virus; PCR.

INTRODUCTION

The genus Leishmania causes a variety of clinical syndromes ranging from self-healing cutaneous lesions through metastasizing mucocutaneous forms to often lethal visceral manifestations. The severity of clinical manifestations in immunocompetent people depends, mainly, on the Leishmania species involved. Patients with leishmaniasis, associated with acquired immunodeficiency syndrome (AIDS), have revealed unusual immunopathological aspects. Most patients, suffering from Leishmania-HIV co-infections, have shown atypical clinical profiles such as disseminated cutaneous leishmaniasis1,4, usual cutaneous lesions5,7, unusual mucocutaneous leishmaniasis caused by L. braziliensis4, cutaneous involvement due to viscerotrophic parasites2 and visceral involvement due to dermotropic parasites22.

Leishmania has been reported to be an opportunistic parasite, in immunosuppressed patients2 and in immunocompromised individuals such as patients with HIV infection23, who live in endemic areas. The overlap between VL and AIDS has increased as a consequence of the spread of the pandemic AIDS in rural areas and that of VL to urban areas5. Consequently, cases of Leishmania/HIV co-infection are becoming more frequent with important clinical, diagnostic, chemotherapeutic, epidemiological and economic implications. The major features of VL clinical picture are: intermittent fever, spleen enlargement, pallor, fatigue, severe weight loss, loss of appetite and pancytopenia9. The disease is fatal if not treated. Although the incubation period of VL may vary from only 10 days to a year, the usual incubation period ranges from 2 to 4 months6,8.

Unfortunately, VL diagnosis is difficult in such patients, since serologic diagnosis is not sensitive enough and culture of Leishmania parasites is both time-consuming and not always positive. Molecular techniques for typing and diagnosing the etiological agent are very useful for specific therapy. PACHECO et al.11 described the isolation, genotypic and phenotypic characterization of a monoxenous trypanosomatid, found in the bone marrow of an HIV-positive patient, presenting a visceral leishmaniasis-like syndrome. Several authors have described PCR assays to be used as a diagnosis method10,12,13,14,15. The present article describes the case report of a VL patient, by L. braziliensis, and HIV co-infection.

METHODS

Indirect immunofluorescent assay (IFA): IFA, to detect antibodies in serum diluted from 1:40 to 1:640, was performed according to the manufacturer’s instructions, using a commercial kit for the diagnosis of human leishmaniasis (FIOCRUZ/BIO-MANGUINHOS). The serological tests, on different Leishmania species, used weakly positive titers ≤ 1:40 and strongly positive titers ≥ 1:80 as IFA cutoff values.

Parasitological test: Cell culture from bone marrow aspirate material was performed in NNN/LIT medium, supplemented with 10% fetal calf serum and incubated at 24-25 °C for 30 days. For direct examination of bone marrow, the slides were stained with Giemsa and examined under light microscopy.

Research supported by the CNPq, PAPES/FIOCRUZ, CPqRR
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DNA extraction: A volume of 20-30 µl of blood and bone marrow aspirates was transferred onto filter paper “FTA Cards” (Gibco BRL) and air-dried. A small portion of the sample, on FTA paper, was removed using a 2 mm punch and placed in a microcentrifuge tube with 200 µl of lysis buffer (FTA processing reagent-Gibco BRL). The sample was washed for 5 min at room temperature and centrifuged three times at 13000 g for 5 min, after which the supernatant was removed and discarded. A volume of 200 µl TE buffer (10 mM Tris-HCl, 1 mM EDTA, pH 7.5) was added to, incubated for 5 min at room temperature and the supernatant was discarded after centrifugation, and the pellet resuspended in 50 µl TE. The paper-bound DNA was then ready for analysis; the paper punch containing purified DNA was directly used for PCR.

PCR 1: In order to amplify the conserved region of the minicircle molecule (Kinetoplastid mitochondrial DNA; kDNA), oligonucleotides (primer A: 5’-(G/C)(G/C)(C/G)CC(A/C)CTA(T/A)TTACACCAACCCC; and primer B: 5’-GGGGAGGGCGTTCTGCGAA) were used in a hot-start PCR procedure. Briefly, each reaction contained 100 ng of the reverse and forward primers, 200 µM of each deoxynucleoside triphosphate (Pharmacia, Uppsalla, Sweden), 2.5 U of Taq polymerase (Perkin-Elmer, Norwalk, USA) as buffers, as recommended by the manufacturer, and a small part of the DNA paper sample. PCR amplification was carried out in a DNA thermocycler (Perkin-Elmer) using 33 cycles at 94 °C for 30 s; at 50 °C for 30 s, and at 72 °C for 1 min and desnaturation at 93 °C for 10 min at 72 °C.

PCR 2: Specific PCR was carried out using the B1 and B2 primers (B1 – GGTTTGTTGAATATAGTGG and B2 – CTAATTGTCACGGGGAAGG) specific for the L. braziliensis complex. Cycles consisted of annealing at 63 °C for 1 min, extension at 72 °C for 1 min and desnaturation at 93 °C for 30 s. The programmes were run for 35 cycles on a Perkin Elmer thermocycler (Perkin-Elmer, USA), a final extension at 72 °C for 10 min was run.

Agarose gel electrophoresis: Each experiment included a positive control (reference strains of Leishmania) and a negative control (rabbit blood). Aliquots of 10 µl of the PCR reaction were analyzed by agarose gel electrophoresis, at 100V, in the presence of ethidium bromide (0.5 µg/ml). The expected amplification products of 120 bp (PCR 1) and 750 bp (PCR 2) were visualized under UV light.

Positive and negative controls: The reference strains L. (L.) chagasi MHOM/BR/74/PP75 (code PP75), L. (V.) braziliensis MHOM/BR/75/M2903 (code M2903) and L. (L.) amazonensis IFLA/BR/67/PH8 (code PH8) were used as positive controls. The promastigotes of such strains were transferred to the filter paper “FTA Cards” (Gibco BRL), to which rabbit blood was added after air dried. Rabbit blood, alone on filter paper, was used as a negative control. DNA from Leishmania reference strains and rabbit blood were prepared according to the protocols previously described.

Hybridization: PCR 1 products were transferred to nylon membranes with a dot-blot apparatus and hybridized with cloned L. braziliensis minicircles as molecular probe, radio-labeled with [α-32P]dATP, by random hexamer priming. The transferred hybridized was blotted at 65 °C, washed in 0.1X sodium saline citrate at the same temperature, and exposed to X-ray films.

Case report: In January 1998, a 19 month-old girl, from the small town of Martinho Campos, in the Brazilian state of Minas Gerais, was admitted to the General Pediatric Center in Belo Horizonte with fever, pneumonia, lymphadenomegalias and hepatosplenomegalias. The girl’s mother and father, aged 22 and 27 respectively, were both HIV-positive. The girl was delivered by cesarian section and weighed 3000 g at birth, with a body length of 50 cm. The child was breast-fed by the mother until 3 months old. She suffered bouts of pneumonia at 3, 5, 6, 8, 9 and 13 months of age. At 8 months, she received two blood transfusions due to her recurrent medical profile of malnutrition, anemia, fever, pallor, hepatomegalias and splenomegalias. At 19 months, she was diagnosed as HIV-positive and received treatment with Zidovudine (AZT) (120 mg/m² every 8h), Didanosine DDI (100 mg/m² every 12h) and intravenous gammaglobulin. On physical examination, she was chronically ill appearing, febrile and pale. The spleen and liver were palpable, extending to 5 and 8 cm below the coastal margin. Laboratory data revealed platelets (35,600/µL), hemoglobin (8.7 g/dL), normal erythrocyte, leukocytopenia. The patient subsequently experienced respiratory failure, due to AIDS-related pneumocytosis, and died 8 weeks later.

RESULTS

Immunological test: A positive titer was observed for Leishmania antibodies (IFA 1:320). After 2 weeks of intermittent fever, another positive titer for Leishmania antibodies (IFA 1:160) was noted.

Parasitological studies: The results of Leishmania IFA were confirmed by visualization of amastigote forms, from bone marrow aspirate, after Giemsa staining (Fig. 1) and isolation of promastigote forms in culture. Specific therapy was initiated with intravenous administration of antimonial Glucantime® (20 mg/Kg body weight).

Fig. 1 - Smear of the bone marrow aspirate from a child with visceral leishmaniasis indicating presence of amastigotes (2-3 µm in diameter).
PCR for the genus *Leishmania*: Purified DNA from the positive controls and the DNA, isolated from blood and bone marrow aspirate of the patient, also yielded an amplification product (Fig. 2a). The DNA purified from rabbit blood was negative for *Leishmania*, based on PCR analysis. No contamination or inhibition was detected.

The parasite was identified as *L. (V) braziliensis* using blood and bone marrow aspirate, the specific primer B1/B2 and PCR-hybridization analysis.

**DISCUSSION**

*Leishmania*/HIV co-infection is considered to be an emerging disease and a threat in several countries in accordance with WHO data. The global experience, mainly in European countries, is marked by an increasing number of co-infection cases in this decade, leading to modifications concerning the epidemiological profile, presentation and clinical outcome of visceral leishmaniasis in various countries.

Recent changes in the epidemiological profiles, identified by the AIDS and leishmaniasis control programs, in Brazil, such as the ruralization and pauperization of HIV infection and the simultaneous urbanization of leishmaniasis, point to a possible expansion of people at risk of acquiring both infections. In Brazil, the number of cases of *Leishmania*/HIV co-infection is growing in several states, including both the cutaneous and visceral clinical presentation. However, the under-notification cases of co-infection remains a problem with important clinical and epidemiological implications. DA-CRUZ et al. reported an atypical case of AIDS-associated mucocutaneous leishmaniasis, due to *L. braziliensis*. There are also a few reports of HIV-infected mucocutaneous leishmaniasis patients, associated or not with cutaneous lesions. In a recent case report CHEHTER et al. described the involvement of the gastrointestinal tract and dissemination forms of *Leishmania*, in co-infection visceral leishmaniasis and HIV patient. In the present paper, atypical VL was caused by *L. braziliensis* and the patient did not show cutaneous or mucosal lesions. HERNANDEZ et al. also demonstrated *L. braziliensis* as causing VL, in a patient with HIV infection. In our laboratory, we have isolated various parasite samples from inhabitants of Martinho Campos and characterized them as *L. braziliensis*; no other cases of VL have yet been reported from the region.

In the last few years, the number of human cases of visceral and cutaneous leishmaniasis, in the metropolitan region of Belo Horizonte (MRBH), has increased, suggesting a rise in the transmission rates of *Leishmania* species. The clinical incubation period of VL typically ranges from 6 weeks to 6 months but may vary from 10 days to 10 years. The course of the disease is identical in children and adults. It may suddenly begin with high fever, vomiting, diarrhea, poor appetite, weight loss, lassitude and pallor and later produce splenomegaly, hepatomegaly, pancytopenia, lymphadenopathy and hyperglobulinemia. Untreated, the disease is fatal in 90% of cases within 1-3 years.

HIV infection and *Leishmania* infection each induce important similar immunological changes, worsened when there is concomitant infection, such as a Th1 to Th2 response switch. However, the clinical presentation is different. The inhibitory effect of IFN-gamma-inducing factors, such as IL-12 and IL-18, on cell proliferation and INF-gamma production also demonstrated that *Leishmania* infection each induce important similar immunological changes, worsened when there is concomitant infection, such as a Th1 to Th2 response switch. However, the clinical presentation is different. The inhibitory effect of IFN-gamma-inducing factors, such as IL-12 and IL-18, on cell proliferation and INF-gamma production...
reinforce the hypothesis that L. (V.) braziliensis is an opportunistic parasite producing severe manifestations that normally do not occur in immunocompetent individuals.

Specific diagnosis of leishmaniasis and characterization of parasites are, therefore, important prerequisites for treatment of immunocompromised patients in Leishmania-endemic areas, due to their increased risk of morbidity and mortality.

**RESUMO**

**Leishmaniose visceral causada por Leishmania (Viannia) braziliensis em paciente infectado com HIV**

No presente artigo os autores relatam caso de uma criança de 1 ano e 07 meses proveniente do estado de Minas Gerais com leishmaniose visceral causada por *Leishmania (Viannia) braziliensis* e co-infeção HIV. A mãe e o pai da criança de 22 e 27 anos de idade respectivamente, são, vão além, HIV positivo. A criança foi internada no Centro Geral de Pedriatria em Belo Horizonte com febre alta, fadiga, perda de peso e aumento de figado e baço. Foi realizado teste de imunofluorescência indireta para *Leishmania* e detectado título de 1:320. Este resultado foi confirmado com o encontro de amastigotas em aspirado de medula óssea e o crescimento de promastigotas em meios de cultura. Os parasitos foram identificados como *Leishmania (Viannia) braziliensis* utilizando PCR com primer específico para o complexo *L. braziliensis*, e primer genérico seguido de hibridização. Terapia específica para leishmaniose (antimonial de Glucantime) foi administrado por via intravenosa.

**ACKNOWLEDGMENTS**

We thank the General Pediatric Center, Belo Horizonte- FHEMIG/ Brazil for the clinical information of the patient.

**REFERENCES**


Received: 17 August 2001
Accepted: 24 April 2002