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A2 proteins are composed predominantly by a repetitive element, which makes it an attractive antigen for diagnosis. These proteins are preferentially expressed in the amastigote stage of *Leishmania donovani*. The genes coding for A2 are also present in *L. mexicana* strains, but not in *L. major* or *L. braziliensis*. The fusion protein A2-GST has been previously evaluated in ELISA assays with sera of Indian and Sudanese patients with kalaazar and being reactive with 60 and 82% of the tested sera, respectively (GHEDIN et al., 1997). In this study the reactivity of A2, fused to GST or to a tag of histidines, was evaluated with of a large panel of canine kalaazar sera (previously tested by IFA) and also of patients with visceral or tegumentar leishmaniasis, tuberculosis and Hanseniasis. The recombinant proteins were expressed in *Escherichia coli* and purified with glutathione beads or by means of a niquel affinity chromatography. ELISA was performed with either A2-GST, GST, A2-His or total extracts of promastigotes of *L. chagasi* antigens. Due to the high reactivity of kalaazar negative canine sera with GST alone, we found that A2-GST is not adequate to kalaazar serologic tests. However, using the A2-His as antigen, anti-A2 antibodies were detected by ELISA in 88% of the parasitological or IFA positive canine sera tested, in 40% of the sera of patients with visceral leishmaniasis and only in 15% of patients with the tegumentar disease. Using the total extract as antigen we found 95%, 82%, 55% of reactivity, respectively. The reactivity of A2 with sera of patients with *Mycobacterium* infection was 10%. Similar results were obtained by Western blotting analysis of selected sera. Our findings suggest that A2 may be an useful antigen to improve the serodiagnosis of visceral leishmaniasis.

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